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Involvement of CD4⁺ Foxp3⁺ Regulatory T Cells in Persistence of Leishmania donovani in the Liver of Alymphoplastic aly/aly Mice

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Abstract

Visceral leishmaniasis (VL) is a chronic and fatal disease in humans and dogs caused by the intracelluar protozoan parasites, Leishmania donovani and L. infantum (L. chagasi). Relapse of disease is frequent in immunocompromised patients, in which the number of VL cases has been increasing recently. The present study is aimed to improve the understanding of mechanisms of L. donovani persistence in immunocompromised conditions using alymphoplastic aly/aly mice. Hepatic parasite burden, granuloma formation and induction of regulatory T cells were determined for up to 7 months after the intravenous inoculation with L. donovani promastigotes. While control aly/+ mice showed a peak of hepatic parasite growth at 4 weeks post infection (WPI) and resolved the infection by 8 WPI, aly/aly mice showed a similar peak in hepatic parasite burden but maintained persistent in the chronic phase of infection, which was associated with delayed and impaired granuloma maturation. Although hepatic CD4⁺Foxp3⁺ but not CD8⁺Foxp3⁺ T cells were first detected at 4 WPI in both strains of mice, the number of CD4⁺Foxp3⁺ T cells was significantly increased in aly/aly mice from 8 WPI. Immunohistochemical analysis demonstrated the presence of Foxp3⁺ T cells in L. donovani–induced hepatic granulomas and perivascular neo-lymphoid aggregates. Quantitative real-time PCR analysis of mature granulomas collected by laser microdissection revealed the correlation of Foxp3 and IL-10 mRNA level. Furthermore, treatment of infected aly/aly mice with anti-CD25 or anti-FR4 mAb resulted in significant reductions in both hepatic Foxp3⁺ cells and parasite burden. Thus, we provide the first evidence that CD4⁺Foxp3⁺ Tregs mediate L. donovani persistence in the liver during VL in immunodeficient murine model, a result that will help to establish new strategies of immunotherapy against this intracellular protozoan pathogen.


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Introduction

Visceral leishmaniasis (VL) is a chronic and fatal disease caused by the intracellular protozoan parasites Leishmania donovani and L. infantum (L. chagasi), which infect a range of mammalian hosts, including humans, dogs and rodents [1]. Liver, spleen, bone marrow (BM) and lymph nodes are the major sites for parasite growth and disease pathology. Transplantation of infected kidney, liver, heart, lung, pancreas or BM has been shown to cause VL in transplant recipients, indicating lifelong parasite persistence in the viscera [2]. Moreover, malnutrition is a risk factor for the development of VL [3]. Recent experiments in protein energy-, zinc- and iron-deficient mice suggest that this effect is mediated primarily through functional failure of the lymph node barrier and increased early visceralization of the parasites [4–6]. Loss of the control of parasite persistence in VL causes the reactivation of parasites and relapse of the disease is frequent in the immunocompromised patients, in which the number of visceral leishmaniasis cases has been increasing recently [7]. However, the mechanisms underlying the parasite persistence in the immunocompromised condition have not been clearly clarified. To develop effective prophylactic or therapeutic strategies against VL, understanding of the precise immune mechanisms including T-cell functions in the chronic stage of infection is required [8].

The role of secondary lymphoid organs for immune responses to Leishmania infection has not been investigated. The aly/aly mouse is an autosomal recessive natural mutant C57BL/6 strain that carries a point mutation within the gene encoding NF-kB inducing kinase (NIK) [9], which prevents the induction of the non-canonical NF-kB pathway [10]. The aly/aly mice lack all lymph nodes and Peyer’s patches with the abnormal architecture of spleen and thymus and exhibit severely impaired humoral response [9]. This mutant mouse strain has been used to examine...
Author Summary

The protozoan parasite Leishmania donovani is the causative agent of visceral leishmaniasis (VL) with a variety of outcomes ranging from asymptomatic to fatal infection. In the last decade, an increasing number of VL cases in immunocompromised conditions have been reported. Loss of the control of parasite persistence causes relapse of the disease in these patients. To clarify why parasite persistence and disease are caused in an immunocompromised condition, we examined L. donovani infection in aliphosphatic aly/aly mice that completely lack lymph nodes and have disturbed spleen architecture. Although parasites grew in the liver of aly/+ mice for the first 4 weeks post infection (WPI) and parasites were eliminated by 8 WPI, we found that parasites persisted in the liver of aly/aly mice with the ineffective of granuloma formation to kill the parasites. These aly/aly mice showed significant increases in CD4+Foxp3+ regulatory T cells in the liver. Consequently, we treated infected mice with anti-CD25 or anti-FR4 mAb to inhibit the function of Tregs, and found significant reductions in both hepatic Foxp3+ cells and parasite burden. These results clearly demonstrated for the first time that the expansion of CD4+Foxp3+ Tregs is involved in hepatic L. donovani persistence in immunodeficient murine model.

Materials and Methods

Mice, Parasites and Infection

Female ALY® NscJcl aly/aly and aly/+ mice of 6–8 weeks of age were purchased from CLEA Japan, Inc. (Tokyo, Japan). Mice were maintained, inoculated and sacrificed within a safety facility of Hokkaido University.

A virulent line of L. donovani (MHOM/SU/62/2S-25M-C2) [22] was maintained by passage of the frozen stabilized parasites in NNN medium containing 5% defibrinated hemolized rabbit blood. Then, parasites were consecutively sub-passaged in liquid M199 medium supplemented with 15% heat-inactivated fetal calf serum (HIFCS), 25 mM HEPES and 50 μg/ml gentamycin. The stationary growth phase of subcultures with less than five passages was used for mouse inoculation. Mice were infected by injecting stationary phase promastigotes (5 × 10⁵) intravenously via the lateral tail vein and were sacrificed at 1, 2, 4, 8, 12, 16 and 28 weeks post infection (WPI). One group of non-infected animals was used as naive control.

Ethics Statement

This study was carried out under the guidance of the Institute for Laboratory Animal Research (ILAR), All animals were housed in a facility in strict accordance with the recommendations in the Guidelines for the Care and Use of Laboratory Animals of Graduate School of Veterinary Medicine, Hokkaido University, which was based on Fundamental Guidelines for Proper Conduct of Animal Experiment and Related Activities in Academic Research Institutions under the jurisdiction of the Ministry of Education, Culture, Sports, Science and Technology, Japan and approved by the American Association for Accreditation of Laboratory Animal Care (AAALAC) international. The protocol was approved by the Committee on the Ethics of Animal Experiments of Hokkaido University (Permit Number: 10-0009).

Determination of Parasite Burdens by LDU in the Liver and qPCR in Different Tissues

Giemsa-stained impression smears of the liver were prepared and parasite burden was determined as Leishman-Donovan Units (LDU), in which LDU is the number of amastigotes per 1,000 host nuclei, multiplied by the liver weight in gram [23].

Genomic DNA (gDNA) was isolated from different tissues, including liver, spleen, BM, blood, heart, lung, kidney, brain and skin, using the QiAamp® DNA Mini Kit (Qiagen, MA, USA). Real-time quantitative (qPCR) assays were performed on the StepOne™ and the StepOnePlus™ Real-Time PCR Systems (Applied Biosystems, CA, USA), following the manufacturer's instructions. A typical 20-µl reaction mixture contained approximately 100 ng gDNA, 1 × SYBR® Premix Ex Taq™ II (Takara, Tokyo, Japan), 0.4 μM each primer (Table S1) and 1 × Rox™ Reference Dye. All samples were run in triplicate and underwent an initial 30 sec incubation step at 95°C, followed by 40 cycles of 5 sec at 95°C and 30 sec at 65°C for the Leishmania surface protein gp63 gene or 60°C for the mouse brain-derived neurotrophic factor (mBDNF) gene [24,25]. The average threshold cycle of amplification (Ct) values was determined, and standard deviation (SD) of all the reaction was analyzed by the software provided with the instrument. The relative amounts of the gp63 gene were then calculated using standard curve method normalized to the amounts of the mBDNF gene.

Histopathological Analysis of Infected Foci in the Liver

The livers were fixed in 10% neutral phosphate-buffered formalin. Paraffin-embedded organs were cut into 4-μm-thick
sections, followed by staining with hematoxylin and eosin for light microscopy. For the detection of parasites, liver sections were subjected to indirect immunohistochemical staining using *L. infantum*-infected dog serum (1:1000 dilution) [26] and horseradish peroxidase (HRP)-conjugated goat anti-dog IgG heavy and light chain antibody (1:300; Bethyl Laboratories, TX, USA). Peroxidase was visualized using 3,3′-diaminobenzidine (DAB·H₂O₂ (Wako, Tokyo, Japan) and the sections were counterstained with Mayer’s hematoxylin before dehydration and mounting.

Hepatic immune responses were categorized into (1) “No granuloma”: no inflammation with no mononuclear cell (MNC) around the parasitized Kupffer cells; (2) “Immature granuloma”: less than 10 MNCs around the parasitized Kupffer cells; (3) “Mature granuloma”: epithelioid cells and more than 10 MNCs around the parasitized Kupffer cells; and (4) “Involuting granuloma”: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue ‘granuloma’’: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue ‘granuloma’’: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue ‘granuloma’’: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue ‘granuloma’’: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue ‘granuloma’’: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue ‘granuloma’’: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue ‘granuloma’’: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue ‘granuloma’’: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue ‘granuloma’’: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue ‘granuloma’’: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue ‘granuloma’’: devoid of amastigotes and tissue inflammatory nearly resolved [18,23].

Determination of Foxp3-expressing T Cells by Flow Cytometry

Hepatic mononuclear cells were isolated using a 33% (vol/vol) Percoll solution, as described elsewhere [27]. Briefly, livers were minced, pressed through a stainless steel mesh and suspended in RPMI1640 medium (Sigma, MO, USA) supplemented with 3% HIFCS (wash buffer). After washing, the cells were resuspended in 33% Percoll solution containing heparin (100 U/ml) and centrifuged at 800 x g for 30 min to remove liver parenchymal cells. The pellet was treated with an RBC lysis solution (155 mM NH₄Cl, 10 mM KHCO₃, 0.1 mM EDTA), washed and re-suspended in 2.4G2 mAb solution to block theFc receptor before staining with antibody. Antibodies used for FACS included PE-labeled rat anti-mouse CD4 (LST4) (BD Pharmingen, CA, USA), FITC-labeled rat anti-mouse CD8a (Lyt-2) (BD Pharmingen), APC-labeled rat anti-mouse Foxp3 (FJK-16s) (eBioscience, CA, USA) and the proper isotype staining control, according to the manufacturer’s instructions. Flow cytometry analysis of the labeled cells was performed on a FACS Calibur (BD Pharmingen), running the Cell Quest program provided with the instrument. Lymphocytes were identified by forward scatter (FSC) and side scatter (SSC) characteristics, gated and further analyzed with Cell Quest software (BD Pharmingen) or FlowJo software V. 5.7.2 (Tree Star Inc., OR, USA).

Determination of Foxp3-expressing Cells and T Cells by Immunohistochemistry

Immunohistochemical analysis of the 4 μm-thick paraffin-embedded sections of the liver was performed to determine the localization of Foxp3+ Tregs. After deparaffinization and rehydration, heat-induced epitope retrieval (HIER) was conducted by autoclaving at 100°C for 17 min using Target Retrieval Solution (pH 9.0) (Dako, Uppsala, Sweden). Endogenous peroxidase was blocked by incubating sections in 0.3% H₂O₂ in absolute methanol for 30 min at 4°C, followed by flushing with water and incubation with 10% goat serum for 1 h at room temperature (RT) to block crystallized receptor fragments. The sections were incubated overnight with rat anti-mouse/rat Foxp3 mAb, clone FJK-16s (eBioscience), in 1:100 diluted with 0.1% Triton X in PBS (pH 7.4). For negative control sections, PBS was used instead of the primary antibody. After washing three times in PBS (5 min each), sections were incubated in 1:100 biotin-conjugated goat anti-rat IgG (H+L) antibody (Invitrogen, MD, USA) for 30 min at RT. Sections were then washed, which was followed by incubation with streptavidin-peroxidase conjugate (Histofine SAB-PO® Kit) for 30 min at RT. The streptavidin-biotin complex was visualized with DAB·H₂O₂ solution, pH 7.0, for 4 min. Sections were washed in distilled water, and finally counterstained with Mayer’s hematoxylin. The mean counts of Foxp3-expressing cells were assessed microscopically at 400 x magnification by counting a total of 25 consecutive fields. The number of immunoreactive cells was estimated in each hepatic granuloma assembly. Values are expressed as the means of immunoreactive cells present in 25 fields.

Double Immunofluorescence Analysis

Double immunofluorescence staining was also conducted to locate Tregs in the liver. Formalin-fixed and paraffin-embedded liver sections were subjected to the deparaffinization, rehydration, and HIER as described above. After blocking of crystallized receptor fragments with 10% goat serum, sections were incubated overnight with rat anti-mouse/rat Foxp3 mAb (clone FJK-16s; eBioscience) at 4°C. Then, the sections were incubated with FITC-goat anti rat IgG (1:200; Zymed, CA, USA) for 30 min at RT and successively incubated in 10% donkey or rabbit serum to block the crystallized receptor fragments. For double staining of Foxp3-expressing cells and T cells, the sections were incubated with rabbit anti-mouse CD3 mAb (1:200; Nichirei) overnight at 4°C and then with TRITC-donkey anti rabbit IgG (1:200; Abcam, MA, USA) for 30 min at RT. On the other hand, double staining of Foxp3-expressing cells and *L. donovani* amastigotes was conducted using *L. infantum*-infected dog serum (1:1,000) and TRITC-goat anti dog IgG (1:200; Rockland, PA, USA). Finally, the sections were mounted using a Fluoromount™ (DBS, CA, USA) and examined under an IX70 confocal microscope (Olympus, Tokyo, Japan).

Laser Microdissection, RNA Purification and qRT-PCR

Laser microdissection (LMD) was performed in RNase-free conditions as described previously [28]. Cryosections of 7 μm thickness were prepared from the frozen livers of naive and infected mice and embedded in Tissue-Tek OTC compound (Sakura, Tokyo, Japan). The sections were mounted on glass slides pre-coated with LMD films (Meiwafosis, Osaka, Japan) and fixed with absolute methanol for 3 min at 4°C. After staining with 0.5% toluidine blue for 10 sec, approximately 20 “Mature granulomas” were microdissected from each frozen liver sample by using Ls-Pro300 (Meiwafosis).

Total RNA was purified from the frozen whole liver tissue and microdissected “Mature granulomas”, using the RNAqueous® Micro Kit (Ambion, Texas, USA). Expression levels of Foxp3, *TGF*-β and *IL-10* mRNA were determined by quantitative RT-PCR (qRT-PCR) using the PrimeScript™ RT Reagent Kit (Takara) and the relative number of these molecules to 1000 housekeeping *glyceraldehyde-3-phosphate dehydrogenase* (*GAPDH*) was calculated using a standard curve method. The PCR reaction was performed as described above using primers shown in Table S1 [29,30].

Inhibition of CD4+ Foxp3+ Treg Function

At 26 WPI, three *L. donovani*-infected aly/aly mice were intraperitoneally injected three times every other day with 0.5 mg of rat anti-mouse CD25 mAb (clone PC61; Biolegend, CA, USA), 0.05 mg of rat anti-mouse FR4 mAb (clone TH6; Biolegend) or 0.5 mg of rat IgG (Jackson ImmunoResearch, PA, USA)
USA) as a control. The mice were euthanized at 10 days post-antibody injection for examination of host responses as described above.

**Statistics**

Statistical differences between aly/aly mice and aly/+ mice at the indicated time points were tested using Student's *t*-test (Microsoft Excel software) and two-way ANOVA as well as post hoc Bonferroni test (Prism software version 5, GraphPad, CA, USA). All data are presented as the mean ± SE unless otherwise stated. *p* < 0.05 was considered as statistically significant.

**Results**

**Long-term Persistence of *L. donovani* in the Liver of aly/aly Mice**

Long-term persistence after clinical cure of the primary infection is a characteristic feature of many intracellular pathogens, including protozoan parasites of the genus *Leishmania*, but the underlying mechanisms are not fully understood [31]. We measured parasite burdens in the livers of aly/+ and aly/aly mice for up to 28 WPI by two different methods. The number of amastigotes in hepatic impression smears was expressed as LDU (Figure 1A), and relative amounts of *Leishmania* gp63 gene to mouse housekeeping BDNF gene by qPCR (Figure 1B). A typical result of two individual experiments is shown. Data are the mean ± SE for three mice of each strain. *p* < 0.05; **p** < 0.01.

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![Graph A: Long-term persistence of *L. donovani* amastigotes in the liver of aly/aly mice.](image)

![Graph B: Delayed and Impaired Hepatic Granuloma Maturation in aly/aly Mice During *L. donovani* Infection.](image)

Figure 1. Long-term persistence of *L. donovani* amastigotes in the liver of aly/aly mice. Promastigotes of *L. donovani* were intravenously inoculated into aly/+ (*) and aly/aly (○) mice, and at the indicated time points, parasite burdens in the liver were determined as LDU (A), and relative amounts of *Leishmania* gp63 gene to mouse housekeeping BDNF gene by qPCR (B). A typical result of two individual experiments is shown. Data are the mean ± SE for three mice of each strain. *p* < 0.05; **p** < 0.01.

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Tregs and *L. donovani* Persistence in the Liver

Delayed and Impaired Hepatic Granuloma Maturation in aly/aly Mice During *L. donovani* Infection

Efficient granuloma development around infected Kupffer cells is a key event in the control of hepatic *L. donovani* infection [8,16,18]. The infected foci in the liver were examined and made a
quantitative analysis of granuloma formation around the parasitized Kupffer cells. The progression of granuloma formation from “No granuloma” to “Immature granuloma”, “Mature granuloma” and finally “Involuting granuloma” was observed in aly/aly mice as well as aly/+ mice (Figure S1), indicating that aly/aly mice have ability to generate hepatic cell-mediated immunity to some extent as shown in the previous study [32].

The number of infected foci was well correlated with the hepatic parasite loads (Figure 1); the number of foci in aly/+ mice reached a peak at 4 WPI and was drastically reduced at 8 WPI (Figure 2A) and the involuting granuloma was well formed (Figure 2B). However, the number of involuting granuloma in the liver of aly/aly mice were much less than those in aly/+ mice at 4 and 8 WPI (Figure 2C) while the 30–40% of the infection foci with no granulomas was found in the liver of aly/aly mice at 4–16 WPI. These may reflect that effective but insufficient clearance of the parasites in granuloma of aly/aly livers renders persistent release of the parasites, which results in increased proportion of infected Kupffer cells in the later stages.

Figure 2. Delayed and impaired hepatic granuloma maturation in aly/aly mice during L. donovani infection. Host immune responses to each infected focus were quantitatively analyzed in aly/+ (open bars) and aly/aly (filled bars) mice during the course of L. donovani infection. Total number of infected foci was counted from 25 consecutive microscopic fields (A). The proportion of “No granuloma”, “Immature granuloma”, “Mature granulomas” and “Involuting granuloma” of aly/+ (B) and aly/aly (C) mice are estimated. A typical result of two individual experiments is shown. Data are the mean ± SE for three mice of each strain. ND, not determined; * p<0.05; ** p<0.01. doi:10.1371/journal.pntd.0001798.g002

Expansion of CD4⁺Foxp3⁺ Tregs in the Liver During Persistent L. donovani Infection

Foxp3⁺ Tregs influence immunity to viral, bacterial or parasitic infections [33]. To begin to characterize the mechanism by which parasites persist in the liver, we examined whether Tregs expand in the livers of aly/aly mice and where they localize during L. donovani infection.

Flow cytometry analysis of hepatic lymphocytes revealed no expansion of CD8⁺Foxp3⁺ T cells in the liver during L. donovani infection in either strain of mice (Figure 3A). In contrast, CD4⁺Foxp3⁺ T cells were first detected at 4 WPI in both strains of mice. In aly/aly mice, the proportions of CD4⁺Foxp3⁺ T cells to
CD4+ T cells (Figure 3B) as well as the absolute number of CD4+Foxp3+ T cells was higher than those of aly/+ mice especially at 8–16 WPI although the total number of hepatic CD4+ T cells was not significantly different between aly/+ and aly/aly mice (Figure S2).

**Presence of Foxp3+ Tregs Inside Granulomas and Perivascular Neo-lymphoid Areas in the L. donovani-infected Liver of aly/aly Mice**

There have been no reports describing the localization of Foxp3-expressing cells in the liver during VL. To address this, we stained Foxp3 in liver sections of naïve and L. donovani-infected aly/+ and aly/aly mice. Foxp3-expressing cells were localized in the “Immature granuloma” and “Mature granulomas” as well as the perivascular areas of infected aly/aly mice. Furthermore, the density of Tregs increased, especially in the perivascular areas, during the course of infection (Figure 4A and B). Development of such abnormal lymphocyte infiltration or neo-lymphoid aggregates at perivascular areas is a feature found in aly/aly and other alymphoplastic mice [32]. In addition, the frequency of “Mature granulomas” containing more than 5 Tregs increased during infection in aly/aly mice (5% at 4 WPI, 18% at 12 WPI and 39% at 28 WPI), suggesting the accumulation of Tregs at sites of inflammatory foci. On the other hand, Foxp3-positive Tregs were limited to the parenchyma, granulomas and perivascular areas at 4 WPI and hardly detectable in the liver of infected aly/+ mice at 12 WPI (Figure 4A and B).

Double immunofluorescence analysis of hepatic granuloma revealed that Foxp3-expressing cells (green in Figure 5A) and CD3+ cells (red in Figure 5A) were present in the granuloma, and Foxp3+ cells expressed CD3 molecules (Figure 5A-merged image). Some CD3+Foxp3+ cells (yellow arrows in Figure 5A-merged image) were adjacent to the CD3+Foxp3- cells (pink arrows in Figure 5A-merged image). In addition, L. donovani amastigotes (red in Figure 5B) were surrounded by Foxp3+ cells (green in Figure 5B) in the hepatic granuloma. These results suggested that the interaction among parasitized cells (Kupffer cells), CD3+Foxp3+ cell (Tregs) and CD3+Foxp3- cells (non Tregs, probably CD4+ and/or CD8+ effector T cells).

**Association of Foxp3 and IL-10 mRNA Expression in “Mature Granulomas”**

Evidence has accumulated regarding the essential roles of Tregs in the control of a variety of physiological and pathological immune responses, but it is still obscure how Tregs control other lymphocytes at the molecular level [34]. Quantitative RT-PCR was performed for Foxp3, IL-10 and TGF-β mRNA levels in the whole liver and micro-dissected “Mature granulomas” liver tissue samples of L. donovani-infected aly/aly mice. The Foxp3 mRNA expression was increased after infection in the whole liver.
Although the TGF-β mRNA transcripts showed similar levels at 4 and 12 WPI in both tissue samples, the levels of IL-10 mRNA markedly increased in mature granuloma but not whole liver samples at 12 WPI (Figure 6A and B), suggesting that IL-10 may be involved in function of Tregs.

**Enhanced Effector Immune Responses and Declined Parasite Burden in the Liver After Inhibition of Treg Function in L. donovani-infected aly/aly Mice**

Manipulation of Tregs by treatment with antibodies has been used to examine the roles of Tregs in many infectious diseases [33]. Effects of anti-GD25 and anti-FR4 mAb on hepatic...
immune responses in *L. donovani*-infected *aly/aly* mice at 26 WPI were examined. Ten days after injection with anti-CD25 or anti-FR4, reduction in Foxp3 mRNA expression was observed (Figure 7A). This reduced Foxp3 mRNA expression was associated with decreases in parasite burden (Figure 7B) and infected foci (Figure 7C). Instead, the frequency of “Mature granulomas” was increased after treatment with especially anti-FR4 mAb (Figure 7D), suggesting that depletion of Tregs can activate hepatic cellular immune responses and accelerate parasite killing. Furthermore, immunohistochemical analysis
confirmed a reduction in Foxp3-immunoreactive cells in the liver parenchyma, granulomas (Figure 7E) and perivascular neolymphoid areas (data not shown).

**Discussion**

In the present study, *aly/aly* mice were used as an immunodeficient VL murine model and immunohistopathologically investigated during *L. donovani* infection for up to 28 WPI. CD4+Foxp3+ T cells were increased in the granulomas and perivascular areas of the liver in the chronic phase and the impairment of granuloma maturation was observed. The depletion of Tregs by the administration of either anti-CD25 or anti-FR4 mAb resulted in significant reductions in hepatic Tregs, infected foci and parasite burden. To our knowledge, this is the first definitive evidence that CD4+Foxp3+ Tregs are involved in hepatic *L. donovani* persistence in a murine model of VL.

The *aly/aly* mice have been used to examine the role of secondary lymphoid organs on immune responses in various
Figure 7. Effects of anti-Treg antibodies on Foxp3 expression, parasite burden and infected foci in the liver. The *L. donovani*-infected *aly/aly* mice were intraperitoneally injected with anti-CD25 or anti-FR4 mAb at 26 WPI and hepatic immune responses were examined after 10 days of injection. (A) Foxp3 mRNA expression by RT-PCR. (B) Parasite burden expressed by qPCR. (C) The number of infected foci from 25 consecutive microscopic fields. (D) Frequency of mature (black column) and immature (grey column) granuloma, and no response (white column) in the infected foci. (E) Localization of Foxp3-stained cells in the liver parenchyma including “Immature granulomas” and “Mature granulomas”. The brown pigments represent Foxp3-immunoreactive cells. * p<0.05; *** p<0.001.
doi:10.1371/journal.pntd.0001798.g007
infection models. Disruptive architecture of the thymus and spleen could affect the development and expansion of T cells. Several studies using bone marrow chimeras between aly/aly and wild-type mice showed that antiviral CTL responses were clearly improved in the wild-type environment [11]. However, expansion of CD25+CD4+ Treg is impaired in the spleen of aly/aly mice [35]. This suggests that expansion of functional CD4+Foxp3+ Treg in the liver of aly/aly mice during L. donovani infection is likely related to the parasite persistence but not to the structural defects of secondary lymphoid organs although this possibility will be confirmed by BM chimera experiments in future.

The NIK gene mutation may contribute to other immune defects due to the partial blocked NF-κB activation [10,36]. NF-kBp52 knockout mice showed less parasite burden in the liver, perhaps due to less number of B cells (unpublished data; Ato M., Kaye PM). NF-kBp50 (NF-kB1) would be important for TNF/TLR signaling which is involved in canonical TLR/TNFR. NIK is confirmed by BM chimera experiments in future. Secondary lymphoid organs although this possibility will be to the parasite persistence but not to the structural defects of the liver of aly/aly mice during L. donovani infection is likely related to the parasite persistence but not to the structural defects of secondary lymphoid organs although this possibility will be confirmed by BM chimera experiments in future.

In conclusion, we focused on immune responses to the chronic phase of murine VL caused by L. donovani infection in an immunodeficient host. In the last decade when the number of visceral leishmaniasis in immunocompromised patients has been increasing, our data presented herein offered a novel insight into the possibly involvement of CD4+Foxp3+ Tregs in persistent L. donovani infection in the liver of immunodeficient hosts. The manipulation of Tregs may provide a promising immunotherapeutic strategy for VL.

Supporting Information

**Figure S1** Hepatic immune response and granuloma formation in aly/+ and aly/aly mice 2–8 weeks after L. donovani infection. Representative hepatic immune responses to parasitized Kupffer cells were expanded in a chronic VL. Although anti-CD25 mAbs have been used for depletion of Tregs in various experimental cases, administration of anti-FR4 mAb also reduced Treg numbers and provoked effective tumor immunity [37–39]. In the present study, 10 days after the third injection of infected aly/aly mice with anti-CD25 and anti-FR4 mAb, the hepatic parasite burdens were reduced by 88% and 89% of that of control mice, respectively (Figure 7B). Likewise, treatment with either anti-CD25 or anti-FR4 mAb also reduced parasite burdens in the spleen and BM (Figure S3). The reason why anti-FR4 mAb was more effective than anti-CD25 mAb in reducing parasite burden is unknown, but the present study is the first to report effectiveness of anti-FR4 mAb to control systemic infection of L. donovani in mice. Thus, anti-FR4 antibodies may be an alternative measure to manipulate Tregs in chronic VL. However, since anti-CD25 mAb can also affect effector T cells and effective immunity [39], and anti-FR4 mAb can also deplete a small population of CD4+Foxp3+ T cells in the lymph node [38], probably including IL-10-producing conventional CD4+ T cells, further studies of the role of Tregs in VL are required.

**Figure S2** Proportion of CD4+Foxp3+ and CD4+ T cells gated hepatic lymphocytes during the course of L. donovani infection by flow cytometry analysis. Hepatic lymphocytes were isolated from the liver of aly/+ and aly/aly mice at indicated time points after L. donovani infection and stained with anti-CD4, CD8 and Foxp3 antibodies. Percentage of CD4+ T cells (white bars) and CD4+Foxp3+ T cells (black bars) of aly/+ (A) and aly/aly mice (B) are shown.

**Figure S3** Effects of anti-Treg antibody treatment on parasite burden in the spleen and bone marrow. The L. donovani-infected aly/aly mice were intraperitoneally injected with anti-CD25 or anti-FR4 mAb at 26 WPI, and parasite burden in the spleen (A) and bone marrow (B) was estimated by qPCR after 10 days of antibody treatment. *p < 0.05; **p < 0.01.

**Table S1** Target genes and primers for qPCR and RT-PCR used in this study.

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Author Contributions
Conceived and designed the experiments: ST, KI, MA, TS, KK. Performed the experiments: ST, KI, MA, HK. Analyzed the data: ST, KI, MA, HK. Wrote the paper: ST, KI, MA, HK, KK.

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