Periodontal repair following implantation of beta-tricalcium phosphate with different pore structures in class III furcation defects in dogs

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The aim of this study was to investigate the effect of the pore characteristics of β-tricalcium phosphate (β-TCP) on periodontal healing in class III furcation defects in dogs. Two types of β-TCP were prepared for grafting: 1) a tunnel pipe structure with an inner diameter of 300 μm, and 2) continuous pore structure with interconnected macropores. The furcations of thirty mandibular premolar teeth were implanted with each type of β-TCP or were left untreated as control. The dogs were sacrificed 8 weeks post-surgery, and healing was evaluated histologically. Downgrowth of junctional epithelium in the tunnel structure group was significantly more than that in the other two groups (p<0.01). There was significantly more new bone formation and new cementum formation in the tunnel structure group than in the other two groups (p<0.01). These findings suggested that β-TCP with a tunnel pipe structure promotes periodontal healing in class III furcation defects.

Keywords: Bone grafting, Regeneration, Animal model

INTRODUCTION

There is an increasing need for periodontal reconstruction in clinical dentistry. Many researchers have shown that the implantation of biomaterials promotes periodontal regeneration in clinical and animal model studies. Bioceramic characteristics, such as porosity, density, and crystallinity, affect the outcome of periodontal regeneration. Several studies have reported that porous structures enhance bone formation in bone defects. Furthermore, it has been shown that pore size has an effect on vascularization. Pore sizes greater than 140 μm induced earlier vessel formation in biomaterials when compared with pore sizes smaller than 140 μm or dense particles. Also, new vessels arising from the residual tissue can increase the number of vessels in macropores, whereas only a few vessels in a macropore could extend into the adjacent macropore through the small connecting area with pore sizes smaller than 140 μm. Vascularization in the defect ensures an adequate supply of nutrients and delivery of cells and growth factors that support the formation of osseous tissue.

Klenke et al. reported that a porous body with a continuous 100–400 μm pore structure increased new bone formation. Therefore, we hypothesized that vascularization would enable delivery of cells and supply of nutrients and oxygen to the cells during the initial healing stage without the small connecting area between macropores.

Beta-tricalcium phosphate (β-TCP) belongs to the group of bioactive bioceramics of completely synthetic origin and has been used in periodontology as a bone graft substitute since the 1980s. At present, there are various types of β-TCP composed of macropores with small connecting areas. We devised a tunnel pipe structure that maintained a continuous pore of 300 μm. After preparation of cylindrical β-TCP with randomly-arranged openings as the building block, the β-TCP was prepared by sintering the samples.

The purpose of this study was to evaluate the effect of the tunnel pipe structure on periodontal regeneration following implantation of β-TCP in surgically created class III furcation defects in dogs.

MATERIALS AND METHODS

Animals
The experimental protocol (No. 08-0257) followed the guidelines for the care and use of laboratory animals of the Graduate School of Medicine, Hokkaido University. Thirty mandibular premolar teeth of five beagle dogs (female, 1 year old; mean weight, 9.8 kg) were used. The dogs were subjected to plaque control measures.
Fig. 1  Macrostructure and microstructure of tunnel-pipe β-TCP and continuous porous β-TCP.
(a) A cube with 5 mm sides was formed by gathering and connecting irregularly arranged particles of tunnel-pipe β-TCP. (b) The tunnel: inner diameter 300 μm, outer diameter 500 μm and length 1–2 mm. (c) A particle with a diameter of 1,000–2,000 μm and 60% porosity. (d) A macropore size of 100–400 μm with interconnectivity. (e) The tunnel-pipe β-TCP was composed of highly crystalline and single-phase β-TCP, and no other phases were observed.
μm with interconnectivity, a diameter of 1,000–2,000 μm and 60% porosity (Figs. 1c and 1d). This was also made of single-phase β-TCP and the average crystal size was 172 nm8).

Surgery and implantation
The surgical procedures were performed under general anesthesia with medetomidine hydrochloride (5 μg/kg, Domitor®, Meiji Seika, Tokyo, Japan) and ketamine hydrochloride (2.9 mg/kg, Ketaral 50®, Sankyo, Tokyo, Japan) and local anesthesia with lidocaine hydrochloride (2% with 1:80,000 epinephrine, Xylocaine®, DENTSPLY SANKIN, Tokyo, Japan). After reflection of a mucoperiosteal flap, the interradicular bone of all second, third and fourth premolars was completely removed with rotating burs under irrigation and hand micro Ochsebein chisels to create through-and-through furcation defects. The defect height from the cemento-enamel junction (CEJ) to the reduced alveolar crest was 4 mm. The exposed root surfaces were planed with curettes in order to remove the periodontal ligament and cementum. At this point, the teeth were randomly assigned to one of three treatment groups based on a random computer-generated list.

In the T group (n=10), β-TCP was shaped using a diamond bur to fit the furcation space. Then, β-TCP was implanted into the furcation and the remaining space was filled with the β-TCP pipe particles. In the CP group (n=10), the furcation defect was filled with continuous porous β-TCP. The furcation was not treated in the remaining ten teeth, which served as control.

After implantation, the flaps were positioned with the gingival margins of the flaps at the CEJ and sutured.

Wound management
Sutures were removed 10 days after the surgical procedure. Plaque control measures included weekly brushing and application of 0.5% chlorhexidine gluconate solution throughout the healing period.

Histological processing and histometric analysis
The dogs were sacrificed 8 weeks after the surgical procedure. Tissue blocks including teeth, bone and soft tissues were resected. Blocks were fixed in 10% buffered formalin, decalcified in 10% ethylenediamine tetraacetic acid (EDTA, Wako, Tokyo, Japan) solution, trimmed, dehydrated, and embedded in paraffin. Serial sections, 4 μm thick, were prepared in the mesio-distal plane. Sections of the middle portion at bucco-lingual width of the crown were stained with hematoxylin and eosin and tartrate-resistant acid phosphatase (TRAP) stain.

Measurements of each specimen were carefully performed by one examiner (YH), who was blinded to the study protocol. The following measurements were performed using Scion Image® (National Institute of Health) (Fig. 2). For determination of the defect size, the area from the mesial and distal end points of root planing to the fornix was regarded as the bone defect area (BDA). The tissues formed from the apical border in a coronal direction after each treatment were regarded as newly formed tissue. The following parameters were also determined: 1) the percentage of regenerated bone area, calculated by the regenerated bone area/BDA×100; 2) the percentage of space composed of new bone and β-TCP, calculated by the regenerated bone area and remaining β-TCP area/BDA×100; 3) the percentage of epithelium, calculated by the linear extension of root surface covered by epithelial tissue/length of the root planed surface from the mesial end to the distal end (RL)×100; and 4) the percentage of new cementum length, calculated by the total length of regenerated cementum or cementum-like deposit on the root/RL×100.

The number of blood vessels inside the pore was calculated using Scion Image®.
Fig. 3  A representative case from the T group.
(a) Overview: Note the extensive new bone (black arrow) formation. (b) New bone had formed on the outer surfaces of β-TCP near the fornix of the furcation. (c) Several blood vessels (black asterisks) had formed in the center of the pipe tunnel. (d) New connective tissue fibers were oriented parallel or inclined to the root surface and embedded in both new cementum (NC) and new bone. (e) through (h) Numerous multinucleated cells (blue asterisks) were seen throughout the entire furcation. ((a) through (d)) Hematoxylin and eosin. ((e) through (h)) Tartrate-resistant acid phosphatase. Filled arrowhead=the apical end of root-planed surface; d=root dentin.

Fig. 4  A representative case from the CP group.
(a) through (e) Continuous porous β-TCP group: New bone (black arrow) was observed near the residual bone surrounding the β-TCP. (b) Gingival epithelium (E) was observed in the fornix of the furcation. (c) New connective tissue fibers were oriented parallel to the root surface and embedded in both new cementum and new bone on β-TCP. (d) and (e) The new bone was observed on the inner surfaces of β-TCP. In the macropore of β-TCP, several blood vessels were observed. In contrast, one or few vessels were observed in the area connecting macropores (black asterisks). ((a) through (e)) Hematoxylin and eosin. Filled arrowhead=the apical end of root-planed surface.
Data analysis
The mean±SD of each measurement was calculated for each tooth from selected sections. Statistical differences were analyzed using the Kruskal-Wallis test. The number of blood vessels was analyzed using the Mann-Whitney U test. All statistical procedures were performed using SPSS 10.0 J (S.P.S.S. Co. Ltd., Tokyo, Japan)

RESULTS

Clinical observations
Clinical healing after the surgical procedures was uneventful with minimum indications of inflammation throughout the experimental period.

Histological observations
In the T group, long junctional epithelium was not observed in the furcation of 7/10 specimens. New bone was observed up to the fornix of the furcation (Figs. 3a and b). Several blood vessels had formed in the center of each pipe tunnel directed horizontally (Fig. 3c). Numerous multinucleated cells adhered to the β-TCP surface and there was new bone throughout the entire furcation (Figs. 3e–h). New connective tissue fibers oriented parallel or inclined to the root surface and embedded in both new cementum and new bone were observed (Fig. 3d).

In the CP group, long junctional epithelium was observed in the furcation of all specimens (Figs. 4a and b). New bone was observed on the inner and outer surfaces of β-TCP. In the macropore of β-TCP, several blood vessels were observed. In contrast, one or few vessels were observed in the small connecting area between macropores (Figs. 4d and e). New cementum with inserting collagen fibers had formed on the root-planed surface (Fig. 4c).

In the C group (Fig. 5), gingival epithelium was observed in the furcation of all specimens and inflammatory cells appeared in the connective tissue under the epithelium. Limited new bone was observed near the residual bone of the furcation defects. Limited amounts of new cementum with inserting collagen fibers were observed on the root-planed surface.

Histometric analysis
There were no significant differences in bone defect area among the three groups. The epithelium was significantly less in the T group than in the C and CP groups (p<0.01) (Table 1). The regenerated bone area and space composed of regenerated bone and β-TCP were significantly greater in the T group than in the C and CP groups (p<0.01) (Table 1). There was 69.4±6.1% newly formed cementum in the T group, significantly greater than in the CP and C groups (p<0.01) (Table 1). The mean number of inner blood vessels was 16.5±1.8 in the T group and 5.2±0.9 in the CP group. In the CP group, the large pore area was 9.0±0.9 and small pore area was 1.2±0.2. The number of blood vessels was significantly greater in the T group than in the CP group (Table 2).

DISCUSSION

Previous studies have reported varied outcomes of periodontal regeneration therapy in furcation defects27-29). It was speculated that periodontal regeneration might be affected by local factors, such as the width and height of furcation defects and the size of the entrance of the furcation. As the characteristics of the first cells to populate a wound dictate the nature and quality of tissue that forms there, it is important that cells from the periodontal ligament and alveolar bone are allowed to initially recolonize periodontal wounds, rather than cells from soft fibrous connective tissue and gingival epithelium30).

The regenerated bone area in the T group was significantly larger than in the C and CP groups in this study. This could be attributed to a difference in the characteristics of the implant materials. The phase
Table 1  Histometric analysis of periodontal healing (Group mean±SD in percentage)

<table>
<thead>
<tr>
<th>Parameter</th>
<th>T group (n=10)</th>
<th>CP group (n=10)</th>
<th>C group (n=10)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Regenerated bone area</td>
<td>38.6±4.3*†</td>
<td>30.8±3.4</td>
<td>24.2±9.3</td>
</tr>
<tr>
<td>Space composed of regenerated bone and β-TCP</td>
<td>72.8±1.3*†</td>
<td>57.6±9.3‡</td>
<td>24.2±9.3</td>
</tr>
<tr>
<td>Epithelium</td>
<td>6.94±5.6*‡</td>
<td>29.5±8.8</td>
<td>48.2±3.1</td>
</tr>
<tr>
<td>New cementum</td>
<td>69.4±6.1*†</td>
<td>51.6±7.8‡</td>
<td>23.4±11.6</td>
</tr>
</tbody>
</table>

\( n=\) number of sites; \( P \) value by Kruskal-Wallis test.
* Significantly different from CP group (\( p<0.01 \)).
† Significantly different from C group (\( p<0.01 \)).
‡ Significantly different from C group (\( p<0.05 \)).

Table 2  Number of vessels in β-TCP (Group mean±SD)

<table>
<thead>
<tr>
<th>Parameter</th>
<th>T group (n=10)</th>
<th>CP group (n=10)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of vessels</td>
<td>16.5±1.8</td>
<td>5.2±0.9*</td>
</tr>
<tr>
<td></td>
<td></td>
<td>large pore area</td>
</tr>
<tr>
<td></td>
<td></td>
<td>small pore area</td>
</tr>
</tbody>
</table>

\( n=\) number of sites; \( P \) value by Mann-Whitney U test.
* Significantly different from T group (\( p<0.01 \)).
† Significantly different from Micro pore of CP group (\( p<0.01 \)).

purity of the two types of β-TCP used in this study was 100% beta phase and the crystal size was similar. However, the porosity of the tunnel-pipe β-TCP (72%) was greater than that of continuous porous β-TCP (60%). These results confirmed the findings of previous reports which demonstrated that high porosity promoted greater bone formation\(^{31-32}\). The volume of implanted β-TCP remaining within the furcation in the T group was less than that in the CP group, which could explain the difference in the porosity of the implanted material. The amount of β-TCP degradation in the T group was significantly less than that in the CP group. Although higher porosity has been shown to promote resorption of β-TCP and replacement with bone tissue, the results of the present study did not support these findings. Resorption is affected by cellular phagocytosis and surrounding pH; however, β-TCP degradation would have been inhibited because the surface of β-TCP was covered with new bone. When osteoconduction is greater, new bone would cover the surface of β-TCP. Osteoconduction is dependent on not only the porosity but also the pore diameter. In this study, there were significantly more blood vessels in the pore of the β-TCP in the T group than that in the CP group. The tunnel structure might have served as an efficient conductor for vessel invasion. According to Rodriguez et al., vascularization is crucial for the development and repair of most tissues, and is a prerequisite for the healing of bone defects\(^{30}\). Vascularization of the implantation site ensures adequate supply of nutrients, prompt removal of metabolic by-products, and delivery of cells and growth factors that support the formation of osseous tissue. In the T group, a number of vessels could be observed in the center of the tunnel structures, whereas, in the CP group, several vessels were observed in the macropore structures with only one or a few vessels in the small connecting area between macropores. The small number of vessels in the small connecting area would limit the supply of cells to the fornix of the furcation in the CP group. On the other hand, the presence of vessels in the tunnel structure might have supplied cells and supported the formation of osseous tissue towards the fornix of the furcation.

Regarding the downgrowth of gingival epithelium, several studies have reported that the implantation of dense β-TCP in surgically created intrabony and furcation defects may result in the formation of a long epithelial attachment similar to that in a non-implant group\(^{34-38}\). It is suggested that β-TCP might have little effect on the recolonization of the gingival epithelium. However, in this study, downgrowth of gingival epithelium in the T group and CP group was significantly smaller than
in the C group. In addition, downgrowth of gingival epithelium in the T group was significantly smaller than in the CP group. Vascularization of the implantation site in the early healing stage might affect the downgrowth of epithelium in the furcation defects by promoting the formation of connective tissue containing bone, periodontal ligament and gingival connective tissue. The connective tissue formation would inhibit downgrowth of the gingival epithelium in the furcation defects. From this point of view, the tunnel structure would decrease the downgrowth of gingival epithelium in comparison to the CP group.

New cementum formation in the T group was greater than in the CP and C groups. In the T group, new cementum was observed as deposition on the resorption lacunae in part of the root dentin. Because odontoclasts have expression patterns of cathepsin K and matrix metalloproteinase-9 (MMP-9) similar to osteoclasts, the odontoclasts could have been stimulated by the active bone formation and high bone turnover. Götz et al. reported that cementoblasts were involved in the resorption-repair sequence, which is considered to be a coupling process as in bone. Similarly, preparation for new cementum formation might start adjacent to resorption areas. Multinucleated cells adhered to not only the β-TCP surface but also new bone throughout the entire furcation. Consequently, the tunnel structure of β-TCP would also induce new cementum formation.

CONCLUSION

The present findings demonstrate that the tunnel structure of β-TCP induced an increase in cells and blood vessels derived from the residual bone, which enhanced periodontal regeneration in class III furcation defects.

ACKNOWLEDGMENTS

This study was supported by Grants-in-Aid for Encouragement of Young Scientists (22792078 to Dr. E. Saito) and Scientific Research (22592307 to Dr. A. Saito) from the Ministry of Education, Science, Sports and Culture of Japan. The authors report no conflicts of interest related to this study.

REFERENCES

22) Knabe C, Koch C, Rack A, Stiller M. Effect of beta-tricalcium


