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Immunocytochemical Analysis of Pituitary Cells in Pre-spawning Chum Salmon

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ABSTRACT—Pituitary hormones regulate various physiological functions during spawning migration in salmonid. Cytological features of pituitary cells were therefore immunocytochemically examined by use of antisera against homologous hormones in pre-spawning chum salmon (Oncorhynchus keta) caught from Ishikari Bay and the Chitose River in October, 1996. Immunoreactivity and sizes of pituitary cells were determined by a computer-aided image analyzing technique. Immunoreactivity in growth hormone (GH) cells was stronger with enlarged cell sizes in freshwater (FW) fish than in seawater (SW) ones of both sexes. Majority of prolactin (PRL) cells also had significantly stronger immunoreactivity with enlarged cell sizes in FW fish than in SW animals of both sexes. Immunoreactivity in somatolactin (SL) cells was markedly stronger with enlarged cell sizes in FW fish than in SW ones of both males and females. In addition, greater portions of SL cells were strongly stained in FW animals than in SW ones. A greater portion of gonadotropin (GTH) I cells had stronger immunoreactivity with reduced cell sizes in FW fish of both sexes, when compared with SW fish. Conversely, GTH II cells had significantly stronger immunoreactivity with enlarged cell sizes in FW ones of both sexes. In proopiomelanocortin (POMC)-derived hormone producing cells, adrenocorticotropin (ACTH) cells had stronger immunoreactivity in FW animals of both sexes, while cell sizes did not change. In melanotropes, the cells immunoreactive to α-melanophore stimulating hormone (α-MSH) antiserum had stronger immunoreactivity with reduced cell sizes only in FW males, while the cells immunoreactive to β-endorphin antiserum had stronger immunoreactivity with reduced cell sizes in FW fish than in SW animals of both sexes. Implication of these results was discussed along with previous reports on gene expression of pituitary hormone precursors.

INTRODUCTION

The pituitary gland of salmonids serves major endocrine function by secreting a variety of hormones that play vital roles in all aspect of life cycle including somatic growth, sexual maturation, reproduction, spawning migration and environmental adaptation. In teleosts, as in other vertebrates, different types of pituitary cells have been identified using histochemical and immunocytochemical techniques in many species, such as sockeye salmon, Oncorhynchus nerka (McKeown and van Overbeeke, 1971; Nagahama, 1973); chum salmon, O. keta (Nagahama, 1973; Naito et al., 1993a); goldfish, Carassius auratus, medaka, Oryzias latipes, eel, Anguilla japonica (Nagahama, 1973); some teleost species (Ball and Baker, 1969); molly, Poecilia latipinna (Batten and Ball, 1975; Batten, 1986); and seabass, Dicentrarchus labrax (Cambre et al., 1986). These studies reported identification and localization of different types of pituitary cells, and established that each type of pituitary cells synthesizes particular hormone for particular functions.

Among the pituitary hormones, growth hormone (GH), prolactin (PRL) and somatolactin (SL) belong to the same hormone family due to their structural similarity (Ono et al., 1990; Takayama et al., 1991; Rand-Weaver et al., 1991). GH and PRL are present throughout the vertebrates (Kawauchi and Yasuda, 1989a), while SL is unique in teleosts (Rand-Weaver and Kawauchi, 1993; Dores et al., 1996). In teleosts, GH is involved in somatic growth (Donaldson et al., 1979), PRL in freshwater adaptation (Hirano, 1986) and SL in osmoregulation as well as final maturation (Rand-Weaver et al.,...
1992; Rand-Weaver and Swanson, 1993; Olivereau and Rand-Weaver, 1994a; Kakizawa et al., 1995). The levels of GH/PRL/SL family mRNAs were elevated in freshwater (FW) chum salmon compared with seawater (SW) ones during spawning migration (Taniyama et al., 1999). Further, the plasma levels of GH/PRL/SL family hormones were higher in FW fish than in SW ones during homing migration in chum salmon (Kakizawa et al., 1995). Therefore, cytological analyses of GH/PRL/SL family cells are important for considering changes in cellular activity during spawning migration.

Teleosts have two gonadotropins (GTH), referred to as GTH I and GTH II (Suzuki et al., 1988; Kawachi et al., 1989b; Swanson et al., 1991). They are produced separately in distinct cells which differentially expressed the gene for β-sub-unit of each GTH (Naito et al., 1991). GTH I is responsible for early stages of gonadal development like tetrapod follicle-stimulating hormone (FSH), while GTH II stimulates final maturation like tetrapod luteinizing hormone (LH). The levels of GTH I β mRNA did not change during homing migration in chum salmon, while GTH II β mRNA considerably increased in FW fish of both sexes (Kitahashi et al., 1998). The plasma levels of GTH II also increased at the final stage of maturation in teleost fishes, as reported in striped bass (Mylonas et al., 1986; Balm and Anguilieria, 1986; Balm and Pottinger, 1995), while α-MSH cells for skin coloration (Lamacz et al., 1991). Cytological studies of POMC-derived hormone producing cells have done in many teleost species such as rainbow trout (Rodrigues and Sumpter, 1984), Mediterranean yellowtail (Garcia-Hernandez et al., 1997) and primitive actinopterygians (Joss et al., 1990). However, studies on cytological changes of POMC-derived hormone producing cells during spawning migration are very limited. The present study therefore included analyses of ACTH cells and α-MSH cells which also contain β-endorphin immunoreactivity to see whether they have significant roles during spawning migration in salmonids.

In the present study, changes in cytological features of pituitary cells were immunocytochemically examined for better understanding of endocrine events in homing salmon. Pre-spawning chum salmon were obtained from the coastal sea and at the hatchery. Tissue sections were stained with antisera against homologous hormones. Hence, we did not immunostain thyroid-stimulating hormone (TSH) cells, because anti-salmon TSH was unavailable.

### MATERIALS AND METHODS

#### Fish

Pre-spawning chum salmon (Oncorhynchus keta) of both sexes in the final stages of spawning migration were caught at Atsuta, a fishermen’s village facing the Ishikari Bay, and Chitose, a town through which a branch of the Ishikari River runs, in October, 1996. The fish caught at Chitose were maintained at the Chitose Salmon Hatchery (Hokkaido, Japan) until fully matured. The sampling sites are the main key points along the migratory route of Ishikari stock chum salmon. Since the Ishikari is a prominent big river, and a vast number of juveniles are released only from the Chitose Salmon Hatchery, the salmon caught at Atsuta and Chitose can be considered to belong to the same genetic population. The animals caught at Atsuta are referred to as seawater (SW), and those captured at Chitose as fresh-water (FW) salmon. Immediately after anesthetization with 0.02% tricaine-methylsulfonate (MS 222, Sigma), the animals were weighed and measured for body length, and then they were decapitated. The maturity of the gonads was later assessed by the gonadosomatic index (GSI, gonad weight x 100/body weight). The changes in GSI values in the SW and FW fish, an increase in females and a decrease in males, indicate that the final maturation of the gonads in the experimental fish might occur during migration from Atsuta to Chitose (Table 1).

#### Table 1. Body weight (kg), gonadosomatic index, GSI (%) and hepatosomatic index, HSI (%) of seawater (SW) and freshwater (FW) chum salmon of both sexes used in the present study.

<table>
<thead>
<tr>
<th>Sex</th>
<th>Sampling site</th>
<th>N</th>
<th>Body weight (kg)</th>
<th>GSI (%)</th>
<th>HSI (%)</th>
</tr>
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<tbody>
<tr>
<td>Δ</td>
<td>Atsuta (SW)</td>
<td>3</td>
<td>3.9 ± 0.3</td>
<td>5.4 ± 0.3</td>
<td>1.3 ± 0.2</td>
</tr>
<tr>
<td></td>
<td>Chitose (FW)</td>
<td>3</td>
<td>3.2 ± 0.8</td>
<td>4.2 ± 0.4</td>
<td>1.9 ± 0.1</td>
</tr>
<tr>
<td>Δ</td>
<td>Atsuta (SW)</td>
<td>3</td>
<td>2.7 ± 0.3</td>
<td>12.0 ± 1.5</td>
<td>2.6 ± 0.5</td>
</tr>
<tr>
<td></td>
<td>Chitose (FW)</td>
<td>3</td>
<td>3.1 ± 0.1</td>
<td>23.2 ± 1.4</td>
<td>1.2 ± 0.1</td>
</tr>
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#### Tissue preparation

The pituitary glands were collected by the decapitation after anesthetization with 0.02% MS-222 and were fixed in phosphate buffered 4% paraformaldehyde (pH 7.5) at 4°C overnight. The samples were dehydrated through a series of graded ethanol and were embedded in paraplast. Tissue blocks were transversely sectioned at 8 μm thickness. The sections were then divided into eight parallel sets in a manner that peripheral, sub-medial and medial sections should be mounted on single gelatinized slides.

#### Immunocytochemistry

Pituitary cells were immunostained with a Vectastain ABC (Avidin-Biotin-Peroxidase Complex) kit following the manufacturer’s instructions with slight modification. In brief, the sections were deparaffinized, rehydrated through graded ethanol, and were washed in a phosphate-buffered saline (PBS, 0.01 M, pH 7.5). The rehydrated sections were pre-incubated in 1% normal goat serum in PBS in a moist chamber at room temperature (RT) for 30 min. The tissue sections were then incubated with a primary antiserum in the moist chamber at RT for 48 hr. Afterward, they were washed in PBS three times, five minutes each, and were incubated in a solution of biotinylated anti-rabbit IgG at RT for 30 min followed by ABC complex (30 min at RT). After wash in tris-buffer (TB, pH 7.5, 0.01 M), the sections were immersed in a DAB solution (3, 3’ diaminobenzidine tetrahydrochloride, 0.1%; H2O2 , 0.02%; tris-buffer, 0.05 M, pH 7.5) at 20°C for 10 min, rinsed in deionized water, and were coverslipped with permount.

Specificity of antisera (GH, GTH I β and GTH II β) were confirmed by an absorption test using chum salmon GH, GTH I β and GTH II β provided by Prof. H. Kawauchi, Kitasato University, Japan. Immunostaining was completely prevented by absorption with the corre-
**RESULTS**

Localization of pituitary cells

In the pituitary of pre-spawning chum salmon, PRL-ir cells were localized in the rostral pars distalis (RPD) (Fig. 1), organized in a follicular form both in SW and FW animals. GH, GTH I, GTH II and ACTH cells were localized in the proximal pars distalis (PPD), while SL, α-MSH and β-endorphin cells were localized in the pars intermedia (PI) (Fig. 1). The cells in the PPD (GH, GTH I and II) showed scattered distribution pattern instead of distinct glandular form. The distribution pattern of these hormonal cells coincided well with the previous reports cited in Introduction, indicating that the antisera used in the present study specifically stained the particular cell type.

**GH cells**

Frequency distribution histograms of immunoreactivity in GH cells in the males and FW females (Figs 2A, B and D) indicate that GH cells in these pre-spawning chum salmon are composed of a single cell population. However, the histogram of GH cells in the SW females (Fig 2C) shows the presence of two cell populations, weakly stained and strongly stained (tested by Gauss Model, figure not shown), as was seen in the juvenile chum salmon that were reared in a low population density and showed a better growth rate than those in a high population density (Salam et al., 1999). Immunoreactivity in GH cells was stronger (p<0.001) in FW males than in SW males (Figs 2A and B), while the structure of GH cell populations changed in females during the last phases of

The optimal dilutions of primary antisera were determined by a preliminary experiment using a series of serially diluted antisera. The antisera used in the present study were anti-chum salmon GH and PRL, 1:64000; anti-chum salmon SL, 1:10000; anti-chum salmon GTH I and II, 1:16000; anti-kokanee salmon ACTH, 1:8000; anti-α-MSH and anti-β-endorphin, 1:4000. The antisera were diluted with PBS containing 0.5% bovine serum albumin (BSA). Backgrounds of immunostained sections were carefully checked since false staining may occur due to endogenous biotin in animal tissues (Wang and Pevsner, 1999).

The rabbit anti-chum salmon GH antiserum (Lot No. 8502), GTH I (Lot No. 8510) and GTH II (Lot No. 8506) were provided by Prof. H. Kawauchi, Kitasato University, Japan. Anti-chum salmon PRL and SL were prepared by Prof. T. Kaneko, Tokyo University, Japan. Anti-kokanee salmon ACTH, α-MSH and β-endorphin were generously provided by Prof. R. M. Dores, University of Denver, USA.

**Selection of immunostained sections**

Camera-lucida drawings were made using all immunostained sections to measure the sizes of areas immunoreactive to each of applied antisera. For each hormone, the section that contained the largest immunoreactive (ir) area was selected as the representative one of individual animal, and was used for later image analysis. The photomicrographs were taken from the selected section with the reverse film (Fuji film, Sensia II) by use of a x 100 objective lens of multipurpose microscope (Zeiss Axiohot, Germany). Six to eight photomicrographs of ca. 100 μm x 150 μm areas were taken from the anterior, posterior, dorsal, ventral and medial parts of the selected sections to avoid deviation of analyzed cells from the whole population of each type of pituitary cells.

**Image analysis**

The photomicrographs were scanned with a film scanner (Polaroid Scanner, Polaroid Corp., USA) at 1350 dpi for image analysis. All recognizable immunoreactive cells were analyzed from the selected photomicrograph as mentioned above. Immunoreactivity and sizes of all recognizable immunoreactive cells on the scanned film were measured by a computer-aided Image Pro-Plus software (Media Cybernetics, USA). Then, cell sizes were calibrated as μm² from the number of pixels. Magnitudes of immunoreactivity were expressed as arbitrary units of gray scale after subtraction of background from the nearby unstained cells.

**Statistical analysis**

Data are expressed as mean±SEM. After F-test for variance, Welch t-test was used to test the differences between SW and FW animals for immunoreactivity and cell sizes. The correlation between cell sizes and immunoreactivity was analyzed by linear correlation method. When immunoreactive cells shown in a frequency distribution histogram of immunoreactivity or cell sizes seemed to be composed of multiple populations, the presence of the number of peaks in the histogram was analyzed by use of a computer software for Gauss Model (Microcal Origin, Microcal Software Inc., USA).
spawning migration. The weakly stained cell population seen in SW females was markedly decreased in FW females (Figs 2C-D). Such change resulted in a slight but significant increase (p<0.01) in GH immunoreactivity in FW females compared with SW females (Figs. 2C-D). The sizes of GH-immunoreactive (ir) cells were slightly enlarged in FW males (Figs. 2E and F, p<0.01) and females (Figs. 2G and H, p<0.05) than in SW ones.

**PRL cells**

PRL-ir cells in FW fish were more strongly immunostained than those in SW fish (Figs. 3A and B). The skewed patterns

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Fig. 2. Frequency distribution histograms of immunoreactivity (A-D) and cell sizes (E-H) of GH-ir cells in the pituitaries of SW and FW fish of both males and females.
Fig. 3. Photomicrographs of PRL (A and B), SL (C and D), GTH II (E and F) and β-endorphin (G and H) cells in the pituitaries of SW (A, C, E and G) and FW (B, D, F and H) female chum salmon. Note the strongly stained cells in FW animals (right panel). Differences in immunoreactivity of these hormonal cells between SW and FW males were similar as shown in these photomicrographs (figures not shown). Scale bar, 40 µm.
of frequency distribution histograms of PRL immunoreactivity and cell sizes may show that PRL cells are composed of dominant and multiple small populations, although the peaks were single (Figs 4A-H). These PRL cell populations similarly behaved during the last phases of spawning migration. Actually, the distribution histograms of immunoreactivity in FW fish of both sexes concordantly shifted towards the right compared with those in SW fish, resulted in the fact that almost all PRL cells were strongly stained in FW fish (Figs. 4A vs B, and C vs D). In both males and females, PRL-ir cells thus had significantly ($p<0.001$) stronger immunoreactivity in FW fish than in SW ones. In addition, PRL-ir cell sizes enlarged significantly ($p<0.001$) in FW animals than in SW fish of both sexes (Figs. 4E vs F, and G vs H). The distribution histograms of PRL cell sizes shifted towards the right in a coordinated manner, and showed that the pituitaries of FW fish contained greater num-

**Fig. 4.** Frequency distribution histograms of immunoreactivity (A-D) and cell sizes (E-H) of PRL-ir cells in the pituitaries of SW and FW fish of both males and females. Note that greater numbers of PRL cells in FW fish of both sexes are strongly stained.
numbers of larger PRL cells compared with the SW ones (Figs. 4E-H). An analysis by scatter plot distributions further revealed that the numbers of strongly stained larger cells were greater in FW animals regardless of sexes (figure not shown).

**SL cells**

Although the frequency distribution histograms of SL immunoreactivity and cell sizes were skewed, major populations of SL cells similarly increased their immunoreactivity and cell sizes during the upstream migration. Immunoreactivity in SL-ir cells in FW fish was thus stronger than that in SW fish of both sexes (p<0.001) (Figs. 3C and D, 5A-D). Frequency distribution histograms of immunoreactivity indicated that almost all SL cells had stronger immunoreactivity in FW fish (Figs.

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**Fig. 5.** Frequency distribution histograms of immunoreactivity (A-D) and cell sizes (E-H) of SL-ir cells in the pituitaries of SW and FW fish of both sexes. Note that greater numbers of SL cells in FW fish of both sexes are strongly stained.
Fig. 6. Scatter plot distributions between immunoreactivity and cell sizes of SL-ir cells in the SW and FW males (A and B) and females (C and D). Note that larger SL cells in FW fish show stronger immunoreactivity. Standard deviations are shown in the white circle. Arrows indicate the presence of cell populations of strongly stained larger cells.

GTH I cells

GTH Iβ-ir cells had significantly stronger immunoreactivity in FW fish than in SW animals of both sexes (p<0.001) (Figs. 7A-D). Distribution histograms of immunoreactivity indicate the presence of greater numbers of strongly stained GTH I cells in FW fish of both sexes (Figs. 7B and D). The rightward shift of the histogram in FW females was more prominent than that in FW males. Whereas, GTH I cell sizes significantly reduced in FW fish than in SW fish of both sexes (p<0.001) (Figs. 7E-H). The changes in the shapes of histograms of GTH-I cell sizes were the most drastic among all pituitary cells examined in the present study. A scatter plot distribution analysis between immunoreactivity and cell sizes showed that smaller cells predominant in FW animals were strongly immunostained (figure not shown).

GTH II cells

In GTH IIβ-ir cells, immunoreactivity was stronger in FW fish than in SW ones in both sexes (p<0.001)(Figs. 3E and F, 8A-D). Distribution histograms of immunoreactivity showed that a broad distributional pattern with multiple peaks in SW fish was transformed into a single peak Gaussian pattern, which was composed of a greater numbers of strongly stained GTH II cells, in FW fish of both sexes (Figs. 8B and D). GTH IIβ-ir cell sizes also were significantly larger (p<0.001) in FW fish compared with SW ones of both sexes (p<0.001) (Figs. 5E-H). Histograms of cell sizes were broaden towards the right, indicating that the numbers of larger cells were greater in FW animals of both sexes (Figs. 5F and H). Since scatter plot distribution showed that the numbers of strongly stained larger SL cells were greater in FW animals when compared with those in SW fish of both sexes (arrows in Figs. 6B and D), it is apparent that a considerable number of SL cells concomitantly increased their immunoreactivity and sizes.
than in SW ones of both sexes (Figs. 8E-H). Histograms of cell sizes shifted towards higher values in FW fish, indicating the presence of greater numbers of larger cells in FW animals compared with SW ones (Figs. 8F and H). Scatter plot analysis showed that, in contrast to GTH Iβ-ir cells in FW fish which were small and strongly immunoreactive, larger GTH-IIβ-ir cells in FW fish were strongly stained, when compared with GTH IIβ-ir cells in SW fish (figure not shown).

Fig. 7. Frequency distribution histograms of immunoreactivity (A-D) and cell sizes (E-H) of GTH I-ir cells in the pituitaries of SW and FW fish of both sexes. Note that greater numbers of GTH I cells in FW fish of both sexes are strongly stained.
Fig. 8. Frequency distribution histograms of immunoreactivity (A–D) and cell sizes (E–H) of GTH II-ir cells in the pituitaries of SW and FW fish of both sexes. Note that greater numbers of GTH II cells in FW fish of both sexes are strongly stained.

POMC-derived hormone cells
Immunoreactivity in corticotropes (ACTH cells) in the PPD was stronger (p<0.001) in FW fish than in SW ones of both sexes (Figs. 9A–D), while cell sizes did not change either in males or females (Figs. 9E–H).

In the pars intermedia, the localization of α-MSH-ir cells (melanotropes) was nearly the same with that of β-endorphin-ir cells. In FW fish, intermediate cells, which were immunoreactive to α-MSH antiserum or β-endorphin antiserum, showed a similar increase in immunoreactivity (p<0.001), whereas their
cell sizes reduced (p<0.001), as shown in Figs. 10A-H for β-endorphin-ir cells. We considered that the present results on α-MSH-ir cells and β-endorphin-ir cells were obtained from pituitary cells of the same population, although we did not use a double immunostaining technique to confirm the presence of the two hormones in the same individual cells.

DISCUSSION

The present study analyzed GH/PRL/SL, GTHs (I and II)
and POMC-derived hormonal cells in the pituitaries of chum salmon during the final stages of spawning migration. We found that GH/PRL/SL family cells had stronger immunoreactivity with enlarged cell sizes in FW fish than in SW ones. In gonadotropes, we found that GTH I cells had stronger immunoreactivity with reduced cell sizes in FW animals than in SW ones, while GTH II cells had stronger immunoreactivity with enlarged cell sizes in FW animals when compared with SW ones. In POMC-derived hormonal cells, ACTH cells (corticotropes) had slightly higher immunoreactivity in FW animals than in SW ones.
ones, but cell sizes did not change. On the other hand, the melanotropes immunoreactive to α-MSH antiserum or β-endorphin antiserum had stronger immunoreactivity with reduced cell sizes in FW fish than in SW ones of both sexes.

The findings in GH/PRL/SL family cells in homing chum salmon coincide well with the previous reports that the levels of GH, PRL and SL mRNAs increased in FW chum salmon (Taniyama et al., 1999). Similarly, plasma levels of GH, PRL and SL elevated in FW chum salmon (Kakizawa et al., 1995). Since the level of mRNA indicates the transcriptional activity and plasma hormonal levels show secretory activity of pituitary cells, it is plausible that, in pre-spawning chum salmon, over-expressed hormones were stored coincidentally with the elevation of synthetic and secretory activity in pituitary cells that produce GH/PRL/SL family hormones.

Changes in morphology and activity of GH, PRL (Power, 1992) and SL (Oliverereau and Rand-Weaver, 1994b) cells were associated with the different life stages. In juvenile chum salmon, larger GH cells were weakly stained compared with smaller ones (Salam et al., 1999). Since GH cells which extensively secreted hormone in serum-free culture decreased in immunoreactivity with the dilation of rough endoplasmic reticulum (Yada et al., 1991), large but weakly immunoreactive cells should have high secretory activity. Thus, the weakly stained cell population seen in the present SW females may be the remnant of such active GH cells. The slight increase in immunoreactivity of GH cells in FW fish indicates that GH cells were not so stimulated in FW environment during spawning migration as reported in striped bass, Morone saxatilis (Huang and Specker, 1994).

Interestingly, remarkable changes were found in immunoreactivity and cell sizes of PRL and SL cells during spawning migration. In both PRL and SL cells, immunoreactivity was considerably stronger with enlarged cell sizes in FW fish than in SW ones of both sexes. The previous reports showed higher transcriptional activity of PRL and SL cells in FW chum salmon (Taniyama et al., 1999) and higher secretory activity of PRL and SL cells in homing salmon (Kakizawa et al., 1995). Further, in the present study the cellular amounts of hormones in PRL and SL cells were increased. We therefore consider that translational activity was much higher than secretory activity during spawning migration of chum salmon. The stimulation of PRL and SL cells might be influenced in relevance with some physiological functions. In teleosts, many studies suggested that PRL is involved in FW adaptation (Hirano, 1986), while SL has function at least for osmoregulation (Kakizawa et al., 1999) and final maturation (Rand-Weaver and Swanson, 1993; Oliverereau and Rand-Weaver, 1994b; Taniyama et al., 1999).

In gonadotropes, GTH I cells had stronger immunoreactivity with reduced cell sizes in FW fish compared with the SW ones of both males and females, while GTH II cells had stronger immunoreactivity with enlarged cell sizes in FW animals. During earlier stages of sexual maturity, GTH I cells had numerous dilated cisternae of the granular endoplasmic reticulum with a small number of secretory granules, suggesting higher secretory activity of GTH I cells in vitellogenic rainbow trout (Naito et al., 1993b). In addition, GTH I cells showed active synthesis and release of hormone at the early stages of gonadal maturation in salmonid (Nozaki et al., 1990; Naito et al., 1991). Large GTH I cells with weak immunoreactivity in SW animals presumably underwent somehow considerable secretion, while small GTH I cells having strong immunoreactivity in FW animals were regressed without further stimulation, as fish attained near to final maturation. This may be true, because during the final stages of spawning migration, no significant changes in the levels of GTH Iβ mRNA were seen between SW and FW chum salmon of the same stock used in the present study (Kitahashi et al., 1998).

In contrast to GTH I cells, GTH II cells were enlarged with stronger immunoreactivity in FW animals than in SW ones of both male and female chum salmon at the time of final maturation. This finding coincides well with the reports that GTH II cells were intensely stained in mature rainbow trout prior to spawning (Nozaki et al., 1990). In addition, the level of GTH Iβ mRNA was higher in FW fish than those in SW ones during the final stages of spawning migration from SW to FW in chum salmon (Kitahashi et al., 1998). The levels of GTH Iβ mRNA also elevated in rainbow trout during final maturation (Naito et al., 1991; Gomez et al., 1999). Further, plasma levels of GTH II markedly increased during the final stages of gonadal maturation in teleost species, such as in coho salmon (Swanson, 1991b), striped bass (Mylonas et al., 1997) and rainbow trout (Gomez et al., 1999). The plasma levels of 17α,20β-dihydroxy-4-pregnen-3-one were thus dramatically increased in FW chum salmon of both sexes of the same population that were used in the present study (Ota, 1999). Since the magnitude of immunoreaction determines the amount of stored hormone, GTH II cells in FW fish may contain much higher stored hormone than those in SW ones regardless of sexes at the final stage of maturation. We consider that the translational activity of GTH II cells was much higher than secretory activity during the final maturation in salmonids.

In POMC-derived hormone-producing cells, ACTH cells had slightly stronger immunoreactivity without changing the sizes of cells in FW animals, indicating that ACTH cells remain active to secrete hormone since rapid salinity changes can act as stressors in some teleost, such as Oreochromis mossambicus (Balm et al., 1994), and Sparus aurata (Mancera et al., 1994). Such possibility in pre-spawning chum salmon is supported by the fact that plasma cortisol levels were dramatically elevated in the same population of homing chum salmon used in the present study (Ota, 1999).

In melanotropes, the cells immunoreactive to α-MSH antiserum had stronger immunoreactivity with reduced cell sizes in males. This result may indicate that these cells were in rest after the extensive secretion of α-MSH to provide nuptial color, since strong nuptial coloration was seen in FW males. Similar evidence in rainbow trout (Suzuki et al., 1997) supports this idea. Since the changes in α-MSH-ir and β-endorphin-ir cells were nearly the same, it is possible that β-endorphin-ir cells underwent extensive secretion prior to spawning and became
inactivated at the final stage of salmon life cycle. Hane et al. (1966) reported that activity of the adrenal cortex diminished progressively by the time of spawning in pacific salmon (Oncorhynchus tshawytscha). Getting together, we consider that POMC-derived hormone-producing cells are activated to secrete hormones prior to or during spawning migration in salmonid.

ACKNOWLEDGEMENTS

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REFERENCES


Nagahama Y (1973) Histophysiologic studies on the pituitary gland of some teleost fishes, with special reference to the classification of hormone producing cells in the adenohypophysis. Mem Fac Fish Hokkaido Univ 21: 1–63


Oliverve M, Rand-Weaver M (1994b) Immunoreactive somatolactin cells in the pituitary of young, migrating, spawning and spent chinook salmon, Oncorhynchus tshawytscha. Fish Physiol
Biochem 13: 141–151
Ono M, Takayama Y, Rand-Weaver M, Sakata S, Yamasaka T, Nosot
Kawauchi H (1990) cDNA cloning of somatolactin, a pituitary
protein related to growth hormone and prolactin. Proc Natl Acad
Sci USA 87: 4330–4334
Ota Y (1999) Sexually dimorphic expression of neurohypophysial
hormone genes and steroid hormones in salmonid. PhD thesis,
Hokkaido Univ, Japan, pp 61–77
Power DM (1992) Immunocytochemical identification of growth hor-
mone, prolactin and gonadotropin cells in the pituitary of male
plaice (Pleuronectes platessa) during gonadal maturation. Gen
Comp Endocrinol 85: 358–366
Rand-Weaver M, Kawauchi H (1993) Growth hormone, prolactin and
somatolactin: a structural overview. In Biochemistry and Molecular
Biology of Fishes (Hochachka PW, Mommsen TP eds), Elsevier,
Amsterdam, pp 39–56
Rand-Weaver M, Swanson P (1993) Plasma somatolactin levels in
coho salmon (Oncorhynchus kisutch) during smoltification and
sexual maturation. Fish Physiol Biochem 11: 175–182
and characterization of somatolactin, a new protein related to
growth hormone and prolactin, from Atlantic cod (Gadus morhua)
pituitary glands. Biochemistry 30: 1509–1515
Somatolactin, a novel pituitary protein. Purification and plasma
levels during reproductive maturation of coho salmon. J
Endocrinol 133: 393–399
on the pituitary and plasma concentrations of some pro-opiomelanocortin-
related peptides in the rainbow trout (Salmo gairdneri). J Endocrinol
101: 277–284
Salam MA, Ota Y, Ando H, Fukuwaka M, Kaeriyama M, Urano A
(1999) High population density of juvenile chum salmon de-
creased the number and sizes of growth hormone cells in the
pituitary. Zool Sci 16: 945–954
ACTH, α-MSH and cortisol levels in salmonid fishes. Gen Comp
Endocrinol 62: 377–385
Sumpter JP, Scott AP (1989) Seasonal variations in plasma and pitu-
itary levels of gonadotropin in males and females of two strains
of rainbow trout (Salmo gairdneri). Gen Comp Endocrinol 75:
376–388
Suzuki K, Kawauchi H, Nagahama Y (1988) Isolation and character-
ization of two distinct gonadotropins from chum salmon pituitary
glands. Gen Comp Endocrinol 71: 292–301
and POMC genes in rainbow trout (Oncorhynchus mykiss) dur-
ing ontogeny and in response to early physiological challenges.
Gen Comp Endocrinol 107: 341–350
Swanson P (1991) Salmon gonadotropins: Reconciling old and new
ideas. In Proc 4th Intl Symp Reprod Physio Fish (Scott AP,
Sumpter JP, Kime DE, Rolf MS eds), Univ East Anglia Printing
Unit, Norwich, UK, pp 2–7
and characterization of two coho salmon gonadotropins, GTH I
and GTH II. Biol Reprod 44: 29–38
conservation of somatolactin, a presumed pituitary hormone of
the growth hormone/prolactin family, than growth hormone in
teleost fish. Gen Comp Endocrinol 83: 366–374
Changes in the levels of mRNAs for GH/PRL/SL family and pit-1/
GHF-1 in the pituitaries of pre-spawning chum salmon. J Mol
Endocrinol 23: 189–198
tissues: novel functions in the hippocampus and implications for
Yada T, Urano A, Hirano T (1991) Growth hormone and prolactin
gene expression and release in the pituitary of rainbow trout in
serum-free culture. Endocrinology 129: 1183–1192
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