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Four hydrogen-bonding motifs in oligonucleotides

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Abstract

The design and synthesis of imidazo[5',4':4,5]pyrido[2,3-*d*]pyrimidine nucleosides and 1,8-naphthyridine *C*-nucleosides with the aim of developing new base pairing motifs consisting of four hydrogen bonds is described.

Keywords: Nucleoside, *C*-nucleoside, imidazopyridopyrimidine, 1,8-naphthyridine, hydrogen bond, thermal stability, oligonucleotide

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1. Introduction

DNA is the storage and carrier of genetic information in all living organisms, and generally forms a right-handed double helix with a complementary DNA molecule. The structure of the DNA double helix is based on the Watson-Crick hydrogen bonding (H-bonding) of adenine:thymine (A:T pair, two H-bonds) and guanine:cytosine (G:C pair, three H-bonds) base pairings.¹

H-bonding interaction plays a critical role in not only conserving and transmitting genetic information but also in double helix stability. Consequently, much attention has been devoted to the synthesis of artificial oligodeoxynucleotides (ODNs) including unnatural nucleobases, which could form a more stable, higher-ordered structure with DNA, RNA and protein, or DNA itself, with application to biochemistry, biotechnology, and medicinal chemistry. In contrast to the research directed toward thermally stable base pairing between natural and unnatural nucleobases, few studies have been reported on the possibility of alternative base pairing consisting of new H-bonding motifs. For example, Benner *et al.* proposed new base pairing, that is, 6-aminopyrazin-2-one: 5-aza-7-deazaisoguanine (V:J), isoguanine: isocytosine (isoG:isoC), and 2,6-diaminopyrimidine: 1-methyl-pyrazolo[4,3-*d*]-pyrimidine-5,7(4*H*,6*H*)-dione (κ:p) base pairs (Figure 1), and investigated their enzymatic incorporation into DNA and RNA with the aim of "extending the genetic alphabet."²

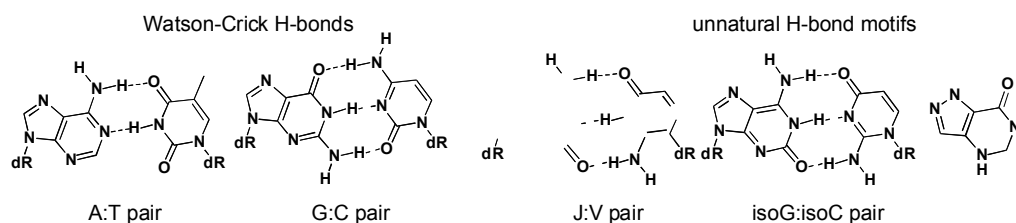


Figure 1. Watson-Crick hydrogen-bonds and examples of unnatural hydrogen-bonds.

We envisioned a new base-pairing motif, which would be more thermally stable than the Watson-Crick H-bonding. We proposed new base pair combinations consisting of four H-bonds, imidazo[5',4':4,5]pyrido[2,3-*d*]pyrimidine-4,7(5*H*,6*H*)-dione (iO^O): 4,7-diamino imidazo[5',4':4,5]pyrido[2,3-*d*]pyrimidine (iN^N) pair and 4-aminoimidazo-[5',4':4,5]pyrido[2,3-*d*]pyrimidine-7(6*H*)-one (iN^O): 7-aminoimidazo[5',4':4,5]pyrido [2,3-*d*]pyrimidine-4(5*H*)-one (iO^N) pair, which are called to be first generation four H-bonding motifs as shown in Figure 2.³

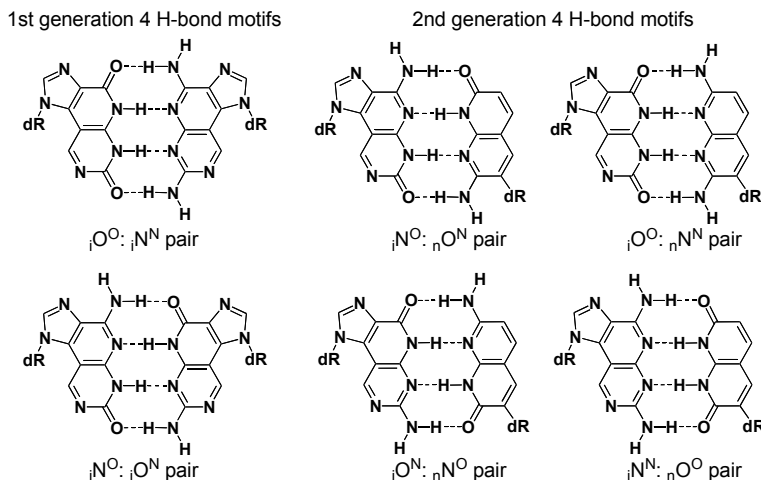


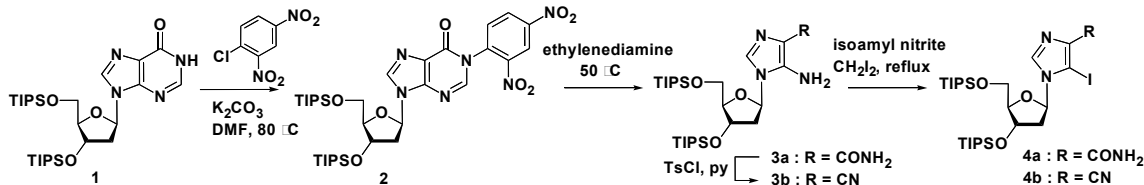
Figure 2. Four hydrogen-bonding motifs.

As second generation motifs, we also designed four sets of naphthyridine:imidazopyridopyrimidine pairs, iN^O :3-substituted 2-amino-7-hydroxy-1,8-naphthyridine (nO^N) pair, iO^N :6-substituted 2-amino-7-hydroxy-1,8-naphthyridine (nN^O) pair, iO^O : 2,7-diamino-1,8-naphthyridine (nN^N) pair, and iO^O : 2,7-dihydroxy-1,8-naphthyridine (nO^O) pair.⁴ These motifs were re-designed to satisfy shape complementarity resembling a pyrimidine:purine base pair in natural DNA. In this report, we summarize the synthesis of these nucleosides and the results of thermal stability when these nucleosides are incorporated into complementary positions of ODNs.

2. Construction of imidazo[5',4':4,5]pyrido[2,3-*d*]pyrimidine nucleosides

2.1 Synthesis of imidazole 2'-deoxyribosides from 2'-deoxyinosine

The most straightforward synthesis of the desired nucleosides was thought to be through intramolecular cyclization of the 5-pyrimidinylimidazole nucleosides, which would be prepared from the Stille coupling reaction of a 5-iodoimidazole nucleoside with an appropriate tributylstannylpyrimidine. The 5-iodoimidazole nucleosides can be synthesized from 2'-deoxyinosine derivative **1**.⁵ The synthesis of the 5-iodoimidazole nucleosides **4a** and **4b** was first carried out as shown in Scheme 1. The 2'-deoxyinosine derivative **1** was treated with 1-chloro-2,4-dinitrobenzene in the presence of K_2CO_3 to give **2**. Conversion of **2** to **3a** was achieved by treatment with ethylenediamine to give **3a**.^{5b} Treatment of **3a** with *p*-toluenesulfonyl chloride in pyridine gave the 5-aminoimidazole-4-carbonitrile derivative **3b**. Compounds **3a** and **3b** were converted to their 5-iodo derivatives by treatment with isoamyl nitrite in diiodomethane to give **4a** and **4b**, respectively.⁶

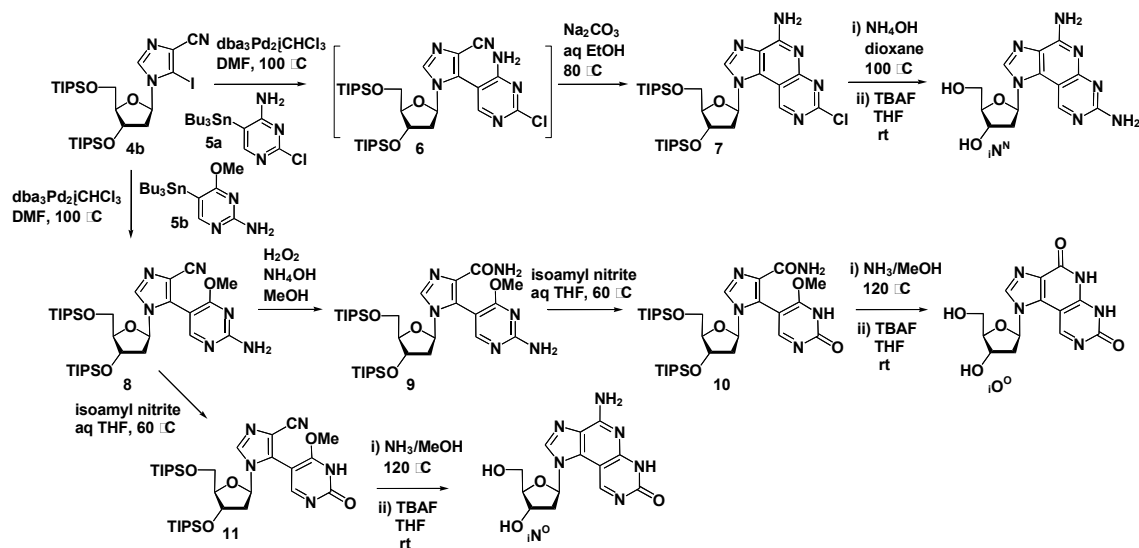


Scheme 1

2.2 Synthesis of imidazo[5',4':4,5]pyrido[2,3-*d*]pyrimidine nucleosides

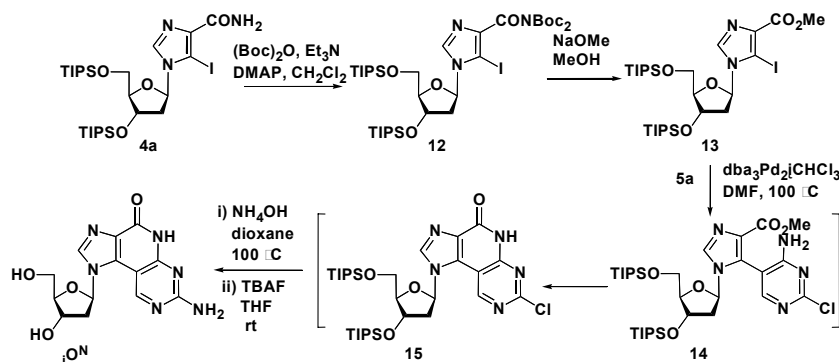
The synthesis of the diamino derivative iN^N was attempted (Scheme 2). When a mixture of the 5-iodoimidazole nucleoside **4b** and the tributylstannyl pyrimidine **5a**⁴ was heated in DMF in the presence of $dba_3Pd_2 \cdot CHCl_3$, two fluorescent spots corresponding **6** and **7** were detected by TLC analysis. Therefore, treatment of the mixture with Na_2CO_3 in aqueous EtOH gave **7**, which was further treated with a mixture of 1,4-dioxane and NH_4OH gave the diamino derivative. Deprotection of the silyl groups with tetrabutylammonium fluoride (TBAF) gave the free nucleoside iN^N .

Preparation of the dioxo derivative iO^O and the amino-oxo derivative iN^O was achieved via the Stille coupling reaction between **4b** and **5b**. When a mixture of **4b** and **5b**⁴ was heated with $dba_3Pd_2 \cdot CHCl_3$, **8** was obtained. To convert **8** into the dioxo derivative iO^O , the 4-cyano group of **8** was hydrolyzed with H_2O_2 in the presence of NH_4OH to give the 4-carboxamide derivative **9**. Hydrolytic deamination of the amino group in **9** by treatment with isoamyl nitrite in THF containing a small amount of H_2O gave **10**.⁷ When the resulting **10** was heated with $NH_3/MeOH$ in a steel container, the dioxo derivative iO^O was obtained after deprotection of the silyl groups. The amino-oxo derivative iN^O was synthesized also starting from **8** using the same procedure, that is, hydrolytic deamination, followed by the intramolecular cyclization and the deprotection.



Scheme 2

The synthesis of the remaining oxo-amino derivative iO^N was next attempted, which was rather troublesome relative to those of the other tricyclic nucleosides iN^N , iO^O and iN^O . The desired iO^N was thought to be obtained via the intramolecular cyclization of **9**, using the same method as for iO^O .



Scheme 3

However, when **9** was treated $NH_3/MeOH$ at 120 °C for 48 h, the desired cyclized product was not obtained, and **9** was recovered in quantitative yield. After several attempts, iO^N was obtained efficiently as illustrated in Scheme 3. First, the 4-carboxamide group of **4a** was converted to the methyl ester according to our previous method.⁸ Thus, treatment of **4a** with di-*tert*-butyl dicarbonate in the presence of Et_3N and DMAP gave **12**, which was then treated with NaOMe in MeOH to give **13**. After the Stille coupling reaction of **13** with **5a** in the presence of $dba_3Pd_2 \cdot CHCl_3$, the resulting crude products, probably including the intermediates **14** and **15**, was treated with a mixture of NH_4OH and 1,4-dioxane at 100 °C for 60 h in a steel container to give the oxo-amino derivative iO^N after further deprotection.

3. Thermal stability of the DNA duplexes containing base pairings between the imidazopyridopyrimidine bases

To introduce the imidazo[5',4':4,5]pyrido[2,3-*d*]pyrimidine nucleosides into ODNs by the phosphoramidite method, these nucleosides must be converted into the corresponding phosphoramidites with appropriate protecting groups on each nucleobase. The phosphoramidite units **16**, **17**, **18**, and **19** shown in Figure 3 were used for the synthesis of ODNs.

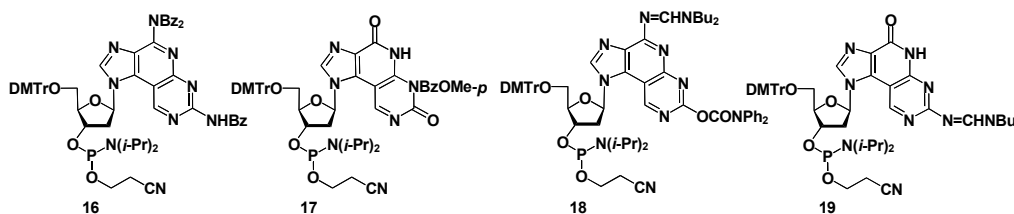


Figure 3

Thermal stability of duplexes, which contained one molecule of the iN^N , iO^O , iN^O , or iO^N in their **X** or **Y** position, was studied by thermal denaturation in a buffer of 0.01 M sodium cacodylate (pH 7.0) containing 0.1 M NaCl (the left column of Table 1). Each profile of the thermal denaturation showed a single transition corresponding to a helix-to-coil transition to give melting temperatures (T_m s).

The melting temperature (T_m s) values and their differences from that of A:T pair(s) at the **X:Y** position of the ODN (ΔT_m s) are shown in Table 1. Contrary to our expectation, when one molecule of the tricyclic nucleosides was incorporated into each strand, the thermal stabilities of the duplexes did not increase. The T_m values of the duplexes were all slightly lower than those of the duplexes containing a natural G:C or A:T pair at the corresponding positions, and no apparent specificity of base pairing was observed.

On the other hand, when three molecules of the tricyclic nucleosides were consecutively incorporated into the center of each ODN (the right column of Table 1), the thermal stabilization of the duplexes due to the specific base pairings was observed. The best result was obtained in the case of the duplex containing the iN^O : iO^N pair ($T_m = 84.0$ °C). The value was between 18.2 and 23.5 °C higher than those of the duplexes containing three consecutive G:C ($T_m = 65.8$ °C) and A:T pairs ($T_m = 60.5$ °C), respectively. Consequently, it was found that the iN^O : iO^N pair stabilized the duplex by about +6 and +8 °C per modification as compared with those of the duplexes containing the G:C and A:T pairs, respectively.

Table 1. Hybridization data of DNA duplex containing X:Y pair(s)

5' -GCACCGAAXAAACCACG-3'				5' -GCACCGAXXXAACCACG-3'			
3' -CGTGGCTT Ψ TTTGGTGC-5'				3' -CGTGGCT $\Psi\Psi\Psi$ TTTGGTGC-5'			
X	Y	T_m (°C) ^a	ΔT_m (°C) ^b	X	Y	T_m (°C) ^a	ΔT_m (°C) ^b
iN^N	iN^N	54.5	\$6.0	iN^N	iN^N	55.7	\$4.8
	iO^O	56.3	\$4.2		iO^O	70.6	+10.1
	iN^O	54.5	\$6.0		iN^O	49.4	\$11.1
	iO^N	56.1	\$4.4		iO^N	51.7	\$8.8
iO^O	iN^N	56.0	\$4.5	iO^O	iN^N	67.1	+6.6
	iO^O	54.3	\$6.2		iO^O	53.5	\$7.0
	iN^O	54.0	\$6.5		iN^O	58.3	\$2.2
	iO^N	54.3	\$6.2		iO^N	62.3	+1.8
iN^O	iN^N	55.1	\$5.4	iN^O	iN^N	46.3	\$14.2
	iO^O	54.0	\$6.5		iO^O	56.0	\$4.5
	iN^O	57.2	\$3.3		iN^O	70.0	+9.5
	iO^N	59.8	\$0.7		iO^N	84.0	+23.5
iO^N	iN^N	55.4	\$5.1	iO^N	iN^N	41.5	\$19.0
	iO^O	53.3	\$7.2		iO^O	57.5	\$3.0
	iN^O	58.4	\$2.1		iN^O	83.8	+23.3
	iO^N	56.9	\$3.6		iO^N	65.1	+4.6
G	C	61.9	+1.4	G	C	65.8	+5.3
A	T	60.5	0	A	T	60.5	0
A	G^c	53.8	\$6.7	A	G^c	47.2	\$13.3
A	A^c	52.4	\$8.1	A	A^c	43.1	\$17.4

^a3 μ M ODN in 0.01 M Na cacodylate (pH 7.0) containing 0.1 M NaCl. ^bThe ΔT_m values were obtained by subtracting data for the T_m possessing **X:Y** = A·T from that for each duplex. ^c mismatch.

The order of thermal stabilizing effects of the base-pair when three consecutively incorporated is as follows: **X:Y** = iN^O : iO^N (T_m = 84.0 °C) \geq iO^N : iN^O (T_m = 83.8 °C) $>$ iN^N : iO^O (T_m = 70.6 °C) $>$ iN^O : iN^O (T_m = 70.0 °C) $>$ iO^O : iN^N (T_m = 67.1 °C) $>$ iO^N : iO^N (T_m = 65.1 °C) $>$ iO^O : iO^N (T_m = 62.3 °C). From these data, it could be concluded that the H-bonding abilities between the imidazopyridopyrimidine bases are essential and affect the thermal stability of the duplex. However, a stacking interaction influence, arising from the extended aromatic surface of the imidazopyridopyrimidine bases, may also contribute to their stabilities.^{9,10}

Therefore, the stacking ability of the imidazopyridopyrimidine bases were compared with natural nucleobases using self-complimentary ODNs containing an unpaired **Z** unit (5'-ZCGCGCG-3'),¹⁰ in which each tricyclic base or natural base was introduced. The order of the stacking ability ($\Delta\Delta G^\circ$) was iN^O (5.6) $>$ iN^N (4.2) $>$ iO^N (3.8) $>$ **G** (2.5) $>$ iO^O (2.4) = **A** (2.4) $>$ **C** (1.3) $>$ **T** (1.0). Together with thermal stability and stacking ability data indicated that not only a number of H-bondings, but also stacking ability contributed the thermal stability of the DNA duplexes containing the imidazopyridopyrimidine bases. However, the base pair between the imidazopyrido-pyrimidine would have a similar C1'-C1' distance between purine:purine base pairs. The distance in the G (anti):A (anti) mismatch was reported to be 12.5 Å,¹¹ while that in a canonical Watson-Crick base pair is 10.5 (\pm 0.2) Å.¹² Therefore, we speculated that thermal

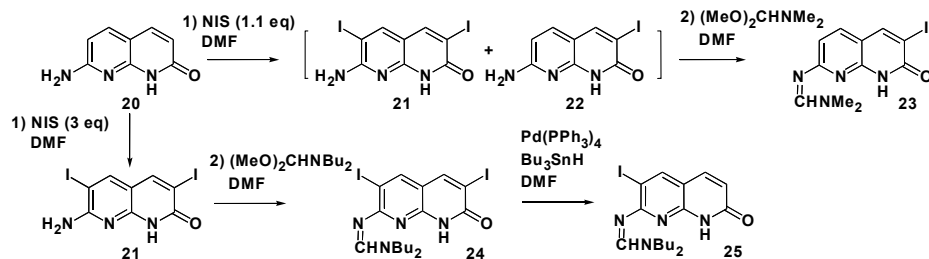
destabilization of the duplex occurs at both sides of these pairs. When one pair of the tricyclic nucleoside was incorporated into the duplex, the destabilization factor arising from disruption of the Watson-Crick base pairs next to the pair would be greater than the stabilization arising from the stable H-bonds. Consequently, the duplex would become less stable (Figure 4A). On the other hand, when the tricyclic nucleosides were consecutively incorporated into the duplex, the duplex would be thermally stabilized to a great extent since the base pairs between the tricyclic nucleosides has stable H-bonds and a strong stacking ability with the adjacent bases. These stabilization factors would be superior to the conformational destabilization around the boundary of the base pairs. (Figure 4B).



Figure 4. Hypothetical DNA structures.

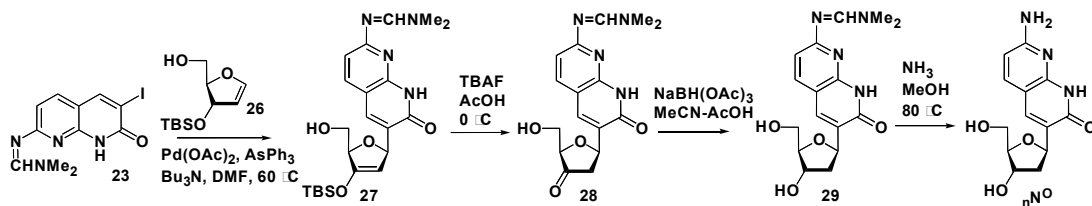
4. Designing and synthesis of 1,8-naphthyridine C-nucleosides

The base pair of the imidazopyridopyrimidines markedly stabilized a duplex when three of the pairs were consecutively incorporated into ODNs. However, incorporation of one pair into ODNs resulted in destabilization of the duplex relative to those containing A:T and G:C base pairs.³ These results showed shape complementarity resembling a pyrimidine:purine base pair in natural DNA is also important to stabilize DNA duplexes along with a number of H-bonds and base stacking. Since the goal of our continuing study is to develop base pairing motifs which stabilize and regulate DNA structures, including a double helix independent mode of incorporation of the new base pair(s), (i.e., one pair, three nonconsecutive pairs, and three consecutive pairs in this study), the novel 1,8-naphthyridine C-nucleosides possessing nN^N , nO^O , nN^O , and nO^N bases were designed.^{3,13} These C-nucleosides are expected to form four sets of naphthyridine: imidazopyridopyrimidine base pairing motifs ($iN^O:nO^N$, $iO^O:nN^N$, $iO^N:nN^O$, and $iN^N:nO^O$) with four H-bonds when these are incorporated into ODNs (Figure 2, 2nd generation). In addition, the new motifs can be regarded as a sort of ring expanded pyrimidine:purine type base pair (possessing C1'–C1' distances similar to the Watson-Crick base pair). Herein, we describe the synthesis of the 1,8-naphthyridine C-nucleosides nN^O , nO^N , nN^N and nO^O and the effects on the thermal stabilities of the ODNs containing the naphthyridine (nN^O and nO^N):imidazopyridopyrimidine (iO^N and iN^O) base pairing motifs.



Scheme 4

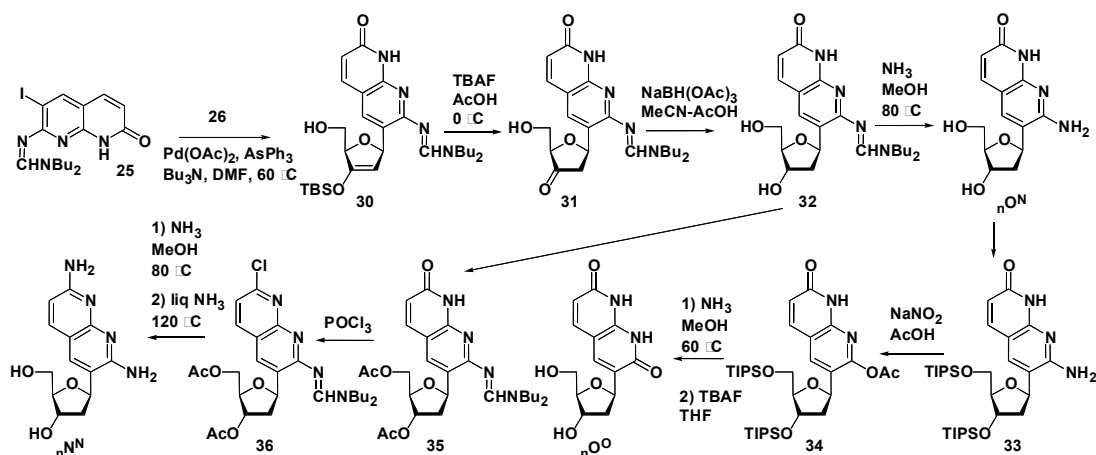
The synthetic route to the target compounds is illustrated in Scheme 4. Starting with 2-amino-7-hydroxy-1,8-naphthyridine (**20**),¹⁴ iodination by *N*-iodosuccinimide (NIS, 1.1 eq), followed by protection of the exocyclic amino group gave the 6-iodo-1,8-naphthyridine derivative **23**, a substrate for the synthesis of nO . For the synthesis of nN , the 3-iodo-1,8-naphthyridine derivative is required. Treatment of **20** with NIS (3 eq), followed by protection of the exocyclic amino group gave the 3,6-diiodo derivative **24**. Conversion of **24** into the 3-iodo derivative **25** was achieved by treatment of **24** with a stoichiometric amount of Bu_3SnH in the presence of $Pd(PPh_3)_4$.¹⁵



Scheme 5

After Heck coupling of the 6-iodo derivative **23** with the glycol **26**¹⁶ [$Pd(OAc)_2$, $AsPh_3$, tributylamine, DMF], followed by deprotection and reduction,¹⁷ the desired **29** was obtained (Scheme 5). In the same manner, the reaction of **25** with **26** afforded **30** (Scheme 6).

Treatment of **29** and **32** with $NH_3/MeOH$ gave the free nucleosides nO and nN , respectively. In Scheme 6, the preparation of the desired nO and nN was also shown. After silylation of nN , **33** was diazotized with $NaNO_2$ in AcOH to give **34**, which was treated with $NH_3/MeOH$, followed by deprotection to afford nO . The desired nN was also prepared from **32**. Chlorination of **35** in $POCl_3$ gave **36**, which was further treated with $NH_3/MeOH$ to remove protecting groups both at the sugar and base moieties. The residue was then heated in liq NH_3 in a steel container to afford nN .



Scheme 6

5. Thermal stability of the DNA duplexes containing base pairings between the naphthyridine and the imidazopyridopyrimidine bases

To incorporate both of the *C*-nucleosides into ODNs, nN^O and nO^N were converted into the corresponding phosphoramidites **38** and **39**, respectively. For the conversion of nO^N , the *N*-benzoyl group was the best choice for the exocyclic amino protecting group, and methyl *N,N*-diisopropylchlorophosphoramidite was used to give **39** because of purification problems when 2-cyanoethyl *N,N*-diisopropylchlorophosphoramidite was used.

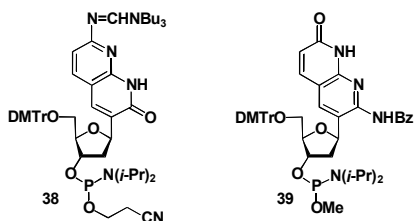


Figure 4

To investigate the base pairing properties of nN^O and nO^N , three classes of complementary duplexes were synthesized. As shown in Table 2, the first class contains one **X:Y** pair in the center of the duplexes (duplex 1). The second one contains three nonconsecutive **X:Y** pairs (duplex 2), and the last one is made up of three consecutive **X:Y** pairs (duplex 3). The thermal stability of all duplexes was measured by thermal denaturation in a buffer of 10 mM sodium cacodylate (pH 7.0) containing 0.001 M NaCl, since in a buffer containing 0.1 M NaCl, some duplexes showed T_m s higher than 95 °C.

As we expected, the pairs of $iO^N:nN^O$ and $iN^O:nO^N$ stabilized the duplex 1 by +9.4 °C and +8.6 °C, respectively. Although the $nN^O:nO^N$ pair stabilized the duplex by +2.3 °C, the $iO^N:iN^O$ pair destabilized the duplex as described above by –3.8 °C. The pronounced effects by $iO^N:nN^O$

and $iN^O:nO^N$ were emphasized in the duplex 2. Both pairs stabilized the duplexes by more than +30 °C, while the effects of $iO^N:iN^O$ and $nN^O:nO^N$ were insufficient despite the expected base pairing motifs with four H-bonds. In the duplex 3, not only the $iO^N:nN^O$ and $iN^O:nO^N$ pairs but also the $iO^N:iN^O$ and $nN^O:nO^N$ pairs stabilized the duplexes much more than G:C and A:T pairs, although the first pairs are generally considered more effective for thermal stability. From these results, it was concluded that the newly designed $iO^N:nN^O$ and $iN^O:nO^N$ base pairs thermally stabilized the duplex by nearly 10 °C more per pair than the A:T pair and 8 °C more than the G:C pair independent mode of incorporation of the new base pair(s) into the ODNs.

Table 2. Hybridization data of DNA duplexes containing **X:Y** pair(s)

duplex	X	Y	T_m (°C) ^a	ΔT_m (°C) ^b
duplex 1 5' -GCACCGAA X AAACCACG-3' 3' -CGTGGCTT Y TTTGGTGC-5'	iO^N	nN^O	57.2	+9.4
	iN^O	nO^N	56.4	+8.6
	iO^N	iN^O	44.0	\$3.8
	nN^O	nO^N	50.1	+2.3
	G	C	49.1	+1.3
	A	T	47.8	0
duplex 2 5' -GC X CCGAAXAAACC X CG-3' 3' -CGYGGCTT Y TTTGGYGC-5'	iO^N	nN^O	82.2	+34.4
	iN^O	nO^N	80.9	+33.1
	iO^N	iN^O	53.3	+5.5
	nN^O	nO^N	48.9	+1.1
	G	C	56.7	+8.9
duplex 3 5' -GCACCGA XXX AACCACG-3' 3' -CGTGGCT YYY TTGGTGC-5'	iO^N	nN^O	80.2	+32.4
	iN^O	nO^N	81.0	+33.2
	iO^N	iN^O	70.4	+22.6
	nN^O	nO^N	68.1	+20.3
	G	C	55.2	+7.4

^a3 μ M ODN in 0.01 M Na cacodylate (pH 7.0) containing **0.001 M NaCl**. ^bThe ΔT_m values were obtained by subtracting data for the T_m possessing **X:Y** = A:T from that for each duplex.

This effect is presumably caused by the four H-bonds and the stacking effect of the expanded aromatic surfaces with shape complementarity to a pyrimidine:purine base pair (Figure 5). Although some shift in the base pairing phase from the usual pyrimidine:purine base pairing would occur to complete the base pairing of $iO^N:nN^O$ and $iN^O:nO^N$ (see Figure 2), the effect of this shift should be negligible for the thermally stable duplex formation since both pairs stabilized the duplex independent mode of incorporation.

In conclusion, the ODNs containing the base pairing motifs $iO^N:nN^O$ and $iN^O:nO^N$ formed extremely thermally stable duplexes by cooperative effects of the four H-bonds, the stacking effects and shape complementarity. In addition, these motifs are specific, so that these would be versatile in stabilizing and regulating a variety of DNA structures.

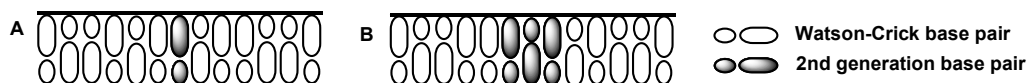


Figure 5. Hypothetical DNA structures.

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