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CALLUS FORMATION AND PROTOPLAST ISOLATION IN RICE (*Oryza sativa* L.) AND ITS RELATED SPECIES

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Introduction

Rice is one of the most important staple foods in world and various kinds of traditional breeding methods have played an important role in rice improvement. In addition, some new techniques such as tissue cultures and gene engineering are essential to raise the efficiency of rice breeding. In culture techniques, the success of callus formation, protoplast isolation and plant regeneration from the various tissues or organs are required for the universal genotypes.

Especially protoplast isolation and plant regeneration from the calli using a wide range of materials are indispensable for the use of somaclonal selection and production of somatic hybrids and cybrids.

In this paper the authors examined the effects of various genotypes involving 12 species of the genus *Oryza* for callus formation and protoplast isolation.

Materials and Methods

The genome constitution of species and the number of strains used are shown in Table 1. Dehulled seeds were surface sterilized using 70% alcohol for 1 minute, then by 10% commercial bleach for 2-3 minutes, followed by washing 2-3 times with sterilized distilled water. 10 seeds per strain were planted on the N6 medium supplemented with 4.4mg/l 2,4-D, 50g/l sucrose and 10g/l agar, and incubated in the darkness at 27°C. Calli from the seeds were subcultured every 10 days. Callus growth was assessed by measuring the diameter of callus 50 days after seed planting. The interspecific variation of callus growth was evaluated by Duncan's multiple range test. In this calculation, intraspecific variance were estimated by means of strains within the same species except for *O. nivara* which was estimated from variance among individuals.

Seed calli (0.5g) at 90 days after plating for the 40 species were added to 10ml

Table 1. 12 species of the genus, *Oryza* used for the experiments

| Species | Genome | No. of strains |
|-------------------------|-------------------------------|----------------|
| <i>O. sativa</i> | AA | 63 |
| <i>O. glaberrima</i> | A ^a A ^g | 8 |
| <i>O. rufipogon</i> | AA | 38 |
| <i>O. punctata</i> | BBCC, BB | 5 |
| <i>O. minuta</i> | BBCC | 5 |
| <i>O. officinalis</i> | CC | 8 |
| <i>O. australiensis</i> | EE | 5 |
| <i>O. latifolia</i> | CCDD | 5 |
| <i>O. gradiglumis</i> | CCDD | 5 |
| <i>O. brachyantha</i> | FF | 5 |
| <i>O. barthii</i> | A ^a A ^g | 5 |
| <i>O. nivara</i> | AA | 1 |
| Total | | 153 |

Table 2. Enzyme solution for protoplast isolation

| | |
|---------------------------|-------|
| Driselase | 1% |
| Cellulase "Onozuka" RS | 2% |
| Cellulase "Onozuka" R-10 | 2% |
| Macerozyme R-10 | 2% |
| Hemicellulase | 1% |
| Pectoryase Y-23 | 0.1% |
| Potassium dextran sulfate | 0.2% |
| Albumin | 0.05% |
| Sorbitol | 0.35M |
| Mannitol | 0.35M |
| MES | 20mM |
| MgCl ₂ | 5mM |
| pH | 5.6 |

enzyme solution (Table 2) and incubated in the darkness on a reciprocal shaker at 50 rpm for 6 hours at 28°C. The protoplast enzyme mixture was filtered through a 42 μ m nylon filter. Filtrated protoplasts were used for microscopic observation for counting.

Yields of protoplast were calculated by hemacytometer. The experiments were repeated nine times, and the average number of nine replications was transformed to a number of protoplasts per one gram of fresh weight of calli.

Results

1. Callus induction and growth

Callus induction was achieved in the strains of 12 species except for a strain of *O. rufipogon* and a strain of *O. latifolia* as shown in Table 3. In the cultivated species, *Oryza sativa* L., a considerable range of variation was observed in the callus growth both in Japonica and Indica type. It is noted that the mean diameter of calli in Japonica was significantly larger than that in Indica after 50 days of culture. In addition, the callus growth of African rice (*Oryza glaberrima*) was nearly equivalent to that of Indica. The wild species indicated an inferior growth as compared with the cultivated species. According to Duncan's multiple range test (Table 4), the interspecific variation of callus growth was classified into 4 or 5 groups.

2. Protoplast isolation

An isolation of intact protoplasts was achieved in 15 strains of Japonica, 12 strains of Indica in *Oryza sativa*, 2 strains of *O. officinalis* and *O. barthii* and 16 strains of *O. rufipogon*. Different protoplast yields in Japonica strains were obtained (Table 5). It is noted that the some superior cultivars such as Koshi-

Table 3. Comparison of callus growth among 12 species

| Species | No. of strains | No callus formation | Diameter of callus(mm) | | | | | | | | | | Mean | |
|-------------------------|----------------|---------------------|------------------------|----|----|----|----|----|----|---|---|---|------|-----|
| | | | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | | 10 |
| <i>O. Sativa</i> | 63 | | 1 | 3 | 10 | 5 | 14 | 15 | 4 | 6 | 3 | 2 | 5.4 | |
| Japonica | 30 | | | | 1 | 1 | 7 | 8 | 2 | 6 | 3 | 2 | 6.7 | |
| Indica | 33 | | 1 | 3 | 9 | 4 | 7 | 7 | 2 | | | | 4.3 | |
| <i>O. glaberrima</i> | 8 | | | | 2 | 1 | 2 | 2 | 1 | | | | 4.9 | |
| <i>O. rufipogon</i> | 38 | 1 | 1 | 7 | 19 | 8 | | 2 | | | | | 3.0 | |
| <i>O. punctata</i> | 5 | | | 3 | 2 | | | | | | | | 2.4 | |
| <i>O. minuta</i> | 5 | | 3 | 2 | | | | | | | | | 1.3 | |
| <i>O. officinalis</i> | 8 | | 5 | 2 | 1 | | | | | | | | 1.3 | |
| <i>O. australiensis</i> | 5 | | 1 | 1 | 3 | | | | | | | | 2.5 | |
| <i>O. latifolia</i> | 5 | 1 | 1 | 3 | | | | | | | | | 1.7 | |
| <i>O. gradiglumis</i> | 5 | | | 1 | 2 | 2 | | | | | | | 3.0 | |
| <i>O. brachyantha</i> | 5 | | 2 | 3 | | | | | | | | | 1.1 | |
| <i>O. barthii</i> | 5 | | | 1 | 2 | 1 | | 1 | | | | | 3.6 | |
| <i>O. nivara</i> | 1 | | | | 1 | | | | | | | | 3.0 | |
| Total | 153 | 2 | 4 | 10 | 26 | 42 | 17 | 16 | 20 | 5 | 6 | 3 | 2 | 2.7 |

Note: Diameter after 50 days from plating.

Table 4. Duncan's multiple range test for the interspecific variation

| Species | Mean diameter of callus (mm) | Duncan's test (p=0.05) |
|-----------------------------|------------------------------|------------------------|
| <i>O. sativa</i> (Japonica) | 6.7 | a* |
| <i>O. glaberrima</i> | 4.9 | b |
| <i>O. sativa</i> (Indica) | 4.3 | bc |
| <i>O. barthii</i> | 3.6 | cd |
| <i>O. rufipogon</i> | 3.0 | de |
| <i>O. nivara</i> | 3.0 | de |
| <i>O. gradiglumis</i> | 3.0 | de |
| <i>O. australiensis</i> | 2.5 | def |
| <i>O. punctata</i> | 2.4 | efg |
| <i>O. latifolia</i> | 1.7 | fgh |
| <i>O. officinalis</i> | 1.3 | gh |
| <i>O. minuta</i> | 1.3 | gh |
| <i>O. brachyantha</i> | 1.1 | h |

Note *: Means designated by the same letter show no significance.

hikari, Shiokari and Kinmaze showed a high yield of protoplasts. In Indica type, only one glutinous strain (Thaiglu. rice) from Thailand indicated the highest yield than the other strains (Table 6). In the strains from wild species, the protoplast yields varied between 0 and 4.0×10^9 protoplasts per g fresh weight of calli (Fig. 1).

Table 5. Relation between callus growth and protoplast yield in Japonica strains of *O. sativa* L.

| Name of strains | Diameter of callus (mm) | No. of protoplasts (x10 ⁹ /gfw) |
|-----------------|-------------------------|--|
| Norin 10 | 9.2 | 1.1 |
| Kochihibiki | 8.4 | 0.1 |
| Kochikaze | 8.2 | 0.2 |
| Todoroki-wase | 7.6 | 0.6 |
| Reimei | 6.6 | 0.4 |
| Jukkoku | 6.3 | 0.3 |
| Reihou | 6.1 | 0.4 |
| Shiokari | 5.8 | 3.1 |
| Akibare | 5.4 | 0.4 |
| Houyoku | 5.4 | 0.6 |
| Kinmaze | 4.9 | 2.0 |
| Tangin-bouzu | 4.9 | 3.2 |
| Akihikari | 4.5 | 1.3 |
| Koshihikari | 3.8 | 4.0 |
| Hokuriku 100 | 3.4 | 0.4 |
| Mean | 6.0 | 1.2 |

Table 6. Relation between callus growth and protoplast yield in Indica strains of *O. sativa* L.

| Name of strains | Diameter of callus (mm) | No. of protoplasts (x10 ⁹ /gfw) |
|-----------------|-------------------------|--|
| Calrose 76 | 7.0 | 1.4 |
| Thai-glu. rice | 6.7 | 6.1 |
| Assam III | 6.3 | 0.1 |
| Calrose | 5.9 | 0.4 |
| Dalashaita | 4.5 | 0.0 |
| Surjumkhi | 4.5 | 0.0 |
| Bhutmuri 36 | 4.2 | 0.04 |
| Dular | 3.5 | 0.1 |
| Hu-nan-zao | 3.1 | 0.2 |
| Mitsutari | 3.0 | 0.2 |
| Charnock | 2.4 | 0.03 |
| Mushakdanti | 2.2 | 0.02 |
| Mean | 4.4 | 0.7 |

between callus growth and protoplast yield (Fig. 1). But the same tendency was seen in Indica strains. For *O. officinalis* and *O. barthii*, the number of strains used for protoplast isolation were only two, however the tendency of relation was almost similar with that of *O. rufipogon*.

Discussion

Intervarietal variation in callus induction from rice seeds was examined using Japonica, Indica and their hybrids. According to MAEDA⁹⁾, there was no difference between Japonica and Indica for the nature and growth rate of callus. However, the present results indicated a superiority in Japonica type, supporting the findings by ABE and SASAHARA¹⁾. In addition, there was a wide range of variation in the both Japonica and Indica type. Therefore, it is possible to screen the strain showing a high callus formation in their respective types. In the anther culture, MIAH *et al.*¹¹⁾ elucidated the inheritance mode of callus formation and suggested the possibility of improvement in callus induceability using the Indica genotypes.

The callus formation and plant regeneration in African rice (*Oryza glaberima*) were also reported (FATOKUN and YAMADA⁶⁾). In this experiment, there was no significant difference in callus development between Indica and African rice. Most of the strains in wild species succeeded in callus formation though the callus growth was rather inferior. Therefore, it is necessary to improve certain media suitable for these species.

Protoplast isolation in rice succeeded by using seed calli or suspension cells (YAMADA *et al.*¹⁹, COULIBAR *et al.*⁴, FUJIMURA *et al.*⁷) and TORIYAMA and HINATA¹⁵), by anther calli (CAL *et al.*³, WAKASA *et al.*¹⁷, TORIYAMA and HINATA¹⁵) and young leaves or mesophyll cells (DEKA and SEN⁵, SHIH *et al.*¹³). The authors isolated protoplasts from the seed calli induced from both cultivated and wild species. It is noted that Japanese familiar cultivars, such as Koshihikari, Kinmaze and Shiokari showed a high yielding ability of protoplasts. In Indica, a glutinous strain from Thailand produced the highest yield of protoplast in this experiment. In the wild species, *Oryza rufipogon*, there were a small number of strains possessing a high yielding ability. In general, there is no intimate relation between the callus growth and protoplast yield.

The plant regeneration from isolated rice protoplasts were reported by many researchers (FUJIMURA *et al.*⁷, YAMADA *et al.*¹⁹, TORIYAMA *et al.*¹⁶), ABDULAH *et al.*²) and COULIBALY *et al.*⁴). But there were quite a few reports for protoplast culture from wild species in *Oryza* (HAYASHI *et al.*⁸). Now, we have been attempting to culture protoplasts from wild species and a detailed examination is now progressing.

Since the formation of somatic hybrids through protoplast fusion was demonstrated between rice and soybean (NIIZEKI *et al.*¹²), and between cultivated rice and four wild *Oryza* species (HAYASHI *et al.*⁸), this technique will be effectively used for rice breeding for the transfer of male sterile cytoplasm and favorable characters from related species through somatic hybrids or cybrids.

Summary

In vitro culture methods is quite important techniques for rice breeding. Especially callus induction and protoplast isolation are essentially basic techniques for the use of somatic hybrids or cybrids.

The induction and growth of seed calli was investigated examining 153 strains of *Oryza sativa* L. and 11 species of the genus *Oryza*. A basal N6 medium containing 4.4 mg/l 2,4-D, 50 g/l sucrose and 10 g/l agar was used for callus induction from seeds. Most of the materials except a strain of *O. rufipogon* and a strain of *O. latifolia*, showed successful callus induction. A remarkable difference in callus growth was observed among the strains of wild species as well as among those of cultivated species. Seed calli from the two cultivated species, *O. sativa* and *O. glaberrima* showed better growth than the other species in *Oryza*.

Protoplast isolation from seed calli was conducted by using an enzyme solution under conditions of 6 hours at 28°C. The protoplast isolation from the callus was achieved in 15 Japonica strains, 10 Indica strains, one strain of *O. officinalis*, two strains of *O. barthii* and 16 strains of *O. rufipogon*. In the relation between callus growth and protoplast yield, the strain showing prolific callus was not always superior in the yield of protoplasts.

Literature Cited

1. ABE, T. and T. SASAHARA : Variations in callus formation from seeds in *Japonica*, *Indica*, their hybrids and large grain varieties in rice. *Japan. J. Breed.* **32** : 53-60. 1982
2. ABDULAH, R. and E. COCKING : Efficient plants regeneration from rice protoplasts through somatic embryogenesis. *Biotechnology* **4** : 1087-1090. 1986
3. CAI, Q., Y. QIAN, Y. ZHOU and S. WU : A further study on the isolation and culture of rice (*Oryza sativa* L.) protoplasts. *Acta Bot. Sinica.* **20** : 97-102. 1987
4. COULIBALY, M.Y. and Y. DEMARLY : Regeneration of plantlets from protoplasts of rice, *Oryza sativa* L. *Z. Pflanzenzuchtg.* **96** : 79-81. 1986
5. DEKA, P.C. and S.K. SEN : Differentiation in calli originated from isolated protoplasts of rice (*Oryza sativa* L.) through plating technique. *Mol. Gen. Genet.* **145** : 239-243. 1976
6. FATOKUM, C.A. and Y. YAMADA : Variations in callus formation and plant regeneration in African rice *Oryza glaberrima* Steud. *J. Plant Physiol.* **117** : 179-183. 1984
7. FUJIMURA, T., M. SAKURAI, H. AKAGI, T. NEGISHI and A. HIROSE : Regeneration of rice plants from protoplasts. *Plant Tissue Culture Lett.* **2** : 74-75. 1986
8. HAYASHI, Y., J. KYOZUKA and K. SHIMAMOTO : Hybrids of rice (*Oryza sativa* L.) and wild *Oryza* species obtained by cell fusion. *Mol. Gen. Genet.* **214** : 6-10. 1988
9. MAEDA, E. : Varietal difference in callus formation of rice seeds under sterile culture. *Proc. Crop Soc. Japan.* **36** : 233-239. 1956
10. MAEDA, E. : Isolation of protoplast from seminal roots of rice. *Proc. Crop Soc. Japan.* **40**:397-398. 1971 (In Japanese)
11. MIAH, M.A.A., E.D. EARLE and G.S. KHUSH : Inheritance of callus formation ability in anther culture of rice, *Oryza sativa* L. *Theor. Appl. Genet.* **70** : 113-116. 1985
12. NIIZEKI, M., M. TANAKA, S. AKADA, A. HIRAI and K. SAITO : Callus formation of somatic hybrid of rice and soy bean and characteristics of the hybrid callus. *Japan. J. Genet.* **60** : 81-92. 1985
13. SHIH, C-Y., T-C. TSENG and C-S. HWA : Subcellular changes during protoplast isolation of *Oryza sativa* L. *Bot. Bull. Acad. Sinica.* **20** : 50-57. 1979
14. TORIYAMA, K. and H. HINATA : Cell suspension and protoplast culture in rice. *Plant Sci.* **41** : 179-183. 1985
15. TORIYAMA, K. and H. HINATA : Panicle culture in liquid media for obtaining anther calli and protoplasts in rice. *Japan. J. Breed.* **35** : 449-452. 1985
16. TORIYAMA K. and K. HINATA : Haploid and Diploid plant regeneration from protoplast of anther culture in rice. *Theor. Appl. Genet.* **73** : 16-19. 1986
17. WAKASA, K., K. KOBAYASHI and H. KANEDA : Colony formation from protoplasts of nitrate reductase deficient rice cell lines. *Japan. J. Breed.* **34** (Suppl. 2) : 28-29. 1984 (In Japanese)
18. YAMADA, Y., Z-Q. YANG and D-T. TANG : Regeneration of rice plants from protoplast. *Rice Genetics Newsletter* **2** : 94. 1985
19. YAMADA, Y., Z-Q. YANG and D-T. TANG : Plant regeneration from protoplast-derived callus of rice (*Oryza sativa* L.). *Plant Cell Reports* **5** : 85-88. 1986