<table>
<thead>
<tr>
<th>Title</th>
<th>PARASITES OF THE JAPANESE BADGER, MELES MELES ANAKUMA TEMMINCK, ESPECIALLY ON ISTHMIOPHORA MELIS (SCHRANK, 1788) LUHE, 1909</th>
</tr>
</thead>
<tbody>
<tr>
<td>Author(s)</td>
<td>Kamiya, Haruo; Suzuki, Yoshitaka</td>
</tr>
<tr>
<td>Citation</td>
<td>Japanese Journal of Veterinary Research, 23(4): 125-130</td>
</tr>
<tr>
<td>Issue Date</td>
<td>1975-10</td>
</tr>
<tr>
<td>DOI</td>
<td>10.14943/jjvr.23.4.125</td>
</tr>
<tr>
<td>Doc URL</td>
<td><a href="http://hdl.handle.net/2115/2070">http://hdl.handle.net/2115/2070</a></td>
</tr>
<tr>
<td>Type</td>
<td>bulletin</td>
</tr>
<tr>
<td>File Information</td>
<td>KJ00002371230.pdf</td>
</tr>
</tbody>
</table>

HOKKAIDO UNIVERSITY
PARASITES OF THE JAPANESE BADGER, MELES MELES ANAKUMA TEMMINCK, ESPECIALLY ON ISTHMIOPHORA MELIS (SCHRANK, 1788) LÜHE, 1909

Haruo Kamiya and Yoshitaka Suzuki*

Department of Parasitology
Faculty of Veterinary Medicine
Hokkaido University, Sapporo, Japan

(Received for publication, July 11, 1975)

A trematode species, Isthmiophora melis (SCHRANK, 1788) LÜHE, 1909, was found in the small intestine of a badger, Meles meles anakuma TEMMINCK, captured in Gifu, central Japan, on June 10, 1974. The differentiation among I. melis, Euparyphium beaveri, YAMAGUTI, 1958, and Echinostoma hortense ASADA, 1926, was discussed. A coccidian parasite, Eimeria sp., was also found in the mucosal epithelium of the jejunum. Furthermore, an unidentified nematode was observed in the jejunal mucosa.

INTRODUCTION

There are a few reports about the parasites of the badger, Meles meles anakuma, in Japan, such as Toxocara canis and Tetragomphius melis by YAMAGUTI (1941) and OHBAYASHI et al. (1974), respectively. On the other hand, Isthmiophora melis (SCHRANK, 1788) LÜHE, 1909, was found only in the weasel (Mustela sibirica) by YOSHIDA et al. (1932) and in the dog by YAMASHITA (1939) in Japan. In this paper, we compare I. melis with several allied echinostomoid species, and deal with a coccidian parasite, Eimeria sp., and an unidentified nematode recognized in histological section preparations of the small intestine.

MATERIALS AND METHODS

The worms were obtained from a small intestine preserved in 10% formalin solution. Some whole-mounted specimens were stained with Semichon’s carmine, and paraffine sections for microscopy were stained with hematoxylin-eosin.

* Present address: Department of Veterinary Anatomy, Faculty of Agriculture, Gifu University, Gifu, Japan
1) *Isthmiophora melis* (SCHRANK, 1788) LÜHE, 1909

(Trematoda: Echinostomatidae)

The number of worms obtained from the duodenum, jejunum, and ileum was 4, 57, and 14, respectively. Some of them were tightly attached to the mucosa by their head collar and suckers. The measurements were based on 10 matured specimens.

Description: Body elongate, leaf-shape, abruptly tapering anterior to acetabulum, more gradual in hind body. Matured worms 4.778~5.957 mm in length, with the maximum width at or just back of acetabulum, 0.897~1.248 mm. Cuticle 0.016~0.020 mm thick. Cuticular spines, stout, well developed, showing tilelike appearance, extending to acetabulum dorsally and to testis ventrally. Oral sucker subterminal, elliptical, 0.176~0.218×0.196~0.248 mm in size. Head collar well developed, 0.326~0.391 mm in diameter. Collar spines 27 in number, tightly attached to head collar. Crown of spines double, dorsally uninterrupted. Dorsal spines 0.049~0.075 mm long by 0.013~0.016 mm wide. Marginal spines not abruptly pointed 0.062~0.082 mm long by 0.013~0.022 mm wide. Prepharynx scarcely recognizable. Pharynx well developed, elongated oval, 0.111~0.163×0.196~0.228 mm in size. Narrow esophagus, short 0.130~0.163 mm long. Bifurcation of esophagus immediately in front of acetabulum. Narrow ceca extending just at posterior end. Acetabulum well developed, oval, 0.507~0.650×0.546~0.754 mm in size, situated one-fourth into the anterior part. Distance from anterior end to middle of acetabulum 0.806~0.910 mm long. Testes tandem, usually contiguous in each other, situated, approximately in the equatorial position. Anterior testis always wider than length, but posterior testis always longer than width. Testis varies in shape with some notches. Size of anterior testis and of posterior testis 0.440~0.637×0.339~0.545 mm and 0.326~0.544×0.481~0.624 mm, respectively. Cirrus sac big, oval in shape, situated between bifurcation of esophagus and about middle of acetabulum, 0.196~0.325×0.388~0.598 mm in size. Genital pore at just behind esophageal bifurcation. Ovary regular in outline, spherical to oval, located on right side of anterior testis, 0.241~0.313×0.186~0.303 mm in size. The most proximal portion of uterus forms seminis receptaculum uterium; true seminal receptacle obscure. Uterus short with few eggs, about 25~120 in number. Eggs 0.114~0.130×0.052~0.069 mm in size. Eggs deposited in the villi of intestine. Mehlis' gland well developed. Distance from posterior end of acetabulum to anterior margin of fore-testis, 0.819~1.040 mm. Follicular vitellaria extending from lateral side of anterior margin of ovary to posterior end of ceca. Vitelline reservoir posterior
Parasites of Japanese badger

and dorsal to Mehlis' gland, receiving vitelline ducts from each side of body just along anterior margin of fore-testis. Vitellaria extracecal up to posterior to hind-testis, and then intercecal. Excretory pore subterminal and dorsal.

Pathological findings: In spite of numerous body spines of the trematode, the damage produced in the mucosa of the intestine is minimal. The destruction of the mucosa, however, was observed at the portion of attachment of head collar and suckers, especially the acetabulum. The cellular reactions were scarcely recognizable.

Discussion: I. melis was reported in the weasel (Mustela sibirica) and the dog of Japan by Yoshida et al. (1932) and Yamashita (1939), respectively. The seminal receptacle in this species, however, was not clearly recognizable. The measurement corresponded well to that of the latter. On the other hand, Donges (1967 & 1968) considered that Euparyphium suinum Ciurea, 1921, and E. spiculator (Dujardin, 1845) were synonyms of I. melis, and that the differentiations of measurement among these species corresponded to the host species. Furthermore, Hsu (1940) considered that E. jassyensis Leon et Ciurea, 1922, from a human case, was a synonym for I. melis.

For E. melis by Beaver (1941), Euparyphium beaveri was established by Yamaguti (1958) as a new species based on the collar spines being abruptly tapered at the outer end, the cirrus being heavily spined, and the eggs being definitely narrower than I. melis. The egg size of the present specimens is similar to that of E. beaveri, but the former differs from the latter in the shape of collar spines, body spines, testes, etc. The difference of egg size between the present specimens and that of Donges (1967) may be due to the methods of fixation used.

Lumsden & Zischke (1961) redescribed E. beaveri from Mustela vison, but their specimens apparently differ from the present ones in the shape of body, testes and the distribution of testes, etc. In this examination we identified the present specimens as I. melis, considering the above-mentioned characteristics. For more detailed differentiation between the two species, we are of the opinion that it is necessary to study the specimens of adult and larva at various developmental stages obtained by experimental infection.

In addition, we compared I. melis with Echinostoma hortense Asada, 1926, since both species possess 27 collar spines. I. melis, however, is easily distinguished from E. hortense in body shape, in the shape and size of collar spines, and in the diameter of head collar, the outline of testis, etc.

2) Eimeria sp. (Sporozoa: Eimeriidae)

Eimeria sp. were detected in the jejunal mucosa. Numerous oocysts 0.020 ~0.021×0.014~0.017 mm in size, a small number of macrogametocytes, and
a few microgametocytes were observed in the epithelium. Macrogametocytes with many basophilic and a few eosinophilic granules, 0.015–0.022×0.013–0.020 mm in size. Microgametocytes with many microgametes, about 0.020 mm in diameter.

Discussion: Two species of Eimeriidae, *Eimeria melis* KOTLAN & Pospesch, 1933, and *Isospora melis* (KOTLAN & Pospesch, 1933) PELLÉRDY, 1955, are known to be found in the badger, *Meles meles* LINNAEUS (PELLÉRDY, 1965). The size of oocysts is 0.017–0.024×0.013–0.017 mm in the former and 0.026–0.034×0.020–0.027 mm in the latter. The coccidian parasite in this study probably belongs to genus *Eimeria*, but final identification must be made in the future after detailed investigations of various developmental stages.

Appendix Nematoda gen. sp. was recognized in histological section preparations. The worms were detected in the mucosal epithelium of the jejunum. The diameter of the worm in the intestinal portions was 0.025–0.030 mm. Cellular reactions to the worms were not observed in the surrounding tissue. It was difficult to clarify the proper taxonomical portion at present, due to limited materials.

**Acknowledgements**

The authors wish to express their gratitude to Asst. Prof. M. OHBAYASHI of this Department for his kind direction and review. Further thanks are offered to Mr. H. INUMA, Gifu Municipal Office, for providing the badger examined.
References

1) BEAVER, P. C. (1941): J. Parasit., 27, 35
2) DÖNGES, J. (1967): Z. ParasitKde., 29, 1
3) DÖNGES, J. (1968): Ibid., 30, 179
4) HSÜ, H. F. (1940): Chin. med. J., 58, 552
5) KOTLÁN, A. & POSPESCH, L. (1933): Parasitology, 25, 102
8) PELLÉRDY, L. (1955): Acta vet. hung., 5, 421
EXPLANATION OF PLATE

Figs. 1~6  *Isthmiophora melis* (SCHRANK, 1788)

Fig. 1  General view

Fig. 2  Head collar with unclear double rows of collar spines and body spines showing a tilelike appearance; ventral view

Fig. 3  Collar spines showing clear double rows; dorsal view

Fig. 4  Morphological variations of anterior testes

Fig. 5  Morphological variations of posterior testes

Fig. 6  Section of small intestine showing the mucosal destruction through attachment by acetabulum

   Hematoxylin-eosin (H.-E.) stain

Figs. 7 & 8  *Eimeria* sp., section preparations

Fig. 7  Small intestine showing numerous oocysts  H.-E. stain

Fig. 8  Small intestine showing macrogametocytes  H.-E. stain

Fig. 9  Small intestine showing cross sections of unidentified nematode