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<td>Author(s)</td>
<td>Wakamatsu, Jun-ichi; Okui, Jun; Hayashi, Nobutaka; Nishimura, Takanori; Hattori, Akihito</td>
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<tr>
<td>Citation</td>
<td>Meat Science, 77(4): 580-586</td>
</tr>
<tr>
<td>Issue Date</td>
<td>2007-12</td>
</tr>
<tr>
<td>Doc URL</td>
<td><a href="http://hdl.handle.net/2115/32300">http://hdl.handle.net/2115/32300</a></td>
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Zn protoporphyrin IX is formed not from heme but from protoporphyrin IX.

Jun-ichi WAKAMATSU*, Jun OKUI, Nobutaka HAYASHI, Takanori NISHIMURA, Akihito HATTORI

Division of Bioresource and Product Science, Graduate School of Agriculture, Hokkaido University, Sapporo, Hokkaido 060-8589, Japan

*Corresponding author.
Tel.: +81 11 706 2547; Fax.: +81 11 706 2547
E-mail address: jwaka@anim.agr.hokudai.ac.jp (J. Wakamatsu)

Meat Science Laboratory, Division of Bioresource and Product Science, Research Faculty of Agriculture, Hokkaido University
N-9 W-9, Kita-ku, Sapporo, Hokkaido 060-8589, Japan
We examined the effects of exogenous myoglobin, a bivalent chelator, and nitrite on Zn protoporphyrin IX (ZPP) formation by using model systems. ZPP was formed in a model solution without addition of exogenous myoglobin. After incubation, the amount of ZPP in a model solution was increased but that of heme was not decreased compared with the amounts before incubation. Protoporphyrin IX (PPIX) instead of ZPP also accumulated in a model solution with addition of EDTA, but the amount of heme was not reduced. These results suggested that ZPP was not formed by the Fe-Zn substitution in heme but was formed by the insertion of Zn into PPIX, which was formed independently. The fact that the effects of various factors in model systems with/without addition of a bivalent chelator were similar suggested that ZPP formation was strongly affected by PPIX formation. Inhibition of PPIX formation by nitrite might be the reason for the low levels of ZPP in cured meats.

Keywords: Zn protoporphyrin IX; protoporphyrin IX; color; dry-cured meat products; nitrite; nitric oxide
1. Introduction

Dry-cured meat products have been produced and consumed throughout history by a diversity of cultures in different parts of the world. Parma ham, one of the Italian dry-cured hams, is made from heavier pigs (> 150 kg in liveweight) and is processed for at least 12 months without addition of nitrite/nitrate (Toldrá, 2002). It has been reported that Parma ham has a red porphyrin derivative that is not nitrosylmyoglobin or oxymyoglobin and is extractable by water, acetone/water (75%/25%) and tetrahydrofuran (Morita, Niu, Sakata & Nagata, 1996; Møller, Adamsen & Skibsted, 2003; Parolari, Gabba & Saccani, 2003). The pigment was purified and identified to be Zn protoporphyrin IX (ZPP) by ESI-HR-MS analysis (Wakamatsu, Nishimura & Hattori, 2004). Moreover, the presence of zinc in the red pigment was directly demonstrated by scanning electron microscopy/energy dispersive X-ray microanalysis (SEM-EDX) (Wakamatsu, Ito, Nishimura & Hattori, 2007). Møller, Adamsen, Catharino, Skibsted, & Eberlin (2007) showed by using ESI-MS and TOF-MS analysis that zinc protoporphyrin IX is present not only in Parma ham but also in Iberian ham.

Effects of various factors on ZPP formation have been investigated using a model system, and it has been shown that oxygen inhibited ZPP formation and that ZPP was formed in the absence of microorganisms (Wakamatsu, Okui, Nishimura & Hattori, 2004). It has also been reported that the amount of ZPP and protoporphyrin IX (PPIX) were increased in pork and turkey during anaerobic conditions (Veberg et al., 2006). By use of autofluorescence of ZPP, it has been revealed that ZPP was distributed in not only lean meat but also fat tissue by using near-UV purple LED irradiation and an image analysis (Wakamatsu, Odagiri, Nishimura & Hattori, 2006).

On the other hand, exogenous myoglobin was added into a model solution that was established by Wakamatsu, Okui, Nishimura & Hattori (2004). The addition of exogenous myoglobin may affect ZPP formation. Moreover, the amount of ZPP formed...
has only been measured by fluorescence intensity, and the amount of heme has not been
determined at all. In order to elucidate the mechanism by which ZPP is formed, these
porphyrins must be quantitatively determined. A recent study has shown that ZPP is
present not only in Parma ham but also in Iberian ham but that ZPP content in meat
products cured with nitrite is very low (Adamsen Møller, Laursen, Olsen & Skibsted,
2006). It is still not clear why ZPP content in cured meat products is much lower than
that in dry-cured ham without addition of nitrite.

In this study, to elucidate the mechanism by which ZPP is formed, we investigated
the effects of various factors on ZPP formation by using a simpler model system with no
addition of exogenous myoglobin and a new model system in which PPX was formed by
the addition of ethylenediaminetetraacetate (EDTA). Then we measured the contents
of heme, ZPP and PPIX before and after incubation in these model solutions and
investigated stoichiometrically the mechanism by which ZPP or PPIX was formed.
Additionally, we investigated the inhibitory effect on ZPP formation of nitrite, which is
generally used in cured meat products.
2. Materials and methods

2.1. Materials

Pork loin samples (n = 3) were purchased from various retail markets. Myoglobin (from horse skeletal muscle), penicillin G potassium, streptomycin sulfate and gentamicin sulfate were purchased from Wako Pure Chemical Industries, Ltd. (Osaka, Japan). Acetone, acetic acid, acetate ethyl, methanol and ammonium acetate were purchased from Kanto Chemical Co. Inc. (Tokyo, Japan). Methanol was of HPLC grade and other chemicals were of analytical grade.

2.2. Model system

Model solutions were prepared as described by Wakamatsu, Okui, Nishimura & Hattori (2004). Experimental designs are shown in Table 1. Pork loin was homogenized with 2 volumes of distilled water using a homogenizer at 10,000 rpm for 1 min. Antibiotics were added to the model solutions to final concentrations of 100 units/ml for penicillin G potassium, 0.1 mg/ml for streptomycin sulfate and 0.05 mg/ml for gentamicin sulfate. The solutions were put into gas-impermeable bags and incubated at 25 °C for 5 days in darkness using an environmental chamber. An anaerobic condition was obtained by using a commercial kit (Oxygen Absorbing System; I.S.O. Inc., Yokohama, Japan) consisting of an oxygen absorber (A-500HS, I.S.O. Inc.) and an oxygen indicator tablet. The color of the oxygen indicator tablet changes from violet to pink when the oxygen concentration becomes less than 0.1%.

2.3. Fluorescent analysis
Extraction by acetone and fluorescent analysis were carried out as described by Wakamatsu, Okui, Nishimura & Hattori (2004) with minor modification. After extraction by acetone, the fluorescent spectra of the extracts were measured from 450 to 700 nm at 420/410 nm for excitation using a spectrofluorophotometer (RF-5300PC, Shimadzu Corp., Kyoto, Japan). Fluorescence intensity at 590 nm for excitation at 420 nm was regarded as the amount of ZPP formed, and fluorescence intensity at 630 nm for excitation at 410 nm was regarded as the amount of PPIX formed. All operations were carried out in darkness as much as possible.

2.4. Quantitative analysis of ZPP, PPIX and heme

The contents of ZPP, PPIX and heme were determined by HPLC as described by Guo, Lim & Peters (1991) with some modification. Porphyrins were extracted by acetic acid/acetate ethyl (1:4, v/v) as described by Smith, Doran, Mazur & Bush (1980) with some modification. One milliliter of model solution was mixed vigorously with 9 ml of acetic acid/acetate ethyl (1:4, v/v), and the mixture was held on ice for 30 min. After centrifugation (3,000 rpm, 15 min), the supernatant was collected. This extraction operation was carried out twice. The resulting solution was mixed with an equal volume of methanol/ammonium acetate (86:14, v/v, pH 5.16). The sample was filtered through a 0.45-µm filter (Minisart RC4, Sartorius AG, Goettingen, Germany). An STR ODS-II column (4.6 x 150 mm, Shinwa Chemical Industries Ltd., Kyoto, Japan) was used for separation of porphyrin, and the separation was carried out by isocratic elution using methanol/ammonium acetate (86:14, v/v, pH 5.16) at a flow rate of 0.6 ml/min at 35°C. Forty microliters of each sample was injected. The detection of ZPP and PPIX was carried out at excitation and emission wavelengths of 420/400 and 590/630 nm, respectively. Heme was monitored at 400 nm absorption. All operations of extraction
were carried out in darkness as much as possible.

2.5. Statistical analysis

Results are expressed as mean values of three independent trials in each individual. Data were analyzed using one-way analysis of variance (ANOVA) with Scheffé's test. Statistics were calculated using Microsoft Excel 2002 and a statistic add-in software (Excel Toukei 2002 for Windows, Social Survey Research Information Co., Ltd., Tokyo, Japan). A probability of $P<0.05$ was considered statistically significant.
3. Results and discussion

First, in order to investigate the effects of exogenous myoglobin in our established experimental model, the effect of the amount of added myoglobin on the amount of ZPP formed was examined (Fig. 1). When exogenous myoglobin was added at 0.1%, the amount of ZPP formed was maximal. With further addition of myoglobin, the amount of formed ZPP gradually decreased. The amount of ZPP formed was significantly increased with the addition of 0.1% exogenous myoglobin. The cause of the increase is unclear and it may be because myoglobin used in this study was metmyoglobin or included impurities. However, ZPP was formed despite no addition of myoglobin. Therefore, this result suggested that exogenous myoglobin is not always essential for the formation of zinc protoporphyrin IX and ZPP. It is also possible that ZPP is formed only from pork components. Since the addition of exogenous myoglobin in a model solution might make it difficult to elucidate the mechanism by which ZPP is formed, the mechanism should be investigated in a simpler model system with no addition of exogenous myoglobin. However, the cause of increase in ZPP formation by the addition of 0.1% exogenous myoglobin was not verified.

Next, we established a simpler model system composed of pork homogenate and antibiotics without addition of exogenous myoglobin and investigated the effects of various factors on ZPP formation (Fig. 2). Oxygen inhibited ZPP formation (Fig. 2A) and the amount of ZPP formed was increased with increase in the amount of pork added (Fig. 2B). The amount of ZPP formed rapidly increased during the first 3 days of incubation and increased slightly after the third day (Fig. 2C). The amount of ZPP formed was also increased with increase in incubation temperature within the range of temperatures examined (4, 15, 25 and 35°C) (Fig. 2D). The amount of ZPP formed peaked at about pH 5.5 and decreased considerably at lower or higher pH (Fig. 2E).
The results showing that ZPP formation depended on many factors suggested the involvement of an enzyme. The most notable dependence is the pH dependence. ZPP formation peaked at 5.5, which is very close to the ultimate pH of meat. The optimal pH range was narrower than those for general enzymes. Although the cause is not clear, this pH range might be an important clue to elucidate the mechanism by which ZPP is formed.

On the other hand, it has been reported that little ZPP was present in meat products cured with nitrite and/or nitrate (Adamsen Møller, Laursen, Olsen & Skibsted, 2006). Therefore, the influence of added nitrite was investigated by using the model system (Fig. 2F). The formation of ZPP was inhibited significantly by addition of nitrite of more than 10 µM (0.69 mg/L). Since the concentration at which ZPP formation was inhibited was much lower than the amount of nitrite added to common meat products, it was speculated that low ZPP content in meat products cured with nitrite and/or nitrate is caused by the inhibition of ZPP formation by nitrite. However, the cause is still unclear and future studies are needed.

In previous studies (Wakamatsu, Nishimura & Hattori, 2004; Wakamatsu, Okui, Nishimura & Hattori, 2004), the amount of ZPP formed was determined by fluorescence intensity and was not determined quantitatively. Heme content was also not measured. Therefore, we determined the quantities of ZPP and heme in the model solutions. As shown in Fig. 3, although the amount of ZPP had significantly increased after 5-day incubation, a decrease in the amount of heme was not observed. PPIX was not detected before and after incubation. Therefore, the total amount of ZPP and heme was increased compared with the total amount before incubation. If ZPP was just formed from heme with substitution of Zn for Fe, the sum total amount of heme and ZPP would not be increased. This result therefore suggested that ZPP was not formed by Fe-Zn substitution in heme but was independently formed in the model during incubation. At the last step of heme biosynthesis, heme was formed by inserting Fe
into PPIX (Ferreira, 1999; Dailey, Dailey, Wu, Medlock, Rose & Wang, 2000; Dailey, 2002). It is therefore possible that ZPP is formed by insertion of Zn into PPIX generated independently. Thus, the chelation of zinc by a chelator was predicted to inhibit the formation of ZPP and to result in the accumulation of PPIX.

Next, we observed the fluorescence spectrum of acetone extract after incubation in the model solution with addition of EDTA, a bivalent metal chelator (Fig. 4). A strong peak at 630 nm was observed in the model solution with addition of EDTA. The maximum excitation wavelength of the fluorescence peak was about 410 nm, shorter than the excitation wavelength of ZPP (420 nm) in agreement with results of a previous study (Shepherd & Dailey, 2005). This fluorescence peak coincided with that of PPIX (Shepherd & Dailey, 2005; Veberg et al., 2006). Thus, since the chelation of zinc by EDTA inhibited the formation of ZPP, ZPP seems to be formed by insertion of Zn into PPIX.

Next, in order to determine whether PPIX was formed from the demetalation of heme or independently, porphyrin contents in the model solutions with and without addition of EDTA were determined after incubation (Fig. 5). Although the addition of EDTA into a model solution significantly inhibited ZPP formation and the amount of PPIX was increased significantly, there was no change in heme content ($P>0.05$). The increase in PPIX content and the fact that the amount of heme did not change strongly suggested that PPIX was formed not from heme but from another substrate. EDTA was added into a model solution, but only a small amount of ZPP was present in the solution. This is due to the presence of ZPP originally contained in pork (see Fig. 3).

We also investigated the effects of various factors on the formation of PPIX in simple model systems with addition of EDTA. Oxygen tended to inhibit the formation of PPIX (Fig. 6A). PPIX was increased with increase in pork content in model solutions (Fig. 6B). The amount of PPIX formed increased rapidly during the first 3 days of incubation and increased slightly after the third day (Fig. 6C). The amount of PPIX
increased with increase in incubation temperature (Fig. 6D) and was maximal at about pH 5.5 (Fig. 6E). The addition of more than 10 µM of nitrite inhibited ZPP formation significantly (Fig. 6F). All of these patterns of PPIX formation are very similar to those of ZPP formation (Fig. 2). Thus, these results indicated that ZPP in a model solution was formed by the insertion of Zn into PPIX, which was formed not from heme but independently. It is therefore possible that ZPP is also formed by the same pathway in Parma ham. PPIX is formed from protoporphyrinogen IX (proto'gen) by protoporphyrinogen oxidase (PPO; EC 1.3.3.4) in the heme biosynthesis system (Dailey, 2002). The enzyme activity of mammalian PPO has been shown to be maximal at pH 8.6 to 8.7 (Poulson, 1976), much higher than results obtained in the present study. On the other hand, the inhibition of PPIX formation by oxygen was reduced in model systems with addition of EDTA, but there was no significant difference between them. There are two forms of PPO, an oxygen-dependent form and an oxygen-independent form (Dailey, 2002), and they may contribute to PPIX formation. Our results revealed that nitrite inhibited the formation of not only ZPP but also PPIX. Anaerobic oxidation of proto'gen to PPIX has been demonstrated in extracts of several microorganisms by nitrite or nitrate as an electron acceptor (Jacobs & Jacobs, 1976; Klemm & Barton, 1985). Since the inhibition of PPIX formation by nitrite is not still clear, further studies are needed to elucidate the contribution of proto'gen and PPO to ZPP/PPIX formation. The patterns of ZPP and PPIX formation were similar in model systems. The pH curves with maximum peaks at about 5.5 are noteworthy, and they might serve as a clue to elucidating the mechanism by which ZPP/PPIX is formed.

Furthermore, our results revealed that nitrite inhibited the formation of not only ZPP but also PPIX. Color formation in cured meat products is caused mainly by the reaction of endogenous myoglobin in meat with nitric oxide (NO) (Sakata, 2000; Fox, 1966). NO is produced from added nitrite (or nitrate) as the curing agent. NO has a high affinity for non-heme iron proteins (Fujii & Yoshimura, 1996) and copper proteins
NO is a paramagnetic molecule and has a high affinity for metal ions other than heme (Fujii and Yoshimura, 1996). Consequently, many metalloenzymes are inactivated by NO. Iron-sulfur (Fe-S) proteins have a very high affinity for NO. Mammalian ferrochelatase contains a [2Fe-2S] cluster (Ferreira, Franco, Lloyd, Pereira, Moura, Moura, & Huynh, 1994; Dailey, Dailey, Wu, Medlock, Rose & Wang, 2000), and the cluster is labile and sensitive to destruction by NO (Sellers, Johnson, & Dailey, 1996). It has been reported that ferrochelatase as a zinc chelatase participate in ZPP formation from myoglobin in pork loin extract and porcine heart extract (Ishikawa, Yoshihara, Baba, Kawabuchi, Sato, Numata & Matsumoto, 2006a; Ishikawa, Yoshihara, Baba, Kawabuchi, Sato, Numata & Matsumoto, 2006b). Although the participation of ferrochelatase is still not clear, the inactivation of metalloenzyme by NO may be involved in the inhibition of PPIX formation.

The results of this study might serve as a clue to elucidating the mechanism by which ZPP/PPIX is formed. Since the formation of PPIX had a strong influence on that of ZPP, it is necessary to investigate the steps prior to PPIX formation in more detail.
This study demonstrated that ZPP in a model solution was not formed by Fe-Zn substitution in heme but was formed by the insertion of Zn into PPIX, which was formed independently. It was shown that ZPP formation was strongly influenced by PPIX formation. The fact that there is little ZPP in cured meat products is because ZPP cannot be formed as a result of inhibition of PPIX formation by nitrite. A full understanding of ZPP formation in Parma ham awaits elucidation of the formation of PPIX or its precursor.

Acknowledgments

This study was supported partially by a Grant-in-Aid for Scientific Research (16780184) from the Ministry of Education, Culture, Sports, Science and Technology of Japan.


Poulson, R. (1976). The enzymic conversion of protoporphyrinogen IX to protoporphyrin

Fig. 1. Effect of exogenous myoglobin addition on the formation of ZPP. Model solutions with addition of various amounts of exogenous myoglobin were incubated for 5 days at 25°C anaerobically. The fluorescence intensity of acetone extracts was measured. Bars represent the standard deviation of the means (n=3). Values bearing different letters are significantly different ($P < 0.05$).

Fig. 2. Effects of oxygen (A), pork content (B), incubation day (C), temperature (D), pH (E) and nitrite (F) on the formation of ZPP in model solutions with no addition of exogenous myoglobin. The fluorescence intensity of acetone extracts was measured after incubation. Bars represent the standard deviation of the means (n=3). Values in the same examination bearing different letters are significantly different ($P < 0.05$).

Fig. 3. Heme ($\text{■}$) and ZPP ($\text{■}$) concentrations in model solutions without addition of exogenous myoglobin before and after incubation. Porphyrins extracted by acetic acid/acetic ethyl were measured by HPLC. Bars represent the standard deviation of the means (n=3). n.s.: not significant (vs before incubation).

Fig. 4. Fluorescence pattern of acetone extract of model solutions with (broken line, Ex. 410 nm) and without (solid line, Ex. 420 nm) addition of EDTA after 5 days at 25°C.

Fig. 5. Heme ($\text{■}$), ZPP ($\text{■}$) and PPIX ($\square$) concentrations in model solutions with and
without addition of EDTA after 5 days at 25°C. Porphyrins extracted by acetic acid/acetic ethyl were measured by HPLC. Bars represent the standard deviation of the means (n=3). n.s.: not significant (vs with addition of EDTA).

Fig. 6. Effects of oxygen (A), pork content (B), incubation day (C), temperature (D), pH (E) and nitrite (F) on the formation of ZPP in the model solution with addition of EDTA. The fluorescence intensity of acetone extracts was measured after incubation. Bars represent the standard deviation of the means (n=3). ab<sup>c</sup>cd<sup>e</sup>. Values in the same examination bearing different letters are significantly different (P<0.05). n.s.: not significant.

Table 1. Experimental designs by using a model system
Fluorescence intensity (Ex/Em: 420/590 nm)
Mb content (%)

Exogenous myoglobin content (%)

Figure 1
Figure 2

A: Bar graph showing nitrite levels under anaerobic and aerobic conditions. The graph indicates significantly higher nitrite levels in anaerobic conditions compared to aerobic conditions.

B: Graph showing the relationship between pork content (%) and fluorescence intensity (Ex/Em 420/590 nm). The fluorescence intensity increases with increasing pork content.

C: Graph showing the fluorescence intensity over days. The fluorescence intensity increases significantly from day 0 to day 10.

D: Graph showing the relationship between temperature (°C) and fluorescence intensity. The fluorescence intensity increases with increasing temperature.

E: Graph demonstrating the fluorescence intensity in relation to pH. The fluorescence intensity peaks at pH 5 and decreases as pH increases.

F: Graph illustrating the relationship between nitrite (μM) and fluorescence intensity. The fluorescence intensity decreases as nitrite levels increase.
Figure 3

Concentration (nmol/g meat)

Day 0

Day 5
Figure 6

A. Comparison of nitrite levels under anaerobic and aerobic conditions.
B. Relationship between pork content (%) and nitrite levels.
C. Fluorescence intensity over days with different treatments.
D. Effect of temperature on nitrite levels.
E. pH levels showing a peak at day 5.
F. Nitrite levels vs. pH, showing a decrease with increasing pH.
Table 1. Experimental designs by using a model system

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| Antibiotics were added to all model solutions to final concentrations of 100 units/ml for penicillin G potassium, 0.1 mg/ml for streptomycin sulfate and 0.05 mg/ml for gentamicin sulfate.

<sup>a</sup>; 0, 0.05, 0.1, 0.5, 1%; <sup>b</sup>; no adjusted; <sup>c</sup>; 0, 5, 10, 25, 50%; <sup>d</sup>; 0, 1, 3, 5, 10 day; <sup>e</sup>; 4, 15, 25, 37ºC; <sup>f</sup>; 0, 5, 10, 50, 100 µM