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Author(s)
SATO, Gihei; MATSUBARA, Syuzo; ETOH, Shun-ichi; KODAMA, Hiroshi

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CULTIVATION OF SAMPLES OF HATCHER CHICK FLUFF,
FLOOR LITTER AND FECES FOR THE DETECTION OF
SALMONELLA INFECTION IN CHICKEN FLOCKS*

Gihei SATO, Syuzo MATSUBARA, Shun-ichi ETOH
and Hiroshi KODAMA
Department of Epizootiology
Faculty of Veterinary Medicine
Hokkaido University, Sapporo, Japan
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Salmonella status on a farm was investigated by the cultivation of samples of
hatcher chick fluff, floor litter and feces from a small number of chickens of
breeder flocks or progeny flocks. Five lots of progeny chicks were found to be
infected with Salmonella typhimurium or Salmonella enteritidis on the basis of
the detection of the organisms from litter and/or feces, and 2 of the 5 lots gave
Salmonella-positive fluffs. The hatcher-fluff-culturing method could not detect S.
typhimurium infection in the breeder chickens from which hatching eggs were
obtained.

The effect of the import of chicks carrying Salmonella on the Salmonella status
of chickens in Japan was discussed.

INTRODUCTION

It is well known that bacteriological examination of dead embryos is useful
in finding Salmonella carriers in stock birds, or in finding Salmonella infections
in progeny from the same hatch. Moreover, examination of dead embryos seems
to give information on Salmonella contamination of incubators. MIURA et al.
indicated that bacteriological examination of hatcher chick fluff samples, especially
from a series of hatches, may be used for the measurement of Salmonella
contamination in commercial hatcheries, in place of the examination of dead-in­
shell chick embryos. Recently SATO et al. described that Salmonella newington
infection of breeder chickens and of their progeny could be detected on a farm
not by the examination of dead embryos, but by that of fluff samples.

SNOEYENBOS et al.10) found that isolation of Salmonella from either floor- or
nest-litter samples was applicable as the index of flock infection. A formula
of the examination procedure to be used for field work has been given9). SNOEYENBOS et al.11) applied the litter-sample culturing to identify and maintain
Salmonella-free commercial chicken breeding flocks.

* Supported in part by a Grant (600038 and 460038) from the Ministry of Education
This paper describes isolation of Salmonella from samples of hatcher chick fluff, floor litter or feces from breeding chickens and their progeny on a farm. It will also give information on the difference of dependability among the 3 sampling methods for monitoring the Salmonella status of the chicken flocks.

**Materials and methods**

**Sampling and cultivation of hatcher chick fluff**

Eggs from different breeder flocks of New Hampshire chickens and those of a meat-type on the farm were set in separate incubators. One lot of hatching eggs consisted of those set simultaneously into 3~8 incubators. A fluff sample was collected from each incubator after removal of chicks. A dose of about 0.2 g of the sample was inoculated into about 15 ml of selenite broth and incubated for 18 hrs at 37°C. A loopful of the broth culture was streaked on MacConkey agar plate. After the first isolation of Salmonella from fluff sample in late 1970, the broth inoculated was incubated for 24 hrs at 43°C and subcultured on brilliant-green agar plate. In 1971, SBG Sulfa broth (Eiken) was used in place of selenite broth. Several Salmonella-like colonies from each sample were examined serologically and biochemically.

**Sampling and cultivation of floor litter or feces**

Progeny broiler chickens: Each lot of chicks collected from different incubators was divided into 2~4 flocks. Each flock consisting of 2,000~7,000 chicks was kept in one pen. Floor litter samples composed of fine dry feces were collected into plastic bags. Two samples were taken from representative sites of each pen, 2 times on the ages of 2 and 10 weeks in 1969 and early 1970.

From late 1970, floor litter and newly passed feces of a small number of chickens were sampled simultaneously, as a rule. One or 2 floor litter samples were collected from each pen, 1~4 times on the ages of 1~10 weeks. Pieces of feces were sampled with sterile chopsticks from each bird into sterile tube. On an average 5 chickens (1~9 chickens) were selected for sampling arbitrarily from each flock.

A dose of 5 or 10 g of the litter samples was inoculated in tenfold volume of SBG Sulfa broth and incubated at 43°C. Incubation period was 48 hrs until early 1970. Then it was changed to 24~30 hrs. The broth culture was streaked on brilliant-green agar plate. Several Salmonella-like colonies were checked from each sample by routine method. Fecal sample (about 5 g) from each bird was cultured into about 20 ml of SBG Sulfa broth and mashed with a glass rod. Further procedure was the same as for litter samples.

Breeder chickens: Each breeder flock occupied one pen. As a rule, 2 floor litter samples containing rice straws and feces were collected from each pen 1 to 6 times at 1~4 weeks intervals from November of 1970 to March of 1971. At the same time, newly passed feces were sampled from about 5 birds per flock as indicated in table 3. Cultivation was done by the procedure described above.

**Results**

Relationship between Salmonella isolation from hatcher fluffs and infection
TABLE 1  Relationship between Salmonella isolation from hatcher chick fluff and infection of chicken flocks

<table>
<thead>
<tr>
<th>YEAR</th>
<th>FLUFF</th>
<th>SAMPLES FROM CHICKS HATCHED</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>floor litter</td>
</tr>
<tr>
<td>1968</td>
<td>0 *1</td>
<td>ND*2</td>
</tr>
<tr>
<td></td>
<td>230 samples (from 39 lots)</td>
<td></td>
</tr>
<tr>
<td>1969</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>117 (18)</td>
<td>208 samples (from 52 flocks of 19 lots)*5</td>
</tr>
<tr>
<td></td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>1970*3</td>
<td>3 (2)</td>
<td>88 (22–7)*6</td>
</tr>
<tr>
<td></td>
<td>271 (60)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>0</td>
<td>88 (22–7)*6</td>
</tr>
<tr>
<td>1971*4</td>
<td>0</td>
<td>7 (2–1)</td>
</tr>
<tr>
<td></td>
<td>33 (11)</td>
<td>40 (28–14)</td>
</tr>
</tbody>
</table>

*1: No. positives/No. samples tested  
*2: Not done  
*3: S. enteritidis was isolated.  
*4: S. typhimurium was isolated.  
*5: Sampled from Sept. to Dec.  
*6: Sampled from Jan. to Feb.  
*7: Sampled from late Nov.
TABLE 2  Examples of Salmonella detection from the samples of broiler chickens

<table>
<thead>
<tr>
<th>LOT NO.</th>
<th>FLUFF</th>
<th>FLOCK NO.</th>
<th>1 WK*1</th>
<th>2</th>
<th>4</th>
<th>5</th>
<th>7</th>
<th>10</th>
<th>SEROTYPE OF ISOLATES</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>floor litter</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>feces</td>
<td>f. l</td>
<td>f. l</td>
<td>f. l</td>
<td>f. l</td>
<td>f. l</td>
<td>f.</td>
</tr>
<tr>
<td>70-46</td>
<td>2/6*2</td>
<td></td>
<td>1*3</td>
<td>4</td>
<td>4/5</td>
<td>4/5</td>
<td>4/5</td>
<td>4/5</td>
<td>4/5</td>
</tr>
<tr>
<td>70-55</td>
<td>0/3</td>
<td>1*3</td>
<td>3</td>
<td>5/5</td>
<td>5/5</td>
<td>5/5</td>
<td>5/5</td>
<td>5/5</td>
<td>5/5</td>
</tr>
<tr>
<td>71- 4</td>
<td>0/3</td>
<td></td>
<td>1*3</td>
<td>4/5</td>
<td>4/5</td>
<td>4/5</td>
<td>4/5</td>
<td>4/5</td>
<td>4/5</td>
</tr>
</tbody>
</table>

*1: Age of chickens at which samples were taken.
*2: No. positives/No. samples tested
*3: Flocks of the meat-type chickens
*4: Not sampled
Detection of Salmonella infection in chicken flocks

Cultivation of hatcher chick fluffs had been made to monitor Salmonella infection of the flocks. Since late 1968, breeder replacement chicks of a meat type have been introduced periodically from a farm in an eastern state of the United States of America. In order to check the introduction of Salmonella by the imported chickens, floor litter samples from broiler chickens which were hatched from the breeders and from New Hampshire breeder chickens which had been kept on the farm were examined in late 1969 and early 1970. No Salmonella was isolated as indicated in table 1.

Fluffs from 3 hatches of 2 lots gave *Salmonella enteritidis* twice at interval of 4 weeks in September and October of 1970. Then bacteriological examination of fecal and litter samples from broiler flocks was started. As shown in table 1, samples of litter or feces tested indicated high frequency of Salmonella isolation in the lots containing Salmonella-positive fluffs. On the other hand, also in 3 lots not containing Salmonella-positive fluffs, Salmonella infection of chicken flocks was detected by the cultivation of litter and/or feces. Clinical problems due to the Salmonella infection were not recorded. Table 2 shows examples of Salmonella isolation from litter or fecal samples from broiler chickens. Chickens of the lots not containing Salmonella-positive fluffs appeared to be infected slightly with Salmonella, though sampling was not made regularly.

**TABLE 3 Detection of Salmonella from floor litter or feces of breeder chickens**

<table>
<thead>
<tr>
<th>FLOCK NO.</th>
<th>SOURCE OF CHICKENS</th>
<th>IN NOV. OF 1970</th>
<th>SAMPLES*1</th>
<th>SEROTYPE OF ISOLATES</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>New Hampshire chickens produced in the farm</td>
<td>19</td>
<td>500</td>
<td>Removed</td>
</tr>
<tr>
<td>2</td>
<td>&quot;</td>
<td>15</td>
<td>780</td>
<td>0/4 (0/2)*2 0/15 (0/3)*2</td>
</tr>
<tr>
<td>3</td>
<td>&quot;</td>
<td>11</td>
<td>550</td>
<td>0/3 (0/2) 0/16 (0/3)</td>
</tr>
<tr>
<td>4</td>
<td>&quot;</td>
<td>11</td>
<td>550</td>
<td>0/3 (0/2) 0/16 (0/3)</td>
</tr>
<tr>
<td>5</td>
<td>&quot;</td>
<td>7</td>
<td>560</td>
<td>0/4 (0/2) 0/19 (0/4)</td>
</tr>
<tr>
<td>6</td>
<td>Meat-type chickens imported from U. S. A.</td>
<td>16</td>
<td>800</td>
<td>Removed</td>
</tr>
<tr>
<td>7</td>
<td>&quot;</td>
<td>13</td>
<td>360</td>
<td>0/2 (0/1) 0/15 (0/3)</td>
</tr>
<tr>
<td>8</td>
<td>&quot;</td>
<td>10</td>
<td>600</td>
<td>1/6 (1/3) 3/20 (2/4)</td>
</tr>
<tr>
<td>9</td>
<td>&quot;</td>
<td>8</td>
<td>700</td>
<td>0/6 (0/3) 0/20 (0/5)</td>
</tr>
<tr>
<td>10</td>
<td>&quot;</td>
<td>6</td>
<td>940</td>
<td>2/12 (2/6) 21/41 (8/8)</td>
</tr>
<tr>
<td>11</td>
<td>&quot;</td>
<td>3</td>
<td>1,100</td>
<td>10/13 (6/6) 16/38 (6/7)</td>
</tr>
</tbody>
</table>

*1: Sampled at intervals of 1~4 weeks from Nov. of 1970 to Mar. of 1971
*2: No. positives/No. samples tested (No. positives/No. sampling times)
*3: One isolation

*S. typhimurium*  
*S. enteritidis*  
*S. blockley*  
*S. typhimurium*  
*S. enteritidis*
Thus, fluff-sample culturing could indicate in advance Salmonella infection in 2 of the 5 lots of progeny chickens, in which Salmonella was detected by the floor litter and/or fecal cultivation.

Detection of Salmonella from samples of floor litter and feces from breeder chickens

As shown in table 3, Salmonella was not isolated from the domestic chickens, but only from the imported breeders, though sample size from the domestic birds was small. There was a flock (No. 10) in which Salmonella was isolated more frequently by the fecal culture compared with the litter-sample culturing. Since, unfortunately, flocks 1 and 6 had been removed when the examination for Salmonella was started, it remained uncertain whether or not *S. enteritidis* isolated from fluffs and chicks originated from the breeder chickens. Eggs from flocks 8 and 10, in which *Salmonella typhimurium* was isolated, had been hatched, but no Salmonella was detected from fluff samples of the hatches corresponding. On the other hand, the organism was isolated from a broiler chicken (Lot 714 in table 2). Thus, so far as *S. typhimurium* is concerned, fluff-sample culturing gave no information on Salmonella status of the breeder chickens and their progeny.

*S. typhimurium* cultures obtained during this study belonged to the phage type la (Anderson's old scheme\(^1\)) and to the biotype 10\(^1\).

**DISCUSSION**

In this study, chicks of the lots containing Salmonella-positive fluffs were contaminated heavily with the same Salmonella. However, hatchery fluff cultivation could not always detect flock infection with Salmonella. MORRIS et al. reported that hatchery samples including air samples indicated much lower frequency of Salmonella isolation compared with samples from flocks of breeders and broilers. In addition, SNOEYENBOS et al.\(^1\) described that they could not isolate Salmonella from fluffs of 18 hatches, in which hatching eggs were obtained from Salmonella-infected breeding chickens, and a great part of flocks of their progeny chicks were infected with the same Salmonella as in the breeders. However, the fluff-culturing method appears to be most suitable to monitor continuously the Salmonella status of hatchery environment. Investigation on improvement of the method, for example, cultivation of large dose of fluffs, is needed.

The data of this study indicated that the floor-litter culturing was efficient to know Salmonella infection of chicken flocks. Moreover, it should be noted that the cultivation of feces from a small number of birds gave much information on Salmonella in the flocks.

As indicated in table 3, *S. typhimurium* and other Salmonella were isolated from the imported breeder chickens. *S. typhimurium* had not yet been isolated from any material on this farm\(^7\). It was probable that the imported meat-type
chickens carried the Salmonella, though record on clinical problem during the quarantine was not available. A report\(^2\) indicated that the same meat-type chickens which were imported from the United States of America in the spring of 1970 had \textit{S. typhimurium} infection during and after the quarantine.

According to the survey by SAKAZAKI & NAKAYA, a total of 1,680 Salmonella cultures belonging to 25 serotypes were isolated from chickens and eggs during the period from 1946 to 1963 in Japan. The serotypes were arranged in descending of isolation frequency as follows; \textit{Salmonella pullorum} (31.8\%), \textit{Salmonella senftenberg} (22.7\%), \textit{Salmonella thompson} (8.5\%), \textit{Salmonella give} (8\%), \textit{Salmonella bareilly} (6.1\%), \textit{Salmonella potsdam} (5.4\%), \textit{Salmonella new brunswick} (4.6\%) and so on. Recently SHIMIZU described Salmonella isolation from chicks which were imported mainly from the United States of America and died at the time of arrival and during the quarantine for the first 2 weeks of life. During 4 years from 1965 to 1968, 303 Salmonella cultures of 20 serotypes were isolated. Eleven of the 20 types had not yet been isolated from chickens in Japan. The cultures included \textit{S. typhimurium} (32\%), \textit{Salmonella infantis} (22.4\%), \textit{Salmonella heidelberg} (20.5\%), \textit{S. thompson} (4.6\%), \textit{Salmonella blockley} (2.6\%), \textit{Salmonella oranienburg} (2.6\%), \textit{S. enteritidis} (2.6\%) and so on. No isolation of \textit{S. pullorum} was recorded. Among the serotypes described above, \textit{S. infantis}, \textit{S. heidelberg} and \textit{S. blockley} were the newly added ones. Rearing of imported poultry flocks contaminated with Salmonella other than \textit{S. pullorum} has not been restricted in Japan, though clinical problem occurred on the flocks during the quarantine. During the period from 1965 to 1968, about 8.8 million chicks (mainly breeders) were imported. According to a recent survey\(^6\) on the reports of Salmonella isolation from domestic chickens, type distribution of Salmonella of chicken origin in Japan has changed remarkably for the past several years, almost corresponding to the distribution in imported chicks shown by SHIMIZU. Excluding \textit{S. pullorum}, \textit{S. typhimurium} or \textit{S. thompson} has been dealt most frequently in the reports. Isolation of \textit{S. infantis} and \textit{S. enteritidis} increased in frequency. \textit{S. blockley} infection was observed among domestic chickens.

Thus, it should be considered more seriously that foreign chicks carrying Salmonella have been playing a role in changing the Salmonella status in Japan.

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