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Extrahypothalamic Projection of Immunoreactive Vasotocin Fibers in the Brain of the Toad, *Bufo japonicus*

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**ABSTRACT**—Extrahypothalamic projection of vasotocin (AVT) fibers in the brain of the toad (*Bufo japonicus*) was examined immunohistochemically by the avidin-biotin-peroxidase complex (ABC) method. Immunoreactive AVT perikarya are localized in the nucleus preopticus pars magnocellularis. The AVT neurons send their immunoreactive varicose fibers to many discrete brain regions, such as the limbic cortex, the thalamus, the optic tectum and the lower brain stem, in addition to the neurohypophysis. A dense network of AVT fibers was found in the septal nuclei and the anterior part of the preoptic nucleus. AVT fibers which run postero-dorsad project to the nucleus posterocentralis thalami, the nucleus posterodorsalis tegmenti mesencephali, and the nucleus isthmi. Meanwhile, AVT fibers which run through in the dorsal infundibular region and then the mesencephalic reticular formation are distributed in the medulla oblongata. These findings suggest that AVT acts as a neuromodulator or a local hormone in the toad brain.

**INTRODUCTION**

It is well established that, in anuran amphibians, vasotocin (AVT) has both antidiuretic and vasopressor effects. In addition, AVT shows pronounced effects on reproductive behavior in *Rana pipiens* [1] and *Taricha granulosa* [2]. In *T. granulosa*, AVT may be involved in control of sexual behavior by acting neurons in the central nervous system [3]. We have shown in the toad brain that vasotocin neurons project their varicose axons into the anterior part of the preoptic nucleus (APON) which is considered to be the triggering center for male mate calling behavior in anuran amphibians [4]. These results suggest that AVT neurons transmit APON neurons peptidergic information concerned with initiation of sexual behavior.

In mammalian brains, the distributions of various neurohormones including arginine-vasopressin (AVP) are not confined only in the hypothalamic-neurohypophyseal system. They are widely distributed throughout in discrete brain loci [5]. Ultrastructural studies showed that axon endings of immunoreactive (ir) luteinizing hormone-releasing hormone (LHRH) fibers [6] and ir-AVP ones [7] form synapses and synaptoid contacts with other neurons. Further, varicose LHRH fibers form en passant synapses in the preoptic area of the guinea pig [6]. Therefore, it is probable in amphibian brains that, in addition to the APON and the neurohypophysis, AVT neurons send their fibers to many extrahypothalamic regions.

In this study, we examined immunohistochemically the extrahypothalamic distribution of AVT fibers in the toad brain to learn whether AVT neurons project their axons to the loci which are related to control of mating behavior. Further, we have tried to elucidate phylogenetically fundamental distributional pattern of AVT in the vertebrate brain, since the amphibian brain is considered to show fundamental structural organization of the vertebrate brain [8].

**MATERIALS AND METHODS**

Adult toads (*Bufo japonicus*) of both sexes, body weight ranging from 117 to 303 g, and body
length (snout to vent) from 11.9 to 14.1 cm, were used. These animals were either obtained from an animal supplier in late September or were captured at the breeding season. Three animals, which were obtained from an animal supplier in late November, were used for immunohistochemical staining of thick frozen sections.

**Immunohistochemical procedure**

Distribution of AVT was immunohistochemically localized in paraffin sections and thick frozen sections that were cut either transversally, sagittally or horizontally to determine exact loci where ir-AVT fibers were found. As for immunohistochemical staining of vasotocin in the paraffin sections of the toad brain, details of fixation, tissue preparation and immunohistochemical procedure, in which the Vectastain ABC kit (Vector) was used, have been described previously [9, 10].

For the staining of thick frozen sections, the brains were fixed by transcardial perfusion with a fixative solution containing 1% glutaraldehyde, 2% paraformaldehyde and 4% sucrose in 0.1 M phosphate buffer (pH 7.4). The brains were removed, postfixed in the same fixative at 4°C overnight, and were washed in 0.1 M phosphate buffer. Frozen sections were cut at 50 μm, and were washed in 0.1 M phosphate buffered saline (PBS-T) at room temperature for 30 min, incubated in the avidin-biotin-peroxidase complex in PBS-T for 30 min. After a brief incubation with the primary antiserum, the sections were washed in PBS-T at room temperature for 30 min, incubated with biotinylated anti-rabbit Ig-G for 1 hr, and were washed in PBS-T. Afterward, the tissue sections were incubated in the avidin-biotin-peroxidase complex in PBS-T for 30 min. After a few rinses, they were incubated in DAB solution including 0.05% 3,3'-diaminobenzidine (Sigma) and 0.01% hydrogen peroxide for 10 min, washed briefly in phosphate buffer, and were mounted on slide glasses with 40% ethanol containing 0.75% gelatin. They were then dehydrated, and were cover-slipped with Permount (Fisher). The tests for specificity of immunohistochemical staining followed the previous study [4, 9].

Nissl stained tissue sections were referred to for describing precise localization of ir-AVT. Nomenclatorial usage in this paper is basically those in *Rana pipiens* [11] and *Bufo japonicus* [9].

**RESULTS**

Distribution of ir-AVT perikarya and fibers

As was described previously [4, 9], ir-AVT perikarya are localized in the ventral (VMC) and dorsal (DMC) magnocellular parts of the preoptic nucleus (Figs. 1 and 5). Beaded or varicose ir-AVT fibers were widely distributed among the discrete extrahypothalamic loci in the limbic system and the brain stem (Figs. 2-4). They were not found in the dorsal and anteroventral regions of the telencephalon. The extrahypothalamic ir-AVT projections can be classified roughly into three groups according to their destinations (see Fig. 5). Neither notable seasonal variation nor sexual difference was found in the distribution of ir-AVT fibers in this study.

**Projection to the telencephalon**

(Figs. 5-8)

A part of ir-AVT fibers emanating from the VM project to the postero medial region of the telencephalon, principally to the nuclei medialis septi and lateralis septi. Ir-AVT fibers are sent out to these loci mainly through the white matter including the medial forebrain bundle which surrounds the neuronal cell mass of the APON. A part of ir-AVT fibers emanating from the VM project to the postero medial region of the telencephalon, principally to the nuclei medialis septi and lateralis septi. Ir-AVT fibers are sent out to these loci mainly through the white matter including the medial forebrain bundle which surrounds the neuronal cell mass of the APON.
found in the telencephalon have a varicose form, however, they rarely show Herring bodies which are frequently observed in the magnocellular preopticoneurohypophyseal neurosecretory system. 

Projection to the thalamus and the tectum (Figs. 5, 8-9). 

Many ir-A VT fibers which project to the brain stem arise from ir-A VT neurons in the VMC. They initially proceed laterad into the white matter in the preoptic region. Then, they turn their destination caudad to the direction of the lower brain stem with many ir-A VT fibers that project to the neurohypophysis. Thereafter, the fibers projecting to the brain stem diverge from the preopticoneurohypophyseal tract around the dorsal infundibular region. A few beaded ir-A VT fibers are localized in the nucleus infundibularis dorsalis (Fig. 2). Many ir-A VT fibers run down to the mesencephalic tegmentum. They project to the nuclei posterodorsalsis tegmenti mesencephali and isthmi, and further to the griseum centrale rhombencephali. A considerable number of fine beaded fibers gather together to form a plexus in the region anterior to the nucleus isthmi (Fig. 3).

Projection to the brain stem (Figs. 5, 10-11). 

Ir-A VT fibers which project to the brain stem diverge from the preopitcineurohypophyseal tract around the dorsal infundibular region. Some fibers which emanate from this plexus proceed dorsad to the stratum griseum tecti. Many ir-A VT fibers which run posteriorly through the white matter in the floor of the mesencephalon were also found (Fig. 4). These fibers, which diverge from the fiber group destined to the isthmic region at the level caudal to the dorsal infundibular region, reach the rhombencephalic reticular formation, and proceed further toward the spinal cord.

DISCUSSION

The present study showed that ir-A VT fibers are distributed widely among many extrahypothalamic loci in the toad brain. Such regions are the limbic cortex, the thalamus, the optic tectum, the isthmus region and the lower brain stem. The distributional pattern of extrahypothalamic ir-A VT fibers in the toad brain seems to be homologous to those described in the brains of other vertebrate classes. In the rat and the monkey, vasopressin neurons, the mammalian counterpart of AVT neurons, project their immunoreactive fibers to the hippocampus, the septum, the amygdala and the preoptic area. They
project further to the thalamus, the superior nuclei [12]. A similar distributional pattern of ir-A VT fibers was observed in the brain of the lizard *Gekko gecko* [13] and the eel *Anguilla japonica* (Fujiwara et al., unpublished). A radioimmunoassay study of microdissected brain areas of rough-skinned newts also showed a similar distributional pattern of AVT and vasopressin are fundamentally homologous in all vertebrate species.

Immunoelectron microscopic studies demonstrated the presence of synapses containing neurohypophysial hormones in the rat brain [7, 15]. In the previous study, we showed that LRH and AVT fibers may contact synaptically with APON neurons [4]. It is therefore highly probable that, in the toad brain, the extrahypothalamic AVT fibers form ordinary and/or en passant synapses with neurons in the locus where AVT fibers were localized. As was discussed in our previous paper [4], beaded or varicose ir-A VT fibers traveling through the white matter may form such synapses with dendrites of many central neurons, because in the amphibian brain many neurons located in the medial cell mass develop their dendritic fields in the adjacent white matter [8, 16]. Since neurohypophysial hormones could excite unit-spike activity of neurons in the rat supraoptic and paraventricular nuclei [17, 18], the eel preoptic nucleus [19] and the toad APON (Fujita and Urano, unpublished), vasotocin may facilitate activity of many central neurons as a neuromodulator or a local hormone. The latter possibility is supported by the fact that AVP of 10^{-7} M, comparable to effective doses of AVP for peripheral targets, could excite rat paraventricular neurons [18]. However, physiological roles of vasotocinergic transmission in the amaran brain are not clear at present, although an involvement in reproductive behavior has been suggested as is described in the introduction.

The present study showed that vasotocin fibers are distributed in the loci concerned with reproductive behavior. Such regions are the limbic cortex, the preoptic area, the optic tectum and the central gray. In the limbic cortex, a considerable number of ir-A VT fibers were found in the septal nuclei, and also in the nucleus of the diagonal band of Broca. These regions contain the majority of ir-LRH neurons in the toad brain [4, 10]. A similar projection of vasopressin fibers was found in the organum vasculosum lamina terminalis in the mammalian brain where many ir-LRH perikarya are localized [12]. In the eel, vasotocin fibers were localized in the preoptic region (Fujiwara et al., unpublished). Meanwhile, ir-LRH fibers project to the VMH in the toad brain [20]. These observations suggest that the LHRI-ergic and vasotocinergic neurosecretory systems are mutually connected, and that they interact on each other for controlling sexual behavior.

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REFERENCES