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Construction of an infectious cDNA clone for Omsk Hemorrhagic fever virus, and characterization of mutations in NS2A and NS5

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Abstract

Omsk hemorrhagic fever virus (OHFV) is a member of the tick-borne encephalitis serocomplex of flaviviruses, and causes hemorrhagic disease in humans. In this study, an infectious cDNA of OHFV was constructed to investigate the molecular mechanisms involved in OHFV pathogenesis for the first time. Our cDNA clone was capable of producing infectious virus which is genetically identical to the parental Guriev strain, and the recombinant virus showed similar biological properties to the parental virus including growth kinetics and virulence characteristics. While characterizing the cDNAs, fortuitous mutations at NS2A position 46 and NS5 position 836 were found to affect viral production. By using a viral replicon expressing luciferase, it was shown that both of the mutations produced a defect in RNA replication and that the NS5 mutation induced a temperature-sensitive phenotype, indicating the importance of these residues in RNA replication. This infectious cDNA will be a useful tool to study the replication and pathogenesis of OHFV.

Keywords: Omsk hemorrhagic fever; flavivirus; infectious cDNA; viral replication.

Abbreviations

OHF: Omsk hemorrhagic fever

OHFV: Omsk hemorrhagic fever virus

TBE: tick-borne encephalitis

LGTV: Langat virus

ALKV: Alkhurma virus

KFDV: Kyasanur Forest disease virus

C: core

prM: premembrane

E: envelope

NS: non-structural

HDV-RZ: hepatitis delta virus ribozyme

MOI: multiplicity of infection

RdRp: RNA-dependent RNA polymerase

EM: energy minimization

rms: root mean square

ts: temperature-sensitive

NLS: nuclear localization sequence

1. Introduction

Omsk hemorrhagic fever (OHF) is a disease caused by Omsk hemorrhagic fever virus (OHFV), which belongs to the tick-borne encephalitis (TBE) serocomplex, genus *Flavivirus*, family *Flaviviridae*. OHFV was first isolated in 1947 from a human presenting with hemorrhagic fever. OHFV is endemic to a fairly localized region of Siberia within the Omsk and Novosibirsk Oblasts in Russia (Burke and Monath, 2001). OHFV is transmitted via the bite of its primary tick vector, *Dermacentor reticulatus*. The transmission cycle of OHFV involves water voles (*Arvicola terrestris*) and muskrats (*Ondatra zibethica*), and many other animals within endemic area can be infected with OHFV (Busygin, 2000; Kharitonova and Leonov, 1985).

The TBE complex includes tick-borne encephalitis virus (TBEV), Powassan virus, Langat virus (LGTV), Louping ill virus, OHFV, Alkhurma virus (ALKV), and Kyasanur Forest disease virus (KFDV) (Buchen-Osmond, 2003). Although the TBE complex is largely represented by viruses causing encephalitis, OHFV, ALKV and KFDV are known to cause hemorrhagic disease. Unlike ALKV and KFDV, OHFV causes a hemorrhagic disease in humans with few neurological effects (Burke and Monath, 2001). Human OHFV infection results in fever, headache, myalgia, dehydration, and hemorrhage. The mortality rate for OHF is estimated to be 0.4%–2.5% (Kharitonova and Leonov, 1985). In the mouse model, OHFV causes disease with few neurological signs compared to neurotropic tick-borne flaviviruses and has also demonstrated significantly different tissue localization indicating potential differences in host cell interactions (Holbrook et al., 2005). However, the specific viral and host response mechanisms involved in OHFV pathogenesis are not well understood.

The flavivirus genome consists of a positive-polarity, single-stranded RNA of approximately 11 kb, which encodes three structural proteins, i.e., the core (C), premembrane (prM), and envelope (E)

proteins, and seven non-structural (NS) proteins (NS1, NS2A, NS2B, NS3, NS4A, NS4B, and NS5), within a single long open reading frame (Chambers et al., 1990). The 5'- and 3'-UTRs have predicted secondary structures that are implicated in viral replication, translation, and packaging of the genomes (Gritsun et al., 1997; Proutski et al., 1997; Rauscher et al., 1997).

Infectious cDNA clones have been generated for multiple flaviviruses, and they provide a useful platform on which to investigate the genetic determinants of flavivirus virulence. There are several reports of infectious cDNA clones for TBE serocomplex, TBEV and LGTV (Campbell and Pletnev, 2000; Gritsun and Gould, 1998; Hayasaka et al., 2004; Mandl et al., 1997; Pletnev, 2001). However, an infectious clone has not been developed for OHFV. In this study, we report the construction and characterization of an infectious cDNA clone of OHFV. Furthermore, while characterizing this cDNA, several fortuitous mutations in NS proteins were shown to attenuate viral replication and reduce virulence in mice.

2. Material and methods

2. 1. Cell and virus

BHK cells were grown in MEM supplemented with 8 % FBS. OHFV strain Guriev was obtained from the World Reference Collection for Emerging Viruses and Arboviruses (WRCEVA) and used for construction of the infectious cDNA clone.

2. 2. Plasmid constructions

Total cellular RNA was extracted from OHFV (strain Guriev)-infected BHK cells using Trizol (Invitrogen). OHFV RNA was reverse-transcribed with specific oligonucleotide primers using Superscript II reverse transcriptase (Invitrogen). The fragments of OHFV cDNA were amplified by *Platinum Taq* polymerase (Invitrogen) using specific oligonucleotide primers (see Fig. 1), resolved

by gel electrophoresis and purified using the Qiaquick gel extraction kit (Qiagen).

The resulting fragments were digested with restriction endonucleases and cloned into the low copy plasmid pACNR provided by Dr. Peter Mason (Ruggli et al., 1996) as depicted in Fig. 1. The oligonucleotide used to amplify the 5' UTR included a T7 promoter recognition site and an additional G preceding the first base of the viral genome. A synthetic oligonucleotide was used to add *Cla* I site at nucleotides 2436-2441. This silent mutation was engineered to permit the ligation of the fragment containing the structural protein gene. A hepatitis delta virus ribozyme (HDV-RZ)/bacteriophage T7 terminator fragment (Mason et al., 2002) was fused to the 3' end of the viral genome to create synthetic run-off transcripts that contained a 3' terminus identical to the viral RNA. The resulting cloned plasmids (designated as OHF-IC) were isolated by standard techniques and sequences were checked at the University of Texas Medical Branch (UTMB) Protein Chemistry Laboratory and compared to the sequence of the original OHFV Guriev sequence (accession No. AB507800). Sequencing identified three mutations encoding amino acid changes at NS2A-46, NS5-65 and NS5-836.

To repair the mutations found in the OHF-IC plasmids, the fragments between nucleotide 3612-3685 (*Hpa* I – *Sph* I site), 7094-8778 (*Bam* HI – *Kpn* I site), 9488-10295 (*Nde* I – *Apa* I site) were amplified by RT-PCR and subsequently cloned into pCR2.1 plasmid (TA cloning kit; Invitrogen). The virus-specific sequence of each intermediate cloning product was checked by sequence analysis. These intermediate plasmids containing the correct sequence were cut by restriction enzymes described above and used to substitute the regions containing the mutations.

Subgenomic OHFV replicons expressing luciferase gene (OHF-REP-luc (Yoshii and Holbrook, 2009)) were used to analyze the effect of the mutations on viral genome replication. Fragments from the mutated OHF-IC plasmids were amplified using the same primers sets described above.

The PCR product from the fragments containing mutations was cloned into the replicon plasmid to generate replicons containing the specific mutations found in OHF-IC.

2. 3. RNA synthesis-transfection

The RNAs were synthesized as described previously (Yoshii et al., 2005). Briefly, the plasmids were prepared for run-off transcription by digestion with *Xba*I restriction endonuclease, and the resulting template DNAs were in vitro transcribed using the mMESSAGE mMACHINE T7 Kit (Ambion) in a 20- μ l reaction mixture that contained an additional 1 μ l of 20 mM GTP solution. After transcription at 37°C for 2 h, the template DNAs were removed by DNase I digestion at 37 °C for 15 min. The RNA was precipitated using lithium chloride, washed with 70 % ethanol, resuspended in RNase-free water, quantitated by spectrophotometer, and stored at –80 °C in aliquots. The synthesized RNA was transfected into BHK cells using TransIT-mRNA (Mirus Bio) by a slight modification of the manufacturer’s protocol.

2. 4. Detection of OHFV-antigen (IFA)

Cells were fixed with 3% paraformaldehyde, and permeabilized with 0.2% Triton X-100. After blocking with 2% BSA, the cells were incubated with Rabbit polyclonal antibodies against TBEV E proteins which is cross-reactive to OHFV E proteins (Yoshii et al., 2004), and then treated with Alexa 555-conjugated anti-rabbit IgG antibodies (Invitrogen). The images were viewed and recorded using fluorescence microscopy.

2. 5. Viral titration and growth curves

Stock preparations of recombinant OHFV from OHF-IC were produced by passaging the virus harvested from the supernatant of RNA-transfected cells once in BHK cells. The sequence of the RNA of the recovered stock virus was confirmed to be identical to that of the each OHF-IC plasmid

by sequence analysis after RT-PCR. For titrations, cell monolayers prepared in multi-well plates were incubated with serial dilutions of virus for 1 h, and then overlaid with MEM containing 2 % FBS and 1.5 % carboxymethyl cellulose (CMC; Sigma) and incubated for 5 days. After incubation, the cells were fixed and stained with 0.25 % crystal violet in 10 % buffered formalin. Plaques were counted and expressed as PFU/ml. For growth curves, BHK cells were infected at a multiplicity of infection (MOI) of 0.01. After virus adsorption for 1 h, the inocula were removed, and the cells were washed with PBS and incubated in MEM containing 2 % FBS. The media was harvested at 24, 48, 72, 96 h post-infection and stored at -80°C .

2. 6. Luciferase assay

For preparation of cell extracts for luciferase assays, BHK cells were washed with PBS and lysed by the addition of cell culture Reporter lysis buffer (Promega), followed by the incubation of cells at room temperature for 10 min. The cell extracts were then harvested and stored at -80°C . Luciferase assays were carried out using Luciferase Assay System (Promega) according to manufacturer instructions, and luminescence was determined using a Microplate Luminometer.

2. 7. Virulence in mice

Eight-week-old female C57BL/6J mice (Charles River Japan, Inc.) were challenged with 1,000 pfu of each virus subcutaneously in the dorsal region. The physical conditions of the mice were observed and the body weight was measured daily. All procedures were according to the guidelines of the Animal Care and Use Committee of the Hokkaido University.

2. 8. Molecular modeling

A homology model of OHFV RNA-dependent RNA polymerase (RdRp) domain was constructed based on the crystal structure of the dengue virus RdRp domain (PDB code, 2J7W,

sequence identity: ca 61%). MODELLER 9v6 (Eswar et al., 2003) was used for homology modeling of OHFV RdRp domains. After one hundred models of the RdRp domain were generated, a model was chosen by the MODELLER objective function value. After addition of hydrogen atoms, the model was refined by energy minimization (EM) using CHARMM force field with the Discovery Studio 2.1 software package (Accelrys, San Diego, CA). Steepest descent followed by conjugate gradient minimizations was carried out until the root mean square (rms) gradient was less than or equal to 0.05 kcal/mol/Å. The generalized Born implicit solvent model (Still et al., 1990; Tsui and Case, 2000) was used to model the effects of solvation. The molecular model was finally evaluated by using PROCHECK (Laskowski et al., 1993) and VERIFY-3D (Eisenberg et al., 1997). The model structure was displayed by PyMOL (DeLano Scientific LLC) (DeLano, 2002).

3. Results

3. 1. The construction of the full-length infectious clone of OHFV

The overall strategy to construct the full-length infectious clone of OHFV strain Guriev is outlined in Fig 1. The viral RNA was extracted from infected BHK cells, and individual dsDNA fragments were amplified by RT-PCR, as shown Fig 1. The six individual fragments were readily assembled into the low-copy plasmid pACNR, which has been used successfully to construct stable infectious clones in several flaviviruses (Bredenbeek et al., 2003; McElroy et al., 2005; Rossi et al., 2005). The complete genome sequence of this plasmid (designated OHF-IC-ori) was determined and compared with the parent virus. There were 10 nucleotide differences with three resulting in amino acid changes, one in NS2A and two in NS5 (Table 1). An additional nucleotide change was intentionally designed to create a *Cla*I site (C to G at nt 2439). This restriction site was used for the cloning of the DNA fragment between nt 135 and 2441. The other mutations were caused by PCR steps during the amplification of each fragment. These mutations were repaired by substitution as

described in *Materials and Methods*, and the infectious clone of OHFV, which is genetically identical to the parent virus Guriev, was obtained and designated as OHF-IC-pt. These OHF-IC plasmids were found to be stable during transformations into *E. coli* and large-scale plasmid production. After several bacterial passages, the sequence of the plasmid was identical.

3. 2. Generation of OHFV from infectious clone

XbaI-linearized OHF-IC template was used for *in vitro* RNA transcription using T7 RNA polymerase and the resulting full-length OHFV transcripts were transfected into BHK cells. At 5 days post-transfection, the transfected cells were fixed and the expression of virus proteins was analyzed by immunofluorescence assay. Most of the cells transfected with the transcripts from OHF-IC-pt showed a perinuclear signal when stained with an E-specific antibody (Fig 3A). Infectious virus could be harvested from the supernatants of the transfected cells, and a cytopathic effect was observed in these cell cultures. However no infectious virus could be recovered from cells transfected with RNAs from OHF-IC-ori, and no virus antigens were observed in the cells by immunofluorescence assay (Fig 3B). These data indicated two things: the production of viral proteins in the cells transfected with RNAs from OHF-IC-pt was not merely the result of translation of the input RNA, and the mutations in OHF-IC-ori affected the production of viral proteins and infectious viruses.

The growth properties of OHF-IC-pt derived virus and wild-type OHFV were analyzed by monitoring the release of virus after infection. BHK cells were infected at m.o.i. of 0.01 with OHFV. Virus was harvested at 24 h intervals and the yield was quantified by plaque assay (Fig 4). There was slight difference in the yields of virus at 24 h post-infection, but the resulting growth curves indicate similar growth properties between the parent virus and recombinant virus from OHF-IC-pt. The slight difference at 24 h post-infection may derive from the quasispecies of the parent virus stock because the virus was not plaque purified.

3. 3. Characterization of NS mutant

To determine how the mutations in OHF-IC-ori affected the viral production from the OHFV infectious cDNAs, we prepared several infectious cDNAs that incorporated the mutations identified in OHF-IC-ori (Fig 2). The OHF-IC-NS5₆₅₋₈₃₆, OHF-IC-NS5₆₅ and OHF-IC-NS5₈₃₆ plasmids have the NS5 mutations, and OHF-IC-NS2A₄₆ has the NS2A mutation. Recombinant viruses were recovered from these infectious cDNAs, and the growth curves were generated as described above.

No significant differences were observed between the growth curves of OHF-IC-pt and OHF-IC-NS5₆₅ (Fig 4), indicating that the amino acid difference at NS5 position 65 (L to P) did not affect virus growth in BHK cells. Meanwhile, the growth of OHF-IC-NS2A₄₆, OHF-IC-NS5₆₅₋₈₃₆ and OHF-IC-NS5₈₃₆ was restricted relative to OHF-IC-pt and OHF-IC-NS5₆₅. These results indicate that the mutations at NS2A position 46 (L to H) and NS5 position 836 (D to G) limited the virus growth in cell culture.

To further investigate the effect of the NS2A and NS5 mutations on virus production, we prepared a replicon of OHFV containing the luciferase reporter gene with or without the NS mutations (see Materials and Methods and Fig. 2). These replicon RNAs were transfected into BHK cells, and luciferase activities were measured at 6 h and 3 days post-transfection. There was no difference between the luciferase activities of the replicons at 6 h post-transfection (Fig. 5A), indicating that the mutations had no effect on the initial translation of reporter gene from transfected replicon RNA. At 3 days post-transfection, the luciferase activity was lower in the lysate from the replicons with mutations at NS2A position 46 or NS5 position 836 at 37 °C (Fig. 5B). In contrast, there was no decrease in luciferase activities was detected in the lysates from the replicon with and without the mutation at NS5 position 836 at 30 °C. Luciferase activity in cells transfected with the replicon containing the NS2A-46 mutation remained diminished in cells incubated at 30°C. Similar temperature sensitivity of NS5-836 was observed in the virus production at 30 °C (supplementary

figure 1). These data indicate that the mutations at NS2A-46 and NS5-836 reduce viral replication by reducing RNA replication and that the NS5-836 mutation produces a temperature-sensitive (ts) defect in RNA replication.

3. 4. Pathogenicity of infectious clone-derived viruses in mouse model

In our previous studies, we described that laboratory mice infected with OHFV showed clinical signs and pathology similar to reports of human infection (Holbrook et al., 2005; Tigabu et al., 2009). The pathogenicity of recombinant viruses was examined in the mouse model. Five adult C57BL/6 mice were infected subcutaneously with 1,000 p.f.u. of each virus and survival was recorded for 28 days (Fig. 6).

OHF-IC-pt virus which is genetically identical to parental OHFV Guriev strain produced 100 % mortality of mice (mean survival time 13.8 ± 1.92). All mice showed general signs of illness such as hunched posture, ruffled fur, and general malaise, but did not have significant or consistent indications of neurologic disease. After onset of disease, they showed a sharp decrease in body weight beginning at 9 and 10 dpi. When examined postmortem, all mice had conjunctival suffusion with crusting and some had bowel hemorrhage. These results were consistent with our previous data in which mice infected with parental OHFV Guriev strain had viscerotropic disease with limited signs of neurological disease (100 % mortality and mean survival time 12.8 ± 2.49)(Tigabu et al., 2009).

The OHF-IC-NS5₆₅ virus had similar virulence to the OHF-IC-pt virus (100 % mortality and mean survival time 14.2 ± 1.92) indicating that the amino acid difference at NS5 position 65 did not affect the biological properties of OHFV confirming observations from *in vitro* studies. However, the OHF-IC-NS2A₄₆, OHF-IC-NS5₆₅₋₈₃₆ and OHF-IC-NS5₈₃₆ virus showed decreased virulence. The OHF-IC-NS5₆₅₋₈₃₆ and OHF-IC-NS5₈₃₆ virus killed three mice, but survival time was increased while the OHF-IC-NS2A₄₆ virus was completely attenuated. The NS5-836 mutation was retained in

the dead mice inoculated with The OHF-IC-NS5₆₅₋₈₃₆ and OHF-IC-NS5₈₃₆ virus. All surviving mice inoculated with the OHF-IC-NS2A₄₆, OHF-IC-NS5₆₅₋₈₃₆ and OHF-IC-NS5₈₃₆ virus showed no signs of illness and no weight loss. These results indicate that the mutations at NS2A position 46 and NS5 position 836 affect OHFV virulence in association with the lower viral replication. All surviving mice inoculated with OHF-IC-NS2A₄₆, OHF-IC-NS5₆₅₋₈₃₆ and OHF-IC-NS5₈₃₆ had neutralizing antibody against OHFV (≥ 320 in 50% reduction), indicating that the virus was able to replicate at the initial stage of infection without causing a disease.

4. Discussion

This is the first report of generation of an infectious clone of OHFV. Infectious clones can be a valuable tool for studying the molecular biology of virus replication, virus structure, virulence determinants, and vaccine development. There is a common difficulty in the construction of full-length clones of flaviviruses, because the plasmids containing a full-length cDNA of these viruses are often unstable during propagation in *E. coli*. Therefore, by using low-copy-number plasmids and specific bacterial hosts, stable full-length infectious clones have been developed for several flaviviruses (Gritsun and Gould, 1998; Hayasaka et al., 2004; Kinney et al., 1997; Mandl et al., 1997; Shi et al., 2002; Yamshchikov et al., 2001; Yun et al., 2003). In this study, the low-copy-number plasmid pACNR was used for the construction of the full-length OHFV cDNA. This vector has been used to construct stable infectious clones of several flaviviruses and pestiviruses (Bredenbeek et al., 2003; McElroy et al., 2005; Mendez et al., 1998; Ruggli et al., 1996). The OHF-IC plasmids were stable during passaging in bacteria, indicating that this infectious clone can be useful for genetic manipulations.

The Guriev strain, which was isolated from human blood, was selected to construct a full-length

infectious clone. We previously demonstrated the similarity between human and murine infection with OHFV (Holbrook et al., 2005; Tigabu et al., 2009). The OHFV-infected mice had no indication of neurological problems, and had conjunctival suffusion that has also been reported in human cases. The recombinant virus which is genetically identical to the parental Guriev strain showed similar biological properties to the parental virus, including growth kinetics and virulence characteristics. These results indicate that an efficient reverse genetics system has been established for OHFV.

Our results have identified two amino acid codon substitutions associated with attenuation of the virus production, i.e., Leu46 to His in the NS2A, and Asp836 to Gly in NS5. These substitutions decreased the efficiency of RNA replication, leading to limited virus propagation and decreased virulence in mice.

Flavivirus NS2A is a small, hydrophobic, membrane associated protein involved in RNA replication. It was reported that NS2A binds with high specificity to the 3' untranslated region (UTR) of viral genomic RNA and to other components of the replication complex (Mackenzie et al., 1998). In addition, NS2A is considered to play roles in modulating the host-antiviral interferon response (Liu et al., 2004; Liu et al., 2006; Liu et al., 2005; Munoz-Jordan et al., 2003) and assembly/ secretion processes of virus particles (Kummerer and Rice, 2002; Leung et al., 2008; Liu et al., 2003). Although the exact membrane topology of NS2A is yet to be determined, NS2A of OHFV has been predicted to span the membrane of the endoplasmic reticulum five times by several transmembrane domain prediction programs (TMHMM (Krogh et al., 2001), and TMPred (Hofmann and Stoffel, 1993)). The Leu46 residue in NS2A is located in the conserved hydrophobic residues of the predicted 2nd transmembrane region. It is possible that the Leu46 to polar His substitution affects the membrane spanning domain and the interaction with other transmembrane domains of NS2A or other membrane-associated viral proteins. A change in membrane associated protein topology could lead to a defect in the replication properties of NS2A such as the formation of the replication complex by binding with viral RNA and other components of the replication

complex.

NS5 is the largest (104kDa) of the flavivirus proteins, and three functional domains have been identified in NS5: a *S*-adenosylmethionine methyltransferase-like domain in the N-terminal region (Egloff et al., 2002; Koonin, 1993; Ray et al., 2006), a centrally located nuclear localization sequence (NLS) (Forwood et al., 1999; Kapoor et al., 1995), and an RNA-dependent RNA polymerase (RdRp) domain in the C-terminal region (Bartholomeusz and Wright, 1993; Koonin, 1991). The Asp836 residue in NS5 is located in the RdRp domain. In the homology model of OHFV RdRp domain based on the crystal structure of the dengue virus RdRp domain, this residue is within the α 25 helix in the Thumb domain (Yap et al., 2007). In position 836 of the NS5 RdRp domain, negatively-charged amino acids aspartic acid or glutamic acid are highly conserved among most flaviviruses. The molecular model suggests that the negatively-charged Asp836 might form a salt bridge with the positively-charged Lys720 in Motif E of the RdRp domain (Fig. 7). Molecular mechanics calculations also showed that the Asp836 to Gly substitution significantly reduced the interaction of the residue at position 836 with the Lys720 and Motif E (Supplementary Table1). These results led to the hypothesis that the reduction of interaction due to Asp836 to Gly substitution causes structural fluctuation, especially in Motif E. Motif E forms an antiparallel β -sheet wedged between the palm domain and several α -helices of the thumb domain. In several studies, it has been shown that some of the residues in Motif E are involved in the GTP-binding site and which have essential roles in *de novo* initiation of RNA synthesis in *Flaviviridae* polymerases (Choi et al., 2004; Lai et al., 1999; Yap et al., 2007). It is possible that residue Asp836 is important for the structural stability of Motif E, and that the structural fluctuation of Motif E caused by the Asp836 to Gly substitution leads to a reduction in the efficiency of *de novo* initiation of RNA synthesis. In general, the conformational fluctuation of proteins is associated with their temperature. Therefore, this thermal fluctuation could be a possible cause of the temperature-sensitive property observed in the replicon studies (Fig 5). Temperature dependence has been reported at the initiation,

but not elongation, phase of *de novo* RNA synthesis by dengue virus RdRp (Ackermann and Padmanabhan, 2001). These data support the involvement of the interaction between Asp836 and Motif E in *de novo* initiation of RNA synthesis.

In summary, we have constructed an infectious cDNA clone of OHFV and demonstrated the utility of this clone in the research of OHFV pathogenesis. We have also identified previously unknown mutations in NS2A and NS5 that appear to play important roles in OHFV RNA synthesis.

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Figure legends

Figure 1

Construction of the full-length infectious clone of OHFV. Six cDNA fragments synthesized by RT-PCR were assembled to form the full-length cDNA clone of OHFV (OHF-IC). Restriction sites used to construct OHF-IC are shown at the bottom. A silent mutation (shown in uppercase) was engineered to create a *Cla* I site (*). The complete OHFV cDNA is positioned under the control of the T7 promoter. A hepatitis delta virus ribozyme (HDV-RZ)/bacteriophage T7 terminator fragment was fused to the 3' end of the viral genome (see Materials and Methods).

Figure 2

Schematic representation of the genome of OHFV showing all of the amino acid coding difference between strain Guriev and constructed full-length clones (OHF-IC) and OHFV replicons used to analyze the effect of NS2A and NS5 mutations. Bold type has been used to designate the amino acids of the consensus sequence in Guriev (see text and Table 1).

Figure 3

Immunofluorescence staining of BHK cells transfected with the *in vitro* transcript of OHF-IC-pt (A) and OHF-IC-ori (B). Cells were fixed at 5 days post-transfection and stained with MAb 1H4.

Figure 4

Growth curves of parental Guriev, OHF-IC-pt, OHF-IC-NS5₆₅, OHF-IC-NS5₈₃₆, OHF-IC-NS5₆₅₋₈₃₆, and OHF-IC-NS2A₄₆. A Monolayer of BHK cells was infected with the individual viruses at a multiplicity of infection (MOI) of 0.01. At each time point, the media was harvested and virus titers were determined by plaque assay in BHK cells.

Figure 5

Effect of NS mutations on OHFV RNA replication. BHK cells were transfected with luciferase-expressing OHFV subgenomic replicon RNAs (OHF-REP-luc) with or without the NS2A or NS5 mutations, and incubated at 37 ° C or 30 ° C . Luciferase activities were measured at 6h (A) 72 h (B) post-transfection. Luciferase activities are expressed in Raw Light Units (RLU). Asterisks show the statistically significant difference compared to OHF-REP-luc-pt by Student T test (P<0.05).

Figure 6

Survival of mice inoculated with OHF-IC. Mice were inoculated subcutaneously with 1,000 p.f.u. of OHF-IC-pt, OHF-IC-NS5₆₅, OHF-IC-NS5₈₃₆, OHF-IC-NS5₆₅₋₈₃₆, and OHF-IC-NS2A₄₆.

Figure 7

Location of amino acid substitutions at Asp836 on a homology model of the OHFV RNA-dependent RNA polymerase domain in NS5 based on that of dengue virus (PDB 2J7W). The zoomed region in the right panel shows the proposed interaction of charged side chains between Asp836 and Lys720.

Figure 1

OHF-IC

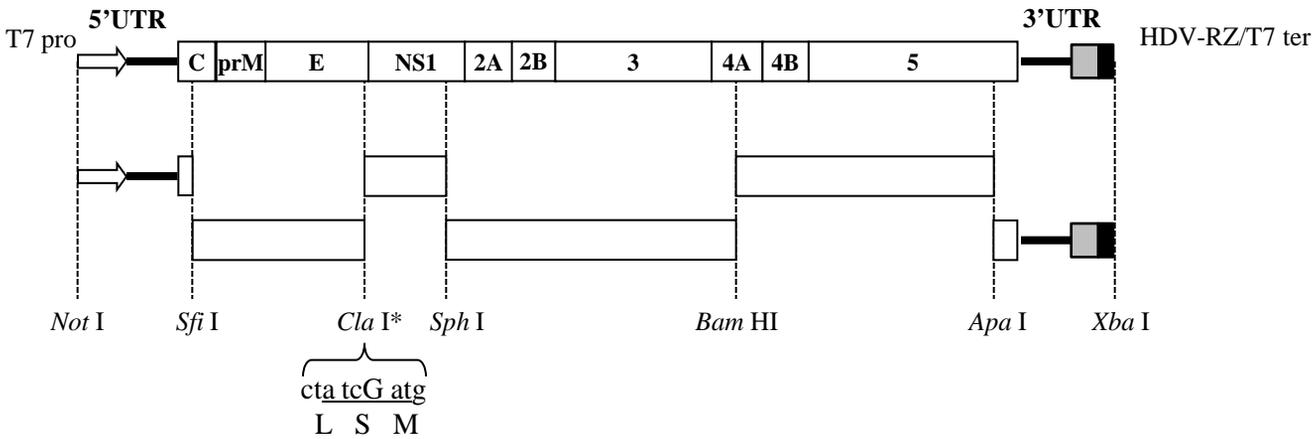
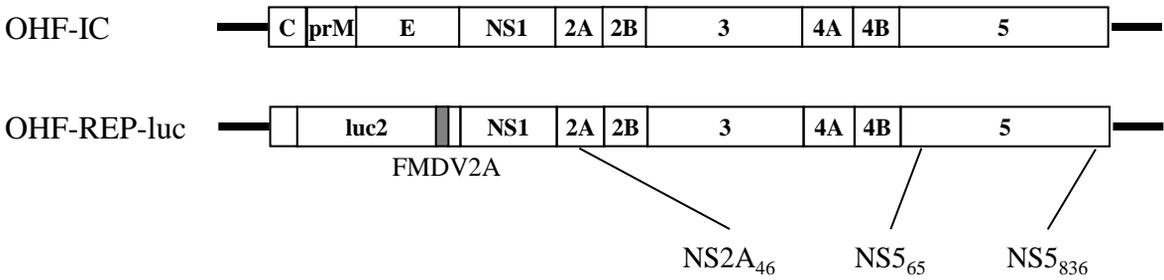


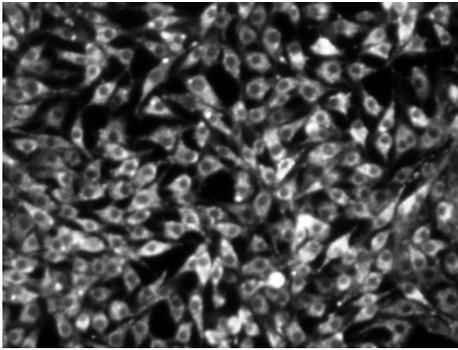
Figure 2



Parental virus (strain Guriev)	<u>L</u>	<u>L</u>	<u>D</u>
OHF-IC-ori	H	P	G
OHF-IC-NS5 ₆₅₋₈₃₆ OHF-REP-luc-NS5 ₆₅₋₈₃₆	<u>L</u>	P	G
OHF-IC-NS5 ₆₅ OHF-REP-luc-NS5 ₆₅	<u>L</u>	P	<u>D</u>
OHF-IC-NS5 ₈₃₆ OHF-REP-luc-NS5 ₈₃₆	<u>L</u>	<u>L</u>	G
OHF-IC-NS2A ₄₆ OHF-REP-luc-NS2A ₄₆	H	<u>L</u>	<u>D</u>
OHF-IC-pt OHF-REP-luc	<u>L</u>	<u>L</u>	<u>D</u>

Figure 3

A. OHF-IC-pt



B. OHF-IC-ori



Figure 4

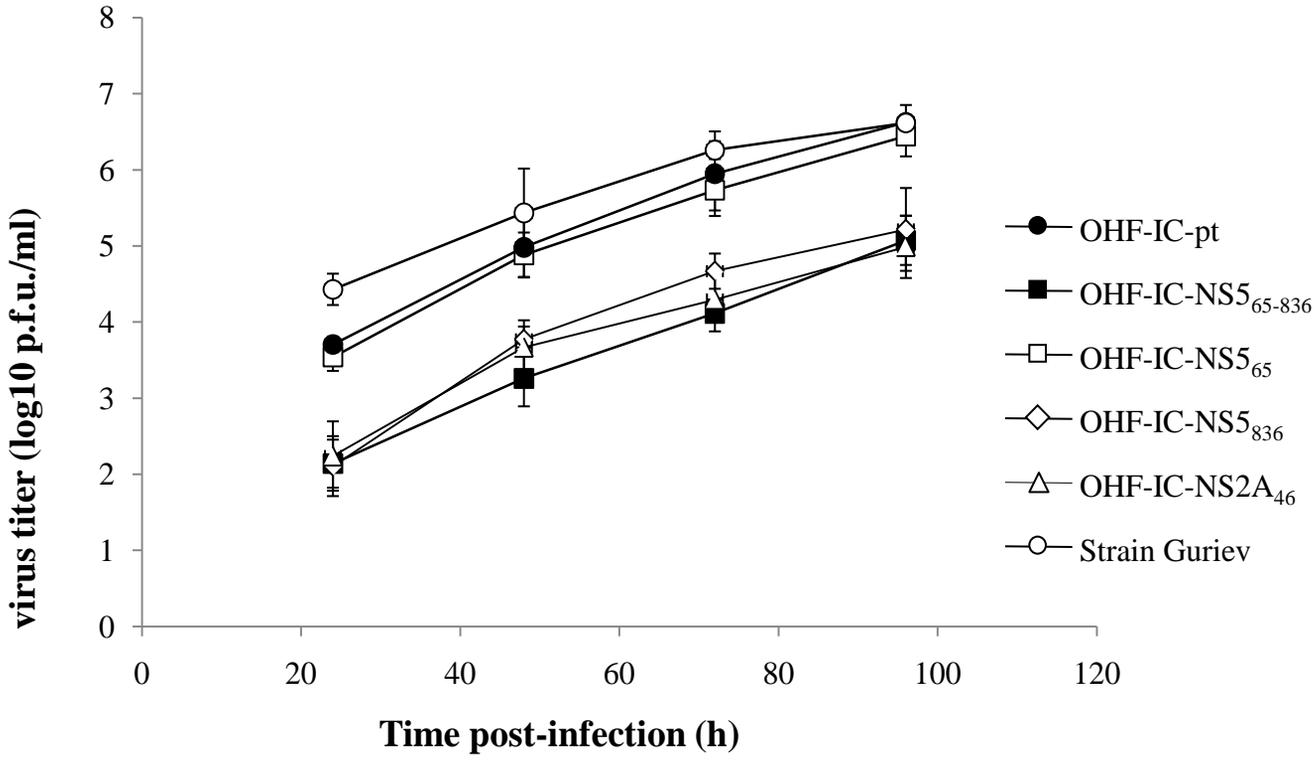


Figure 5

A) 6 hour

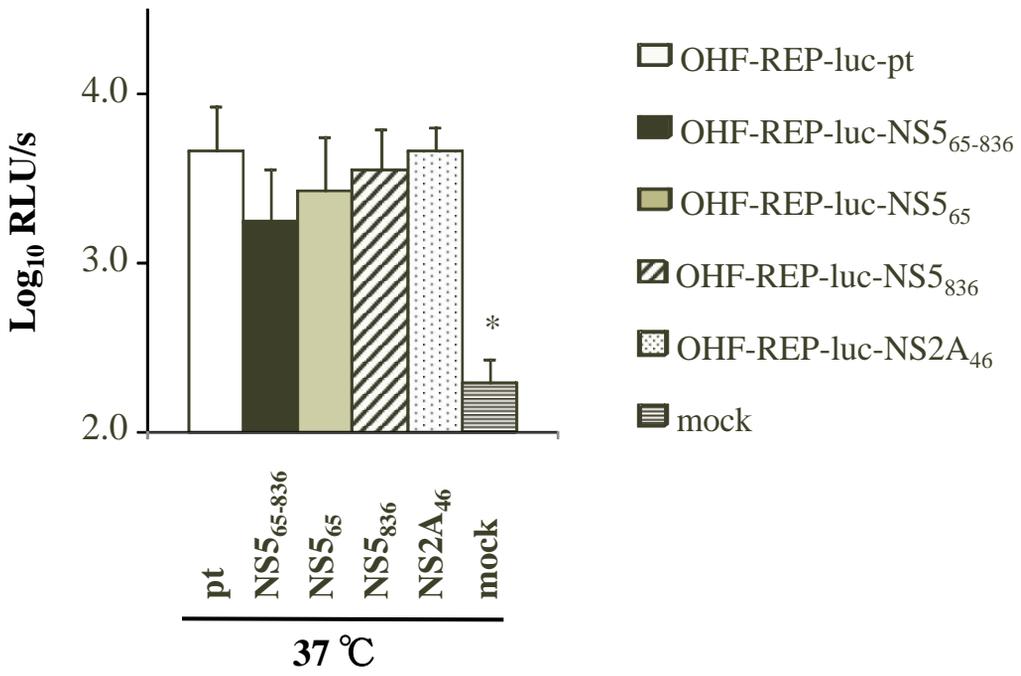


Figure 5

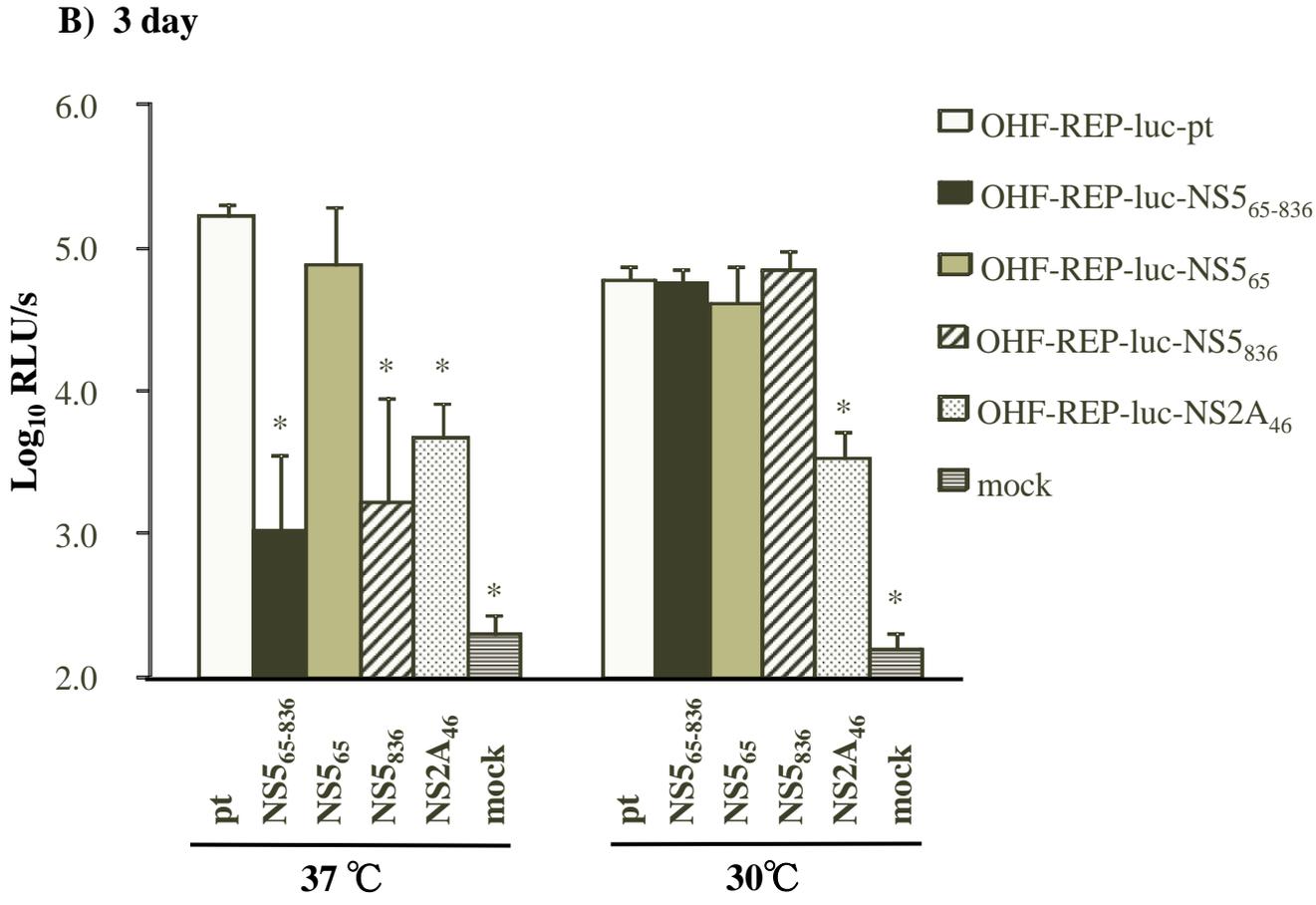


Figure 6

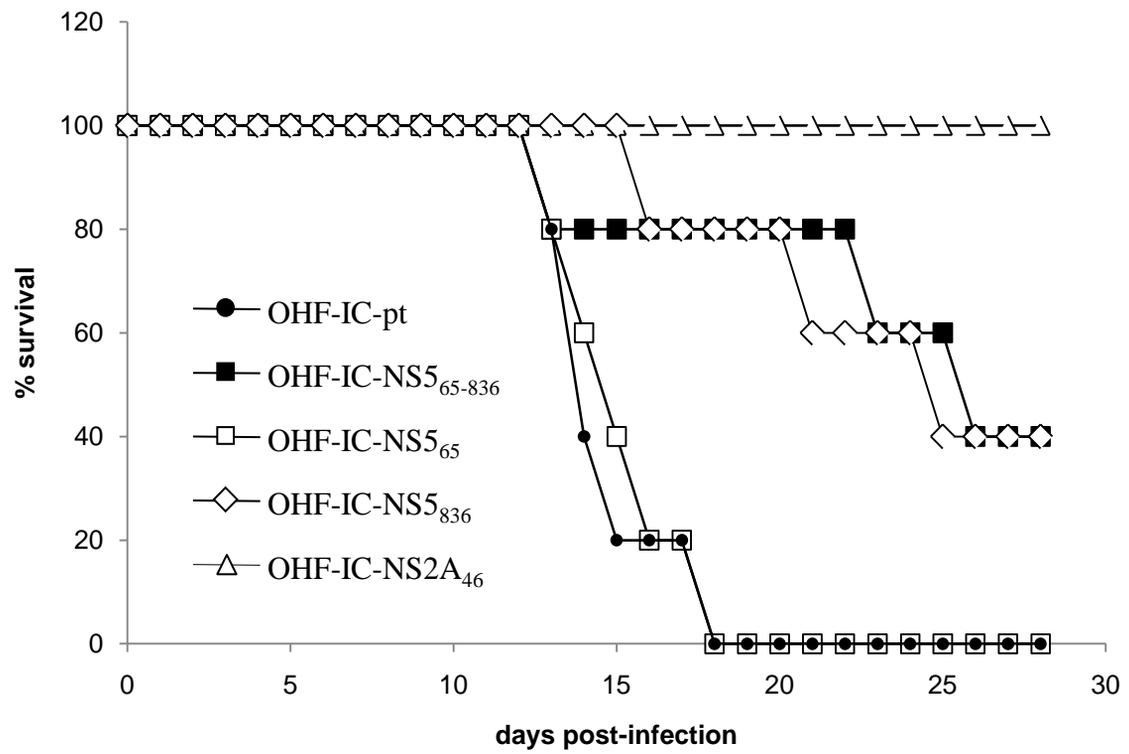


Figure 7

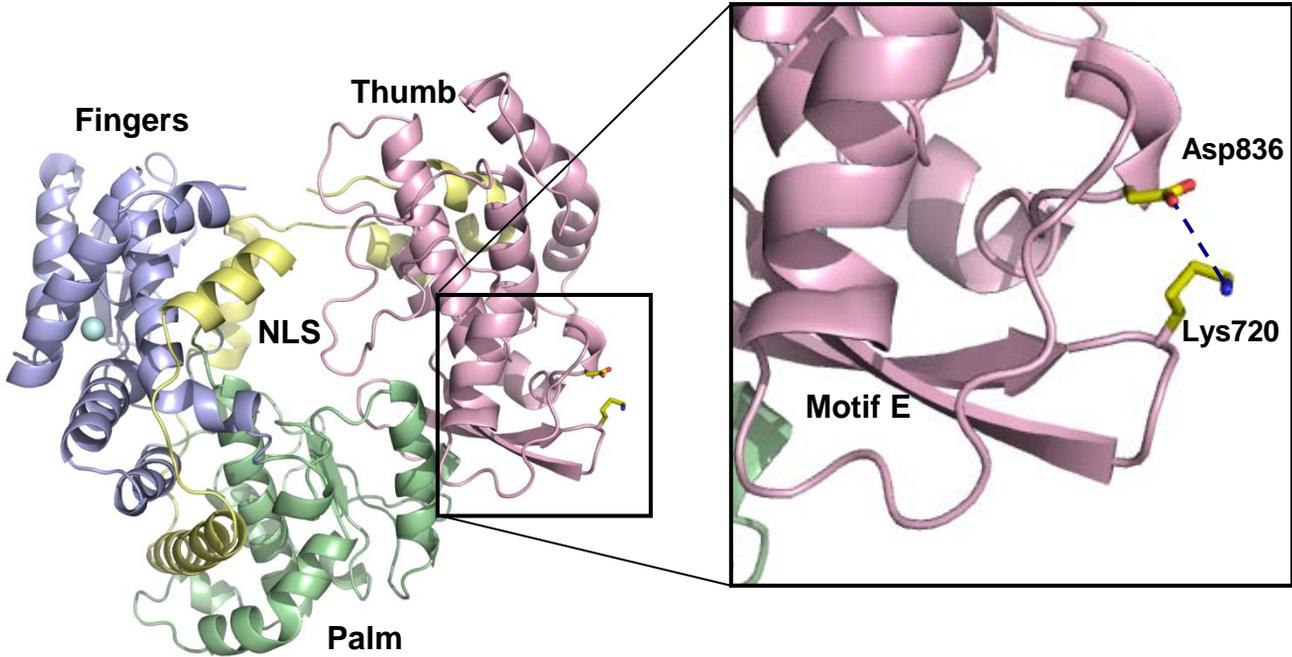


Table 1. Summary of sequence difference between the infectious cDNA OHF-IC-ori and parental OHFV strain Guriev

Base position ^a	Strain Guriev	OHF-IC-ori	Amino acid	Location
297	A	G	-	
2367	G	A	-	
2439	C	G ^b	-	
3653	T	A	L → H	NS2A ₄₆ ^c
6231	C	T	-	
6591	A	T	-	
6678	T	C	-	
7859	T	C	L → P	NS5 ₆₅
7983	A	G	-	
10172	A	G	D → G	NS5 ₈₃₆

^a Nucleotide position and sequence are based on OHFV strain Guriev (Accession no. AB507800)

^b This silent mutation (shown in uppercase) was engineered to create a *Cla* I site (*).

^c The numbers indicate the amino acid position in each protein