



Title	Population structure, egg production and gut content pigment of large grazing copepods during the spring phytoplankton bloom in the Oyashio region
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1 **Population structure, egg production and gut content pigment of large grazing**  
2 **copepods during the spring phytoplankton bloom in the Oyashio region**

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11

12 Abstract

13 As a basis for analyzing development of six large grazing copepods (*Eucalanus bungii*,  
14 *Metridia pacifica*, *M. okhotensis*, *Neocalanus cristatus*, *N. flemingeri* and *N. plumchrus*)  
15 in the Oyashio region, quasi-daily twin-NORPAC net (0.33 and 0.10 mm mesh) hauls  
16 were taken through the upper 150 m and 500 m at a station southwest of Hokkaido  
17 before (9-14 March) and after (6-30 April) the onset of the phytoplankton bloom in  
18 2007. Based on additional fresh specimens collected from 0-150 m, egg production of  
19 *E. bungii*, *M. pacifica* and *M. okhotensis*, and gut pigments of late copepodid stages in  
20 each species were evaluated. Total zooplankton biomass was greater from 10 April  
21 onward by a factor of 2- to 8-fold the previous levels. This increase of the 0-150 m  
22 biomass was caused by development of *Neocalanus* spp. copepodids and upward  
23 migration of resting *E. bungii*. Egg production of *E. bungii* peaked on 18 April, while  
24 abundance of its nauplii and C1 peaked on 20 and 25 April, respectively. Sex ratio and  
25 C6-female gonad maturation index of *E. bungii* showed new recruitment to C6 during  
26 20-30 April, likely derived from a population that over-wintered as C3 or C4. Egg  
27 production and hatchability of *M. pacifica* and *M. okhotensis* were highly variable and  
28 no temporal trend was detected. Comparison with field abundance data for *Metridia*  
29 spp. suggests that our estimates of egg production and hatchability are too low, despite  
30 care with experimental conditions. All the *Neocalanus* species utilize the bloom as  
31 energy for juvenile growth. *Neocalanus cristatus* developed from C2 through C4, and  
32 stage duration of C3 was estimated to be 24 days. *Neocalanus flemingeri* also  
33 developed from C1 through C3, and stage durations of C1 and C2 were estimated to be  
34 7-9 days. *Neocalanus plumchrus* occurred in small numbers from mid-April onward.  
35 The stage duration estimates for *Neocalanus* spp. are similar to those reported from the

36 high-nutrition southeastern Bering Sea shelf. Gut pigment variation clearly showed  
37 nocturnal feeding by *Metridia* spp., while no diel changes in gut pigment were  
38 recognized for *E. bungii* or *Neocalanus* spp. The diel changes in gut pigment of  
39 *Metridia* spp. were related to their diel vertical migrations. The calendar of sequential  
40 responses of copepods to the phytoplankton bloom is summarized.

41 Keywords: Copepods, Egg production, Gut pigment, Life cycle, OECOS

42

43 Introduction

44 Life cycles of large- and medium-sized, grazing copepods (*Neocalanus cristatus*,  
45 *Neocalanus flemingeri*, *Neocalanus plumchrus*, *Eucalanus bungii* and *Metridia*  
46 *pacifica*) have been studied extensively at Station P in the eastern subarctic Pacific by  
47 U.S. scientists during the 1980s (Miller et al., 1984; Batchelder, 1985; Miller and  
48 Clemons, 1988).

49 In the western subarctic Pacific, Japanese scientists have pursued intensive  
50 studies on life cycles of dominant mesozooplankton in the Oyashio region since 1996.  
51 A time series of mesozooplankton data at Site H (areas covered between  
52 41°30'-42°30'N and 145°00'-146°00'E) in the Oyashio region was developed by  
53 sampling at monthly intervals over one full year by vertical hauls with closing nets  
54 through 5 discrete strata between the surface and 2000 m. Whenever possible, some  
55 short-term reference stations were set in the central and eastern subarctic Pacific, Bering  
56 Sea, Okhotsk Sea, and Japan Sea during the program, and the results were compared  
57 with those from Site H. As a result, information about life cycles (spawning season,  
58 development pattern, generation time, longevity, etc.) has accumulated rapidly on a  
59 number of copepod species in the Oyashio region: i.e., *N. cristatus*, *N. flemingeri*, *N.*  
60 *plumchrus* (Kobari and Ikeda, 1999, 2001a, 2001b), *E. bungii* (Shoden et al., 2005),  
61 *Metridia okhotensis*, *M. pacifica* (Padmavati et al., 2004), *Gaidius variabilis*,  
62 *Heterorhabdus tanneri*, *Paraeuchaeta elongata*, *Paraeuchaeta birostrata*, *Paraeuchaeta*  
63 *rubra*, *Pleuromamma scutullata* (Yamaguchi and Ikeda, 2000a, 2000b, 2001) and  
64 *Oncaea grossa*, *Oncaea parila*, *Triconia borealis*, *Triconia canadensis* (Nishibe and  
65 Ikeda, 2007). There are differences in copepod phenology between the Gulf of Alaska  
66 and the Oyashio: *E. bungii* had a 1-year life cycle at Site H in contrast to its 2-year life

67 cycle at Station P (Shoden et al., 2005), and *M. pacifica* completed two generations at  
68 Site H in contrast to three at Station P (Padmavati et al., 2004).

69 Low-frequency sampling (e.g. one sample per month), mentioned above, was  
70 sufficient to show the overall picture of mesozooplankton life cycles for species with  
71 generation lengths of one year or more. However, precise calculation of population  
72 production of these mesozooplankton requires high-frequency data on their  
73 developmental sequences. For example, all large to medium-sized copepods achieve  
74 rapid development from early to late copepodid stages during the phytoplankton bloom  
75 that typically lasts ca. 3 months (April-June). As seen in our low-frequency sampling,  
76 abundance peaks of 3 to 4 copepodid stages occurred simultaneously, that is, in the  
77 sample results for the same month, making it difficult to estimate development time  
78 of any stage. High-frequency sampling (1 sample per day) during OECOS was  
79 intended to solve that problem.

80 The OECOS project (Oceanic Ecodynamics Comparison in the Subarctic  
81 Pacific), sponsored by PICES, aims to evaluate detailed dynamics of physical, chemical  
82 and biological oceanography in the Alaskan Gyre and Oyashio region (Miller and Ikeda,  
83 2006). From the results of the OECOS study, we can characterize short-term changes  
84 in mesozooplankton biomass, population structure, egg production and gut pigment of  
85 large- to medium-sized grazing copepods during the spring phytoplankton bloom of  
86 2007 in the Oyashio region. We interpret the data in terms of the species-specific  
87 sequential responses of copepods to the phytoplankton bloom.

## 88 **Materials and methods**

### 89 *Field sampling*

90 During cruises of T/S *Oshoro-Maru* (March) and R/V *Hakuho-Maru* (April), 22 tows of  
91 twin-NORPAC nets (0.10 and 0.33 mm mesh, 45 cm diameter; Motoda, 1957) with  
92 flowmeters were made from 0-150 and 0-500 m at Station A-5 (42°00'N, 145°15'E)  
93 during 9-14 March and 6-30 April 2007 (Fig. 1a, b). Sampling was done in daytime  
94 (08:00-10:00 local time) and nighttime (19:00-21:00 local time), and the whole samples  
95 were immediately preserved in 5% borax-buffered formalin. To collect fresh, live  
96 specimens, 22 tows of a ring net (0.33 mm mesh, 80 cm diameter) with a large bottle for  
97 its cod end were made from 0-150 m during day and night on the same date. Using  
98 live specimens, temporal changes in egg production, gut pigment and individual mass  
99 were examined (methods below). CTD casts were also made three times per day.  
100 Water samples for chlorophyll-*a* were collected from 0, 5, 10, 20, 30, 40, 50, 75 and 100  
101 m, then filtered through Whatman GF/F filters, and measured fluorometrically after  
102 extraction with dimethyl-formamide (Suzuki and Ishimaru, 1990).

### 103 *Total zooplankton biomass*

104 In the land laboratory, zooplankton samples from the 0.33 mm mesh NORPAC net  
105 (0-150 m and 0-500 m) were split with a Motoda splitting device (Motoda, 1959), and a  
106 one-half aliquot was used for measurement of wet mass by microbalance (Mettler  
107 PM4000, precision 0.01 g).

### 108 *Population structure*

109 Copepodids C1-C6F/M of *E. bungii*, *M. pacifica*, *M. okhotensis*, *N. cristatus*, *N.*  
110 *flemingeri* and *N. plumchrus* were enumerated in the samples collected with 0-500 m  
111 NORPAC nets. Prior to analysis, we examined subsamples (1/10 – 1/100 of total

112 volume, made with a wide-bore pipette) of both 0.33 and 0.10 mm mesh net samples.  
113 Then we compared the abundance based on 0.10 and 0.33 mm mesh nets and  
114 summarized the ratio of abundance between them (Table 1). Ratios for most of the late  
115 copepodid stages were near 1 (no differences between the two nets), while some early  
116 copepodid stages showed substantially greater factors (0.10 mm > 0.33 mm mesh). For  
117 the stages showing a high factor (>5: nauplii of *E. bungii*, C1, C2 and C3 of *M. pacifica*,  
118 Table 1), we used the abundance data based on 0.10 mm mesh nets. For the other late  
119 copepodid stages, we used the abundance data based on 0.33 mm mesh nets.

120 For *E. bungii*, the naupliar stages were easily distinguished from the other  
121 species. Its naupliar stages were identified and enumerated based on the descriptions  
122 of Johnson (1937). Also for *E. bungii*, gonad maturation stages of C6F were  
123 enumerated (gonad maturation stages I-VII) based on the description by Miller et al.  
124 (1984). Since the gonad developmental stage V (two full rows of ova that appear  
125 ready for spawning) of Miller et al. (1984) could be separated into two (two full rows of  
126 ova and three or more rows of ova), we enumerated them separately as gonad  
127 developmental stages V-1 and V-2, respectively.

### 128 *Egg production and hatching*

129 Fresh adult females of *E. bungii*, *M. pacifica* and *M. okhotensis* collected with the ring  
130 net at night were used for on-board egg production experiments. Three liters of  
131 surface seawater were collected with a bucket, filtered with a GF/F filter and well  
132 aerated by shaking in 500 ml bottles. Fresh adult females were transferred into plastic  
133 chambers (ca. 300 ml) with 0.33-mm mesh bottoms (to prevent egg cannibalism by  
134 females) inside the 500 ml bottles. In each chamber, three (*E. bungii*) or ten (*M.*

135 *pacifica*) adult females were added. The bottles with adult females were held for one  
136 day in an incubator set at 3°C (integrated mean temperature for 0-100 m during April,  
137 Fig. 1c). After 24h, females in the chamber were removed. The remaining seawater  
138 was examined under a stereomicroscope to seek eggs. When found they were counted  
139 and transferred into 5-mL multi-well plates filled with chilled, filtered seawater. Eggs  
140 in the multi-well were also incubated at 3°C and checked daily for hatching. Egg  
141 incubation lasted ca. 1 week.

142 Egg production rates were expressed as eggs female<sup>-1</sup> day<sup>-1</sup> and, multiplying  
143 these values with female abundance (ind. m<sup>-3</sup>), as eggs m<sup>-3</sup> day<sup>-1</sup>. Two types of egg  
144 morphology were recognized: eggs with solid (normal) and eggs with thin (abnormal)  
145 outer membranes. We counted them separately, presenting the percentage of normal  
146 eggs in the total. Hatchability of eggs (%) was calculated for the normal eggs. The  
147 total recruitment of nauplii to the population (ind. m<sup>-3</sup> day<sup>-1</sup>) was calculated by  
148 multiplying egg production (eggs m<sup>-3</sup> day<sup>-1</sup>) by the proportion of normal eggs and  
149 proportion hatching.

#### 150 *Gut pigment*

151 Using fresh specimens collected both day and night, pigments contained in the guts of  
152 late copepodid stages of dominant copepods were examined. Portions of the samples  
153 collected with the ring net were poured into a 1-l pitcher, and 10% v/v soda (saturated  
154 CO<sub>2</sub> in water) was added to prevent gut evacuation and decomposition of pigment.  
155 Fresh samples were examined under a stereomicroscope and C6F of *E. bungii*, *M.*  
156 *pacifica* and *M. okhotensis* and C5 of *N. cristatus*, *N. flemingeri* and *N. plumchrus* were  
157 sorted out. Batches of two (*N. cristatus* C5) to five (*M. pacifica* C6F) specimens were

158 immersed in N,N-dimethylformamide, stored in dark, cold conditions overnight to  
159 extract chlorophyll and phaeopigments. After extraction of pigment, chlorophyll and  
160 phaeopigments were measured with a Turner Designs fluorometer. Chlorophyll and  
161 phaeopigment amounts were summed and expressed as ng pigment ind.<sup>-1</sup> (cf. Mackas  
162 and Bohrer, 1976).

### 163 *Individual mass*

164 To evaluate temporal changes in individual body composition, C6F of *M. pacifica* and  
165 C5 of *N. cristatus* were sorted from the daytime live samples. Specimens were  
166 incubated in filtered seawater for three hours to evacuate their gut contents, rinsed with  
167 distilled water, transferred into pre-weighed aluminum pans and stored in a freezer at  
168 -80°C. In the land laboratory, wet masses (WM) were determined ( $\pm 1 \mu\text{g}$ ) with a  
169 microbalance (Mettler Toledo MT5), then samples were freeze-dried and dry masses  
170 (DM) were determined. Dried specimens were incinerated at 480°C for 5 h, and ash  
171 weight was determined. From these data, water content (WATER, %WM) and ash-free  
172 dry mass (AFDM, %DM) were estimated using the following equations:

$$173 \quad \text{WATER} = 100(\text{WM} - \text{DM}) / \text{WM} \text{ and } \text{AFDM} = 100(\text{DM} - \text{Ash}) / \text{DM}.$$

## 174 **Results**

### 175 *Hydrography*

176 Through the study period, integrated mean temperature and salinity in the 0-100 m  
177 stratum varied between 1.5-6.0°C and 33.16-33.65, respectively (Fig. 1c).  
178 Temperature and salinity varied in parallel. They were high in March, then had two  
179 minima on 7 and 22 April. Standing stock of chlorophyll-*a* varied between 0.2 and 7.6

180  $\text{mg m}^{-3}$ . It was low in March, and then had two maxima on 7 and 22 April. This  
181 temporal change pattern in chlorophyll-*a* was inverse to those of temperature and  
182 salinity. Temporal changes of temperature and salinity were governed by the mixing  
183 ratio of two water masses: one was low-temperature and less saline Coastal Oyashio  
184 Water, which carried high chlorophyll-*a*. The other was high-temperature and more  
185 saline modified Kuroshio Water, which carried lower chlorophyll-*a*. Details of  
186 temporal changes in these two water masses during the study period are reported by  
187 Kono and Sato (this issue).

#### 188 *Zooplankton biomass*

189 Zooplankton wet mass was low during 8-14 March, increased by 10 April, and then  
190 remained high during 10-30 April (Fig. 2a, b). This increase in zooplankton mass from  
191 March to April was ca. 8-fold in the upper 150 m and ca. 2-fold for the upper 500 m.  
192 This can only happen if the stock of the 0-500 m layer mostly becomes concentrated in  
193 the 0-150 m layer, while the overall abundance only doubles. Day vs. night  
194 differences were only evident for 0-150 m in March, but were not observed for 0-500 m  
195 throughout the study period (Fig. 2a, b). Depth distributions of zooplankton mass  
196 changed drastically around 10 April. Before that most (means  $\pm 1\text{sd}$ :  $92 \pm 3\%$  for day  
197 and  $82 \pm 5\%$  for night) of the biomass was seen in the 150-500 m stratum. After 10  
198 April, zooplankton biomass in the shallower 0-150 m depth was greater than from  
199 150-500 m (mean  $\pm 1\text{sd}$ :  $59 \pm 10\%$  for day and  $65 \pm 17\%$  for night) (Fig. 2c, d).

#### 200 *Population structure*

201 *Eucalanus bungii* - In March 2007 the stock was composed of C3-C6 females and

202 C6males (C6F/M), but no nauplii or C1-C2 were collected (Fig. 3a, b). The dominant  
203 stage was C3 comprising 38-51% of the copepodid population. First copepodids (C1)  
204 were first observed on 15 April. Total copepodid abundance increased gradually after  
205 that and peaked on 25 April, when the C1 and C2 dominated (50% and 29% of the  
206 copepodids) (Fig. 3a).

207 Nauplii of *E. bungii* were observed after 6 April with only N1-N4 present  
208 during 6-9 April (Fig. 3b). Naupliar stage composition was stable after 12 April, and  
209 N4 was the dominant stage ( $31 \pm 2\%$ : mean  $\pm 1$ sd during 12-30 April). The peak of  
210 naupliar abundance ( $1440 \text{ ind. m}^{-3}$ ) was observed on 20 April, 5 days before the peak of  
211 copepodid abundance (Fig. 3b).

212 The sex ratio of *E. bungii* was near 1:1 for C4 (females were  $57 \pm 20\%$  of the  
213 population) and for C5 ( $56 \pm 19\%$ ) (Fig. 3c). The average ratio was skewed to females  
214 in C6 ( $79 \pm 17\%$ ). Sex ratio of C6 *E. bungii* showed short-term changes: some males  
215 (ca. 25% of adults) occurred during March-12 April, while only females were collected  
216 during 15-20 April. Then males occurred again, about 25% of adults, during 24-30  
217 April (Fig. 3c).

218 Gonads of C6F *E. bungii* were mostly in immature stages I-II during March  
219 (Fig. 3d). The dominance of stage I decreased to 61% of C6F by 14 March. On 6  
220 April, the proportion of spawning females (stage V-1 and V-2) was small, but their  
221 frequency then increased within one or two days. Actively spawning females were a  
222 majority ( $30 \pm 6\%$  V-1 and  $34 \pm 13\%$  V-2) during 8-18 April, and then after 20 April the  
223 proportion of stages I-IV increased again ( $57-72\%$  of C6F) (Fig. 3d).

224 *Metridia* spp. – The population structure of *M. pacifica* was dominated by C6 on 9-10

225 March, after which dominance of C6 declined and abundance of C1 increased (Fig. 4a).  
226 Abundance of *M. pacifica* on 6-7 April was extremely low (9-16 ind. m<sup>-3</sup>). After that  
227 abundance of *M. pacifica* was stable, varying between 51 and 91 ind. m<sup>-3</sup> (mean ±1sd:  
228 72 ±14 ind. m<sup>-3</sup>), and their population structure was also stable through April: C1  
229 dominated (49 ±11%) followed by C2 (14 ±7%), C3 (16 ±6%), C4 (6 ±3%), C5 (6 ±3%)  
230 and C6 (9 ±4%) (Fig. 4a).

231 Abundance of *M. okhotensis* (3-30 ind. m<sup>-3</sup>) was lower than that of *M. pacifica*  
232 (Fig. 4b). After 20 April, their abundance peaked and was slightly greater than before.  
233 Its population structure was dominated by C5 (34 ±19%), followed by C6 (24 ±13%)  
234 and C4 (21 ±10%); thus these late copepodid stages composed 79% of the population  
235 and showed no strong temporal variation (Fig. 4b).

236 *Neocalanus* spp. - Abundance of *N. cristatus* was greater in March than in April (Fig.  
237 5a). Their abundance can be expressed by a regression:  $Y = 7.9 - 0.065X$ , where  $Y$  is  
238 abundance (ind. m<sup>-3</sup>) and  $X$  is julian day starting from 1 March ( $r^2 = 0.25$ ,  $p < 0.05$ ).  
239 Population structure of *N. cristatus* was dominated by C1-C3 in March, by C3 on 6-7  
240 April and by C4 and C5 on 20-30 April. Thus, dominant stages shifted from early  
241 copepodid stages to late copepodid stages.

242 Abundance of *N. flemingeri* was low in March and increased through April (Fig.  
243 5b). Population structure of *N. flemingeri* passed through successive stages: a peak of  
244 C1 was observed on 8 April, of C2 on 18 April and of C3 on 25 April. The proportion  
245 of C5 and C6 remained extremely low for *N. flemingeri* through the end of April (Fig.  
246 5b).

247 Abundance of *N. plumchrus* was extremely low in March (Fig. 5c). It was

248 present in April, but its abundance was consistently lower than that of *N. flemingeri*.  
249 The population structure of *N. plumchrus* during 15-30 April was dominated by C1 (30  
250  $\pm 11\%$ ), C2 (33  $\pm 12\%$ ) and C3 (17  $\pm 7\%$ ) (Fig. 5c).

### 251 *Egg production*

252 Egg production of *E. bungii* was zero in March and remained low (<50 eggs female<sup>-1</sup>  
253 day<sup>-1</sup>) during 6-9 April (Fig. 6a). Abnormal eggs with a weak membrane structure  
254 were also observed. Both normal and abnormal egg production rates increased during  
255 6-12 April; after that production of abnormal eggs was rare (Fig. 6a). In early April,  
256 both the proportion of normal eggs and the hatchability of normal eggs were low.  
257 Those increased during 6-16 April, after which normal eggs were 93  $\pm 6\%$  of egg  
258 production and hatchability was 83  $\pm 5\%$  of normal eggs (Fig. 6a). Recruitment of  
259 nauplii female<sup>-1</sup> d<sup>-1</sup> was low during 6-9 April, and then much higher during 11-30 April  
260 (mean  $\pm 1$ sd: 88  $\pm 22$  ind. female<sup>-1</sup> d<sup>-1</sup>). Total recruitment of nauplii to the water  
261 column *E. bungii* population was also greater during 11-30 April (206  $\pm 124$  ind. m<sup>-3</sup>  
262 day<sup>-1</sup>), and peaked on 18 April (447 ind. m<sup>-3</sup> day<sup>-1</sup>) (Fig. 6a).

263 Egg production of *M. pacifica* fluctuated greatly through the study period (Fig.  
264 6b). It also produced abnormal eggs with weak membranes, more of them (76%) than  
265 normal eggs (24% of total egg production). Hatchability of normal eggs also  
266 fluctuated around a mean of 67%. Temporal variability of percentage of normal eggs  
267 and hatchability of normal eggs were correlated (Fig. 6b). Individual recruitment  
268 peaked on 30 April with 18 ind. female<sup>-1</sup> day<sup>-1</sup>, and population recruitment also peaked  
269 then (37 ind. m<sup>-3</sup> day<sup>-1</sup>).

270 Egg production of *M. okhotensis* was observed from 17 April onward (Fig. 6c).

271 Their percentage of normal eggs was  $88 \pm 13\%$ , but hatchability fluctuated. Their  
272 individual recruitment peaked on 20 April at  $7.3 \text{ ind. female}^{-1} \text{ day}^{-1}$ , and population  
273 recruitment peaked in same period at  $22 \text{ ind. m}^{-3} \text{ day}^{-1}$  (Fig. 6c).

#### 274 *Gut pigment*

275 Diel changes in gut content pigment of late copepodid stages separated into two types:  
276 (1) *E. bungii* C6F, *N. cristatus* C5 and *N. flemingeri* C5 showed low gut pigment in  
277 March and no day-night differences in gut pigment during April (except for *N. cristatus*  
278 C5 during 6-10 April) (Fig. 7a, d, e). (2) *M. pacifica* C6F and *M. okhotensis* C6F  
279 showed significantly higher gut pigment at night than in daylight ( $p < 0.01$ , U-test) and  
280 had similar gut pigment levels at night in March and April (Fig. 7b, c). Mean gut  
281 pigment varied with species, highest for *N. cristatus* C5 ( $133\text{-}197 \text{ ng Chl. ind.}^{-1}$ ) and  
282 lowest for *M. pacifica* C6F ( $2.3\text{-}6.2 \text{ ng Chl. ind.}^{-1}$ ), a function of their body sizes (Table  
283 2).

#### 284 *Individual mass and body composition*

285 The WM, DM and ash of *M. pacifica* C6F were  $0.94 \pm 0.17$ ,  $0.16 \pm 0.03$  and  $0.015$   
286  $\pm 0.002 \text{ mg ind.}^{-1}$ , respectively (means  $\pm 1\text{sd}$ ) (Fig. 8a). Its water and AFDM contents  
287 were  $82.1 \pm 3.2\%$  of WM and  $90.3 \pm 2.0\%$  of DM, respectively (Fig. 8b). Regressions  
288 of compositional components vs. date showed that water and AFDM of *M. pacifica*  
289 were uncorrelated with date (Table 3).

290 The WM, DM and ash of *N. cristatus* C5 were  $20.0 \pm 2.3$ ,  $3.99 \pm 1.89$  and  
291  $0.30 \pm 0.04 \text{ mg ind.}^{-1}$ , respectively (mean  $\pm 1\text{sd}$ ) (Fig. 8c). Its Water and AFDM contents  
292 averaged  $81.5 \pm 7.1\%$  of WM and  $90.1 \pm 4.9\%$  of DM, respectively (Fig. 8d).

293 Regressions vs. date showed that AFDM content of *N. cristatus* C5 was significantly  
294 increasing with date, with water content decreasing with date (Table 3).

## 295 **Discussion**

### 296 **Zooplankton biomass**

297 Total zooplankton biomass in the upper 500 m showed little diel variability, while that  
298 in the 0-150 m layer was greater at night during March 2007 (Fig. 2a, b). The  
299 magnitude of this diel change in biomass was a factor of ca. 2.5 (N>D), which  
300 corresponds to the values Steinberg et al. (2008) reported from Station K2 in the  
301 western subarctic Pacific. According to Goldblatt et al. (1999), diel changes in  
302 zooplankton biomass were present at Station P in summer, while not present during  
303 winter-spring. Thus, the presence (or not) of diel changes in zooplankton biomass in  
304 the upper waters seems to vary with season and location. No diel changes of total  
305 zooplankton biomass above 500 m implies that the magnitude of DVM of  
306 mesozooplankton in this region is less than 500 m, which corresponds to the results of  
307 Yamaguchi et al. (2004).

308 Comparing March to April, we observed several temporal changes in total  
309 zooplankton biomass: (i) total zooplankton stocks increased after 10 April by a factor of  
310 8 for 0-150 m and 2-fold for 0-500 m, (ii) diel changes in zooplankton biomass ceased  
311 after 10 April (Fig. 2a), and (iii) more than half of the zooplankton biomass was in the  
312 upper 150 m after 10 April (Fig. 2c, d). All of these might be caused by two factors:  
313 surface stocks of *Neocalanus* species developed into larger, late copepodid stages and  
314 upward migration of resting stocks of *E. bungii* and *M. okhotensis* occurred then  
315 (Yamaguchi et al., this issue). Recently, Ikeda et al. (2008) summarized seasonal

316 changes in biomass of large and medium-sized copepods in two depth strata (0-250 and  
317 250-2000 m) at Site H in the Oyashio region. In the 250-2000 m layer, peak biomass  
318 of these copepods occurs in early September, decreasing gradually until April of the  
319 next year, and then increases again until August-October (cf. Fig. 3 of Ikeda et al., 2008).  
320 Thus, the period of the present study (late March to early May) corresponds to the  
321 season when most of the dominant copepods are concentrated in the near-surface layer,  
322 and biomass in the deep layer is the lowest in the year. The new results confirm the  
323 spring portion of that pattern (Fig. 2c, d).

#### 324 **Population structure**

##### 325 *Eucalanus bungii*

326 The life cycle of *E. bungii* in the Oyashio region was studied by Tsuda et al. (2004) and  
327 Shoden et al. (2005). According to Tsuda et al. (2004), life cycle timing of *E. bungii* is  
328 two months ahead of the annual schedule in the Alaskan Gyre, and this earlier timing is  
329 likely related to the timing and magnitude of primary production. Overwintering  
330 stages in the Oyashio are C3-C6F, and the resting C5 molt to C6 males and females in  
331 February and March, respectively (Shoden et al., 2005). Dominance of C3-C6 and no  
332 occurrence of nauplii or C1-C2 during March in this study (Fig. 3a) correspond well to  
333 the previously reported life cycle schema. Sex ratio was nearly equal (F:M= 1:1) for  
334 C4 and C5 and ca. 3:1 for C6 during March to 10 April (Fig. 3c). This skewed sex  
335 ratio in C6 may reflect the shorter longevity of C6M, since they remain at 250-500 m  
336 without feeding (Miller et al., 1984; Shoden et al., 2005). Mating perhaps occurs at  
337 depth, slightly ahead of the phytoplankton bloom, and only mated C6F ascend to the  
338 surface layer in April for spawning (Shoden et al., 2005). Gonad maturation of C6F

339 was a dominant process in March with immature ovarian stages I and II present.  
340 Ovaries matured rapidly after the phytoplankton bloom started around 7 April (Fig. 3d).  
341 A similarly rapid response to phytoplankton abundance has also been reported for  
342 *Eucalanus californicus*, which can be dormant as both adult females and C5s, with  
343 winter females responding relatively rapidly to elevated food and temperature  
344 conditions; they begin feeding and producing eggs within 2-3 days (Ohman et al., 1998).  
345 During 10-20 April, gonad maturation of C6F had advanced and most ovaries were in  
346 reproductive stages V-1 and V-2 (Fig. 3d), which corresponds to the previously reported  
347 reproductive timing of this species (Tsuda et al., 2004; Shoden et al., 2005).

348 Occurrence of *E. bungii* nauplii was observed from 7 April onward, although  
349 there was no clear developmental sequence detected for naupliar stage composition (Fig.  
350 3b). This indicates that continuous recruitment of nauplii occurred through April.  
351 The peak abundance of nauplii occurred on 20 April (Fig. 3b), and that of C1 was on 25  
352 April (Fig. 3a). At Station P in the Alaskan Gyre, nauplii peaked sharply in abundance  
353 on 19 July, C1 and C2 peaked on 1 August, and all had advanced to the third copepodid  
354 stage by September (Miller et al., 1984). Thus the development period in the Oyashio  
355 region as observed in this study was much earlier than that in the Alaskan Gyre.

356 The remarkable point of this study is that newly recruited C6 of *E. bungii* were  
357 observed during 20-30 April (Fig. 3c, d). C6M were expected to have short longevity,  
358 since they do not feed in the current year. The energy of C6M is accumulated in the  
359 previous year, and is then utilized to maintain them as a resting population of C5M in  
360 deep layers. They molt to C6M without feeding, produce spermatophores and mate in  
361 their second spring. When C5F molt to C6F, they migrate upward and feed on  
362 phytoplankton, obtaining energy for reproduction and maintenance of their population.

363 Thus the complete absence of C6M during 16-20 April in this study (Fig. 3c) might  
364 have been caused by their death after a period of mating activity.

365 Then, C6M of *E. bungii* were observed again during 24-30 April, and the sex  
366 ratio was ca. 3:1 (F:M), similar to those in March-early April (Fig. 3c). In terms of  
367 C6F gonad maturation, dominance of reproductive stages V-1 and V-2 decreased  
368 suddenly on 20 April, and the proportion of immature gonad stages I-IV increased  
369 during 20-30 April (Fig. 3d). Dominance of immature ovaries in this period is  
370 anomalous, because a massive phytoplankton bloom was occurring in the epipelagic  
371 layer at that time. This phenomenon is best explained as new recruitment to the C6F  
372 population in this period, as well as that of C6M. Since the first C6F population is  
373 composed by the population which overwinters as C5F (molting to C6F in March,  
374 Shoden et al., 2005), these newly recruited C6F are considered to have over-wintered as  
375 C3 and C4F. The over-wintering C3 and C4 likely migrated into shallower layers in  
376 early April (Yamaguchi et al., this issue), developed through C4-C5 during early-mid  
377 April, and then reached C6F during 20-30 April (Fig. 3c, d).

378 A similar two-phase reproductive mode of *E. bungii* is also reported for the  
379 populations in the Alaskan Gyre. According to Miller et al. (1984), *E. bungii*  
380 reproduced in the mixed layer in early May and in early July. The first event was a  
381 spawning by females that had previously spawned and then had returned to diapause.  
382 The second, heavier spawning (more females, more eggs per female) was by newly  
383 matured females from stocks that had over-wintered as C5. The flexibility of life cycle  
384 is considered to be a special characteristic of eucalanid copepods. Several authors  
385 have characterized such patterns as “bet-hedging” tactics: the young from a given  
386 mother having more than one chance to encounter sustaining conditions. Apparently,

387 *E. bungii* in the subarctic Pacific is an “event-driven” species, capable of responding on  
388 a short-term basis to favorable environmental circumstances. Similar flexibility in  
389 life-cycle timing has been observed for *E. californicus* in the coastal NE Pacific (Ohman  
390 et al., 1998) and for *Rhincalanus gigas* in the Southern Ocean (Ward et al., 1997), which  
391 varies in generation length from 1 year to 2 years.

### 392 *Metridia pacifica*

393 Life cycles of *M. pacifica* and *M. okhotensis* in the Oyashio region were studied by  
394 Padmavati et al. (2004). Although recruitment of *M. pacifica* occurs throughout the  
395 year in the top 250 m of the water column, two pronounced generations have been  
396 observed. The first is characterized by rapid development during the spring  
397 phytoplankton bloom (generation length: 2-3 months), and the second by slow  
398 development (9-10 months) with over-wintering at C5 in deeper layers (up to  
399 1000-2000 m) (Padmavati et al., 2004). Stage structure of *M. pacifica* in this study  
400 was dominated by C6 in March and by early copepodid stages in April (Fig. 4a), which  
401 corresponds well to previous life cycle studies in the Oyashio region (cf. Fig. 10a of  
402 Padmavati et al., 2004).

403         During March, pigment was present in the guts of *M. pacifica* (Fig. 7b), and  
404 there were both egg production (Fig. 6b) and active diel vertical migration (Yamaguchi  
405 et al., this issue). All of these indicate that *M. pacifica* was not in a resting phase.  
406 Since the rest period of *M. pacifica* is variable with location (cf. Hirakawa and Imamura,  
407 1993), perhaps it should be termed quiescence not diapause. In the Oyashio region,  
408 the resting stages are predominantly C5M or C6F with immature ovaries (Padmavati et  
409 al., 2004). This resting stage structure implies that this species is ready to start

410 reproduction once food is supplied. Much earlier occurrence of *M. pacifica* C1 (Fig.  
411 4a) than of *E. bungii* (Fig. 3a) reflects this stage structure of *M. pacifica*.

#### 412 *Metridia okhotensis*

413 In contrast to *M. pacifica*, the life cycle of *M. okhotensis* is estimated to be that C1  
414 produced during one year over-winter and develop to C5 through the phytoplankton  
415 bloom of the next year. Those C5 over-winter, molt to C6 and reproduce during the  
416 phytoplankton bloom of the following year; thus, the generation length of *M. okhotensis*  
417 is estimated as 2 years (Padmavati et al., 2004). The over-wintering C5 emerge from  
418 diapause and start diel vertical migration in early April (Yamaguchi et al., this issue).  
419 They feed, molt to C6F and then reproduce. Because of the time required for this final  
420 development, the start of their reproduction, as shown by our data, was later than that of  
421 *M. pacifica* (Fig. 6b, c).

#### 422 *Neocalanus cristatus*

423 The life cycle of *N. cristatus* is reported to be similar for its entire distribution range in  
424 the subarctic Pacific (Tsuda et al., 2004). Reproduction occurs below 500 m during  
425 October to December, with a resultant peak of C1 in the near-surface layer during  
426 January to February. Populations develop through C5 by about June (Miller et al.,  
427 1984; Kobari and Ikeda, 1999; Tsuda et al., 2004). Development from C2 to C4 during  
428 March to April corresponds well to the life cycle results from the Oyashio region  
429 (Kobari and Ikeda, 1999; Tsuda et al., 2004). Since our OECOS study period  
430 (March-April) was at the annual minimum of resting stock below 250 m (cf. Fig. 3 of  
431 Ikeda et al., 2008), population analysis from 0-500 m might be useful to estimate

432 developmental time of *Neocalanus* spp. in the epipelagic layer.

433           Following a procedure like the cumulative stage composition analyses of  
434 Miller and Nielsen (1988) and Miller (1993), stage developmental times were analyzed  
435 for *Neocalanus* spp. (Fig. 9). For *N. cristatus*, significant regressions were observed  
436 for shifting proportions of the stock between C1-C2, C2-C3, C3-C4 and C4-C5 ( $p < 0.05$ ,  
437 Fig. 9a). Assuming the interval between half of the population entering one stage and  
438 half entering the next is the stage development time, we obtained *N. cristatus* data for  
439 C2 (-17 March), C3 (17 March-11 April) and C4 (11 April-) (Fig. 9a). Thus we  
440 estimate the duration of *N. cristatus* C3 is 24 days. This roughly corresponds to the  
441 stage duration reported from the southeastern Bering Sea shelf (20 days) (Vidal and  
442 Smith, 1986).

443           Since most of the population of *N. cristatus* developed from C2 to C4 during  
444 the study period, their abundance regression ( $Y = 7.9 - 0.065X$ , where  $Y$  is abundance  
445 [ $\text{ind. m}^{-3}$ ] and  $X$  is julian day starting from 1 March, cf. Fig. 3a) may reflect mortality.  
446 If we assume that the regression is the effect of mortality, the mortality rate of *N.*  
447 *cristatus* is calculated as  $0.82\% \text{ day}^{-1}$  ( $=0.065/7.9$ ). This value corresponds to the  
448 lower mortality rate of pre-diapause copepodid stages reported in the upper layers in the  
449 eastern subarctic Pacific ( $0.75\text{-}2.0\% \text{ day}^{-1}$ ; Miller, 1993; Mackas et al., 1998).

#### 450 *Neocalanus flemingeri*

451 The life history of *N. flemingeri* varies with location, particularly in respect to the  
452 copepodid stage during diapause in deep layers. Rest as fertilized C6F with an  
453 immature ovary was originally reported for the population at Station P in the Alaskan  
454 Gyre (Miller and Clemons, 1988), while rest of C4 and of C6F with immature ovaries

455 are reported for the population in the Japan Sea (Miller and Terazaki, 1989). Both  
456 resting stages (C4 and C6F with immature ovary) are confirmed to be present also in the  
457 Oyashio region (Tsuda et al., 1999; Kobari and Ikeda, 2001a). The body sizes of *N.*  
458 *flemingeri* C4 and C5 in the Oyashio region are reported to be bimodal, which has been  
459 suggested to result from differences in generation length (Tsuda et al., 1999) or from  
460 sexual dimorphism (Kobari and Ikeda, 2001a).

461         Throughout the study period, all the copepodid stages of *N. flemingeri* were  
462 observed, with an abundance peak of the total on 25 April (Fig. 5b). The proportional  
463 composition of the copepodid stages showed a clear sequence with time: C1 peaked on  
464 9 April, C2 on 18 April and C3 on 25 April (Fig. 9b). If we assume that the intervals  
465 between peaks were stage development times, stage durations of C1 and C2 were ~9  
466 days (9 April-18 April) and ~7 days (18 April-25 April), respectively. There are no  
467 other field developmental data for *N. flemingeri* with which to compare these results.  
468 We can, however, compare our results to published values for the similarly sized sibling  
469 species, *N. plumchrus*. At Station P in the Alaskan Gyre, stage duration of its C3 and  
470 C4 during May 1984 have been reported to be 24.0 and 24.8 days, respectively (Miller  
471 and Nielsen, 1988), while that of C3 during May 1988 was closer to 13.4 days (Miller,  
472 1993). For more nutrition-rich regions, stage durations of each copepodid stage of *N.*  
473 *plumchrus* have been reported to be 8-16 days in the southeastern Bering Sea shelf  
474 (Vidal and Smith, 1986) and 12.4-14.1 days in the northern Gulf of Alaska (Liu and  
475 Hopcroft, 2006). The stage duration we observed, 7-9 days, corresponded to that in  
476 the southeastern Bering Sea. Since the chlorophyll-*a* observed during this study  
477 (maximum 34  $\mu\text{g l}^{-1}$ ) was similar to that in the southeastern Bering Sea, copepodid  
478 development of *Neocalanus* spp. with replete nutrition appears to require just over a

479 week in each stage. According to Dagg (1991), development of *N. plumchrus* from C1  
480 to C5 requires 91 days in the Alaskan Gyre vs. only 46 days in the southeastern Bering  
481 Sea. In addition, the average body size of a C5 *N. plumchrus* from the Bering Sea is  
482 more than twice that of an individual from Ocean Station P (Dagg, 1991). Thus the  
483 developmental rate of *Neocalanus* spp. varies with location, apparently governed by  
484 available nutrition.

#### 485 *Neocalanus plumchrus*

486 The life cycle of *N. plumchrus* is known to vary in reproductive timing in different  
487 subarctic areas of the Pacific (cf. Miller and Clemons, 1988), while the season of  
488 development in the surface layer is similar among areas (Miller and Clemons, 1988;  
489 Tsuda et al., 1999; Kobari and Ikeda, 2001b). Throughout the region, surface  
490 occurrence of *N. plumchrus* is consistently much later than that of *N. flemingeri* (cf.  
491 Tsuda et al., 1999). Developmental periods of the two species (winter-spring for *N.*  
492 *flemingeri* and spring-summer for *N. plumchrus*) imply species-differences in lipid  
493 accumulation pattern; it starts in early copepodid stages of *N. flemingeri*, but only  
494 occurs in late copepodid stages of *N. plumchrus* (Tsuda et al., 2001). This temporal  
495 separation between the two species was again observed in the present study. In March,  
496 *N. plumchrus* was extremely low in abundance from 0-500 m the water column, with  
497 small numbers occurring from 7 April onward (Fig. 5c). In the Oyashio region,  
498 abundance of *N. plumchrus* is reported to increase during May-June (Tsuda et al., 1999;  
499 Kobari and Ikeda, 2001b). Assuming this seasonality was repeated in 2007, the very  
500 small surface stocks of *N. plumchrus* in this study were expected.

501 **Egg production**

502 *Eucalanus bungii*

503 Egg production rate of *E. bungii* was in good agreement with their field population; that  
504 is, the general levels of copepodid and naupliar stage abundance (Fig. 3a, b) were  
505 similar to the water column rate of egg production (Fig. 6a). Egg production rate of *E.*  
506 *bungii* was reported as  $129 \pm 26$  eggs female<sup>-1</sup> day<sup>-1</sup> (range: 88-170) during spring in the  
507 southeastern Bering Sea (Vidal and Smith, 1986). That corresponds to our present  
508 estimates for the Oyashio (Fig. 6a). The abnormal eggs of *E. bungii* were  
509 characterized by a weak outer membrane, no cell division and finally bursting into oil  
510 droplets. Abnormal eggs were observed mainly during the early phase of egg  
511 production (before 10 April) and were extremely rare from 15 April onward (Fig. 6a).  
512 We suggest that they are unfertilized eggs. The fractions of normal eggs and of  
513 successful hatching were low at the initiation of egg production (6 April), then rapidly  
514 improved from 15 April onward (Fig. 6a). This timing corresponds well to the  
515 proportional data for C6F gonad maturity stages (Fig. 3d).

516 The recruitment rate to the field *E. bungii* population (ind. m<sup>-3</sup> day<sup>-1</sup>: egg  
517 production x proportion of normal egg x hatchability x C6F abundance in the field)  
518 peaked on 18 April (bottom panel of Fig. 6a). Naupliar abundance in the field peaked  
519 on 20 April (Fig. 3b). Thus, timing of egg production corresponded well to the field  
520 abundance data for newly hatched young. Considering these facts, negative effects on  
521 copepod hatching success of the diatoms abundant during the spring bloom (the  
522 ‘paradox’ of diatom-copepod interaction, Ianora et al., 2003) could not have been  
523 particularly severe for *E. bungii* at A5 during our study. The fractions of abnormal  
524 eggs and the low hatching success of *E. bungii* improved through April during the

525 massive diatom bloom (Fig. 6a). This also suggests that the initial egg abnormality  
526 and low hatchability were caused by spawning of immature ova or fertilization failures.

### 527 *Metridia pacifica*

528 Concerning egg production of *Metridia* spp., a discrepancy between high abundance  
529 and low egg production and hatchability rates has been documented (Halsband-Lenk,  
530 2005; Hopcroft et al., 2005; Plourde and Joly, 2008). Suggested possible causes of the  
531 low egg production and hatchability of *Metridia* spp., at least in experiments, include  
532 egg cannibalism by C6F, insufficient size of incubation containers and damage to eggs  
533 when they sink through the egg separator mesh (Plourde and Joly, 2008). Egg  
534 production, fraction of normal eggs and hatchability of *M. pacifica* were highly variable  
535 in our experiments, and no constant trend was detected along the time course of the  
536 study (Fig. 6b). The peak of normal egg production in this study (30.1 eggs female<sup>-1</sup>  
537 day<sup>-1</sup>) corresponds well to that observed by Hopcroft et al. (2005) (30 eggs female<sup>-1</sup>  
538 day<sup>-1</sup>), suggesting that the incubation conditions of this study were not particularly  
539 deficient. The results of Hopcroft et al.'s (2005) were obtained with a specially  
540 designed incubation chamber (spawning tower).

541         Given the magnitude of population recruitment (bottom panel of Fig. 6b),  
542 based on the field abundance of copepodid stages (Fig. 4a), the experimental egg  
543 production and hatchability estimates in this study are not great enough. Reproduction  
544 must be more successful in the ocean. In the present study, C6F incubations were done  
545 with plastic chambers (volume ca. 300 ml) with an attached 0.33 mm mesh on the  
546 bottom. Ten C6F were incubated for 24 hours in well-oxidized, filtered seawater at  
547 3°C in the dark. Thus the volume of the container per individual is 30 ml (=300/10).

548 This volume was smaller than the 45 ml used by Plourde and Joly (2008), but larger  
549 than the 15 ml of Halsband-Lenk (2005) and Hopcroft et al. (2005). The container  
550 volume undoubtedly represents an additional factor explaining the large difference  
551 observed in clutch size and egg production rate between treatments with and without  
552 egg separation in Halsband-Lenk (2005) and Hopcroft et al. (2005). Plourde and Joly  
553 (2008) also suggest that freshly laid eggs could be damaged by sinking through the egg  
554 separator mesh, causing abnormal development and death. Thus, the strongly  
555 fluctuating egg production and hatchability data of *M. pacifica* in the present study are  
556 of questionable value. As an alternative explanation, since *M. pacifica* has several  
557 cohorts in one sampling period, there could be great differences in feeding history,  
558 gonad maturation and reproductive condition. These individual differences in  
559 reproductive condition may mask a clear synchronization of reproduction like that seen  
560 in *E. bungii* mentioned above.

#### 561 *Metridia okhotensis*

562 Reproduction of *M. okhotensis* is thought to be carried out by adults that over-wintered  
563 as C5, and then molted to C6 at the onset of the spring phytoplankton bloom (Padmavati  
564 et al., 2004). Initiation of reproduction by *M. okhotensis* in 2007 (beginning 17 April,  
565 Fig. 6c) was later than that of *M. pacifica*. Diel vertical migration behavior of *M.*  
566 *okhotensis* is only observed in spring (Padmavati et al., 2004; Takahashi et al., 2008),  
567 and reproduction of this species is limited to spring only in the northern Gulf of Alaska  
568 (Hopcroft et al., 2005). This information suggests that *M. okhotensis* spends most of  
569 the year resting as C5, and previously resting C5 initiate DVM in spring (Takahashi et  
570 al., 2008; Yamaguchi et al., this issue). Feeding and then molting to C6 in spring,

571 followed by reproduction is the pattern confirmed by the OECOS data set.

## 572 **Gut pigment**

573 Diel changes in gut pigment (greater at night) were only detected in the present study  
574 for *M. pacifica* and *M. okhotensis* (Table 2). Individuals of these relatively small  
575 copepods contain more phytoplankton pigment at night in March, than individuals of  
576 the much larger *Neocalanus* spp. or *E. bungii* (Fig. 7b). The diel cycling indicates that  
577 *M. pacifica* undertook a nocturnal ascent to feed, while the large *N. cristatus* and *E.*  
578 *bungii* did not show clear DVM (Yamaguchi et al., this issue) or strong diel feeding  
579 cycles (but see below for *N. cristatus*). Gut pigment in *Neocalanus* spp. is known to  
580 be determined by the availability of phytoplankton (cf. Dagg et al., 2006). For  
581 *Neocalanus* spp. and *E. bungii*, the lower gut pigment values in March than that in April  
582 in this study (Fig. 7a, d, e) are considered to have been mainly governed by the temporal  
583 changes in environmental chlorophyll *a* (greater in April, Fig. 1c). In contrast,  
584 nighttime gut pigment of *M. pacifica* did not vary between March and April (Fig. 7b).  
585 Since March was pre-bloom and April was during the massive bloom, similar gut  
586 pigment contents of *M. pacifica* between March and April are surprising. Their active  
587 DVM and swimming behavior are keys to the mechanism by which they apparently fed  
588 at the same rates in March and April, despite large differences in available  
589 phytoplankton. Bearing this in mind, in April when the food conditions were sufficient,  
590 *M. pacifica* may have met its food quota with less activity than was required in March.  
591 This could have enabled the cessation of DVM behavior of both *M. pacifica* and *M.*  
592 *okhotensis* (except C6F) in late April (Yamaguchi et al., this issue). Terminating DVM  
593 under conditions of sufficient food was reported for summer populations of *Metridia*

594 spp. in the Oyashio region by Hattori (1989).

595           Interestingly, diel changes in gut pigment were observed for *N. cristatus* before  
596 10 April, while pigment was similar between day and night after 12 April (Fig. 7d).  
597 Since large *N. cristatus* C5 contain large amounts of gut pigment (cf. Table 2),  
598 coloration of their guts may be easily recognized by visual predators (cf. Tsuda et al.,  
599 1998). This could be why *N. cristatus* C5 sometimes feeds mainly at night in this  
600 region (Saito, 1996; Takahashi et al., 2008). During the phytoplankton bloom, there  
601 would have been greater attenuation of light intensity, which would make  
602 pigment-loaded copepods more difficult to see. Thus, there may have been little  
603 advantage in April for diel feeding cycles or daytime reduction in gut pigment of *E.*  
604 *bungii* or *Neocalanus* spp. toward the end of our study (Fig. 7a, d, e). Ingestion of  
605 highly concentrated food both day and night implies a doubling food intake each day.  
606 A plausible explanation for the similar life cycle patterns seen for *Neocalanus* spp.  
607 living at Site H and at Station P is that temperature and food abundance act oppositely  
608 to produce matched timing of the life cycle (Kobari and Ikeda, 1999, 2001a, 2001b).  
609 That is, the negative effect of lower temperature on the development of *Neocalanus* spp.  
610 at Site H is compensated by the positive effect of higher food concentrations in the same  
611 area.

#### 612 **Individual mass**

613 There are previous reports of temporal changes of individual mass in subarctic copepods.  
614 For example, lipid changes in resting *N. plumchrus* were shown by Evanson et al.,  
615 (2000) and Campbell et al. (2004), and lipid consumption is implied by energy  
616 requirement estimates for resting *N. cristatus* reported by Ikeda et al. (1990, 2004).

617 All of these studies concern the fate of stored energy, and the sampling intervals were  
618 about one month. Thus no previous studies have been made on individual mass  
619 increment during a phytoplankton bloom. In body composition (water and AFDM  
620 content), there were no correlations with time for *M. pacifica*, while decreasing water  
621 content and increasing AFDM with time were observed for *N. cristatus* (Fig. 8b, d;  
622 Table 3). These differences in individual mass accumulation patterns may be related to  
623 their life cycle patterns. Thus multiple cohorts without resting by *M. pacifica* only  
624 require small amounts of accumulated lipids, while one-year generations with prolonged  
625 diapause require *N. cristatus* to accumulate large lipid stores.

## 626 **Conclusion**

627 There is increasing evidence that the patterns of zooplankton productivity are changing  
628 over time, probably in response to inter-decadal ocean climate variability (cf. Chiba et  
629 al., 2006). These changes include 2-3 fold shifts in total biomass, 30-60 day shifts in  
630 seasonal timing and 10-25% changes in average body length (Miller et al., 1992;  
631 Mackas et al., 1998; Tadokoro et al. 2005; Kobari et al., 2007). Bearing this in mind,  
632 the direct comparison of life cycle timing between locations is likely meaningless.  
633 However, it is useful to evaluate the sequence of copepod responses to phytoplankton  
634 blooms at a given location. This information is useful for predicting a future  
635 zooplankton community in terms of the matches or mismatches of each species with the  
636 seasonal events. Sequential responses of copepods to phytoplankton blooms are  
637 summarized in Fig. 10. Through this study, the reproductive and developmental  
638 timing of large copepods during the spring phytoplankton bloom was evaluated. For *E.*  
639 *bungii*, domination by C3-C6F/M in March is considered to be the last phase of their

640 diapause. In early April, their gonads matured rapidly, followed by a peak of  
641 reproduction. Peak naupliar abundance was observed in the middle of April. This  
642 reproduction was mainly governed by the specimens that over-wintered as C5 and C6.  
643 New recruitment of C6 that over-wintered as C3 and C4 started in the middle of April.  
644 For *M. pacifica*, the stock was mainly composed of C1, while egg production and  
645 hatchability were low. They cease DVM at the end of April, except for adult females  
646 (Yamaguchi et al., this issue). For *M. okhotensis*, no DVM was observed in March.  
647 C5F/M started DVM in early April, then ceased by the end of April, except for adult  
648 females, similarly to *M. pacifica*. No *Neocalanus* species undertook DVM, and a clear  
649 developmental sequence was observed for *N. cristatus* and *N. flemingeri*. In brief,  
650 *Neocalanus* species that utilize the spring bloom as energy for growth did not vary their  
651 vertical distribution during this period. In contrast, *E. bungii* and *Metridia* spp. that  
652 utilize the spring bloom as energy for reproduction can change their vertical distribution,  
653 DVM pattern, and gonad maturation drastically during the spring bloom. The results  
654 point out that the time-scales of phenology in these copepods can be stretched or  
655 compressed depending on conditions. Especially *E. bungii* but also *Neocalanus* spp.  
656 have the ability to adapt, as evidenced by the variability of life cycle timing across a  
657 wide range of North Pacific environments.

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662

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829 Diel and ontogenetic variations in vertical distributions of large grazing  
830 copepods during the spring phytoplankton bloom in the Oyashio region.

831 Deep-Sea Research II this issue.

832

833 Figure captions

834 Fig. 1. Location of the Oyashio region in the western subarctic Pacific (a), and of  
835 sampling station A-5 in the Oyashio region (b), and temporal changes in  
836 integrated mean temperature, salinity and chlorophyll *a* in the upper 100 m  
837 depths during 9 March to 30 April 2007 (c). Depth contours are shown in (b).

838 Fig. 2. Temporal changes in zooplankton wet biomass in the 0-150 m (a) and 0-500 m  
839 (b) water column in the Oyashio region both day and night during 9-14 March  
840 and 6-30 April 2007. Night:day (N:D) ratios are shown, and horizontal dashed  
841 lines indicate the position of N:D=1. Temporal changes in depth composition  
842 of zooplankton biomass in the 0-150 m and 150-500 m during day (c) and night  
843 (d) during the same period.

844 Fig. 3. *Eucalanus bungii*. Temporal changes in abundance (circles) and composition  
845 of copepodid stages (a), of naupliar stages (b), sex ratio (C4-C6) (c) and gonad  
846 maturation phases of C6F (d) in the Oyashio region during 9-14 March and 6-30  
847 April 2007. Horizontal lines in (c) indicate means of each copepodid stage.  
848 For gonad maturation of C6F (d), eight categories were applied.

849 Fig. 4. *Metridia pacifica* (a) and *M. okhotensis* (b). Temporal changes in abundance  
850 (circles) and composition of copepodid stages in the Oyashio region during 9-14  
851 March and 6-30 April 2007.

852 Fig. 5. *Neocalanus cristatus* (a), *N. flemingeri* (b) and *N. plumchrus* (c). Temporal  
853 changes in abundance (circles) and composition of copepodid stages in the  
854 Oyashio region during 9-14 March and 6-30 April 2007.

855 Fig. 6. *Eucalanus bungii* (a), *Metridia pacifica* (b) and *M. okhotensis* (c). Time  
856 series of egg production rate estimates (upper panels), proportion of normal eggs

857 and hatchability (second panels), individual recruitment (ind. female<sup>-1</sup> day<sup>-1</sup>)  
858 (third panels) and population recruitment (ind. m<sup>-3</sup> day<sup>-1</sup>) (bottom panels) in the  
859 Oyashio region during 9-14 March and 6-30 April 2007.

860 Fig. 7. *Eucalanus bungii* C6F (a), *Metridia pacifica* C6F (b), *M. okhotensis* C6F (c),  
861 *Neocalanus cristatus* C5 (d) and *N. flemingeri* C5 (e). Temporal changes in gut  
862 pigment content of individuals collected in the upper 150 m during day (open  
863 circles) and night (solid circles) in the Oyashio region during 9-14 March and  
864 6-30 April 2007. Vertical bars indicate standard deviations.

865 Fig. 8. *Metridia pacifica* C6F (a, b) and *Neocalanus cristatus* C5 (c, d). Temporal  
866 changes in individual body mass (WM, DM and Ash) of specimens collected  
867 during daytime in the upper 150 m (a, c) and their water content (%WM) and  
868 AFDM content (%DM) (b, d) in the Oyashio region during 9-14 March and 6-30  
869 April 2007. Vertical bars indicate standard deviations. For body composition,  
870 a significant relationship with time was recognized only for *N. cristatus* (d).

871 Fig. 9. *Neocalanus cristatus* (a) and *N. flemingeri* (b). Temporal changes in  
872 cumulative stage composition of *N. cristatus* (a) and proportions of each stage in  
873 *N. flemingeri* (b) in the Oyashio region during 9-14 March and 6-30 April 2007.  
874 Horizontal dashed line in (a) indicates position of 50% in each of younger and  
875 older stages, and arrows in (b) indicate peaks of each copepodid stage.

876 Fig. 10. Schematic diagram showing reproductive and developmental timing of large  
877 copepods (*Eucalanus bungii*, *Metridia pacifica*, *M. okhotensis*, *Neocalanus*  
878 *cristatus*, *N. flemingeri* and *N. plumchrus*) in the Oyashio region during the  
879 spring phytoplankton bloom in 2007.

Table 1. Comparison in abundance which collected with 0.10 (XX13) and 0.33 (GG54) mm mesh of Twin NORPAC net during spring phytoplankton bloom in the Oyashio region.

Values are means of ratio of XX13:GG54 (both abundance in ind. m<sup>-3</sup>). Stages substantially greater abundance was observed for XX13 were shown with underlines.

Species	Stage						
	Nauplii	C1	C2	C3	C4	C5	C6
<i>Eucalanus bungii</i>	<u>79.4</u>	1.2	0.7	1.2	1.2	0.8	1.4
<i>Metridia pacifica</i>		<u>43.9</u>	<u>11.3</u>	<u>5.2</u>	1.6	1.6	1.7
<i>Neocalanus cristatus</i>		0.9	3.6	1.9	1.5	1.0	
<i>Neocalanus flemingeri</i>		1.2	1.0	1.0	0.6	0.9	0.4
<i>Neocalanus plumchrus</i>		1.0	0.8	0.6	1.5	0.7	

Table 2. Day and night comparison on gut pigment of late copepodid stages of dominant copepods in the Oyashio region during 9 March-30 April 2007. Values are mean±1sd. \*\*:  $p < 0.01$ , ns: not significant. Details of data see Fig. 7.

Species (stage)	Gut pigment (ng Chl. ind. <sup>-1</sup> )		U -test
	Day	Night	
<i>Eucalanus bungii</i> (C6F)	20.8±6.4	25.0±8.9	ns
<i>Metridia pacifica</i> (C6F)	2.3±2.4	6.2±2.5	**
<i>Metridia okhotensis</i> (C6F)	6.7±3.2	55.1±29.2	**
<i>Neocalanus cristatus</i> (C5)	133.6±61.3	197.5±66.1	ns
<i>Neocalanus flemingeri</i> (C5)	15.7±8.5	25.9±6.9	ns

Table 3. Correlation coefficient between date (julian day) and individual body masses (wet mass [WM], dry mass [DM], ash and ash-free dry mass [AFDM]) or body composition (water and AFDM) of *Metridia pacifica* C6F and *Neocalanus cristatus* C5 in the Oyashio region during March-April 2007. \*:  $p < 0.05$ , \*\*:  $p < 0.01$ , \*\*\*:  $p < 0.001$ , ns: not significant.

Species (stage)	Body mass				Chemical contents	
	WM	DM	Ash	AFDM	Water	AFDM
<i>Metridia pacifica</i> (C6F)	0.60**	0.69**	0.42 <sup>ns</sup>	0.68**	-0.30 <sup>ns</sup>	0.43 <sup>ns</sup>
<i>Neocalanus cristatus</i> (C5)	0.64**	0.72***	0.37 <sup>ns</sup>	0.72***	-0.73***	0.80***

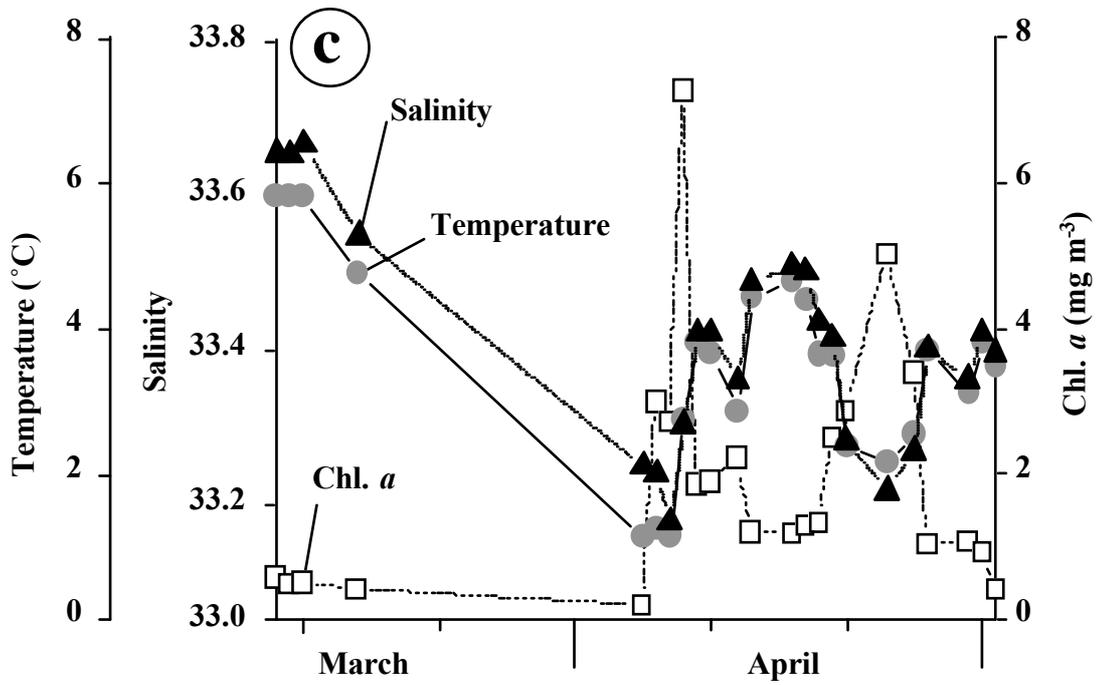
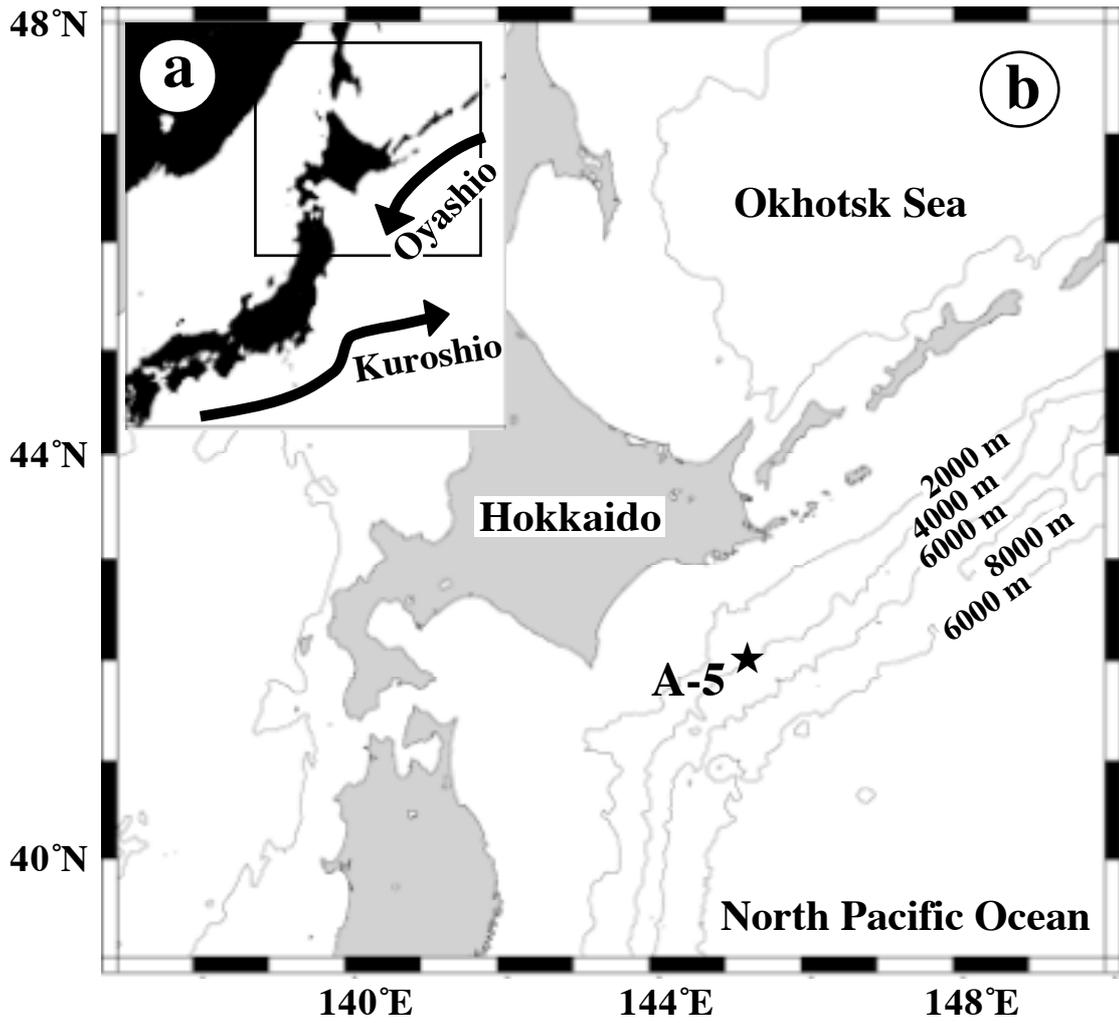


Fig. 1 (Yamaguchi et al.)

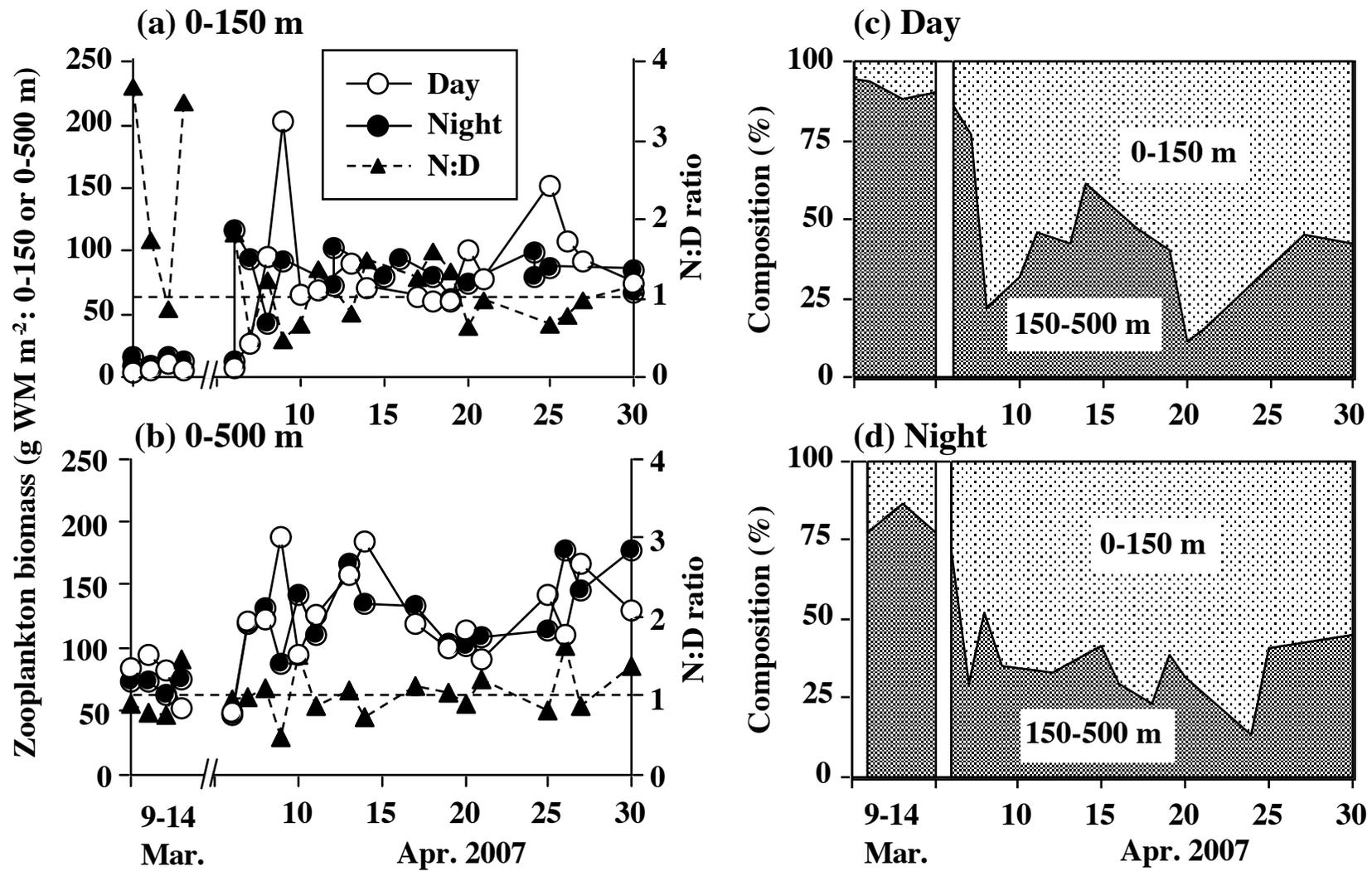


Fig. 2 (Yamaguchi et al.)

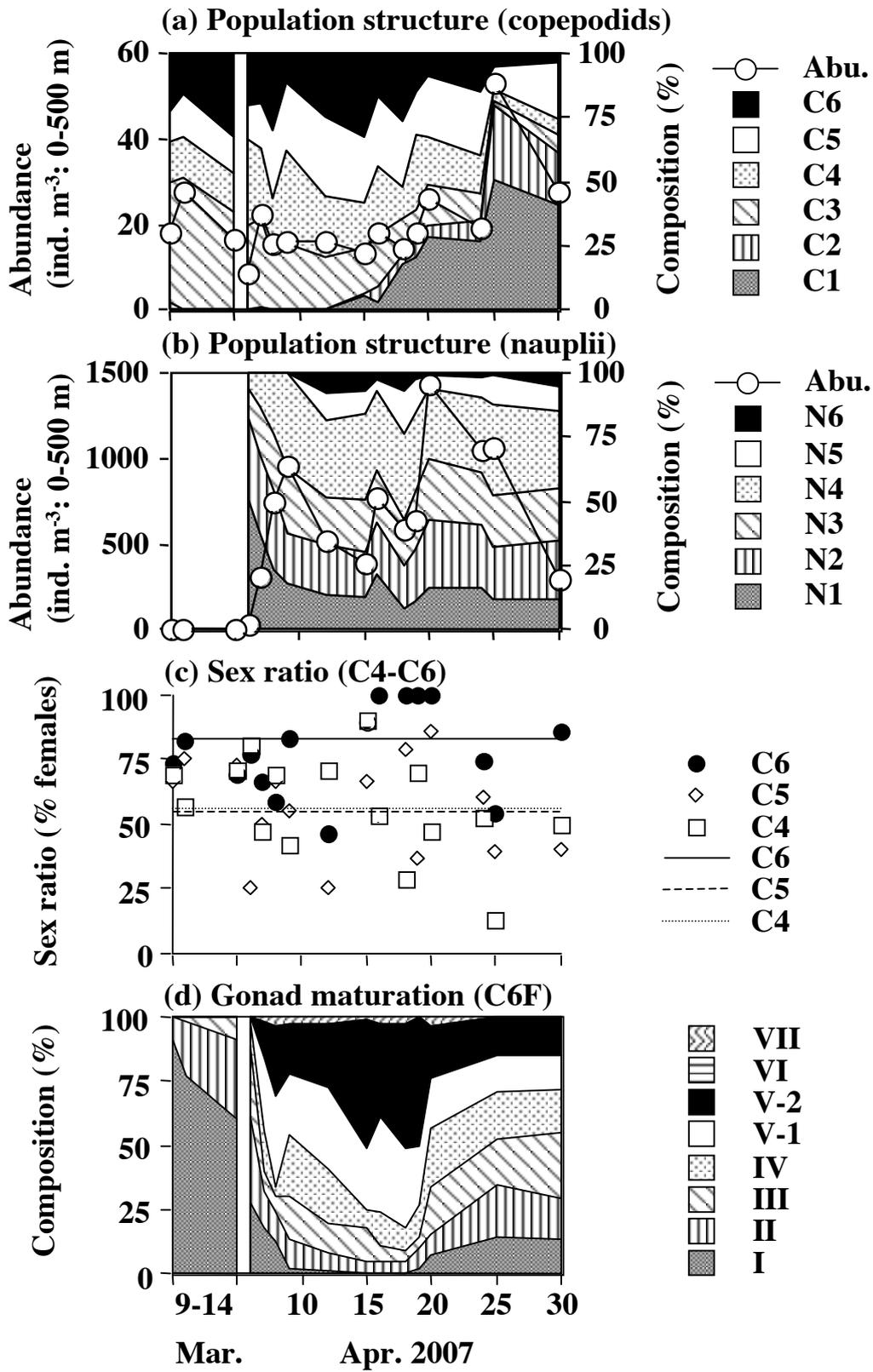


Fig. 3 (Yamaguchi et al.)

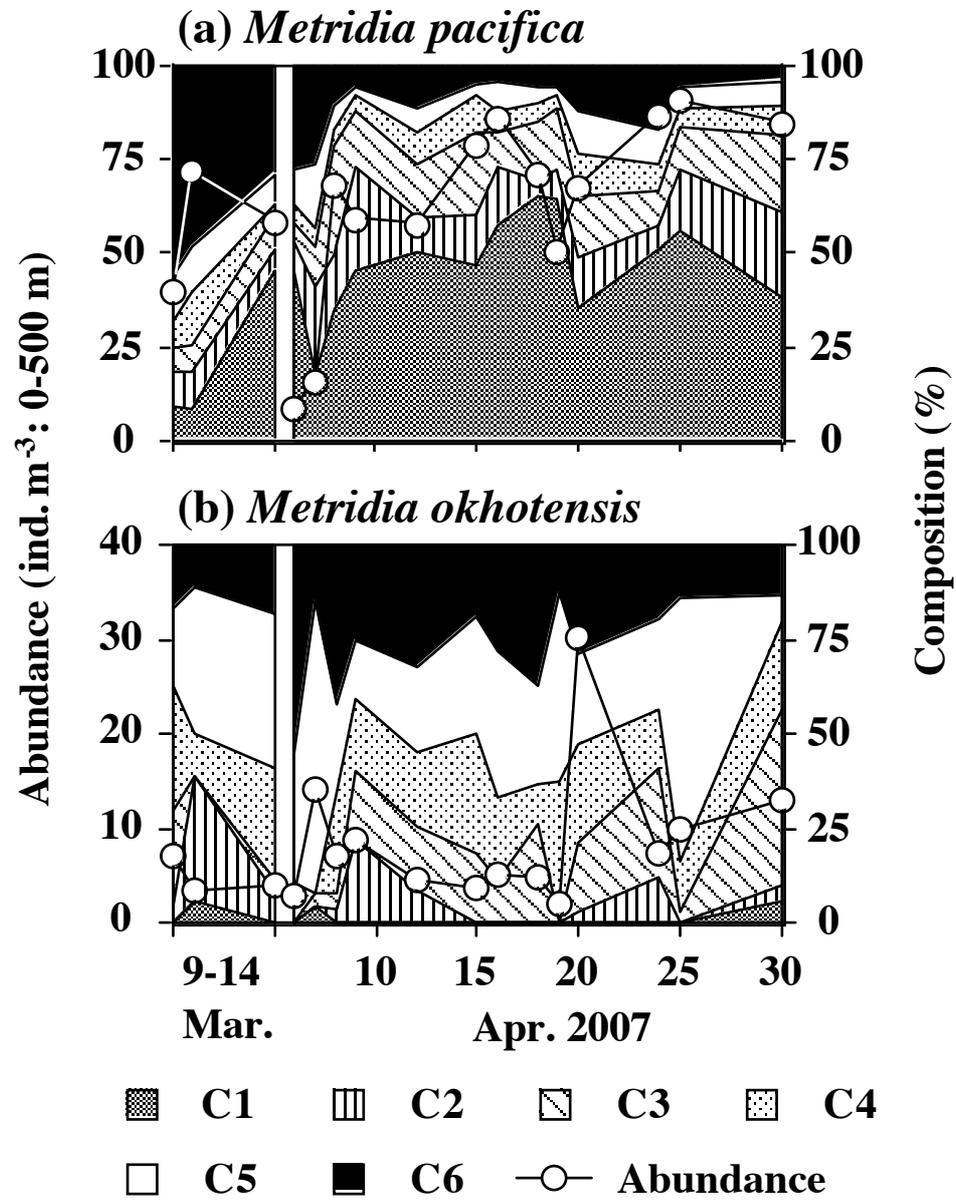


Fig. 4 (Yamaguchi et al.)

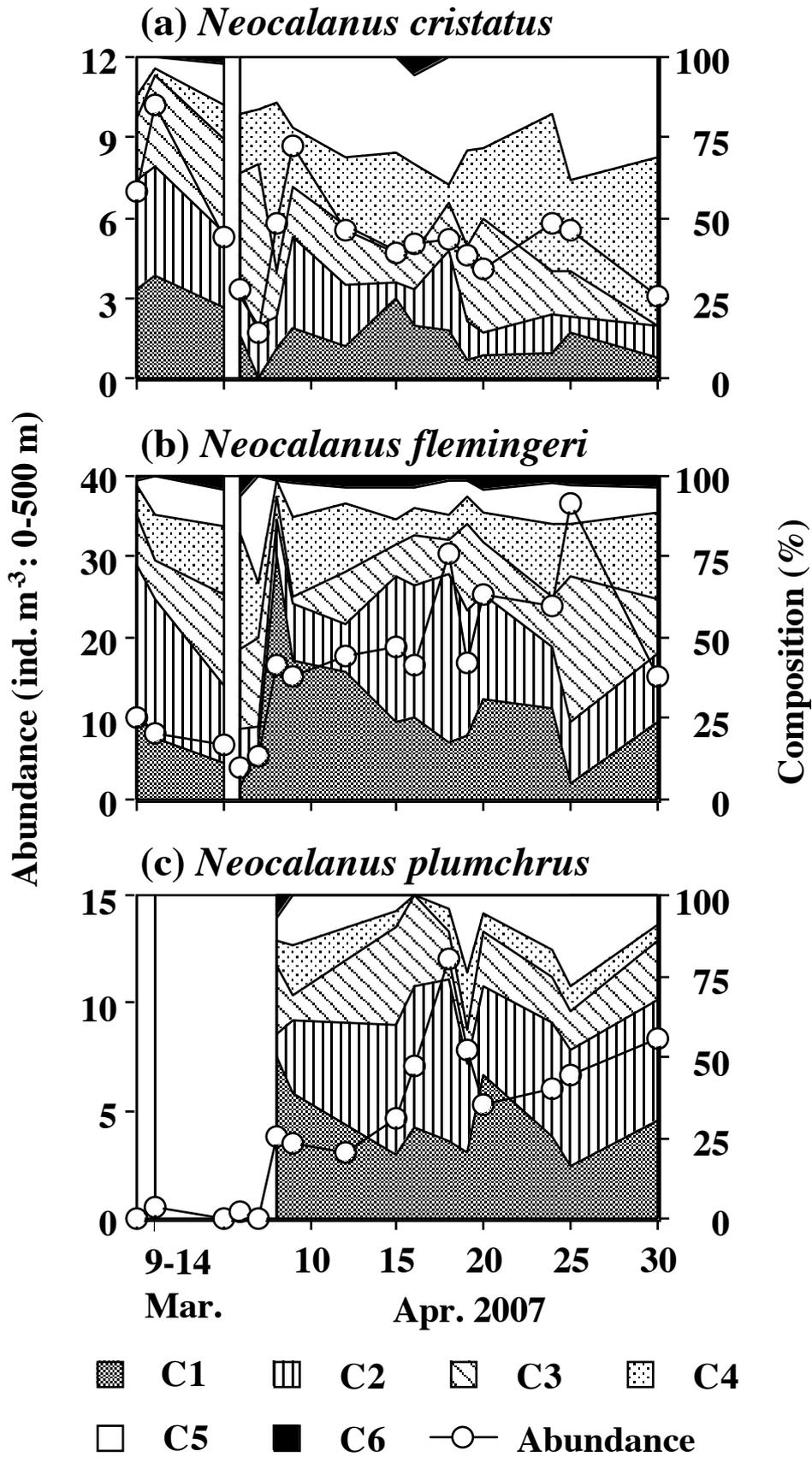


Fig. 5 (Yamaguchi et al.)

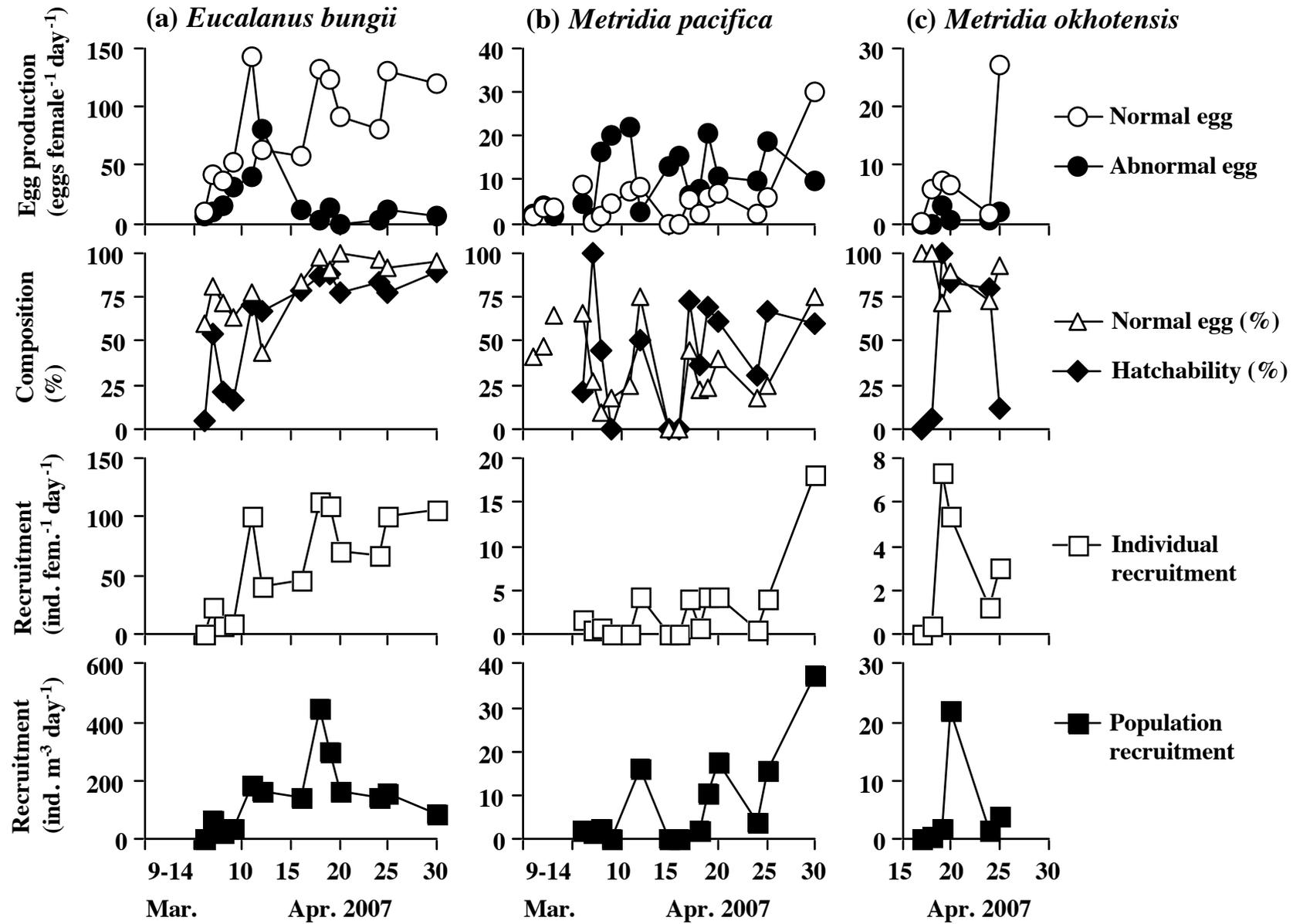


Fig. 6 (Yamaguchi et al.)

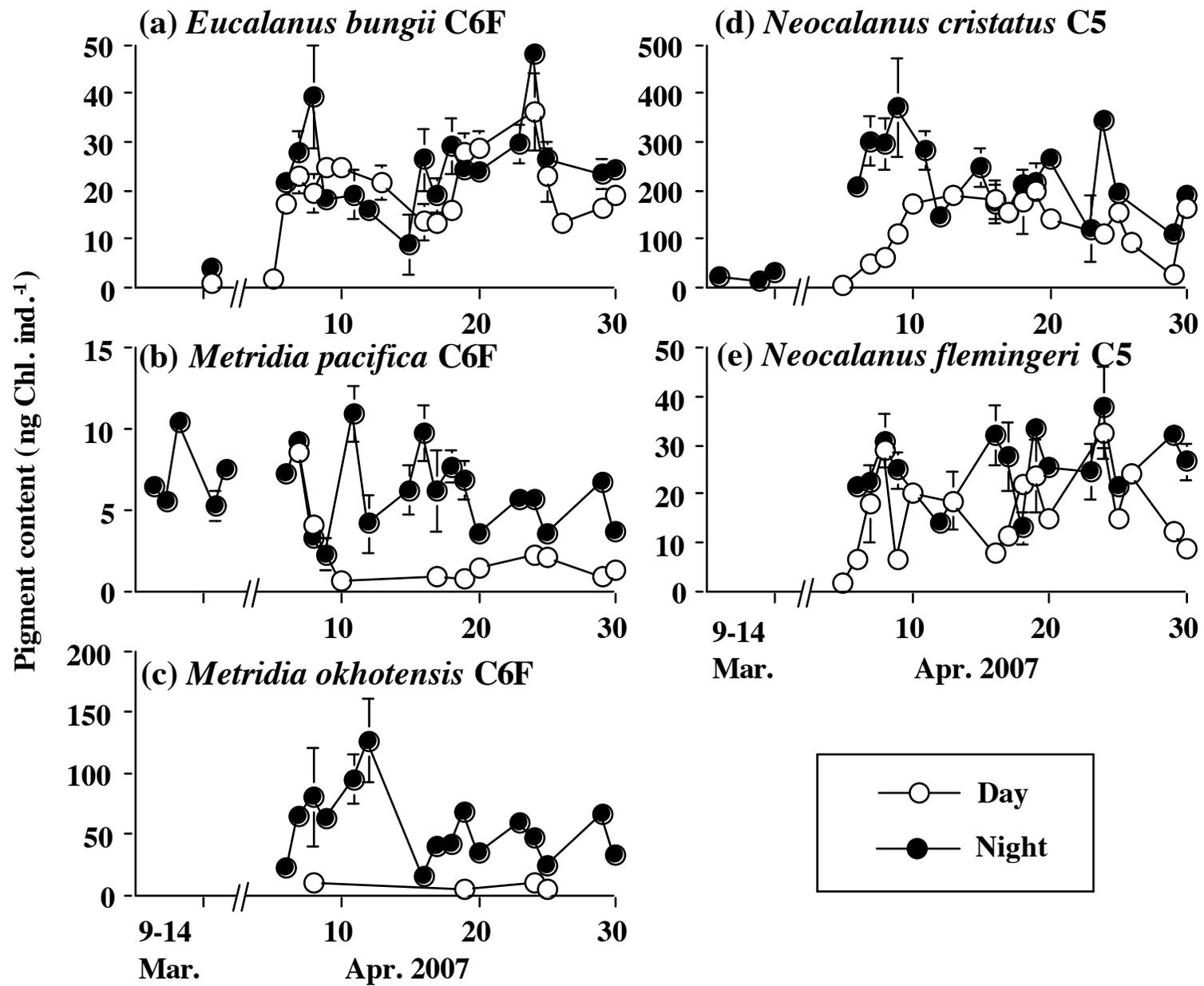


Fig. 7 (Yamaguchi et al.)

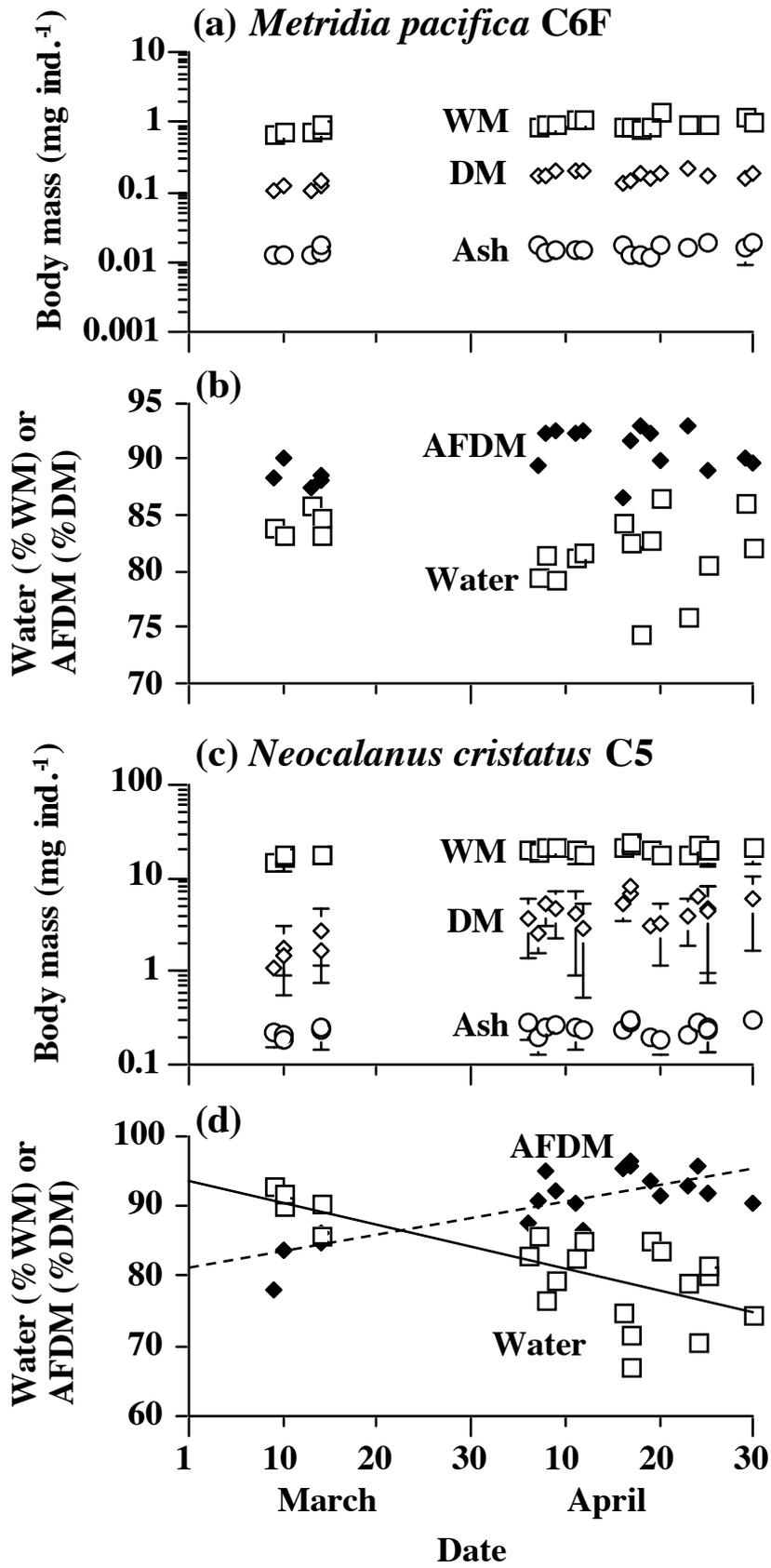


Fig. 8 (Yamaguchi et al.)

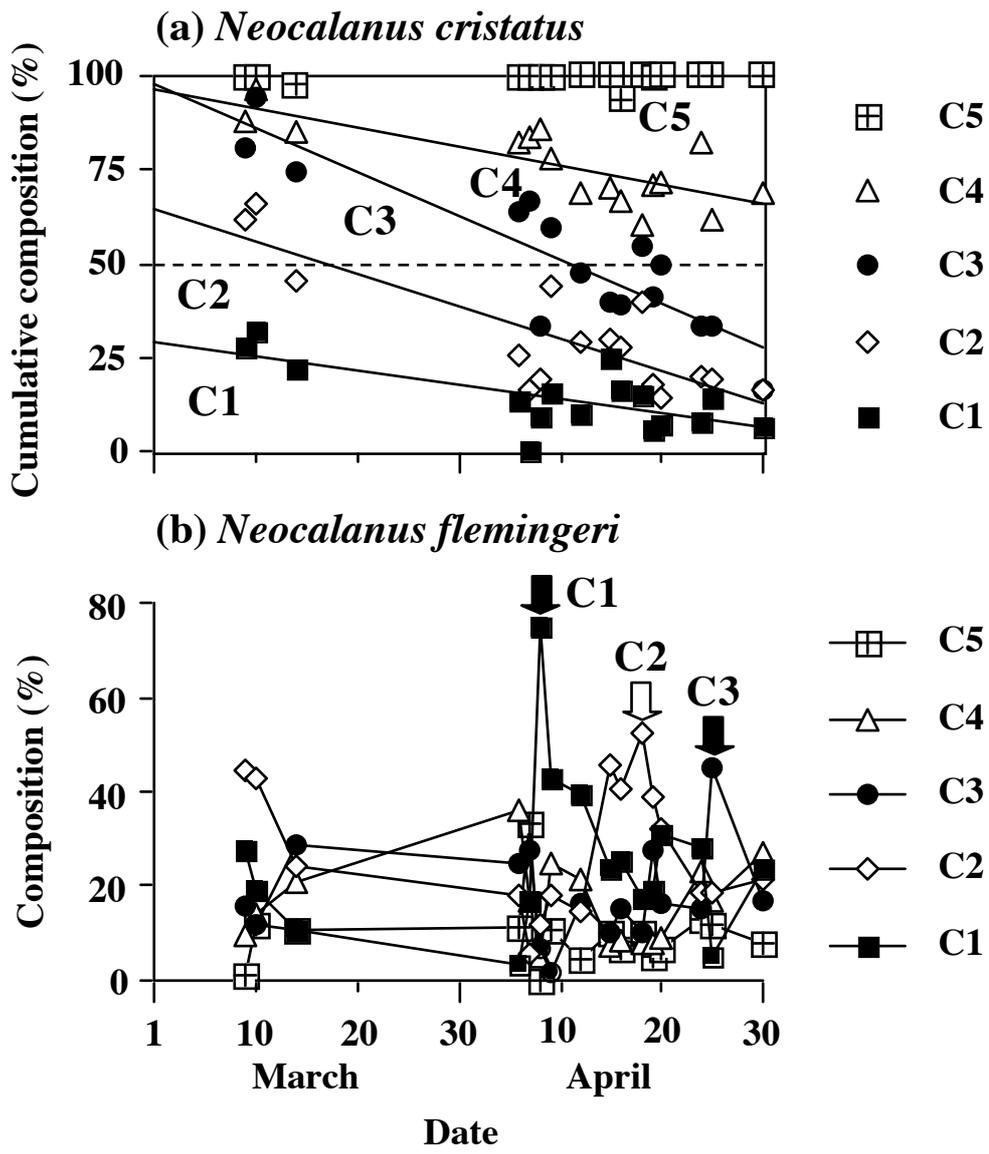


Fig. 9 (Yamaguchi et al.)

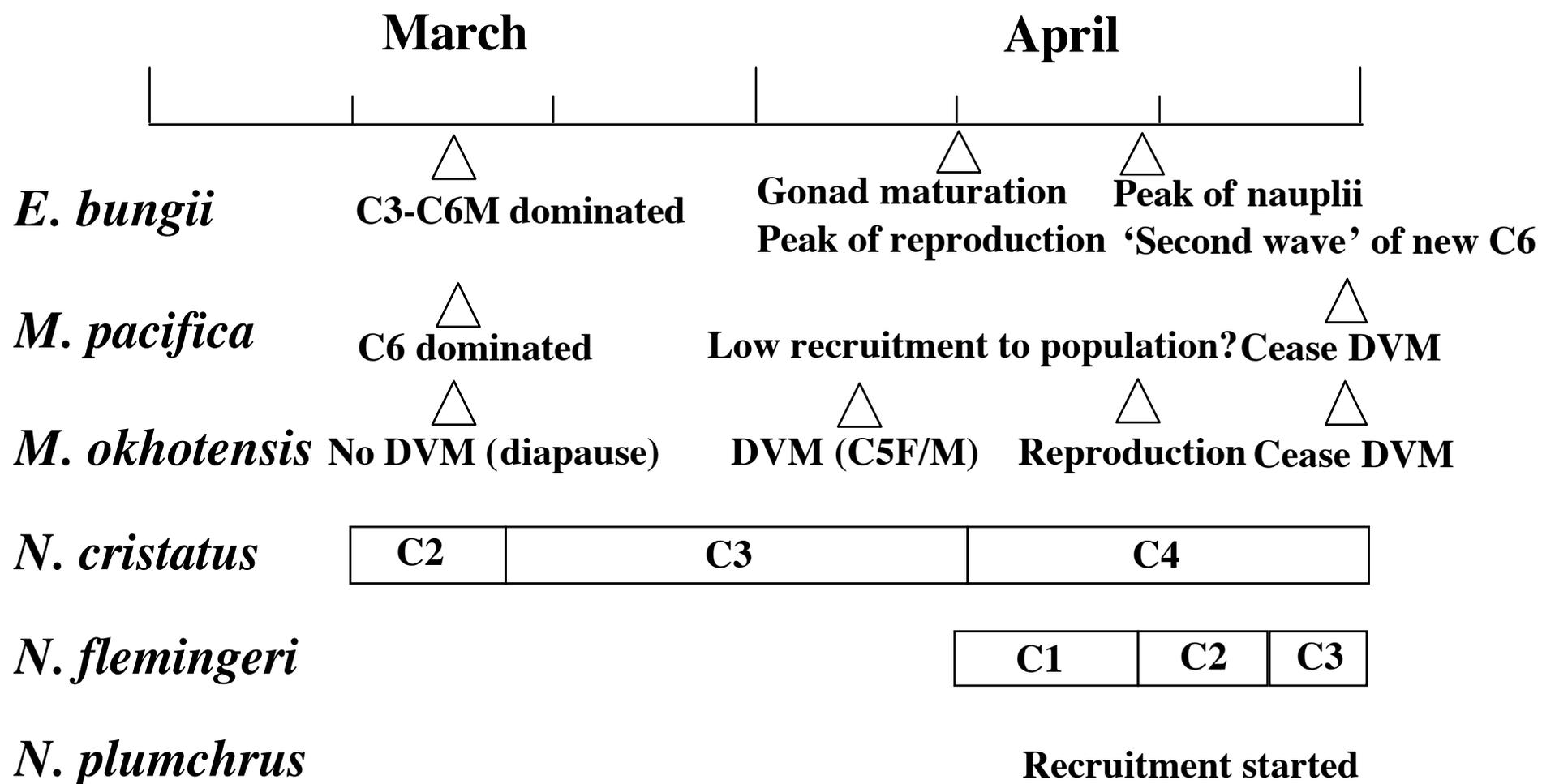


Fig. 10 (Yamaguchi et al.)