



Title	Disinfection of Water for Aquaculture
Author(s)	Kasai, Hisae; Yoshimizu, Mamoru; Ezura, Yoshio
Citation	Fisheries science, 68(Supplement 1), 821-824 https://doi.org/10.2331/fishsci.68.sup1_821 Proceedings of International Commemorative Symposium, 70th Anniversary of The Japanese Society of Fisheries Science
Issue Date	2002-11
Doc URL	http://hdl.handle.net/2115/38580
Rights	© 2002 公益社団法人日本水産学会; © 2002 The Japanese Society of Fisheries Science
Type	article
Note	International Commemorative Symposium, 70th Anniversary of The Japanese Society of Fisheries Science. 1-5 October 2001. Yokohama, Japan.
File Information	yoshimizu-149.pdf



[Instructions for use](#)

DISINFECTION OF WATER FOR AQUACULTURE

HISAE KASAI,¹ MAMORU YOSHIMIZU¹ AND YOSHIO EZURA¹

Laboratory of Biochemistry and Biotechnology, Division of Marine Bioscience, Graduate School of Fisheries Science, Hokkaido University, Minato 3-1-1, Hakodate, Hokkaido 041-8611, Japan.

SUMMARY: Disinfection of water for aquaculture is critical for preventing the introduction and spread of infectious disease. A pathogen-free water source is essential for success in aquaculture. Typical treatment systems make use of high efficiency sand filters to clarify the water before treatment with ultraviolet (UV) light or ozonization. Fish pathogens are divided into two groups based on their sensitivity to UV and total residual oxidants (TROs) produced by ozonization of seawater. Hypochlorite produced by electrolysis of seawater (salt water) showed bactericidal and viricidal effects. This method can easily treat large volumes of water, and is suitable for disinfecting wastewater before discharging.

KEY WORDS: disinfection, aquaculture, fish pathogen, UV, ozone, oxidant, electrolization

INTRODUCTION

Water supplies for seed production and aquaculture often provide an efficient means for the introduction and spread of infectious diseases. A pathogen-free water source is essential for success in aquaculture. Surface waters commonly used in aquaculture come from coastal waters or rivers and may contain some fish pathogens and such open water supplies should not be used without treatment. Disinfection of wastewater before discharging is necessary to avoid the pathogen contamination in the environment. In this study, we examine the effect of ultraviolet (UV), oxidant produced by ozonization of seawater and hypochlorite produced by electrolysis of seawater. Additionally the disinfectant effects of the three methods for a hatchery water supply and

wastewater were compared and the survival rate of cultured fish that were reared using water treated with these methods was assessed.

UV susceptibility of fish pathogens and the effects of UV treatment on hatchery water

The disinfectant effects of UV irradiation were examined on cell suspensions of 4 species of fish pathogenic bacteria and *Escherichia coli*, using a punched agar medium disk covered with 10 strains of aquatic fungi and 14 strains of cell free fish pathogenic viruses. Of the viable bacterial cells of Gram negative bacteria and Gram positive bacteria, 99.9% or more were killed by UV irradiation at the dose of 4.0×10^3 and $2.0 \times 10^4 \mu W \cdot sec/cm^2$, respectively.⁴⁾ The hyphae of aquatic fungi showed relatively lower susceptibility to UV irradiation, levels that inhibited the growth of hyphae were 1.5×10^5 to $2.5 \times 10^5 \mu W \cdot sec/cm^2$.⁵⁾ Six fish rhabdoviruses, three fish herpesviruses and two fish iridovirus were found to be sensitive to UV irradiation. The dose that resulted in a 99 % or more infectivity decrease (ID₉₉) was observed at the dose of 1.0×10^3 to $3.0 \times 10^3 \mu W \cdot sec/cm^2$. Susceptibility of two birnaviruses, a fish reovirus and a fish nodavirus was found to be low. ID₉₉ measured 1.5×10^5 to $2.5 \times 10^5 \mu W \cdot sec/cm^2$ (Fig.1).⁶⁾

The infectivity of infectious hematopoietic necrosis virus (IHNV), in virus-contaminated river water and pond water, as measured by the molecular filtration method was 0.56 and 5.6 TCID₅₀/l, respectively. UV treatment of river water with $10^4 \mu W \cdot sec/cm^2$ UV dose prevented an IHN outbreak.⁷⁾ Furthermore, UV treatment of the hatchery water supply also decreased the viable bacterial counts and fungal infection rates of salmonid eggs.⁸⁾

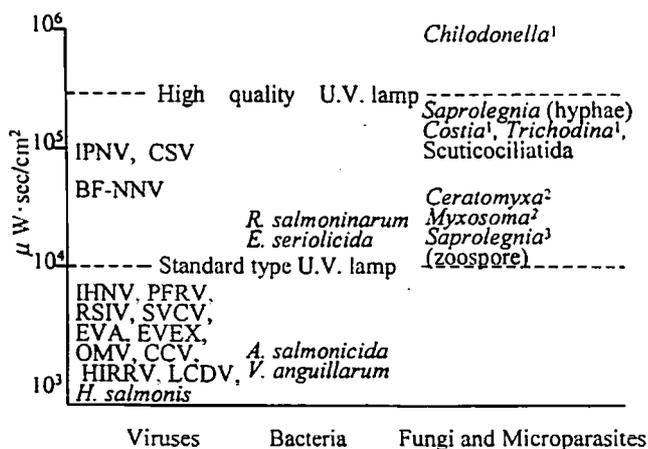


Fig. 1. U.V. susceptibility of fish pathogens.

¹ Vlasenko, M. I. (1969) ² Hoffman, G. L. (1974)

³ Normandeau, D. A. (1968)

Table 1. Effect of total residual oxidants (TROs) concentrations produced by ozonization of seawater on infectivities of fish pathogens

Fish pathogens	TROs concentration (mg/l)	Treatment time (sec)	Reduction rate (%)	Initial number (Log.)
Yellowtail ascites virus (YAV)	0.5	60	>99	4.3 ¹
hirame rhabdovirus (HIRRV)	0.5	15	>99	5.6 ¹
Infectious pancreatic necrosis virus (IPNV)	0.5	60	>99	4.1 ¹
Infectious haematopoietic necrosis virus (IHNV)	0.5	15	>99	4.1 ¹
<i>Oncorhynchus masou</i> virus (OMV)	0.5	15	>99	3.1 ¹
Chum salmon virus (CSV)	0.5	60	>99	4.1 ¹
<i>Vibrio anguillarum</i> NCMB6	0.5	15	>99.9	5.6 ²
<i>Enterococcus serioricida</i> 538	0.5	15	>99.9	5.8 ²
<i>Aeromonas salmonicida</i> ATCC 14174	0.5	15	>99.9	5.1 ²
<i>Aeromonas hydrophila</i> IAM 1018	0.5	15	>99.9	4.6 ²
Scuticociliatida BR-9001	0.8	30	>99.9	5.5 ³

¹ Initial viral infectivity (TCID₅₀/ml). ² Initial viable bacterial number (CFU/ml). ³ Initial viable number.

Disinfectant effect of oxidant produced by ozonization of sea water on fish pathogens

Treatment of natural seawater with ozone produced oxidant that showed a disinfectant effect. Total residual oxidant (TROs) produced in seawater were stable for 1 h or more. Disinfectant effect of TROs against fish pathogenic organisms was observed at a dose of 0.5 mg/l for 15 to 30 s or 0.1 mg/l for 60 s,

and killed more than 99.9 % of bacterial cells of *Vibrio anguillarum*, *Enterococcus serioricida*, *Aeromonas salmonicida*, *A. hydrophila* and *E. coli*, and inactivated 99 % or more of IHNV, hirame rhabdovirus (HIRRV) and *Oncorhynchus masou* virus (OMV). To inactivate or kill more than 99 % of yellowtail ascites virus (YAV), infectious pancreatic necrosis virus (IPNV), chum salmon virus (CSV), and a Scuticociliatida (ciliata), higher doses of 0.5 to 1.0 mg/l for 1 min were required (Table 1).^{9,10}

TROs showed toxicity for fish. Barfin flounder *Verasper moseri* and herring *Clupea pallasii* died after 16 and 2 h exposure to TROs of 0.1 and 0.5 mg/l, respectively. However, Japanese flounder could be cultured in ozonized seawater after the TRO were removed by charcoal (Fig.2), resulting in survival rates similar to fish cultured in UV treated or non-treated seawater.¹¹⁾

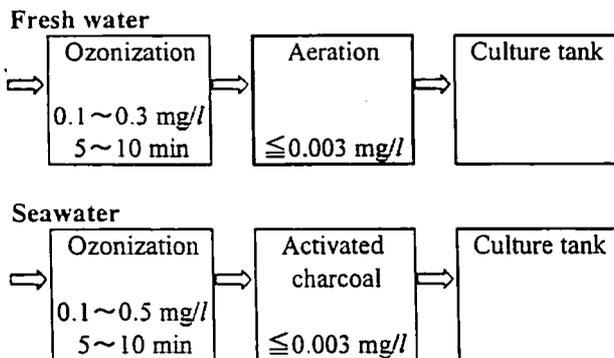


Fig. 2. Ozonization of water for aquaculture.

Disinfectant effects of electrolyzed salt water on fish pathogenic bacteria and viruses

The bactericidal and viricidal effects of hypochlorite produced by electrolysis of salt water were examined against pathogenic bacteria and viruses of fish.

Table 2. The chlorine concentration produced by electrolysis of salt water and treatment time required to reduce the viability of bacteria and the infectivity of viruses by 99.9 %

Pathogens	Chlorine concentration (mg/l)	Treatment time (min)	Initial number (Log.)	Reduction rate (%)
<i>Vibrio anguillarum</i> NCMB6	0.07	1	6.7 ¹	>99.99
<i>Aeromonas salmonicida</i> ATCC 14174	0.06	1	6.6 ¹	99.96
<i>Escherichia coli</i> O-26	0.14	1	6.6 ¹	99.98
Yellowtail ascites virus (YAV)	0.45	1	4.5 ²	99.92
hirame rhabdovirus (HIRRV)	0.34	1	4.5 ²	99.97

¹ Initial viable bacterial number (CFU/ml). ² Initial viral infectivity (TCID₅₀/ml).

Table 3. Effects of U.V. irradiation, ozonization and electrolyzation on the viability of bacteria in hatchery waste-seawater

Methods	Flow rate (m ³ /h)	Treatment	Viable counts (CFU/ml)	Reduction rate (%)
Non-treated	2.0	-	4.7×10 ⁴	-
U.V. irradiation	2.0	1.0×10 ⁵ μW·sec/cm ²	1.3×10 ⁰	99.99
Ozonization	2.0	TROs 0.5 mg/l, 1 min	3.0×10 ⁰	99.99
Electrolization	2.0	Chlorine 0.6 mg/l, 1 min	4.1×10 ²	99.13

Sodium chloride solutions, ranging from 0.5 to 3 % were electrolyzed and the concentration of chlorine produced was measured. Similar concentrations of chlorine were produced when 1.0 % or higher NaCl solution and seawater were electrolyzed.¹²⁾ A 3 % solution of sodium chloride containing pathogenic bacteria or virus was electrolyzed and the organisms were exposed to chlorine. Greater than 99.9 % of *V. anguillarum* and *A. salmonicida* cells were killed when the bacteria were exposed to 0.1 mg/l chlorine for 1 min. YAV and HIRRV were 99.9 % or greater inactivated after treatment with 0.45 mg/l chlorine for 1 min (Table 2).¹³⁾

The bactericidal and viricidal effects of hypochlorite produced by electrolysis (3.5 m³/h, 0.1 A) were greater than that of the chemical reagent. The purity of the sodium chloride used influenced the effects of production of hypochlorite. Sodium chloride obtained as a super grade chemical reagent was more effective than food-grade sodium chloride. However, a sufficient disinfectant effect was observed in electrolyzed seawater, a treatment which may have an application in aquaculture. To use electrolyzed seawater for culture, the chlorine has to be removed with charcoal because of its toxicity.

Disinfection of wastewater

The bactericidal effect of hypochlorite produced by a continuous flow electrolyzer on hatchery wastewater was investigated. The number of viable bacteria in the wastewater was reduced more than 99 % when the water was treated with chlorine at a concentration of 0.6 mg/l for 1 min, and over 99.9 % of the bacteria

cells were killed when treated with 1.28 mg/l for 1 min (Table 3). In another experiment, 2.0 m³/min of hatchery wastewater was electrolyzed and the produced hypochlorite that was mixed with the remaining, 16.5 m³/min wastewater. Viability of bacteria was reduced greater than 99 % after treatment with 0.5 mg/l of chlorite for 1 min. The bactericidal effect of electrolysis was almost the same as that of ultraviolet irradiation (1.0 × 10⁵ μW·sec/cm²) or ozonization (TROs 0.5 mg/l, 1 min) of seawater. Electrolyzation can be used to treat a large volume of wastewater compared with the ultraviolet irradiation or ozonization.¹⁴⁾

Disinfection of water for hatchery water supplies and survival rate of cultured fish

The effects of the three disinfection methods on bacteria in the hatchery water supplies are shown in Table 4. All methods resulted in a reduction of 96.6 to 99.8 % after the treatment. Survival rate of Japanese flounder *Paralichthys olivaceus* and barfin flounder

Table 4. Viability of bacteria in hatchery water supply after treatment using different disinfection methods

Method	Treatment time (min)	Viable counts (CFU/ml)	Reduction rate (%)
Control	-	2.2×10 ²	-
U.V. irradiation	1	7.3×10 ⁰	96.6
Ozonization ²	5	5.0×10 ⁻¹	99.8
Electrolization ³	1	1.1×10 ⁰	99.5

¹ U.V. dose: 1.0×10⁵ μW·sec/cm².

² TROs: 0.5 mg/l. ³ Chlorine: 0.5 mg/l

Table 5. Survival rate of Japanese flounder cultured in U.V. irradiated, ozonized, electrolyzed or non-treated seawater

Survival rate	U.V. irradiated ¹	Ozonized ²	Electrolyzed ³	Non-treated
Japanese flounder ⁴	71.5	71.8	NT	67.4
Barfin flounder ⁵	NT	34.3	32.6	25.7

¹ U.V. dose: 1.0×10⁴ μW·sec/cm².

² Ozonization: 1.0 mg/l TROs for 8.5 min for Japanese flounder, 0.5 mg/l for 5 min for Barfin flounder.

³ Electrolyzation: 0.5 mg/l chlorine for 5 min.

⁴ Tank size: 0.5 t, Number of fish: 2000, Duration: 49 days, Feeding: 1 time/day.

⁵ Tank size: 0.5 t, Number of fish: 6500-7100, Duration: 30 days, Feeding: 2 times/day.

cultured in UV irradiated, ozonized and electrolyzed seawater are shown in Table 5. No statistically significant differences in survival rates were found between the three groups of fish cultured with treated water.

Ozonized and electrolyzed seawater have been demonstrated to be effective for disinfecting equipment used in aquaculture^{15,16)} and ozonized seawater is effective for disinfecting fertilized barfin flounder eggs contaminated with viral nervous necrosis virus.¹⁷⁾ Therefore, ozonization and electrolyzation of seawater seem to be effective methods for disinfection of the water for fish culture.

CONCLUSION

Gram negative bacteria and fish rhabdoviruses, herpesviruses and iridoviruses were killed when UV irradiated at the dose of $10^4 \mu W \cdot \text{sec}/\text{cm}^2$. Standard, inexpensive UV lamps can irradiate at that dosage and may be suitable for hatcheries or culturing stations that have problems caused by these microorganisms. This would be the best method for disinfection of UV susceptible pathogens. Water contaminated with Gram-positive bacteria, fish birnaviruses, fish reoviruses, fish nodaviruses and aquatic fungi that showed lower susceptibility should be disinfected with ozonization, electrolyzation or high quality UV lamps.

Disinfection of wastewater is necessary to prevent pathogenic contamination of the environments. Electrolyzation is easy to scale up and can be used to treat a large volume of water, thus making it a suitable method for disinfecting wastewater.

ACKNOWLEDGMENT

This research was supported in part by Grant in Aid for Scientific research (B)(2) No.09556041 and (B)(2) 13556027 under the Ministry of Education, Science, Sports and Technology, Japan.

REFERENCES

1. Vlasenko M.I. Ultraviolet rays as a method for the control of diseases of fish eggs and young fishes. *Problems of Ichthyol.* 1969; 9: 697-705.
2. Hoffman G.L. Disinfection of contaminated water by ultraviolet irradiation, with emphasis on whirling disease (*Myxosoma cerebralis*) and its effect on fish. *Trans. Amer. Fish. Soc.* 1974; 3: 541-550.
3. Noormandau D.A. Progress report, Project F-14-R-3, State of New Hampshire, 1968.
4. Kimura T, Yoshimizu M, Tajima K, Ezura Y, Sakai M. Disinfection of hatchery water supply by ultraviolet (U.V.) irradiation- I. Susceptibility of some fish-pathogenic bacterium and microorganisms inhabiting pond waters. *Nippon Suisan Gakkaishi* 1976; 42: 207-211.
5. Kimura T, Yoshimizu M, Tajima K, Ezura Y. Disinfection of hatchery water supply by ultraviolet (U.V.) irradiation- II. U.V. susceptibility of some fish pathogenic fungi. *Fish pathol.* 1980; 14: 133-137.
6. Yoshimizu M, Takizawa H, Kimura T. U.V. susceptibility of some fish pathogenic viruses. *Fish pathol.* 1986; 21: 47-52.
7. Yoshimizu M, Sami M, Kohara M, Yamazaki T and Kimura T. Detection of IHNV in hatchery water by molecular filtration method and effectiveness of U.V. irradiation on IHNV infectivity. *Nippon Suisan Gakkaishi* 1991; 57: 555-560.
8. Kimura T, Yoshimizu M, Atoda M. Disinfection of hatchery water supply by ultraviolet (U.V.) irradiation-III. Effect of disinfection of hatchery water supply by ultraviolet irradiation on hatching rate of salmonid eggs. *Fish pathol.* 1980; 14: 139-142.
9. Yoshimizu M, Hyuga S, Oh M.-J., Ito S, Ezura Y. Disinfectant effect of oxidant produced by ozonization of sea water on fish pathogenic viruses, bacteria and ciliata. In "Diseases in Asian Aquaculture II", Asian Fisheries Society, Manila, Phillipines 1995; pp. 203-209.
10. Itoh S, Yoshimizu M, Ezura Y. Disinfectant effects of low level of total residual oxidants in artificial seawater on fish pathogenic microorganisms. *Nippon Suisan Gakkaishi* 1997; 63: 97-102.
11. Itoh S, Yoshimizu M, Oh M.-J., Hyuuga S, Watanabe K, Hayakawa Y, Ezura Y. Effects of ozonized seawater on bacterial pollution and survival of cultured flounders (*Paralichthys olivaceus* and *Verasper moseri*). *Suisanzoshoku* 1996; 44: 457-463.
12. Kasai H, Watanabe K, Yoshimizu M. Disinfectant effects of hypochlorite produced by batch electrolytic system on fish pathogenic bacteria and virus. *Suisanzoshoku* 2001; 49: 237-241.
13. Kasai H, Ishikawa A, Hori Y, Watanabe K, Yoshimizu M. Disinfectant effects of electrolyzed salt water on fish pathogenic bacteria and viruses. *Nippon Suisan Gakkaishi* 2000; 66: 1020-1025.
14. Kasai H, Watanabe K, Yoshimizu M. Bactericidal effect of continuous flow electrolyzer on hatchery waste-seawater. *Nippon Suisan Gakkaishi* 2001; 67: 222-225.
15. Watanabe K, Yoshimizu M. Disinfection of equipments for aquaculture and fertilized eggs by ozonated seawater. *Fish pathol.* 1998; 33: 145-146.
16. Watanabe K, Yoshimizu M. Disinfection of equipment for aquaculture by electrolyzed seawater. *Nippon Suisan Gakkaishi* 2001; 67: 304-305.
17. Watanabe K, Yoshimizu M. Disinfection of viral nervous necrosis contaminated fertilized eggs by ozonated seawater. *Nippon Suisan Gakkaishi* 2000; 66: 1066-1067.