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Author(s)
Tiwananthagorn, Saruda; Iwabuchi, Kazuya; Ato, Manabu; Sakurai, Tatsuya; Kato, Hirotomo; Katakura, Ken

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Involvement of CD4$^+$ Foxp3$^+$ Regulatory T Cells in Persistence of *Leishmania donovani* in the Liver of Alymphoplastic *aly/aly* Mice

Saruda Tiwananthagorn$^{1,2}$, Kazuya Iwabuchi$^3$, Manabu Ato$^4$, Tatsuya Sakurai$^1$, Hirotomo Kato$^1$, Ken Katakura$^{1*}$

1 Laboratory of Parasitology, Graduate School of Veterinary Medicine, Hokkaido University, Sapporo, Japan, 2 Department of Veterinary Biosciences and Veterinary Public Health, Faculty of Veterinary Medicine, Chiang Mai University, Chiang Mai, Thailand, 3 Department of Immunology, Kitasato University School of Medicine, Kanagawa, Japan, 4 Department of Immunology, National Institute of Infectious Diseases, Tokyo, Japan

Abstract

Visceral leishmaniasis (VL) is a chronic and fatal disease in humans and dogs caused by the intracellular protozoan parasites, *Leishmania donovani* and *L. infantum* (*L. chagasi*). Relapse of disease is frequent in immunocompromised conditions, which is associated with reactivation of parasites and increased early visceralization of the liver. This mutant mouse strain has been used to examine mechanisms of *L. donovani* persistence in immunocompromised conditions. The role of secondary lymphoid organs for immune responses to *Leishmania* infection has not been investigated. The *aly/aly* mouse strain has been used to examine mechanisms including T-cell functions in the chronic stage of infection has not been clearly clarified. To develop effective prophylactic or therapeutic strategies against VL, understanding of the precise immune mechanisms including T-cell functions in the chronic stage of infection is required.

Introduction

Visceral leishmaniasis (VL) is a chronic and fatal disease caused by the intracellular protozoan parasites *Leishmania donovani* and *L. infantum* (*L. chagasi*), which infect a range of mammalian hosts, including humans, dogs and rodents [1]. Liver, spleen, bone marrow (BM) and lymph nodes are the major sites for parasite growth and disease pathology. Transplantation of infected kidney, liver, heart, lung, pancreas or BM has been shown to cause VL in transplant recipients, indicating lifelong parasite persistence in the viscera [2]. Moreover, malnutrition is a risk factor for the development of VL [3]. Recent experiments in protein energy-, zinc- and iron-deficient mice suggest that this effect is mediated primarily through functional failure of the lymph node barrier and increased early visceralization of the parasites [4–6]. Loss of the control of parasite persistence in VL causes the reactivation of parasites and relapse of the disease is frequent in the immunocompromised patients, in which the number of visceral leishmaniasis cases has been increasing recently [7]. However, the mechanisms underlying the parasite persistence in the immunocompromised condition have not been clearly clarified. To develop effective prophylactic or therapeutic strategies against VL, understanding of the precise immune mechanisms including T-cell functions in the chronic stage of infection is required [8].

The role of secondary lymphoid organs for immune responses to *Leishmania* infection has not been investigated. The *aly/aly* mouse is an autosomal recessive natural mutant C57BL/6 strain that carries a point mutation within the gene encoding NF-$\kappa$B inducing kinase (NIK) [9], which prevents the induction of the non-canonical NF-$\kappa$B pathway [10]. The *aly/aly* mouse lack all lymph nodes and Peyer's patches with the abnormal architecture of spleen and thymus and exhibit severely impaired humoral response [9]. This mutant mouse strain has been used to examine...
The protozoan parasite Leishmania donovani is the causative agent of visceral leishmaniasis (VL) with a variety of outcomes ranging from asymptomatic to fatal infection. In the last decade, an increasing number of VL cases in immunocompromised conditions have been reported. Loss of the control of parasite persistence causes relapse of the disease in these patients. To clarify why parasite persistence and disease are caused in an immunocompromised condition, we examined L. donovani infection in aly/aly mice that completely lack lymph nodes and have disturbed spleen architecture. Although parasites grew in the liver of aly/+ mice for the first 4 weeks post infection (WPI) and parasites were eliminated by 8 WPI, we found that parasites persisted in the liver of aly/aly mice with the ineffective granuloma formation to kill the parasites. These aly/aly mice showed significant increases in CD4+Foxp3+ regulatory T cells in the liver. Consequently, we treated infected mice with anti-CD25 or anti-FR4 mAb to inhibit the function of Tregs, and found significant reductions in both hepatic Foxp3+ cells and parasite burden. These results clearly demonstrated for the first time that the expansion of CD4+Foxp3+ Tregs is involved in hepatic L. donovani persistence in immunodeficient murine model.

Materials and Methods

Mice, Parasites and Infection

Female ALY® NsCjcl aly/aly and aly/+ mice of 6–8 weeks of age were purchased from CLEA Japan, Inc. (Tokyo, Japan). Mice were maintained, inoculated and sacrificed within a safety facility of Hokkaido University.

A virulent line of L. donovani (MHOM/SU/62/25-MC2) [22] was maintained by passage of the frozen stabilized parasites in NNN medium containing 5% defibrinated hemolyzed rabbit blood. Then, parasites were consecutively sub-passaged in liquid M199 medium supplemented with 15% heat-inactivated fetal calf serum (HIFCS), 25 mM HEPES and 50 μg/ml gentamycin. The stationary growth phase of subcultures with less than five passages was used for mouse inoculation. Mice were infected by injecting stationary phase promastigotes (5×10⁷) intravenously via the lateral tail vein and were sacrificed at 1, 2, 4, 8, 12, 16 and 28 weeks post infection (WPI). One group of non-infected animals was used as naive control.

Ethics Statement

This study was carried out under the guidance of the Institute for Laboratory Animal Research (ILAR). All animals were housed in a facility in strict accordance with the recommendations in the Guidelines for the Care and Use of Laboratory Animals of Graduate School of Veterinary Medicine, Hokkaido University, which was based on Fundamental Guidelines for Proper Conduct of Animal Experiment and Related Activities in Academic Research Institutions under the jurisdiction of the Ministry of Education, Culture, Sports, Science and Technology, Japan and approved by the American Association for Accreditation of Laboratory Animal Care (AAALAC) international. The protocol was approved by the Committee on the Ethics of Animal Experiments of Hokkaido University ( Permit Number: 10-0009).

Histopathological Analysis of Infected Foci in the Liver

The livers were fixed in 10% neutral phosphate-buffered formalin. Parafin-embedded organs were cut into 4 μm-thick sections and stained with hematoxylin and eosin (H&E). The number of parasites was assessed in two independent experiments. In each experiment, organs from two animals were sacrificed at 1, 2, 4, 8, 12, 16 and 28 weeks post infection (WPI). One group of non-infected animals was used as naive control.
sections, followed by staining with hematoxylin and eosin for light microscopy. For the detection of parasites, liver sections were subjected to indirect immunohistochemical staining using L. infantum-infected dog serum (1:1000 dilution) [26] and horseradish peroxidase (HRP)-conjugated goat anti-dog IgG heavy and light chain antibody (1:300; Bethyl Laboratories, TX, USA). Peroxidase was visualized using 3,3′-diaminobenzidine (DAB-H2O2 (Wako, Tokyo, Japan) and the sections were counterstained with Mayer’s hematoxylin before dehydration and mounting.

Hepatic immune responses were categorized into (1) “No granuloma”: no inflammation with no mononuclear cell (MNC) around the parasitized Kupffer cells; (2) “Immature granuloma”: less than 10 MNCs around the parasitized Kupffer cells; (3) “Mature granuloma”: epithelioid cells and more than 10 MNCs around the parasitized Kupffer cells; and (4) “Involuting granuloma”: devoid of amastigotes and tissue inflammatory nearly resolved [18,23]. The number of infected foci with each tissue response including “No granuloma”, “Immature granuloma”, “Mature granuloma” and “Involuting granuloma” was counted for 25 consecutive microscopic fields per mouse liver at ×400 magnification.

Determination of Foxp3-expressing T Cells by Flow Cytometry

Hepatic mononuclear cells were isolated using a 33% (vol/vol) Percoll solution, as described elsewhere [27]. Briefly, livers were minced, pressed through a stainless steel mesh and suspended in RPMI1640 medium (Sigma, MO, USA) supplemented with 3% HIFCS (wash buffer). After washing, the cells were resuspended in 33% Percoll solution containing heparin (100 U/ml) and centrifuged at 800 × g for 30 min to remove liver parenchymal cells. The pellet was treated with an RBC lysis solution (155 mM NH4Cl, 10 mM KHCO3, 0.1 mM EDTA), washed and resuspended in 2.4G2 mAb solution to block the Fc receptor before staining with antibody. Antibodies used for FACS included PE-labeled rat anti-mouse CD4 (LST4; BD Pharmingen, CA, USA), FITC-labeled rat anti-mouse CD8α (Lyt-2; BD Pharmingen), APC-labeled rat anti-mouse Foxp3 (FJK-16s) (eBioscience, CA, USA) and the proper isotype staining control, according to the manufacturer’s instructions. Flow cytometry analysis of the labeled cells was performed on a FACS Calibur (BD Pharmingen), running the Cell Quest program provided with the instrument. Lymphocytes were identified by forward scatter (FSC) and side scatter (SSC) characteristics, gated and further analyzed with Cell Quest software (BD Pharmingen) or FlowJo software V. 5.7.2 (Tree Star Inc., OR, USA).

Determination of Foxp3-expressing Cells and T Cells by Immunohistochemistry

Immunohistochemical analysis of the 4 μm-thick paraffin-embedded sections of the liver was performed to determine the localization of Foxp3+ Tregs. After deparaffinization and rehydration, heat-induced epitope retrieval (HIER) was conducted by autoclaving at 100°C for 17 min using Target Retrieval Solution (pH 9.0) (Dako, Uppsala, Sweden). Endogenous peroxidase was blocked by incubating sections in 0.3% H2O2 in absolute methanol for 30 min at 4°C, followed by flushing with water and incubation with 10% goat serum for 1 h at room temperature (RT) to block crystallized receptor fragments. The sections were incubated overnight with rat anti-mouse/rat Foxp3 mAb, clone FJK-16s (eBioscience), in 1:100 diluted with 0.1% Triton X in PBS (pH 7.4). For negative control sections, PBS was used instead of the primary antibody. After washing three times in PBS (5 min each), sections were incubated in 1:100 biotin-conjugated goat anti-rat IgG (H+L) antibody (Invitrogen, MD, USA) for 30 min at RT. Sections were then washed, which was followed by incubation with streptavidin-peroxidase conjugate (Histofine SAB-PO® Kit) for 30 min at RT. The streptavidin-biotin complex was visualized with DAB-H2O2 solution, pH 7.0, for 4 min. Sections were washed in distilled water, and finally counterstained with Mayer’s hematoxylin. The mean counts of Foxp3-expressing cells were assessed microscopically at 400× magnification by counting a total of 25 consecutive fields. The number of immunoreactive cells was estimated in each hepatic granuloma assembly. Values are expressed as the means of immunoreactive cells present in 25 fields.

Double Immunofluorescence Analysis

Double immunofluorescence staining was also conducted to locate Tregs in the liver. Formalin-fixed and paraffin-embedded liver sections were subjected to the deparaffinization, rehydration and HIER as described above. After blocking of crystallized receptor fragments with 10% goat serum, sections were incubated overnight with rat anti-mouse/rat Foxp3 mAb (clone FJK-16s; 1:100; eBioscience) at 4°C. Then, the sections were incubated with FITC-goat anti rat IgG (1:200; Zymed, CA, USA) for 30 min at RT and successively incubated in 10% donkey or rabbit serum to block the crystallized receptor fragments. For double staining of Foxp3-expressing cells and T cells, the sections were incubated with rabbit anti-mouse CD3 mAb (1:200; Nichirei) overnight at 4°C and then with TRITC-donkey anti rabbit IgG (1:200; Alciam, MA, USA) for 30 min at RT. On the other hand, double staining of Foxp3-expressing cells and L. donovani amastigotes was conducted using L. infantum-infected dog serum (1:1,000) and TRITC-goat anti dog IgG (1:200; Rockland, PA, USA). Finally, the sections were mounted using a Fluoromount™ (DBS, CA, USA) and examined under an IX70 confocal microscope (Olympus, Tokyo, Japan).

 Laser Microdissection, RNA Purification and qRT-PCR

 Laser microdissection (LMD) was performed in RNase-free conditions as described previously [28]. Cryosections of 7 μm thickness were prepared from the frozen liver of naive and infected mice and embedded in Tissue-Tek OTC compound (Sakura, Tokyo, Japan). The sections were mounted on glass slides pre-coated with LMD films (Meiwafosis, Osaka, Japan) and fixed with absolute methanol for 3 min at 4°C. After staining with 0.5% toluidine blue for 10 sec, approximately 20 “Mature granulomas” were microdissected from each frozen liver sample by using Ls-Pro300 (Meiwafosis). Total RNA was purified from the frozen whole liver tissue and microdissected “Mature granulomas”, using the RNAqueous® Micro Kit (Ambion, Texas, USA). Expression levels of Foxp3, TGF-β and IL-10 mRNA were determined by quantitative RT-PCR (qRT-PCR) using the PrimeScript™ RT Reagent Kit (Takara) and the relative number of these molecules to 1000 housekeeping glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was calculated using a standard curve method. The PCR reaction was performed as described above using primers shown in Table S1 [29,30].

Inhibition of CD4+ Foxp3+ Treg Function

At 26 WPI, three L. donovani-infected aly/aly mice were intraperitoneally injected three times every other day with 0.5 mg of rat anti-mouse CD25 mAb (clone PC61; Biolegend, CA, USA), 0.05 mg of rat anti-mouse FR4 mAb (clone TH6; Biolegend) or 0.5 mg of rat IgG (Jackson ImmunoResearch, PA,
USA) as a control. The mice were euthanized at 10 days postantibody injection for examination of host responses as described above.

Statistics
Statistical differences between aly/+ mice and aly/aly mice at the indicated time points were tested using Student’s t-test (Microsoft Excel software) and two-way ANOVA as well as post hoc Bonferroni test (Prism software version 5, GraphPad, CA, USA). All data are presented as the mean ± SE unless otherwise stated. p<0.05 was considered as statistically significant.

Results
Long-term Persistence of L. donovani in the Liver of aly/aly Mice
Long-term persistence after clinical cure of the primary infection is a characteristic feature of many intracellular pathogens, including protozoan parasites of the genus Leishmania, but the underlying mechanisms are not fully understood [31]. We measured parasite burdens in the livers of aly/+ and aly/aly mice for up to 28 WPI by two different methods. The number of amastigotes in hepatic impression smears was expressed as LDU (Figure 1A), and relative amounts of Leishmania gp63 gene to mouse housekeeping BDNF gene by qPCR (Figure 1B). A typical result of two individual experiments is shown. Data are the mean ± SE for three mice of each strain. * p<0.05; ** p<0.01.

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Figure 1. Long-term persistence of L. donovani amastigotes in the liver of aly/aly mice. Promastigotes of L. donovani were intravenously inoculated into aly/+ (○) and aly/aly (●) mice, and at the indicated time points, parasite burdens in the liver were determined as LDU (A), and relative amounts of Leishmania gp63 gene to mouse housekeeping BDNF gene by qPCR (B). A typical result of two individual experiments is shown. Data are the mean ± SE for three mice of each strain. * p<0.05; ** p<0.01.

Tregs and L. donovani Persistence in the Liver

Delayed and Impaired Hepatic Granuloma Maturation in aly/aly Mice During L. donovani Infection
Efficient granuloma development around infected Kupffer cells is a key event in the control of hepatic L. donovani infection [8,16,18]. The infected foci in the liver were examined and made a
quantitative analysis of granuloma formation around the parasitized Kupffer cells. The progression of granuloma formation from “No granuloma” to “Immature granuloma”, “Mature granuloma” and finally “Involuting granuloma” was observed in aly/aly mice as well as aly/+ mice (Figure S1), indicating that aly/aly mice have ability to generate hepatic cell-mediated immunity to some extent as shown in the previous study [32].

The number of infected foci was well correlated with the hepatic parasite loads (Figure 1); the number of foci in aly/+ mice reached a peak at 4 WPI and was drastically reduced at 8 WPI (Figure 2A) and the involuting granuloma was well formed (Figure 2B). However, the number of involuting granuloma in the liver of aly/aly mice were much less than those in aly/+ mice at 4 and 8 WPI (Figure 2C) while the 30–40% of the infection foci with no granulomas was found in the liver of aly/aly mice at 4–16 WPI. These may reflect that effective but insufficient clearance of the parasites in granuloma of aly/aly livers renders persistent release of the parasites, which results in increased proportion of infected Kupffer cells in the later stages.

**Expansion of CD4+Foxp3+ Tregs in the Liver of aly/aly Mice During Persistent L. donovani Infection**

Foxp3+ Tregs influence immunity to viral, bacterial or parasitic infections [33]. To begin to characterize the mechanism by which parasites persist in the liver, we examined whether Tregs expand in the livers of aly/aly mice and where they localize during L. donovani infection.

Flow cytometry analysis of hepatic lymphocytes revealed no expansion of CD8+Foxp3+ T cells in the liver during L. donovani infection in either strain of mice (Figure 3A). In contrast, CD4+Foxp3+ T cells were first detected at 4 WPI in both strains of mice. In aly/aly mice, the proportions of CD4+Foxp3+ T cells to

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**Figure 2. Delayed and impaired hepatic granuloma maturation in aly/aly mice during L. donovani infection.** Host immune responses to each infected focus were quantitatively analyzed in aly/+ (open bars) and aly/aly (filled bars) mice during the course of L. donovani infection. Total number of infected foci was counted from 25 consecutive microscopic fields (A). The proportion of "No granuloma", "Immature granuloma", "Mature granulomas" and "Involuting granuloma" of aly/+ (B) and aly/aly (C) mice are estimated. A typical result of two individual experiments is shown. Data are the mean ± SE for three mice of each strain. ND, not determined; * p<0.05; ** p<0.01.
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**Figure 3. Expansion of CD4^+ Foxp3^+ Tregs in the liver of aly/aly mice during persistent L. donovani infection.** Flow cytometry analysis of CD4^+Foxp3^+ and CD8^+Foxp3^+ Tregs among the hepatic lymphocytes. (A) Representative data of the gated lymphocytes extracted from the liver at 12 WPI of aly/+ (left) and aly/aly mice (right), stained with anti-Foxp3 and anti-CD4 or anti-CD8 mAb. (B) The proportion of CD4^+Foxp3^+ T cells to CD4^+ T cells in aly/+ (open bars) and aly/aly (filled bars) mice. (C) The total number of CD4^+Foxp3^+ T cells in aly/+ (open bars) and aly/aly (filled bars) mice at 8–16 WPI. A typical result of two individual experiments is shown. Data are the mean ± SE for three mice of each strain. ND, not determined; * p<0.05; *** p<0.001. doi:10.1371/journal.pntd.0001798.g003

**Presence of Foxp3^+ Tregs Inside Granulomas and Perivascular Neo-lymphoid Areas in the L. donovani-infected liver of aly/aly Mice**

There have been no reports describing the localization of Foxp3-expressing cells in the liver during VL. To address this, we stained Foxp3 in liver sections of naïve and *L. donovani*-infected aly/+ and aly/aly mice. Foxp3-expressing cells were localized in the “Immature granuloma” and “Mature granulomas” as well as the perivascular areas of infected aly/aly mice. Furthermore, the density of Tregs increased, especially in the perivascular areas, during the course of infection (Figure 4A and B). Development of such abnormal lymphocyte infiltration or neo-lymphoid aggregates at perivascular areas is a feature found in aly/aly and other alypoplastic mice [32]. In addition, the frequency of “Mature granulomas” containing more than 5 Tregs increased during infection in aly/aly mice (5% at 4 WPI, 18% at 12 WPI and 39% at 28 WPI), suggesting the accumulation of Tregs at sites of inflammatory foci. On the other hand, Foxp3-positive Tregs were limited to the parenchyma, granulomas and perivascular areas at 4 WPI and hardly detectable in the liver of infected aly/+ mice at 12 WPI (Figure 4A and B).

Double immunofluorescence analysis of hepatic granuloma revealed that Foxp3-expressing cells (green in Figure 5A) and CD3^+ cells (red in Figure 5A) were present in the granuloma, and Foxp3^+ cells expressed CD3 molecules (Figure 5A-merged image). Some CD3^+Foxp3^+ cells (yellow arrows in Figure 5A-merged image) were adjacent to the CD3^+Foxp3^- cells (pink arrows in Figure 5A-merged image). In addition, *L. donovani* amastigotes (red in Figure 5B) were surrounded by Foxp3^+ cells (green in Figure 5B) in the hepatic granuloma. These results suggested that the interaction among parasitized cells (Kupffer cells), CD3^+Foxp3^+ cell (Tregs) and CD3^+Foxp3^- cells (non Tregs, probably CD4^+ and/or CD8^+ effector T cells).

**Association of Foxp3 and IL-10 mRNA Expression in “Mature Granulomas”**

Evidence has accumulated regarding the essential roles of Tregs in the control of a variety of physiological and pathological immune responses, but it is still obscure how Tregs control other lymphocytes at the molecular level [34]. Quantitative RT-PCR was performed for Foxp3, IL-10 and TGF-β mRNA levels in the whole liver and micro-dissected “Mature granulomas” liver tissue samples of *L. donovani*-infected aly/aly mice. The Foxp3 mRNA expression was increased after infection in the whole liver.
Although the TGF-β mRNA transcripts showed similar levels at 4 and 12 WPI in both tissue samples, the levels of IL-10 mRNA markedly increased in mature granuloma but not whole liver samples at 12 WPI (Figure 6A and B), suggesting that IL-10 may be involved in function of Tregs.

**Figure 4. Presence of Foxp3+ Tregs inside granulomas in the liver of L. donovani-infected aly/aly mice.** Localization of Foxp3-expressing cells in liver sections from aly/+ mice and aly/aly mice during the course of L. donovani infection was analyzed by immunohistochemistry using anti-Foxp3 mAb. Typical reactions in the liver parenchyma including “Immature granuloma” and “Mature granulomas” (A) and perivascular areas (B) in uninfected naive mice and infected mice at 4 and 12 WPI are shown. The brown pigments represent Foxp3-immunoreactive cells. doi:10.1371/journal.pntd.0001798.g004

**Enhanced Effector Immune Responses and Declined Parasite Burden in the Liver After Inhibition of Treg Function in L. donovani-infected aly/aly Mice**

Manipulation of Tregs by treatment with antibodies has been used to examine the roles of Tregs in many infectious diseases [33]. Effects of anti-GD25 and anti-FR4 mAb on hepatic
immune responses in L. donovani-infected aly/aly mice at 26 WPI were examined. Ten days after injection with anti-CD25 or anti-FR4, reduction in Foxp3 mRNA expression was observed (Figure 7A). This reduced Foxp3 mRNA expression was associated with decreases in parasite burden (Figure 7B) and infected foci (Figure 7C). Instead, the frequency of “Mature granulomas” was increased after treatment with especially anti-FR4 mAb (Figure 7D), suggesting that depletion of Tregs can activate hepatic cellular immune responses and accelerate parasite killing. Furthermore, immunohistochemical analysis

**Figure 5. Localization of Foxp3-expressing cells in L. donovani-infected granuloma of aly/aly mice.** Representative “Mature granuloma” in the liver sections of aly/aly mice at 12 WPI using single and double immunostaining. (A) Foxp3+ cells (green; upper left), CD3+ cells (red; upper right) and a merged image (lower) showed CD3+Foxp3+ (yellow arrows) and CD3+Foxp3− (pink arrows) cells. (B) Foxp3+ cells (green; upper left), Leishmania amastigotes (red; upper right) and a merged image (lower). Arrows indicated the positive stained cells.

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confirmed a reduction in Foxp3-immunoreactive cells in the liver parenchyma, granulomas (Figure 7E) and perivascular neo-lymphoid areas (data not shown).

Discussion

In the present study, aly/aly mice were used as an immunodeficient VL murine model and immunohistopathologically investigated during L. donovani infection for up to 28 WPI. CD4+Foxp3+ T cells were increased in the granulomas and perivascular areas of the liver in the chronic phase and the impairment of granuloma maturation was observed. The depletion of Tregs by the administration of either anti-CD25 or anti-FR4 mAb resulted in significant reductions in hepatic Tregs, infected foci and parasite burden. To our knowledge, this is the first definitive evidence that CD4+Foxp3+ Tregs are involved in hepatic L. donovani persistence in a murine model of VL.

The aly/aly mice have been used to examine the role of secondary lymphoid organs on immune responses in various
Figure 7. Effects of anti-Treg antibodies on Foxp3 expression, parasite burden and infected foci in the liver. The *L. donovani*-infected *aly/aly* mice were intraperitoneally injected with anti-CD25 or anti-FR4 mAb at 26 WPI and hepatic immune responses were examined after 10 days of injection. (A) Foxp3 mRNA expression by RT-PCR. (B) Parasite burden expressed by qPCR. (C) The number of infected foci from 25 consecutive microscopic fields. (D) Frequency of mature (black column) and immature (grey column) granuloma, and no response (white column) in the infected foci. (E) Localization of Foxp3-stained cells in the liver parenchyma including “Immature granulomas” and “Mature granulomas”. The brown pigments represent Foxp3-immunoreactive cells. * $p<0.05$; *** $p<0.001$.

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infection models. Disruptive architecture of the thymus and spleen could affect the development and expansion of T cells. Several studies using bone marrow chimeras between aly/aly and wild type mice showed that antiviral CTL responses were clearly improved in the wild type environment [11]. However, expansion of CD25+CD4+ Treg is impaire in the spleen of aly/aly mice [35]. This suggests that expansion of functional CD4+Foxp3+ Treg in the liver of aly/aly mice during L. donovani infection is likely related to the parasite persistence but not to the structural defects of secondary lymphoid organs although this possibility will be confirmed by BM chimera experiments in future.

The NIK gene mutation may contribute to other immune defects due to the partial blocked NF-κB signaling for activation of dendritic cells and macrophages. NIK TLR signaling which is involved in canonical TLR/TNFR pathway. Perhaps due to less number of B cells (unpublished data; Ato M., Kaye PM). NF-κB may be involved in antiviral immunity [37–39]. In the present study, 10 days after the third injection of infected aly/aly mice with anti-CD25 and anti-CD4 mAb, the hepatic parasite burdens were reduced by 88% and 89% of that control mice, respectively (Figure 7B). Likewise, the spleen of aly/aly mice showed impaired antigen presentation ability. Lower hepatic parasite loads was unexpectedly observed in aly/aly mice than aly/+ mice in the first 4 WPI. This may be due to lower number of the sessile Kupffer cells (unpublished data), but associated with the strong innate immunity as reported during Listeria monocytogenes infection in aly/aly mice [12]. Partial hepatic granuloma progression and neo-lymphoid aggregates in aly/aly mice imply that mice lacking secondary lymphoid tissues can still generate T cell-mediated immune responses to some extent [32].

Although anti-CD25 mAbs have been used for depletion of Tregs in various experimental cases, administration of anti-CD4 mAb also reduced Treg numbers and provoked effective tumor immunity [37–39]. In the present study, 10 days after the third injection of infected aly/aly mice with anti-CD25 and anti-CD4 mAb, the hepatic parasite burdens were reduced by 88% and 89% of that control mice, respectively (Figure 7B). Likewise, treatment with either anti-CD25 or anti-CD4 mAb also reduced parasite burdens in the spleen and BM (Figure S3). The reason why anti-CD4 mAb was more effective than anti-CD25 mAb in reducing parasite burden is unknown, but the present study is the first to report effectiveness of anti-CD4 mAb to control systemic infection of L. donovani in mice. Thus, anti-CD4 antibodies may be an alternative measure to manipulate Tregs in chronic VL. However, since anti-CD25 mAb can also affect effector T cells and effective immunity [39], and anti-CD4 mAb can also deplete a small population of CD4+Foxp3+ T cells in the lymph node [38], probably including IL-10-producing conventional CD4+ T cells, further studies of the role of Tregs in VL are required.

Studies of Tregs in cutaneous leishmaniasis demonstrated the involvement of CD4+CD25+ Tregs in cutaneous leishmaniasis caused by L. major [40,41] and L. amazonensis in mice [42] and by L. braziliensis in humans [43]. Regarding VL, the role of Tregs is uncertain and the primary source of IL-10 is controversial. In the spleen of VL patients in India, CD4+CD25+ Foxp3+ cells were identified as the major producers of IL-10 [44]. In L. infantum-infected BALB/c mice, CD4+CD25+ Foxp3+ cells expanded in a pooled fraction of draining lymph nodes and spleen cells at 7 and 28 days of infection [45]. In L. donovani-infected BALB/c mice, the number of splenic CD4+CD127lowCD25+Foxp3+ T cells expressing higher Foxp3 and IL-10 increased at 21 days of infection [46]. IL-10 production by splenic CD4+CD25+Foxp3- IL10+ T cells, representing type 1 regulatory T (Tr1) cells, was a strong correlate of disease progression in L. donovani-infected C57BL/6 mice [47]. Further analyses using quantitative RT-PCR of IL-10 and Foxp3 transcripts in selected populations of CD25+ and CD25- enriched hepatic CD4+ T cells, and/or by intracellular cytokine staining, will elucidate the issue. Nevertheless, in the present study, Treg and IL-10 augment immunosuppressive effects in hepatic granuloma of L. donovani-infected aly/aly mice. Maintenance of relatively higher expression levels of TGF-β in the chronic phase of the infection in aly/aly mice may be related to the generation and maintenance of CD4+Foxp3+ Tregs [48] rather than the inhibition of granuloma maturation [49].

In conclusion, we focused on immune responses to the chronic phase of murine VL caused by L. donovani infection in an immunodeficient host. In the last decade when the number of visceral leishmaniasis in immunocompromised patients has been increasing, our data presented herein offered a novel insight into the possibly involvement of CD4+Foxp3+ Tregs in persistent L. donovani infection in the liver of immunodeficient hosts. The manipulation of Tregs may provide a promising immunotherapeutic strategy for VL.

Supporting Information

Figure S1 Hepatic immune response and granuloma formation in aly/+ and aly/aly mice 2–8 weeks after L. donovani infection. Representative hepatic immune responses to infected foci in liver sections by staining with HE and immunostaining with anti-Leishmania serum for aly/+ (A–D and E–H, respectively) and aly/aly (I–L and M–P, respectively) mice are shown. Immune responses to parasitized Kupffer cells were categorized into four types; “No granuloma” (A and E, I and M), “Immature granuloma” (B and F, J and N), “Mature granuloma” (C and G, K and O) and “Involuting granuloma” (D and H, L and P). The brown pigments indicate L. donovani amastigotes. The yellow circles indicate L. donovani-infected foci in each type of tissue responses. (PDF)

Figure S2 Proportion of CD4+Foxp3+ and CD4+ T cells gated hepatic lymphocytes during the course of L. donovani infection by flow cytometry analysis. Hepatic lymphocytes were isolated from the liver of aly/+ and aly/aly mice at indicated time points after L. donovani infection and stained with anti-CD4, CD8 and Foxp3 antibodies. Percentage of CD4+ T cells (white bars) and CD4+Foxp3+ T cells (black bars) of aly/+ (A) and aly/aly mice (B) are shown. (PDF)

Figure S3 Effects of anti-Treg antibody treatment on parasite burden in the spleen and bone marrow. The L. donovani-infected aly/aly mice were intraperitoneally injected with anti-CD25 or anti-FR4 mAb at 26 WPI, and parasite burden in the spleen (A) and bone marrow (B) was estimated by qPCR after 10 days of antibody treatment. * p<0.05; ** p<0.01. (PDF)

Table S1 Target genes and primers for qPCR and RT-PCR used in this study. (DOCX)

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References

Author Contributions
Conceived and designed the experiments: ST KI MA TS KK. Performed the experiments: ST KI KK. Analyzed the data: ST KI MA HK. Wrote the paper: ST KI MA HK KK.

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