TITLE

Difference of paternity rates depending on alternative reproductive behaviors in the squid *Loligo bleekeri*

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ABSTRACT

The mating behavior of captive *Loligo bleekeri* and the paternity of the resulting progeny were examined based on behavioral observations and genetic analyses. In this species, there are three mating behaviors (male-parallel, head-to-head, and extra-pair), and two sperm storage sites in females (seminal receptacle and the opening of the oviduct), which suggest that sperm competition occurs. All three mating behaviors were observed, and females mated often with different males, resulting in multiple paternity within 3 of the 4 broods examined. In each brood, the male to mate last and frequently before the female spawned fertilized the most eggs (87-100%). A sneaker male that mated by extra pair copulation sired 8.5% of the eggs in a brood. Some eggs were fertilized by sperm received before the start of the study, indicating that sperm can be stored for at least several days before a spawning. In the broods with multiple paternity,
the paternity patterns differed among egg capsules. Male competition was more intense between similar-sized males than between different-sized males, but body size did not affect the copulative success in the male-parallel position. We found the multiple mating and multiple paternity in *L. bleekeri*, and the paternity rates differed depending on the complex factor with mating position, timing, frequency and duration, relating with alternative reproductive behaviors.

**KEY WORDS**
Multiple paternity, Reproductive strategy, Squid, *Loligo*, Microsatellite, Sperm competition

**INTRODUCTION**

Loliginid squids are important members of nearshore and continental shelf ecosystems, and fished commercially worldwide. The fisheries tend to target spawning aggregations in shallow waters, so information on their reproductive strategies are needed to assess how the fisheries impact the squid populations (Hanlon 1998). One of the commercially most important loliginids in Japan is *Loligo bleekeri* Keferstein. It occurs widely in coastal waters from southern Korea to northern Japan (Roper et al. 1984), has a one-year life span, and spawning season continues over a six-month period (Natsukari & Tashiro 1991). Like other loliginids, *L. bleekeri* is a demersal spawner, and at one spawning, lays up to 30 egg capsules, each containing approximately 50 eggs (Natsukari & Tashiro 1991).

Loliginid squids mate in two main positions: “head-to-head” and “male-parallel” (Drew 1911, Hanlon & Messenger 1996). In the head-to-head position, the male
deposits spermatophores in or near the female’s seminal receptacles, which are located near her mouth. In the male-parallel position, the male deposits spermatophores inside the female’s oviduct in the mantle cavity.

Immature loliginid females mate in head-to-head position and store sperm in their seminal receptacle (Drew 1911). On loliginid spawning grounds, large males form temporary pairs with mature females, which give the males preferred access to the female just before she spawns (Hanlon et al. 1997, Hanlon et al. 2002). These consort males guard females before and after copulation, and while the females spawn, but sometimes large intruders replace the consort males and copulate with the females (Hanlon et al. 1997, Hanlon et al. 2002). Furthermore, extra-pair copulations (EPCs) also occur, in which a small “sneaker males” the male copulates with a female that is paired with another male by attaching spermatophores near the female’s mouth (Hanlon 1996, Hanlon et al. 1997, Hanlon 1998, Hanlon et al. 2002). The spermatozoa are stored until they are released at spawning, so loliginid females can use spermatozoa from several males when they spawn.

When squids spawn, eggs are extruded from the oviduct, and pass through the funnel and near the mouth. The eggs can be fertilized either as they leave the oviduct by spermatozoa placed in the mantle cavity during a male-parallel mating or near the mouth by spermatozoa stored in the seminal receptacle after a head-to-head mating or EPC. This difference in the fertilization mechanism will presumably affect the reproductive success of each copulative behavior and play a role in sexual selection (Birkhead and Møller 1998).

If the reproductive successes of each copulative behavior differ, males will compete to mate in the favorable position. Agonistic behavior between males has been shown to
escalate through several phases from visual signaling through physical contact (DiMarco & Hanlon 1997). In the most escalated phase of a contest, a male will rush at another male and sometimes grab it (King et al. 1999). The relative sizes of males affect the outcome of these contests (DiMarco & Hanlon 1997). When competitive abilities of competing individuals differ, alternative reproductive tactics might increase the fitness of low competitive status males (Gross 1996), and lead to the evolution of phenotypic diversity in males such as large fighters and small sneakers (Moczek & Emlen 2000).

Parentage analysis is a valuable tool to understand reproductive systems including sperm competition. Microsatellite DNA analyses of several loliginid squids have shown that the eggs within individual egg capsules are usually sired by multiple males (Loligo forbesi: Shaw & Boyle 1997, Emery et al. 2001; L. pealei: Buresch et al. 2001; L. vulgaris reynaudii: Shaw & Sauer 2004). In those studies, mating was not observed, so it is not known if or how the mating behavior is related to reproductive success. A combined approach using both behavioral observations in captive condition and genetic analysis can provide important information about the mechanism of sperm storage, release and fertilization for each mating position (sperm storage site). Furthermore, previous studies analyzed only one or two egg capsules for each female even though females generally spawn many egg capsules (~20 to 100 in L. pealeii, Maxwell & Hanlon 2000). The utilization pattern of spermatozoa stored inside the mantle cavity and those stored in the seminal receptacle probably differ among egg capsules, so parentage analysis of all egg capsules spawned is needed to accurately determine the parentage rates of the different mating behaviors.

In this study, we observed the mating behavior of captive Loligo bleekeri and analyzed the paternity of subsequently spawned eggs to determine (1) if mating with
more than one male results in multiple paternity, (2) the parentage rates of the three mating behaviors (male-parallel, head-to-head, and EPCs), (3) if the paternity pattern differs among egg capsules from a single spawning, and (4) if male body size affects copulative success and mate guarding intensity.

MATERIALS AND METHODS

Reproductive behavior

From January to May 2003, squids were caught from inshore set nets in southern Hokkaido to observe their reproductive behavior. To investigate the difference in paternity rates among the alternative reproductive behaviors, we introduced one female and two males into a race-way aquarium (5.5×2.5×1m depth, 13,000L) at the Usujiri Fisheries Station, Field Science Center for Northern Biosphere, Hokkaido University. Each individual was used only once, and one male randomly selected from the two was marked with a tiny plastic ribbon tag on one of its fins for individual identification. A cube-shaped structure made from steel pipes (0.5×0.5×0.5m) and with three sides covered with polyethylene mesh nets was set on the bottom of the aquarium as a spawning substrate.

The mantle lengths (ML) of all individuals were measured before the experiments, and body weights, gonad weights, and accessory gland weights were measured after the experiment. The maturity stages of all individuals were determined according to the maturity stage indices in L. plei (Perez et al. 2002). All males were classified as mature stage; in each male, the testis and spermatophoric organs were enlarged, there were many spermatophores in the accessory gland, and none had an excessively low gonad weight (Table 1). As a result, all males were considered fertile, and none considered to
be exhausted. Muscle tissues were sampled from the gills for genetic analysis. Three trials were conducted, and the body size ratio between the two males (calculated as the ML of the larger male / the ML of the smaller male) was 1.01 in Trial A, 1.18 in Trial B and 1.40 in Trial C (Table 1).

The behavior of all squid in each experiment was recorded on videotape and analyzed from the start of the experiment until one hour after the beginning of spawning. The total observation time was 34h in trial A, 2h in trial B, and 63h in trial C. The time at which each mating occurred and the duration of each male-parallel mating and EPC were recorded (head-to-head mating always lasted <1 second). Agonistic behavior between males was recorded from one hour before the beginning of spawning until one hour after the beginning of spawning. During this period, the numbers of rushes (a sudden approach toward another male) and grabs (the grasping of another male), which is the most escalated behavior in contests between males, were recorded. Red lamps were used for nighttime observations (18:00-06:00 h).

**Microsatellite DNA analysis**

Spawned egg capsules were removed from the aquarium and kept separately in 500ml bottles with circulating water at 12°C until the paralarvae hatched about two months later. The paralarvae were collected daily and stored at −20°C. In trial A, all 47 offspring from five capsules were genotyped. In trial B (30 egg capsules) and trial C (39 egg capsules in two spawning), 10 offspring were randomly selected from each capsule, and 297 offspring (one egg capsule in trial B had only seven eggs) and 390 offspring were genotyped, respectively. Including the nine adults used for behavioral analysis, a total of 47 adults collected from the same location were also genotyped at all loci for population
allele frequency data.

Gill tissue samples dissected from the 47 adults and the whole body of each paralarvae were homogenized by overnight incubation at 50°C in a solution of 100 mM Tris-HCl (pH 8.0), 10 mM EDTA (pH 7.5), 1.4 mM NaCl, 2% n-Hexadecyltrimethylammonium Bromide, 0.2% 2-mercaptopethanol and 150 µg/ml proteinase K. DNA in the samples was extracted with phenol-chloroform and then precipitated in ethanol. Four previously published microsatellite primers (Lb1, Lb3, Lb4, Lb5; Iwata et al. 2003) and one newly developed primer for the present study (Lb6) were used for genotyping (Table 2). All parents were genotyped at the five microsatellite loci, and the three loci at which two males in each trial were found not to share an allele were used for paternity analysis (Table 2). The experimental procedures used during the PCR and visualization of the products followed Iwata et al. (2003). The genotype of each paralarvae was verified by electrophoresing the PCR products through a polyacrylamide gel.

**Paternity analysis**

Deviations of the observed heterozygosities at each locus from the Hardy-Weinberg expectations and genotypic linkage disequilibrium were tested using exact tests with significance determined by the Markov chain method (GENEPOL v3.4; Raymond & Rousset 1995). Expected exclusion probabilities with one known parent, which indicate the probabilities that two randomly selected individuals from population do not have the same genotypes at one or several loci, were calculated using the computer program GERUD1.0 (Jones 2001). If the expected exclusion probability is sufficiently high, the possibility of paternal misidentification and confusion between experimental and field
males is low.

As the females may have mated before they were collected, we used a multi-locus method to estimate the minimum number of fathers and the parentage rate of each male. This analysis was also conducted using GERUD1.0. This program identifies paternal alleles using two offspring alleles and known maternal alleles (if both alleles of an offspring have the same genotype as the mother, then the father shares one or two alleles with the mother and paternal contribution will be ambiguous within the two alleles, and both alleles will be put up as candidates for paternal alleles), and determines the combination of all possible multi-locus paternal genotypes observed in a brood. If the program obtains multiple combinations of multi-locus genotypes with the minimum number of fathers, the likelihood is ranked based on two criteria: (1) the summarized $\chi^2$ values calculated for each father at each locus based on an assumed 1:1 Mendelian inheritance ratio, and (2) the combined expected frequency assuming the Hardy-Weinberg equilibrium obtained from population allele frequencies.

The frequency of the offspring’s maternal genotypes in each brood were tested to determine if the maternal alleles deviated significantly from a 1:1 Mendelian inheritance ratio, if the mother was heterozygous, and if the mother did not share both alleles with the father. All offspring were assigned to fathers using the most probable paternal genotypes. If the combination of paternal multi-locus genotypes coincided completely with the genotype of an experimental male, that male was considered to be the father. The number of offspring sired by each father in each capsule was estimated by multiplying the parentage rate within 10 offspring from each capsule by the number of eggs in the capsule. The total estimated number of eggs sired in all egg capsules was then used to determine the total parentage rate for each male. If multiple paternity was
observed, the deviation of the total parentage rate of each male from equal allocation and the difference of the parentage rate of each male between egg capsules were tested with the $\chi^2$ test.

**RESULTS**

**Reproductive behavior**

In trial A, the female spawned twice during the two days of observation. Before the first spawning, one male (A-M2) mated in the male-parallel position (mating duration = 439 seconds) just before the spawning, and the other one (A-M1) mated twice by brief EPCs (3s and 5s) as A-M2 was mating (Table 4). Before the second spawning, A-M1 mated in the head-to-head position 100 min before the spawning and in the male-parallel position (388s) just before the spawning, while A-M2 mated by extended EPCs (223s and 40s) during the male-parallel copulation by A-M1 (Table 4). The last male to transfer spermatophores inside the female’s mantle cavity was A-M1, and the last male to transfer spermatophores near the seminal receptacle was A-M2. All eggs from the first spawning died in the embryonic stage, so only those from the second spawning were used in paternity analysis. The second spawning comprised 47 eggs in five capsules, and all eggs were fertilized.

In trial B, one male (B-M2) mated in both the male-parallel (23s) and head-to-head positions (Table 4). The female spawned 1060 eggs in 30 capsules, and all but one egg were fertilized.

In trial C, the female spawned twice during the three days of observation. Before the first spawning, the male C-M2 mated in the male-parallel (428s) and head-to-head positions (Table 4). After the first spawning, this same male frequently mated in the
head-to-head position (Table 4). Just before the second spawning, C-M2 and C-M1 alternately mated in the male-parallel position (Table 4), but C-M2 mated for much longer periods (C-M2: 435 and 323s; C-M1: 23s). The last male to transfer spermatophores inside the female’s mantle cavity and near the seminal receptacle before second spawning was C-M2. The first spawning comprised 1237 eggs in 27 capsules, and the second comprised 445 eggs in 12 capsules. The fertilization rates of the first and second spawning were 99.2% and 99.6%, respectively.

The occurrence of rushes and grabs was much higher between similar-sized squid than between different-sized squid (Fig. 1). Between males of similar size (trial A), rushes and grabs were most often performed by males that mated in the male-parallel position. Between males of different size (trial B and trial C), rushes and grabs were performed most often by larger males on smaller males regardless if the larger males successfully copulated in the male-parallel position or not. The frequency of rushes and grabs increased as the size difference between males increased (Fig. 1).

**Paternity analysis**

The numbers of alleles of each locus observed in the 47 adults ranged from 13 to 30, and observed heterozygosities of each locus ranged from 0.787 to 0.938 (Table 2). One locus (Lb5) significantly deviated from Hardy-Weinberg expectations, and the frequency of observed homozygous was more than expected (Table 2). One possible reason for this result is the presence of null alleles (non-amplifying alleles). In trial A, in which locus Lb5 was analyzed, the maternal genotypes was homozygous and all offspring had one of the maternal alleles, indicating that the mother did not have null alleles. One of the candidate fathers was homozygous (AM1). If the father has null
alleles, the offspring’s genotypes will be homozygous with maternal allele. Five offspring were homozygous with the maternal allele in trial A. Considering the correlation between alleles of other multiple loci and assuming the Mendelian inheritance ratio, three of the five offspring were determined to be fertilized by AM2, which had the same allele with one of the maternal genotype, and the remaining two offspring were fertilized by an unknown male, which had the same allele with one of the maternal genotype or a null allele (see later paragraph of results). All homozygous offspring’s genotypes were checked by reamplification at least twice.

Expected exclusion probabilities with one known parent ranged from 0.699 to 0.899, and those calculated from each of the three loci ranged from 0.989 to 0.996 (Table 2). These expected exclusion probabilities were sufficiently high, so the possibility of paternal misidentification and confusion between experimental and field males was extremely low.

All offspring were successfully genotyped, and each shared one of the maternal alleles in every locus. The frequency of offspring’s maternal genotypes in each brood did not significantly differ from a 1:1 Mendelian inheritance ratio in all tested alleles (Table 3). Three of the four broods genotyped had multiple paternities, and the highest male parentage rate in each trial was ≥87%. The minimum estimated numbers of fathers for each brood were three in trial A, three in trial B, two in trial C (first spawning) and one in trial C (second spawning).

In trial A, the last male to mate in the male-parallel position (A-M1) sired 41 offspring (87.23% of the brood), the sneaker male (A-M2) sired four offspring (8.51%), and another male that mated before the start of the experiment (A-3) sired two offspring (4.26%, Table 4). The parentage rate of A-M1 differed significantly from those of A-M2
and A-M3 (df=2, $\chi^2=61.57$, p<0.001), but A-M2 and A-3 did not differ significantly (df=1, $\chi^2=0.67$, p=0.689). Parentage rates of each male did not significantly differ between egg capsules (df=4, $\chi^2=1.89$, p=0.879, the two capsules that contained <5 eggs were excluded from the analysis, Fig. 2).

In trial B, the male that mated for a short time in the male-parallel and head-to-head positions (B-M2) sired 288 of the 297 genotyped eggs (96.97%, the total parentage rates) and the other male (B-M1) sired no eggs. In addition, two males that mated before the start of the experiment (B-3, B-4) sired eight eggs (2.57%) and one egg (0.46%), respectively (Table 4). The parentage rates differed significantly among the three males (df=2, $\chi^2=15.13$, p<0.001), and between B-3 and B-4 (df=1, $\chi^2=0.67$, p=0.689). All eggs in 23 of the 30 capsules were sired by the male that mated in the male-parallel position (B-M2); eggs in the remaining seven capsules were sired by B-M2 and another male. Among these seven egg capsules, the parentage rate of each male significantly differed (df=12, $\chi^2=36.91$, p<0.001, Fig. 3), but the parentage of eggs fertilized by B-M2 in each capsule did not significantly differ (df=6, $\chi^2=6.69$, p=0.361).

In the first spawning of trial C, the male (C-M2) that mated both in the male-parallel and head-to-head position sired 269 of the 270 genotyped eggs (99.75%), and the remaining one egg (0.25%) was sired by a male that mated with the female before the start of the experiment (C-3, Table 4). In the second spawning of trial C, the male that mated for a short time in the male-parallel position (C-M1) sired no eggs, and all genotyped eggs were sired by C-M2, which was the last to mate in both the male-parallel and head-to-head positions (Table 4).

**DISCUSSION**
*Loligo bleekeri* females copulated multiple times with different males, resulting in multiple paternity within broods. All three females used sperm received during mating in the field before they were captured to fertilize their eggs, indicating that sperm stored for several days before spawning were competitive for fertilization. In trial A, a male that copulated in the field before the first spawning (A-3) sired some eggs in the second spawning, indicating that stored sperm would be competitive for fertilization in several spawnings. Multiple paternity have been reported in other loliginid species and one of the males sire the most of eggs in a capsule (Buresch et al. 2001, Emery et al. 2001, Shaw & Sauer 2004). Field observations have shown that large loliginid males guard females after copulation (Hanlon et al. 1997, Hanlon et al. 2002), and in *L. vulgaris reynaudii*, a male that guarded a pre-spawning female had a high parentage rate (Shaw and Sauer 2004). If the last male to mate fertilizes the most eggs, postcopulatory-guarding behavior is an effective strategy for that male to increase his fitness even with the guarding cost (Yamamura 1986). The mechanisms of sperm precedence in loliginid squids are not known, but behavioral observations predict that the final male to mate before a female spawns will sire the most eggs (Hanlon et al. 2002). In present study, the mating prior to the experiment resulted in fairly low paternity, in each trial, the males that sired the most of eggs (87-100%) was the male to mate last and frequently with the female before she spawned. Our results were not enough to support the prediction of the last male precedence, but could not exclude this possibility either. Because in the present study, the last male mated frequently, which could have lead to the high paternity in this study, and the position, duration and frequency of the mating in the field are unknown. For example, both males mated in the male-parallel position alternatively in trial C second spawning, and the last male mated
in the male-parallel position fertilized all eggs. But the last male also mated in the head-to-head position frequently, and the sperm transferred near the seminal receptacle would contribute his paternity in part. A larger sample of females having various mating histories is needed to confirm it.

The parentage rates of the most successful male in each trial (87-100%) were higher than those reported for other loliginids (62-76%, summarized in Shaw & Sauer 2004). There are two possible explanations for this difference. First, the captive conditions in the present study prevented the females from mating with more males and facilitated remating with the same males. Second, the mating duration in the male-parallel position in the present study (mean ± SD: 294 ± 189 seconds) was much longer than that reported in other loliginid species (*Loligo vulgaris reynaudii*: range 2-39s, *L. pealei*: 5-20s, summarized in Hanlon & Messenger 1996). Prolonged copulation (~300s) has previously been reported in *L. bleekeri* (Hamabe & Shimizu 1957) and appears to be characteristic of this species. Males that perform prolonged copulation can reduce sperm competition with other males by reducing access to the female before she spawns (copulatory guarding), diluting the sperm of previous mates (sperm loading), and removing the sperm of previous mates (sperm displacement), all of which will lead to increased fertilization success for the last mated male (Alcock 1994, Parker 1998). *L. vulgaris reynaudii* and *L. pealei* males copulate for a relatively short duration, not due to disturbance by other males but to a change in behavior to guarding of the females after copulation; during this period, the males remain near the female and chase away other males (Hanlon et al. 2002, Hanlon 1996). In contrast, males in *L. opalescens* stay conjoined in the male-parallel mating position (Hanlon et al. 2004). These two mate-guarding strategies have been observed in insect males; the most familiar
examples involve dragonflies and damselflies (Alcock 1994). Two mate-guarding strategies are observed in a dragonfly species, some pairs remain in tandem during the whole oviposition, while the remainder separate before completion of the oviposition and the male continues to guard by remaining near the female and chasing away other males, and the switch in males from mating to guarding behavior is related to the amount of interference from other males experienced during the early stages of tandem oviposition (Convey 1989). The male-competitive intensity can differ between species and/or situations in squid spawning aggregations. When competition among males for females is high, this could lead to prolonged copulation to guard mates more tightly. The captive conditions and experimental design in the present study could have affected the competitive intensity, so this idea has to be considered carefully with in situ observation. However it suggests that captive experiments in which the male-competitive intensity (e.g. operational sex ratio, male body size) is manipulated have great potential for helping understand the mate-guarding strategy. Sperm displacement has not been studied in detail in squids, but behavioral observations indicate it occurs in other cephalopods. Cuttlefish copulate only in the head-to-head position, and males spend more than half of the copulation period blowing water around the female’s seminal receptacles to displace spermatophores from previous mates (Hanlon et al. 1999, Naud et al. 2004). Males of the pygmy octopus Octopus joubini transfer spermatophores into a female’s mantle cavity, and those that mate with females who have recently mated with other males prolong the initial period of copulation before transferring spermatophores, presumably to displace previously deposited sperm using their hectocotylus (Cigliano 1995). In future studies, the reason for prolonged copulation in Loligo bleekeri can best be determined by observing if the copulative
period is prolonged before the transfer of spermatophores (sperm displacement hypothesis), during the transfer (sperm loading hypothesis) or after the transfer (copulatory guarding hypothesis).

In the second spawning in trial A, the sneaker male (A-M2) sired 8.5% of the offspring, which was higher than the rates for males that mated before the start of the experiment (A-3, B-3, B-4 and C-3). However those offspring could have resulted from the male-parallel mating by A-M2 before the first spawning, so the actual effect of sneaking was equal or less than 8.5%. Extra pair copulation has never been observed in *Loligo bleekeri* in the field, but it is frequently observed in the laboratory (20-75% of male-parallel copulation were sneaked at least once; Iwata, unpublished data). Addition, EPCs are observed in other loliginid squids, suggesting that it may be a common behavior in this taxon (*L. vulgaris reynaudii*: Hanlon et al. 1994, Hanlon et al. 2002; *L. pealei*: Hanlon 1996). Lone males use multiple tactics to obtain mates, both by fighting a paired male to replace it as a consort, or by performing extra-pair copulations (Hanlon 1998). In loliginid spawning ground, there are usually more males than females (Hanlon et al. 2004). So the large number of unpaired males will lead to frequently sneaking on the spawning grounds. Hanlon et al. (2002) observed six extra-pair copulations during nine male-parallel copulations in *L. vulgaris reynaudii* in field. Field observations suggest that lone males appear to frequently immigrate to and emigrate from spawning groups (Hanlon et al. 2004). Such movement between reproductive groups is probably adaptive to increase male reproductive success by shortening the time spent in a group and increasing the number of possible mates, suggesting that the fertilization success from one EPC would be low.

The effect of head-to-head copulation on paternity was also unclear, as males that
mated in this position also mated in the male-parallel position. Sperm transferred to opening of oviduct by mating in male-parallel position will be the first to meet when they leave the oviduct during spawning, and will thus have an advantage for fertilization. In addition, *Loligo vulgaris reynaudii* males transfer spermatophores twice during a single male-parallel copulation (Hanlon et al. 2002), so the amount of sperm transferred could be larger in a male-parallel copulation than in a head-to-head copulation, which will presumably increase the fertilization success. This suggests that the fertilization rate of sperm from the seminal receptacle should be low. This idea is supported by the result that the spermatophores transferred to near the seminal receptacle during two EPCs just before spawning sired only 8.5% of the offspring in the second spawning of trial A.

In trial B, the paternity pattern differed among egg capsules; most capsules were sired by a single male that mated both in the male-parallel and head-to-head positions, but several capsules showed multiple paternity (Fig. 3). In trial A, there was no significant difference in the paternity pattern between egg capsules; most had multiple paternity (Fig. 2). We do not have clear evidence to explain the difference in paternity between egg capsules and/or between broods, because we do not know which position the field male mated in. But the difference in paternity not only suggests the necessity of analysis using multiple egg capsules to accurately determine parentage, but also suggests the potential for further study to understand the sperm storage and release mechanism. It is unclear if sperm from several males are mixed or stored in layers in the seminal receptacle. Sperm storage mechanism has not been studied in detail in cephalopods, but in other taxa, sperm stratification occurs in the sperm storage organs. Histology and genetic analysis of mated female snow crabs show that multiple ejaculates are discretely
layered from the most recent to oldest in the spermatheca (Urbani et al. 1998, Saint-Marie et al. 2000). This stratification of recently received sperm on top of previously acquired sperm within the sperm storage organ results in a “last in – first out” system (Birkhead & Hunter 1990), and it might result in a paternity difference between firstly spawned eggs and lastly spawned eggs. An analysis in spawned order would be useful. Furthermore, the sperm usage mechanism is more complex in loliginid squids because they have two sperm storage sites (semenal receptacle and opening of oviduct). Shaw & Sauer (2004) found that in *Loligo vulgaris reynaudii* capsules, the distribution of offspring sired by different males is not random, suggesting that sperm from different males are not mixed within sperm storage organs and/or between the two sperm storage sites. In the present study, the fertilization rates of the most successful male and the other males did not differ significantly among egg capsules with multiple paternity in both trial A and trial B, suggesting that the paternity ratio of the final male to mate in the male-parallel position to the total of previous mates would be constant.

Except for minor sneaker males, loliginid males are larger than females. Such a sexual dimorphism is favored when males obtain size advantage in contests over females, female preferences, and sperm competition (Andersson 1994). Male competition (assessed based on the numbers of rushes and grabs) was more intense between similar sized males than between different sized males, and between different sized males, larger males attacked smaller males and the frequency of attacks increased as the size difference between males increased (Fig. 1). These results suggest that larger male size gives an advantage in contests over females. But the copulative success in the male-parallel position did not depend on the body size (Fig. 1). It is possible that larger males could not disperse other males completely, and which would allow smaller males
to approach females, due to the closed captive conditions in the present study. Large body size might not be needed for males to successfully mate in the male-parallel position. In the giant Australian cuttlefish *Sepia apama*, which shows sexual size dimorphism (males are larger than females) and male size dimorphism, females reject most mating attempts (43-70%) and sometimes reject large males but accept small males (Hall & Hanlon 2002, Naud et al. 2004). Boal (1997) suggested that female preference in the cuttlefish *S. officinalis* is determined by olfactory cues rather than visual cues. However, mating success is not the same as fertilization success. The present study suggests that the individual fertilization success varies depending on the mating position, timing and frequency. Hall and Hanlon (2002) showed that in *S. apama*, large males tend to form pairs with females for longer duration than small ones do. Large body size and high fighting ability would be favored for prolonged copulation and post copulatory guarding.

In summary, this study used both behavioral observations and genetic analysis to show that in *Loligo bleekeri*; there was multiple mating and multiple paternity. The male to mate last and frequently before the female spawned fertilized the most eggs. The paternity rates differed depending on the complex factor with mating position, timing, frequency and duration, relating with alternative reproductive behaviors. Further analysis with larger samples is needed to confirm the prediction that the final male to mate before a female spawns will sire the most eggs. These results suggest that studies of loliginid mating systems have a great deal of potential to help us understand the evolution of alternative reproductive strategies.
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Table 1. *Loligo bleekeri*. Morphological characters of the male squid used in each trial. Gonad weight = testis weight + accessory organ weight, GSI = gonad weight / body weight × 100, Body size ratio = mantle length of the larger male / Mantle length of the smaller male.

<table>
<thead>
<tr>
<th>Male ID</th>
<th>Mantle length (mm)</th>
<th>Body weight (g)</th>
<th>Gonad weight (g)</th>
<th>GSI</th>
<th>Body size ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trial A</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>A-M1</td>
<td>276</td>
<td>218</td>
<td>5.24</td>
<td>2.40</td>
<td>1.01</td>
</tr>
<tr>
<td>A-M2</td>
<td>272</td>
<td>219</td>
<td>4.98</td>
<td>2.27</td>
<td></td>
</tr>
<tr>
<td>Trial B</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>B-M1</td>
<td>253</td>
<td>234</td>
<td>4.98</td>
<td>2.13</td>
<td>1.18</td>
</tr>
<tr>
<td>B-M2</td>
<td>215</td>
<td>113</td>
<td>3.82</td>
<td>3.38</td>
<td></td>
</tr>
<tr>
<td>Trial C</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C-M1</td>
<td>303</td>
<td>173</td>
<td>5.20</td>
<td>3.01</td>
<td>1.40</td>
</tr>
<tr>
<td>C-M2</td>
<td>217</td>
<td>126</td>
<td>2.92</td>
<td>2.32</td>
<td></td>
</tr>
</tbody>
</table>
Table 2. *Loligo bleekeri*. Characteristics of the microsatellite loci used for parentage analysis. $H_O$ = observed heterozygosity, $H_E$ = expected heterozygosity, $P$ = probability of departure from Hardy-Weinberg expected genotype ratios, EEP = expected exclusion probability with a known parent, Circle = used locus for parental analysis in each trial, DDBJ = DNA Data Bank of Japan. A total of 47 individuals were genotyped for each locus.

<table>
<thead>
<tr>
<th>Locus</th>
<th>Sequence</th>
<th>Number of alleles</th>
<th>$H_O$</th>
<th>$H_E$</th>
<th>$P$</th>
<th>EEP</th>
<th>Combination of loci</th>
<th>DDBJ accession no.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lb1</td>
<td>TATGGTTA CACTACCCA C C TACGATAACCATTACA C GACG</td>
<td>16</td>
<td>0.894</td>
<td>0.856</td>
<td>0.407</td>
<td>0.723</td>
<td>○ ○ ○</td>
<td>AB100368</td>
</tr>
<tr>
<td>Lb3</td>
<td>GCCATCCGAA CAACACTTTA TACGTTATACGC TACCAT TT</td>
<td>24</td>
<td>0.936</td>
<td>0.938</td>
<td>0.532</td>
<td>0.874</td>
<td>○ ○ ○</td>
<td>AB100370</td>
</tr>
<tr>
<td>Lb4</td>
<td>CCCACGGGTTT CTGGGTATCCG GGTAAATGTA</td>
<td>13</td>
<td>0.787</td>
<td>0.861</td>
<td>0.139</td>
<td>0.724</td>
<td>– ○ –</td>
<td>AB100371</td>
</tr>
<tr>
<td>Lb5</td>
<td>TTTGACATGTGC CGCGAT ATATGCCCTT TGGCTTGC</td>
<td>13</td>
<td>0.787</td>
<td>0.870</td>
<td>&lt;0.01</td>
<td>0.699</td>
<td>○ – –</td>
<td>AB100372</td>
</tr>
<tr>
<td>Lb6</td>
<td>TTAATGAAA AGATGC GCGCT TACGGGTTT CTTAFT AC GG</td>
<td>30</td>
<td>0.938</td>
<td>0.950</td>
<td>0.903</td>
<td>0.899</td>
<td>– – ○</td>
<td>AB193344</td>
</tr>
</tbody>
</table>

Combined EEP 0.999 0.989 0.990 0.996
Table 3. *Loligo bleekeri*. Frequency of offspring’s maternal genotypes in each brood. *1*: Statistical test did not be conducted because both maternal alleles were same with one of the fathers (A-M1; 154/158) and it could not be confirmed which allele was hereditary from mother. *2*: Statistical test did not be conducted because the mothers were homozygous on the alleles.

<table>
<thead>
<tr>
<th>Trial</th>
<th>Female genotypes</th>
<th>Offspring's maternal genotypes (frequency)</th>
<th>( \chi^2 )</th>
<th>( P )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trial A</td>
<td>Lb1 154/158</td>
<td>154 or 158 (47)*1</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>Lb3 133/175</td>
<td>153 (22)</td>
<td>0.19</td>
<td>0.66</td>
</tr>
<tr>
<td></td>
<td>Lb5 104/124</td>
<td>178 (28)</td>
<td>1.72</td>
<td>0.19</td>
</tr>
<tr>
<td>Trial B first</td>
<td>Lb1 150/150</td>
<td>150 (270)*2</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>Lb3 157/163</td>
<td>157 (137)</td>
<td>0.06</td>
<td>0.81</td>
</tr>
<tr>
<td></td>
<td>Lb5 171/199</td>
<td>171 (135)</td>
<td>0.00</td>
<td>1.00</td>
</tr>
<tr>
<td>Trial B second</td>
<td>Lb1 150/150</td>
<td>150 (120)*2</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>Lb3 157/163</td>
<td>157 (68)</td>
<td>2.13</td>
<td>0.14</td>
</tr>
<tr>
<td></td>
<td>Lb5 171/199</td>
<td>171 (51)</td>
<td>2.70</td>
<td>0.10</td>
</tr>
<tr>
<td>Trial C</td>
<td>Lb1 152/160</td>
<td>152 (157)</td>
<td>0.97</td>
<td>0.32</td>
</tr>
<tr>
<td></td>
<td>Lb3 135/167</td>
<td>135 (158)</td>
<td>1.22</td>
<td>0.27</td>
</tr>
<tr>
<td></td>
<td>Lb4 170/170</td>
<td>170 (297)*2</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>
Table 4. *Loligo bleekeri*. Type and the duration (seconds) of mating behavior that occurred before each spawning, and paternity parentage for each male. MP = male-parallel copulation, EPC = extra pair copulation, HH = head-to-head copulation, ND: eggs in the first spawning in trial A were not analyzed because most died in the embryonic stage.

<table>
<thead>
<tr>
<th>Time until spawning</th>
<th>Mating behavior</th>
<th>Mating behavior</th>
<th>Mating behavior</th>
<th>Mating behavior</th>
<th>Mating behavior</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Trial A First</td>
<td>Trial A Second</td>
<td>Trial B</td>
<td>Trial C First</td>
<td>Trial C Second</td>
</tr>
<tr>
<td></td>
<td>A-M1</td>
<td>A-M2</td>
<td>A-3</td>
<td>B-M1</td>
<td>B-M2</td>
</tr>
<tr>
<td>52h</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
</tr>
<tr>
<td>50h</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
</tr>
<tr>
<td>34h</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
</tr>
<tr>
<td>24h</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
</tr>
<tr>
<td>20h</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
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<tr>
<td>16h</td>
<td>HH</td>
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<td>HH</td>
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<tr>
<td>12h</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
</tr>
<tr>
<td>4h</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
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<td>3h</td>
<td>HH</td>
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<tr>
<td>1h</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
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<tr>
<td>30min</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
</tr>
<tr>
<td>15min</td>
<td>MP(435)</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
<td>HH</td>
</tr>
<tr>
<td>10min</td>
<td>MP(23)</td>
<td>HH</td>
<td>HH</td>
<td>MP(23)</td>
<td>HH</td>
</tr>
<tr>
<td>Paternity (%)</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td></td>
<td>87.2</td>
<td>8.5</td>
<td>4.3</td>
<td>0</td>
<td>96.97</td>
</tr>
</tbody>
</table>

EPC(3) EPC(5) MP(439) EPC(223) EPC(40) MP(428) MP(323)
Fig. 1. *Loligo bleekeri*. Total number of rushes and grabs made by males on the other male present from one hour before beginning of spawning until one hour after beginning of spawning. The body size ratio of the two males was 1.01 in trial A, 1.18 in trial B, and 1.40 in trial C, and M1 was larger than M2 in trial B and C.
Fig. 2. *Loligo bleekeri*. Number of offspring sired by each father for the second spawning in trial A.
Fig. 3. *Loligo bleekeri*. Parentage rates of each father for 30 egg capsules in trial B. Ten eggs were randomly selected and genotyped in each egg capsule, and the number of offspring sired by each father was calculated using the parentage rate and the number of eggs in each egg capsule.