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Primer Sequences and Cross-species Amplification for Parentage Discrimination of Tanganyikan Cichlid Fishes

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Abstract

It was done that isolation and cross-species amplification of microsatellite loci for parentage analysis of the Tanganyikan cichlids. The sequences of the primers and the results of cross-species amplification were reported.

Key words: Cichlidae, Tanganyika, cross-species amplification, microsatellite loci

Since the late 19th century, many ichthyologists have studied the radiation of cichlid fishes in Lake Tanganyika. There are more than 170 species of cichlids in Lake Tanganyika, most of which are endemic (Kohda et al., 1997), and Lake Malawi may contain even more species (Ribbink et al., 1983). We are studying the evolution of allopaternal care in the Tanganyikan cichlids, *Chalinochromis brichardi*, *Neolamprologus meeli*, *Julidochromis ornatus*, and *Telmatochromis temporalis*.

Microsatellite genotyping has proven to be a useful tool for phylogenetic (Lee and Kocher, 1996; Zardoya et al., 1996; Neff et al., 2000) and parentage analysis (e.g. Kellogg et al., 1995; Parker and Kornfield, 1996) of the cichlids. We designed six primer sets to amplify microsatellites in the four Tanganyikan species to search for genetic markers. Four primer sets (Pze1-4) from the Malawian cichlid fish, *Pseudotropheus zebra*, sequenced by van Oppen et al. (1997) were also tested in these four

species. We described the sequences of the primers and the results of test amplifications.

Pieces of muscle from fish collected in Lake Tanganyika were fixed in 70% ethanol. The genomic DNA for the four cichlid species studied was isolated from this tissue. The DNA was extracted with a standard phenol-chloroform procedure after a reaction in lysis buffer. The procedures for the construction of the genomic libraries, isolation of plasmids containing (GT/CA)_n regions, and sequencing the plasmids were the same manner as those described in Munehara and Takenaka (2000). Four loci containing perfect repeats of more than (GT/CA)₁₅ were screened from each genomic library of *C. brichardi* and *N. meeli*, and used to design the PCR primers (Table 1). The level of polymorphism for each locus was estimated using 10-36 individuals from different families in each species. Species and cross-species amplification with our 4 primer

Table 1. Primer sequences of useful microsatellite loci of two cichlid fishes.

Source	Code name	Primer sequence (5'-3')	Accession number
<i>Chalinochromis brichardi</i>	Chb1	TTCTTTCAGGCTCTAGCTTTCC AAGCTCAGAGTCTGCATGTGC	AB012117
<i>C. brichardi</i>	Chb2	ATGTCAATTGAAGGTCCTAACT TCAGCAAACCTCAACTTAA	AB012118
<i>Neolamprologus meeli</i>	Nem1	TGGTGCAATTCAAACCTTTTCTG CCTTTCCTAACTTCTTGGGG	AB012119
<i>N. meeli</i>	Nem2	CACAAACCAAGCAGAGGCTTA CTTAACGAGGTCAAATCTGTTG	AB012120

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Table 2. Polymorphism of some microsatellite loci for three cichlid fishes. Amplifications more than 0.50 in heterozygosity were showed in this table.

Chalinochromis brichardi

Code name	Annealing temp. (°C)	Length of PCR products (bp)	No. of alleles	Heterozygosity		Sample size
				observed	expected	
Chb1	58	88, 92, 110, 114, 116	5	0.60	0.56	30
Nem2	52	134, 136, 142	3	0.70	0.57	30

Neolamprologus meeli

Code name	Annealing temp. (°C)	Length of PCR products (bp)	No. of alleles	Heterozygosity		Sample size
				observed	expected	
Chb2	54	90, 96, 98, 100	4	0.55	0.66	30
Nem1	52	101, 111, 113, 115	4	0.57	0.65	30
Pze4	54	112, 114, 126, 128	4	0.70	0.62	30

Julidochromis ornatus

Code name	Annealing temp. (°C)	Length of PCR products (bp)	No. of alleles	Heterozygosity		Sample size
				observed	expected	
Chb1	61	range of 96–138	15	0.56	0.66	36
Chb2	54	range of 97–170	17	0.69	0.83	36
Nem1	50	range of 103–121	8	0.58	0.49	36
Pze3	50	range of 314–392	17	0.81	0.82	36

Telmatochromis temporalis

Code name	Annealing temp. (°C)	Length of PCR products (bp)	No. of alleles	Heterozygosity		Sample size
				observed	expected	
Pze2	46	range of 152–222	20	0.88	0.93	34
Pze4	54	range of 122–144	11	0.76	0.84	34

sets and cross-species amplification with the 4 primer sets from the Malawian cichlid (van Oppen et al., 1997) were performed in a thermocycler, following the manufacturer's recommendation (Perkin-Elmer). An initial denaturation at 95°C for 5 min was performed before the addition of *Taq* DNA polymerase (TaKaRa Co.). The reaction was carried out for 23 or 25 cycles of 30 s at 94°C, 50 s at the annealing temperature (Table 2), and 80 s at 72°C. The procedures described in Munehara and Takenaka (2000) were used to detect PCR products. The length of the PCR products was estimated with 10 bp or 20 bp ladder markers and PCR products from individuals used for primer design.

Cross-species amplification successfully identified heterozygous alleles of *C. brichardi*, *N. meeli*, *J. ornatus*, and *T. temporalis*, and useful primer sets varied with species (Table 2). Successful cross-species amplifications with Pze 2, 3 and 4 suggested that primers from Tanganyikan and Malawian cichlid fishes could provide useful information for parentage analysis.

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References

- Kellogg, K.A., Markert, J.A., Stauffer, Jr., and Kocher, T.D. (1995). Microsatellite lekking cichlid fishes from Lake Malawi, Africa. *Proceedings of the Royal Society of London, Series B*, **260**, 79–84.
- Kohda, M., Hori, M., Nshombo, M. (1997). Inter-individual variation in foraging behaviour and dimorphism in predatory cichlid fishes. p. 121–136, Kawanabe, H., Hori, M. and Nagoshi, M. (eds.). *Fish communities in Lake Tanganyika*. Kyoto University Press, Kyoto.

- Lee, W.J. and Kocher, T.D. (1996). Microsatellite DNA markers for genetic mapping in *Oreochromis niloticus*. *J. Fish Biol.*, **49**, 169-171.
- Munehara, H. and Takenaka, O. (2000). Estimation of reproductive success of sneakers in a paternal care fish, *Hexagrammos otakii*, using microsatellite variants. *J. Ethol.*, **18**, 101-104.
- Neff, B.D., Repka, J. and Gross, M.R. (2000). Parentage analysis with incomplete sampling of candidate parents and offspring. *Mol. Ecol.* **9**, 515-528.
- Parker, A. and Kornfield, I. (1996). Polyandry in *Pseudotropheus zebra*, a cichlid fish from Lake Malawi. *Env. Biol. Fish.*, **47**, 345-352.
- Ribbink, A.J., Marsh, A.C., Ribbink, C.C. and Sharp, B.J. (1983). A preliminary survey of the cichlid fishes of rocky habitats in Lake Malawi. *South Africa J. Zool.*, **18**, 149-310.
- van Oppen, M.J.H., Rico, C., Deutsch, J.C., Turner, G.F. and Hewitt, G.M. (1997). Isolation and characterization of microsatellite loci in the cichlid fish *Pseudotropheus zebra*. *Mol. Ecol.*, **6**, 387-388.
- Zardoya, R., Vollmer, D.M., Craddock, C., Streelman, J.T., Karl, S. and Meyer, A. (1996). Evolutionary conservation of microsatellite flanking regions and their use in resolving the phylogeny of cichlid fishes (Pisces: Perciformes). *Pro. of the Royal Soc. London, Series B*, **263**, 1589-1598.