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HOKKAIDO UNIVERSITY
Novel Quantitative Assessment of Myocardial Perfusion by Harmonic Power Doppler Imaging During Myocardial Contrast Echocardiography

北海道大学
山田 聡
Novel Quantitative Assessment of Myocardial Perfusion by Harmonic Power Doppler Imaging During Myocardial Contrast Echocardiography

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There are no conflicts of interest.

Short title: Quantification by harmonic power Doppler imaging
STRUCTURED ABSTRACT

Objective: To test the hypothesis that the power of the received signal of harmonic power Doppler imaging (HPDI) is proportional to the bubble concentration under conditions of constant applied acoustic pressure, and to determine whether a new quantitative method can overcome the acoustic field inhomogeneity during myocardial contrast echocardiography (MCE) and identify perfusion abnormalities caused by myocardial infarction.

Methods: The relationship between Levovist concentration and contrast signal intensity (CI) of HPDI was investigated in vitro under conditions of constant acoustic pressure. MCE was performed during continuous infusion of Levovist with intermittent HPDI every sixth cardiac cycle in 11 healthy subjects and 25 patients with previous myocardial infarction. In the apical views myocardial CI (CI\textsubscript{myo}) was quantified in five myocardial segments. The CI from the left ventricular blood pool adjacent to the segment was also measured in dB and subtracted from the CI\textsubscript{myo} (relative CI (RelCI)).

Results: CI had a logarithmic correlation and the calculated signal power a strong linear correlation with Levovist concentration in vitro. Thus, a difference in CI of X dB indicates a microbubble concentration ratio of 10\textsuperscript{X/10}. In normal control subjects, CI\textsubscript{myo} differed between the five segments (p<0.0001), with a lower CI\textsubscript{myo} in deeper segments. However, RelCI did not differ significantly between segments (p=0.083). RelCI was lower (p<0.0001) in the 39 infarct segments (mean (SD) -18.6 (2.8) dB) than in the 55 normal segments (mean (SD) -15.1 (1.6) dB). RelCI differed more than CI\textsubscript{myo} between groups.

Conclusions: The new quantitation method described can overcome the acoustic field inhomogeneity in evaluation of myocardial perfusion during MCE. RelCI represents the ratio of myocardium to blood microbubble concentrations and may correctly reflect myocardial blood volume fraction.
Microbubble concentration within tissue during myocardial contrast echocardiography (MCE) represents the myocardial blood volume per unit volume (that is, myocardial blood volume fraction). [1] [2] The contrast signal intensity (CI) on greyscale B mode harmonic imaging after intravenous injection of ultrasound contrast agents has been suggested to reflect the microbubble concentration,[3] [4] and is used clinically as a quantitative measure to allow estimation of myocardial blood volume. [2] [5][6][7][8] Harmonic power Doppler imaging (HPDI), which has advantages over B mode imaging in opacification of the myocardium, has been used during MCE.[9][10][11][12][13][14] Although animal experiments have shown that the severity of coronary stenosis can be assessed by quantitative analysis of HPDI,[9] [15] some problems remain regarding quantitative assessment of myocardial blood volume with HPDI: the lack of detailed information regarding the relation between microbubble concentration and CI of HPDI; and the inhomogeneity of the insonified ultrasound field during MCE. As CI is dependent on the acoustic pressure,[16][17][18] comparison of myocardial CI (CI\textsubscript{myo}) between myocardial regions, to which different pressures must be applied, is of only limited value.

We hypothesised that the power of the received signal of HPDI is proportional to the bubble concentration under conditions of constant applied acoustic pressure. During MCE, applied acoustic pressure may be similar between a given myocardial region and the adjacent intracavity blood pool. Therefore, CI\textsubscript{myo} can be calibrated by using the CI from the adjacent blood pool to quantify the ratio (myocardium to blood) of microbubble concentrations. This study was performed to test our hypothesis in vitro and to determine whether our new quantitation method can overcome the acoustic field inhomogeneity during MCE and identify perfusion abnormalities caused by myocardial infarction.
METHODS

Microbubble concentration and CI of HPDI in vitro

Microbubble solutions of the ultrasound contrast agent Leovist, consisting of galactose based, air filled microbubbles (Schering AG, Berlin, Germany), were prepared at concentrations of 1, 2, 8, 32, and 128 mg/l by mixing with 1 litre of distilled water in glass beakers. The bubbles thus produced were mixed constantly by a magnetic stirrer. A transducer was fixed to the upper surface of a cylindrical jerry block 5 cm in height fixed in the centre of the beaker. Harmonic Doppler images were recorded by a Sonos 5500 (Philips Medical Systems, Andover, Massachusetts, USA) with a broadband harmonic transducer (S3 probe), with transmission at 1.3 MHz and reception at 2.6 MHz. The displayed dynamic range was 40 dB. The image angle was set at 15°, with a focal point of 6 cm, pulse repetition frequency of 4.6 kHz (maximum), and a medium packet size. The microbubble solutions were exposed to different transmission powers with mechanical indices (MIs) of 0.6 and 1.0. On the basis of the results of preliminary experiments, Doppler gain was adjusted to 20% for MI of 0.6 and 0% for MI of 1.0 to avoid both signal saturation and lack of opacification. To minimise the destruction of microbubbles in the 1 litre solution, the solution was scanned with each MI only once. For each microbubble concentration, five datasets were obtained from different solutions. Data were recorded on 5 inch magneto-optical disks.

Images were analysed digitally by a QuantiCon system (GE Medical Systems, Milwaukee, Wisconsin, USA). The mean CI of HPDI in the user defined region of interest (ROI) was measured in dB, acoustic units (AUs), or as the square of AU (AU²). AU is derived from the value in dB (X) as $255\times10^{(X-\text{DR})/20}$, where DR is the dynamic range. AU ranges from 0-255, and is theoretically proportional to the amplitude of the received signal. Therefore, AU² is theoretically proportional to the power of the harmonic power
Doppler signal. At the deep parts of the microbubble solution, acoustic pressure may be decreased because of attenuation of ultrasound by the presence of microbubbles in the path of the ultrasound beam. Therefore, CI was measured in a rectangular ROI 2.5 mm in height placed just below the border between the jerry and the solution, where the applied pressure was considered constant (fig 1).

Subjects of a clinical study

Eleven healthy subjects (mean (SD) 35 (9) years old, 10 men) and 25 consecutive patients with previous myocardial infarction (mean (SD) 65 (10) years old, 20 men) were enrolled in the present study. Myocardial infarction was confirmed by the presence of a Q wave in two or more ECG leads. Inclusion criteria of the patients were the presence of severe asynergy in the apical two or four chamber view on resting echocardiography, and the presence of defective or reduced perfusion on resting single photon emission computed tomography (SPECT) in the segments corresponding to those with echocardiographic wall motion abnormalities. Subjects with acute myocardial infarction within three months, unstable angina, pregnancy, or lactation were excluded from the present study. Coronary angiography was not a prerequisite for this study. The study protocol was approved by our institutional ethical committee. Informed consent was obtained from all subjects before participation in the study.

Myocardial contrast echocardiography

MCE was performed in normal subjects to determine whether the new quantitation method could overcome the inhomogeneous opacification between myocardial regions. In patients with myocardial infarction, MCE was performed within two weeks of SPECT.

HPDI (transmit, 1.3, reception, 2.6 MHz) was performed with the same ultrasound system. The MI was set at 1.6, with a pulse repetition frequency of 3.7-4.2 kHz (maximum), and a medium packet size. The focus was placed at the mid level of the left
ventricular cavity. Before contrast administration, the upper limit of Doppler gain was determined to avoid motion induced artefacts. The apical four chamber view was used in normal subjects and the apical two or four chamber view was used in each patient with myocardial infarction based on the location of the wall motion abnormality determined on resting echocardiography. Levovist (300 mg/ml) was administered into the right antecubital vein by continuous infusion at a rate of 2.5 ml/min. Two minutes after commencement of the infusion, images of intermittent HPDI were acquired by ultrasound transmission gated to end systole every sixth cardiac cycle. For accurate measurement of the signals from both the myocardium and the blood pool, two types of Doppler gain were used. The higher gain was optimised visually to obtain sufficient opacification of the intracavity blood adjacent to the basal segments and the lower gain was optimised to avoid saturation of signals from the blood pool adjacent to the apical myocardium. Images were recorded on magneto-optical disks.

Analysis of HPDI

Images were analysed by a QuantiCon system. The left ventricular wall in the apical two and four chamber views was divided into five segments: basal inferior/septal, mid-inferior/septal, apical, mid-anterior/lateral, and basal anterior/lateral. An ellipsoidal ROI was placed over every myocardial segment on each image and mean CI\text{myo} was measured in dB. Then the CI in the ROI placed at the left ventricular cavity adjacent to the myocardial ROI (CI\text{blood}) was also measured in dB and subtracted from the CI\text{myo}. This value, relative CI (RelCI), represents the ratio of microbubble concentrations between the myocardium and the adjacent blood pool, if our hypothesis regarding the relation between the bubble concentration and CI is correct. As signals from the blood pool are much stronger than those from the myocardium, saturation of the blood pool should be avoided for accurate measurement of RelCI. Insufficient opacification of the myocardium, which
is often seen in the far imaging field, should also be avoided. To eliminate such
inappropriate conditions, we decided that CI<sub>blood</sub> of 20 to 36 dB was acceptable. If the
CI<sub>blood</sub> was beyond the limits on every image, the segment was excluded from further
analysis.

**SPECT**

In each patient with myocardial infarction, 600 MBq of technetium-99m sestamibi
was injected at rest and images were recorded after one hour. SPECT images were
acquired by a rotating dual detector gamma camera with the detectors mounted at right
angles and fitted with high resolution collimators (VXHR, Vertex; ADAC Laboratories,
Milpitas, California, USA). Sixty four projections of 40 seconds were acquired over a
180° orbit. Data were processed with ramp and low pass filters. After back projection,
tomographic images were reconstructed in the horizontal and vertical long axis views.
Counts within a myocardial segment in each image were normalized to the highest counts
within that image and colour coded. SPECT images were interpreted by two observers
blinded to all other information. The left ventricular wall was divided into five segments
corresponding to those in echocardiographic analysis. Myocardial perfusion was
classified as normal, reduced, and defective.

**Statistical analysis**

Data are expressed as mean (SD). One way analysis of variance was used to
estimate the significance of differences in the indices between the five segments in the
normal subjects. Differences between the two segments were assessed by the paired \( t \) test.
Variables were compared between the normal myocardial segments and the infarct
segments by the unpaired \( t \) test. Differences were considered significant at \( p<0.05 \).
RESULTS

Relation between microbubble concentration and CI

CI in dB increased linearly with microbubble concentration in a logarithmic scale (fig 2A, B). The power of the received signal is theoretically proportional to the bubble concentration, and thus the theoretical slope of the CI in dB plotted against the concentration in a logarithmic scale as $10 \times \log(\text{concentration})$ should be 1.0. Best fitted slopes for MI of 0.6 and 1.0 were 0.84 and 0.96, respectively. There was an excellent linear correlation between bubble concentration and CI in AU^2 (fig 2C, D), indicating that the theoretical relation between bubble concentration and CI is valid for HPDI in conjunction with Levovist. Thus, a difference in CI of X dB under conditions of constant acoustic pressure indicates a ratio of microbubble concentration of $10^{X/10}$—for example, an increase in CI of 3 dB indicates doubling of the ratio of bubble concentration.

Clinical study

Fig 3 shows representative contrast images obtained in a normal control subject. Firstly, CI_{myo} and CI_{blood} were measured in each segment on the image at higher Doppler gain. In the mid-septal and apical segments, CI_{blood} was more than 36 dB. Accordingly, these two segments were measured again on the image at lower Doppler gain.

In all of the 55 segments in the 11 normal subjects CI_{blood} values at either higher or lower Doppler gain were within the defined limits of 20 to 36 dB and no data were excluded from the analysis. The CI_{myo} values measured on images at higher Doppler gain differed between the five segments ($p<0.0001$, analysis of variance): CI_{myo} was lower in the segments located deeper in the ultrasound field (fig 4). However, RelCI did not differ between the five segments ($p=0.083$, analysis of variance) (fig 4) and varied less than CI_{myo} in each segment.

In the 25 patients with previous myocardial infarction, 43 segments were akinetic
or severely hypokineic on resting echocardiography. Twelve of these were basal inferior/septal, nine mid-inferior/septal, 16 apical, five mid-anterior/lateral segments, and one basal anterior segment. In four of these 43 segments (9%; one basal inferior, one basal septal, and two apical) \( \text{Cl}_{\text{blood}} \) was lower than 20 dB even on the image taken at higher Doppler gain, and these segments were excluded from the analysis. \( \text{Cl}_{\text{myo}} \) and \( \text{RelCI} \) were measured in the remaining 39 infarct segments in 24 patients and the values were compared with those in the 55 normal segments in the normal control subjects (fig 5). \( \text{Cl}_{\text{myo}} \) was significantly lower in the infarct segments than in the normal segments but there was considerable overlap between the groups. \( \text{RelCI} \) differed more between the infarct and normal segments than \( \text{Cl}_{\text{myo}} \).

**DISCUSSION**

The relation between microbubble concentration and CI of HPDI was investigated under conditions of constant applied acoustic pressure. The results indicated that the values of CI measured in dB and in \( \text{AU}^2 \) (CI measured in \( \text{AU}^2 \) represents received signal power) are correlated logarithmically and linearly with Levovist concentration, respectively. We used this validated relation to calibrate \( \text{Cl}_{\text{myo}} \) during MCE by \( \text{Cl}_{\text{blood}} \) from the adjacent blood pool to estimate the ratio of myocardium to blood microbubble concentrations. In normal control subjects, myocardial opacification varied between regions because of acoustic field inhomogeneity, whereas the calibrated index \( \text{RelCI} \) varied much less both between myocardial segments and between individual subjects. We also observed reduced \( \text{RelCI} \) in the infarcted myocardium in patients with chronic coronary artery disease. These observations indicated that this new method can overcome the acoustic field inhomogeneity in quantitative evaluation of myocardial perfusion during MCE.
Microbubble concentration and CI of HPDI

Theoretically, relative changes in microbubble concentration produce corresponding changes of equal magnitude in the scattered intensity.[19] Schwarz and colleagues[19] showed previously that pulsed wave Doppler audio intensity (the square of the voltage) is proportional to the relative concentration of Levovist. Although they observed relative changes in microbubble concentration of only 4.6-fold, we observed changes in CI of HPDI corresponding to the relative changes in Levovist concentration of up to 128-fold. Consisting with the results of pulsed wave Doppler experiments reported by Schwarz and colleagues, our findings indicated that the theoretical relation between bubble concentration and CI is also valid for HPDI in conjunction with Levovist.

CI measured in dB, the log compressed value of the power of the received signal as $10 \times \log(\text{power})$, correlated logarithmically with microbubble concentration. Therefore, the difference in CI in dB should be regarded as the ratio of microbubble concentration as follows: $C_1/C_2 = 10^{X_1/10}/10^{X_2/10} = 10^{(X_1-X_2)/10}$, where $C_1$ and $C_2$ are the microbubble concentrations at sites 1 and 2, respectively, and $X_1$ and $X_2$ are CI in dB at sites 1 and 2, respectively. Consequently, the difference in CI in dB can be used on HPDI during MCE to estimate the ratio of microbubble concentrations between two sites where the applied acoustic pressure is similar—for example, an increase in CI of 3 dB indicates a twofold increase in the ratio of bubble concentration.

New quantitation method using HPDI during MCE

The insonified ultrasound field generated during MCE is not homogeneous,[13] which is principally due to attenuation by the presence of microbubbles in the path of the ultrasound beam. In the present study, CI$_{myo}$ was lower in myocardial regions located deeper in the ultrasound field in the normal control subjects. This was thought to be caused by the inhomogeneity of the insonified field, in which the acoustic pressure was
lower in the far field than in the near field. Therefore, it is impossible to estimate the relative microbubble concentration within the myocardium by simply comparing $CI_{\text{myo}}$ between segments. We devised a new quantitation method to resolve this problem by calibrating $CI_{\text{myo}}$ by using $CI_{\text{blood}}$ from the adjacent blood pool in the left ventricle. As the applied acoustic pressure would be almost equal between a myocardial segment and the adjacent blood pool, the myocardium to blood ratio of microbubble concentrations can be estimated by the CI measurements at these two sites. Because the acoustic pressure is higher in the centre and less towards the edges of the image, our quantitative method may not completely compensate for these lateral variations. Although the difference in the pressure between the two close ROIs may be small and the lateral variations are expected to be substantially compensated, this issue will be investigated further in future studies.

The results of the present clinical study indicated that $CI_{\text{myo}}$ on HPDI is quite variable between myocardial segments and that there is considerable interindividual variability of $CI_{\text{myo}}$ from each myocardial segment. On the other hand, $RelCI$ varied to a much lesser degree both between myocardial segments and between individual subjects. Thus, the new quantitative method described here can overcome the changes in myocardial opacification with changes in the applied acoustic pressure. In patients with chronic coronary artery disease, $RelCI$ of the infarcted myocardium was shown to be reduced more notably than $CI_{\text{myo}}$ alone. Our findings suggest that $RelCI$ represents the ratio of microbubble concentrations between the myocardium and intracavity blood pool independently of the applied acoustic pressure.

Theoretically, by using the new index $RelCI$ calculated by subtracting $CI_{\text{blood}}$ from $CI_{\text{myo}}$ in dB, the myocardium to blood ratio of microbubble concentrations can be estimated as $10^{RelCI/10}$. If the microbubble concentration in the blood present in the left ventricular myocardium is equivalent to that in the intracavity blood pool, myocardial blood volume
fraction can be derived as $10^{\frac{\text{RelCI}}{10}} \times 100$ (%) because blood volume fraction of the intracavity blood is 100%. In the present study, the myocardial blood volume fraction at end systole was estimated to be 3.1% from the mean RelCI in the normal control subjects. This value is not much different from the cast myocardial blood volume fraction of 4.5% reported in the pig heart,[4] [20] although morphometric data regarding myocardial blood volume in the human heart are not available. Thus, RelCI may correctly reflect myocardial blood volume fraction.

**Study limitations**

The in vitro and clinical experiments used only a single contrast agent and a single ultrasound apparatus. However, as the properties of each of these are relatively generic, it should be possible to extrapolate the results to other agents or instruments. Although our new quantitative method requires measurement of signals from the myocardium and from the intracavity blood pool, the dynamic range of HPDI is relatively low. Therefore, it was necessary to adjust the Doppler gain and to record two images at different gains to allow analysis of all myocardial segments. Thus, the development of newer imaging modalities with a higher dynamic range is required.
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**FIGURE LEGENDS**

**Figure 1**  Harmonic power Doppler image of Levovist solution. The contrast signal intensity (CI) decreased in proportion to depth of the solution due to ultrasound attenuation by the presence of microbubbles. The region of interest (ROI) was placed just below the border between the jerry block and the solution to measure CI under conditions of constant applied acoustic pressure.

**Figure 2**  Relations between Levovist concentration and CI in dB with mechanical index (MI) of (A) 0.6 and (B) 1.0, and between the concentration and CI in squared acoustic units (AU²) with MI of (C) 0.6 and (D) 1.0. In A and B, the concentration is expressed in logarithmic scale where the value in dB was calculated as $10 \times \log(\text{concentration})$. In addition, CI in dB can be regarded as the log converted value of the power of the ultrasound signal as $10 \times \log(\text{power})$.

**Figure 3**  Representative results of measurement of relative CI (RelCI). Images obtained (A) at the higher Doppler gain of 65% and (B) at the lower Doppler gain of 45% were analysed. The RelCIs were calculated as $-15.9$ dB in the basal septal segment, $-17.3$ dB in the mid-septal segment, $-16.6$ dB in the apical segment, $-16.0$ dB in the mid-lateral segment, and $-16.1$ dB in the basal lateral segment.

**Figure 4**  (A) Myocardial contrast intensity (CI\text{myo}) and (B) RelCI in normal control subjects. Apical, apical segment; Basal L, basal lateral segment; Basal S, basal septal segment; Mid L, mid-lateral segment; Mid S, mid-septal segment. ANOVA, analysis of variance; NS, not significant.
Figure 5  (A) CI$_{myo}$ and (B) RelCI in normal and infarct segments.
Figure 2

A

[Graph showing a linear relationship between concentration (mg/L) and CI (dB) for MI=0.6. The equation is y=0.84x+10.2 with r=0.985, p<0.0001.]

B

[Graph showing a linear relationship between concentration (mg/L) and CI (dB) for MI=1.0. The equation is y=0.96x+16.3 with r=0.989, p<0.0001.]

C

[Graph showing a linear relationship between concentration (mg/L) and CI (AU²) for MI=0.6. The equation is y=26.2x+222 with r=0.957, p<0.0001.]

D

[Graph showing a linear relationship between concentration (mg/L) and CI (AU²) for MI=1.0. The equation is y=176x+1020 with r=0.968, p<0.0001.]
Figure 4

Part A: Cl_{myo} (dB)

- Basal S: 10.5 (4.5)
- Mid S: 18.2 (4.0)
- Apical: 22.9 (2.7)
- Mid L: 11.2 (3.2)
- Basal L: 8.3 (1.8)

Significance levels:
- p < 0.0001
- p < 0.001
- p < 0.01
- p < 0.05

Part B: RelCl (dB)

- Basal S: -15.5 (1.1)
- Mid S: -14.6 (1.2)
- Apical: -14.5 (1.6)
- Mid L: -14.9 (2.0)
- Basal L: -16.1 (1.5)

NS (ANOVA)
Figure 5

(A) CI_{myo} (dB)

- Normal segments: 14.2 (6.4)
- Infarct segments: 11.3 (5.4)

p < 0.05

(B) RelCI (dB)

- Normal segments: -15.1 (1.6)
- Infarct segments: -18.6 (2.8)

p < 0.0001