



Title	Structure of bacterial cellulose synthase subunit D octamer with four inner passageways
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## **Supporting Information**

### **Text**

#### ***Bacterial strains and growth conditions***

*Escherichia coli* (*E. coli*) JM109 and BL21(pREP-4) were grown in Luria-Bertani (LB) broth at 37°C. Isopropyl- $\beta$ -D(-)-thiogalactopyranoside (IPTG) was used as an inducer of genes. When needed, the antibiotics ampicillin (Ap) and kanamycin (Km) were added to the medium at final concentrations of 100 and 25  $\mu$ g/mL, respectively. The above media with 1.5% (w/v) agar were used for cell growth on plates.

#### ***Preparation of His<sub>6</sub>-tagged AxCeSD***

The expression and purification of selenomethionine (SeMet)-substituted AxCeSD with a His<sub>6</sub>-tag at the N-terminus (N\_AxCeSD) were carried out as described previously (1). Using primer pairs corresponding to the specified 5' and 3' termini (Tab. S2), the PCR products were digested with *NcoI/HindIII*, and then cloned into pET-28b (Novagen, San Diego, CA, USA) to construct AxCeSD with a His<sub>6</sub>-tag at the C-terminus. AxCeSD with a His<sub>6</sub>-tag at the N-terminus (N\_AxCeSD) or C-terminus (C\_AxCeSD) was prepared by the His<sub>6</sub>-tag system as described previously (1).

#### ***Crystallization and data collection***

The crystallization and data collection of N\_AxCeSD were performed as described

previously (1). Single crystals of C\_AxCeSD were grown at 20 °C for 1-2 weeks by the hanging drop vapor diffusion method. Each drop was prepared by mixing 2.5  $\mu$ L of protein solution (10 mg/mL) and the same volume of reservoir solution containing 0.2 M  $\text{Li}_2\text{SO}_4$ , 0.1 M phosphate citrate (pH 5.4), and 10% (v/v) isopropanol. The diffraction data were collected on the in-house X-ray diffraction equipment of R-Axis 4<sup>++</sup> (Rigaku, Tokyo, Japan) at -173 °C after crystals were soaked into reservoir solution buffer containing 20 % glycerol. Crystals of C\_AxCeSD-CPT complex were obtained by soaking native C\_AxCeSD crystals in the reservoir solution containing 3 mM CPT for 140 min then moved into cryoprotectant buffer (reservoir solution containing 3 mM CPT and 20 % glycerol) before data collection on beamline BL41XU (SPring-8, Japan). All diffraction data were indexed, integrated, scaled, and merged with the program *HKL2000* (2). Crystallographic parameters and data collection statistics are shown in Table S1..

### ***Small angle X-ray scattering***

Small-angle X-ray scattering (SAXS) measurements were carried out at SPring-8 beamline 40B2 of Japan (3). A wavelength of 1.0 Å was used, and the specimen-to-detector distance was 2 m. The condition of data collection was determined to use 1.75 mg/mL of N\_AxCesD with an exposure time of 60 s at room

temperature. The SAXS data were normalized to the intensity of the incident beam and processed for background subtraction using the standard procedures with the program package *PRIMUS* (4). The  $R_g$  volume and the discrepancies between the calculated and experimental scattering curves were calculated and minimized using the program *CRY SOL* (5) as described previously (6, 7).

### ***Preparation of *axcesD* gene deletion mutant strain (DBCD)***

An *axcesD* gene deletion mutant strain of *A. xylinum* ATCC 23769 was prepared by homologous recombination with the ampicillin resistance gene used as a marker gene. Preparation of a plasmid to delete the *axcesD* gene was performed according to the procedure reported by Saxena *et al.* (8). Deletion of the *axcesD* gene was confirmed by PCR using SP(bcsD) and AP(bcsD) as a set of specific primers (Tab. S2) and Western-blotting analysis. A band of an amplicon with larger molecular weight than that of native *axcesD* gene (Fig. S5a, lane 2) was observed when a genomic DNA from a candidate of *axcesD* gene deletion mutant strain was used as a template of the PCR (Fig. S5a, lane 3), suggesting that an antibiotic-resistant gene was inserted into the genomic DNA of the candidate. A protein band corresponding to AxCeSD was not observed in the sample prepared from the candidate of *axcesD* gene deletion mutant strain (Fig. S5b, lane 3). From these results, we concluded that the *axcesD* gene was

deleted in the candidate strain, which was designated as DBCD.

## References

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operon (acs operon) of *Acetobacter xylinum*: implications for cellulose crystallization. *J Bacteriol* **176**, 5735-5752.

## Figure Legends

### Figure S1. Molecular weight determination of AxCeSD in solution

(a) The result of the gel filtration experiment using a column of Hi-load 26/60 Superdex 200 (Amersham Biosciences) (b) The standard molecules were Vitamin B<sub>12</sub> (13.5 kDa), Myoglobin (17 kDa), Ovalbumin (44 kDa),  $\gamma$ -Globulin (158 kDa), and Thyroglobulin (670 kDa). The calculated molecular weight of AxCeSD in solution was 145.6 kDa, corresponding to AxCeSD octamer.

### Figure S2. Plot of small angle X-ray scattering

The logarithm of the scattering intensity of N\_AxCeSD ( $R_g = 34.4 \text{ \AA}$ ) is plotted by black dots against the momentum transfer  $S = 4\pi \sin\theta/\lambda$ , where  $2\theta$  is the scattering angle and  $\lambda = 1.0 \text{ \AA}$  is the wavelength. The red ( $R_g = 34.4 \text{ \AA}$ ,  $\chi = 0.085$ ) and green ( $R_g = 34.0 \text{ \AA}$ ,  $\chi = 0.059$ ) curves are theoretical data calculated from crystal structure of octamer N\_AxCeSD and full-length octamer N\_AxCeSD model (with His<sub>6</sub>-tag and linker of 5 residues), respectively.

### Figure S3. Interaction between dimers

A ribbon representation of the AxCeSD structure is shown from the side view with each monomer A (labeled with \*), B (labeled with \*'), C (labeled), and D (unlabeled) in blue, red, green, and cyan, respectively. The secondary structures contributing to

dimer – dimer interaction are labeled in the same color of their ribbon model.

**Figure S4.** The CPTs with omit maps from the inner view of the AxCeSD cylinder

The four copies of C\_AxCeSD (A, B, C, and D) are shown in ribbon representation in blue, red, green, and cyan, respectively (the same in all structure figures in this paper).

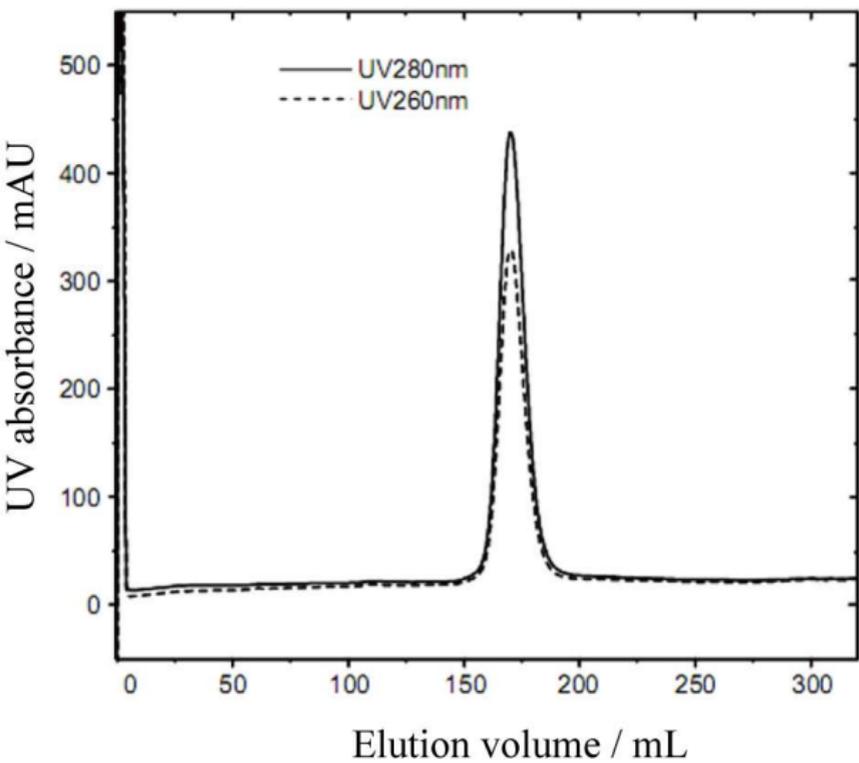
The CPTs are shown as stick models (oxygen atoms: red, carbon atoms: yellow or orange), and omit maps are contoured at  $1.6 \sigma$ . One of two CPTs in a pair is shown in (a) and the other is shown in (b). (c) A pair of CPTs.

**Figure S5** Confirmation of *axcesD* gene deletion

(a) The result of PCR for checking *axcesD* gene deletion by insertion of antibiotic-resistant gene into a genomic DNA. (b) The result of Western-blotting for checking *axcesD* gene deletion. The lanes are same in both experiments, Lane 1: Marker; Lane 2: wild-type; Lane 3: DBCD.

Figure S1

(a)



(b)

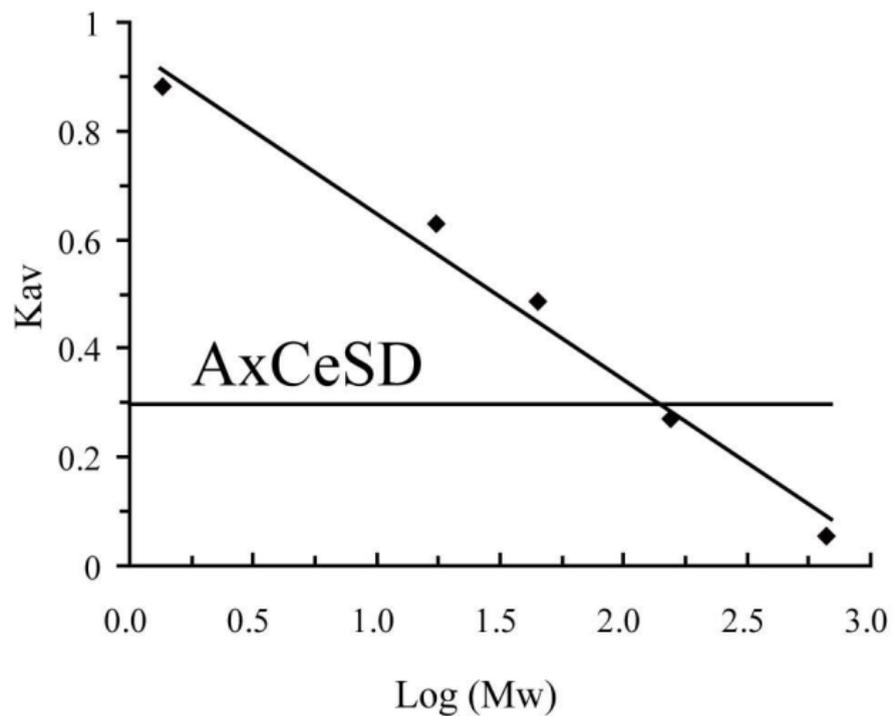


Figure S2

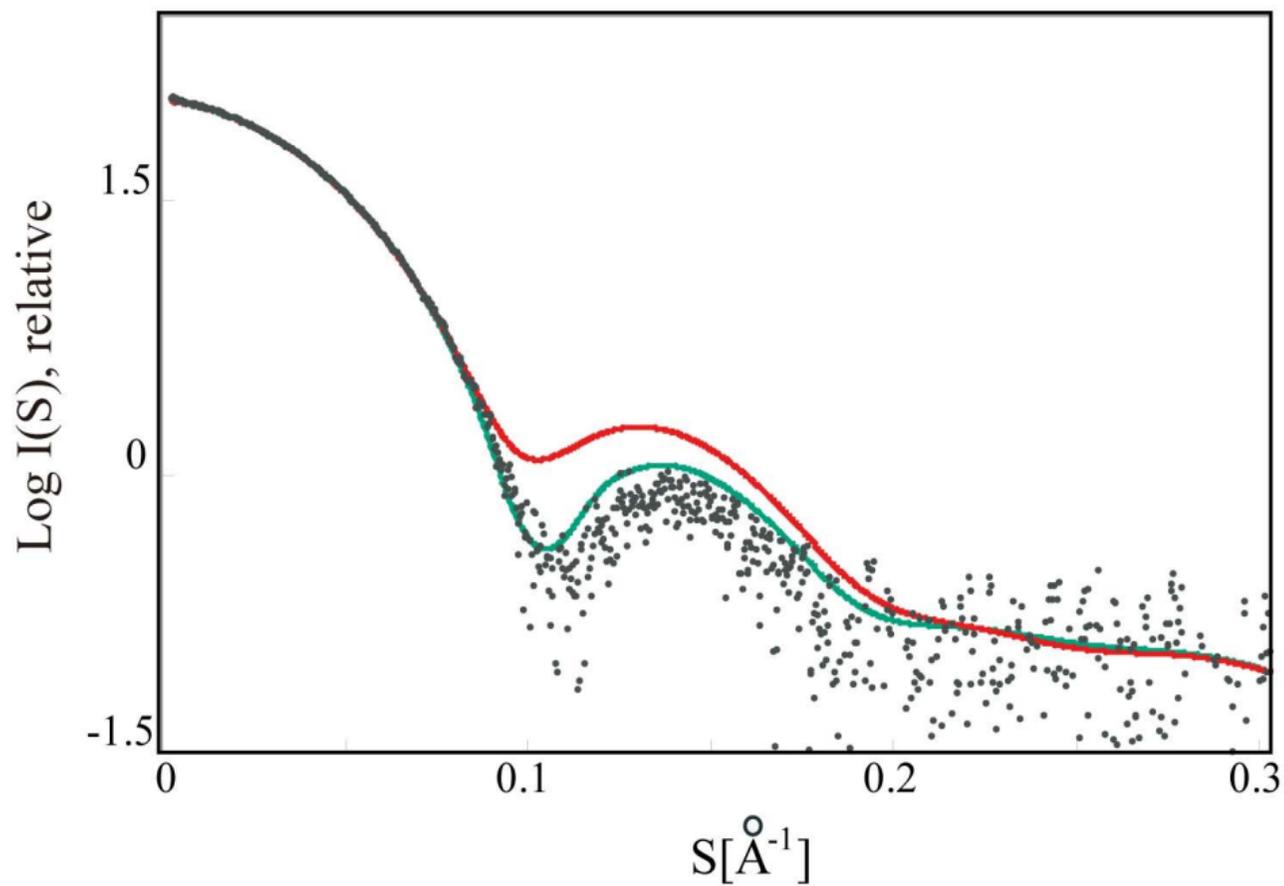


Figure S3

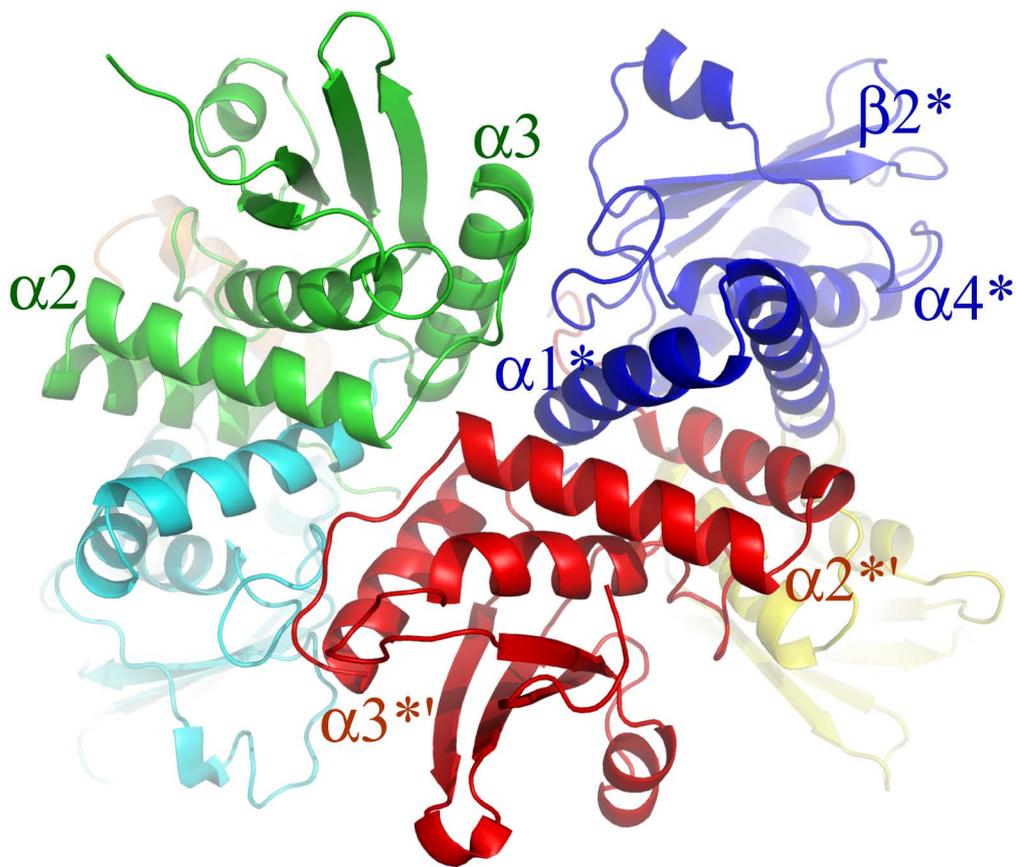
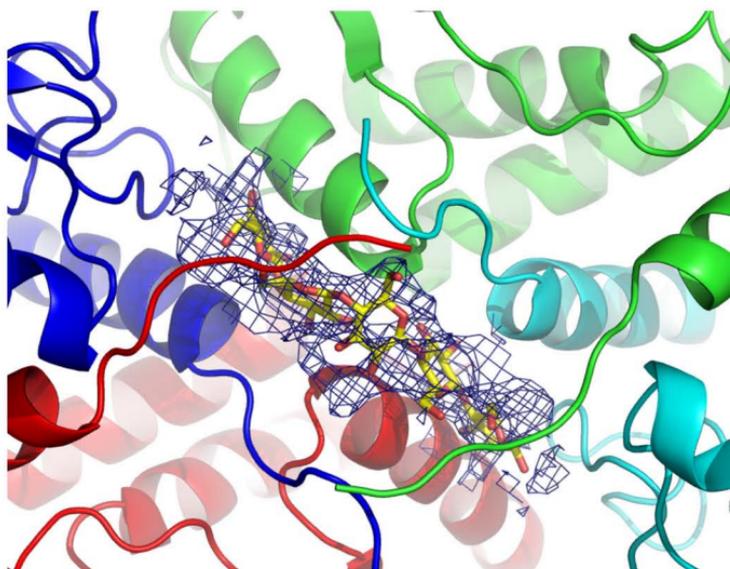
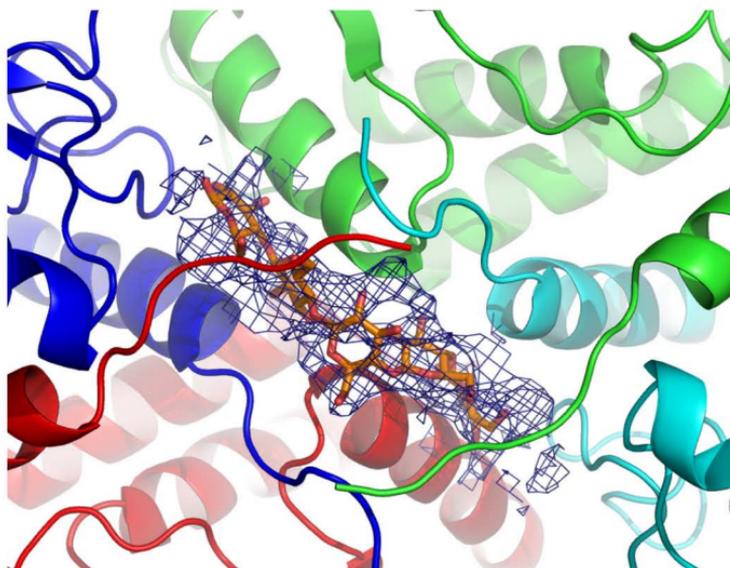


Figure S4

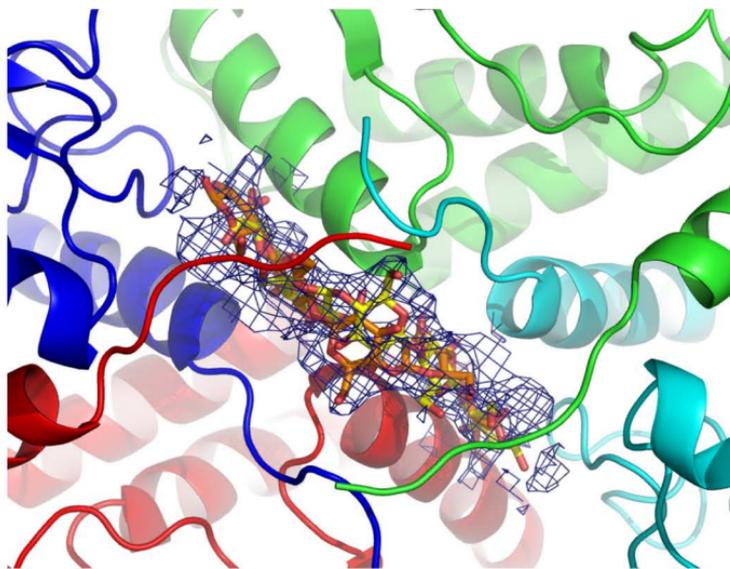
(a)



(b)

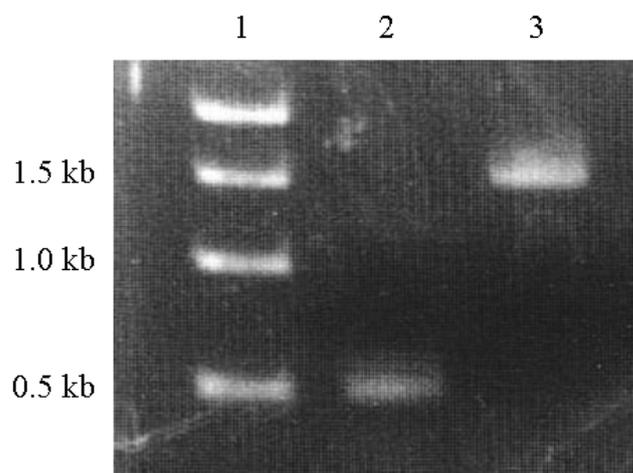


(c)



**Figure S5**

(a)



(b)

