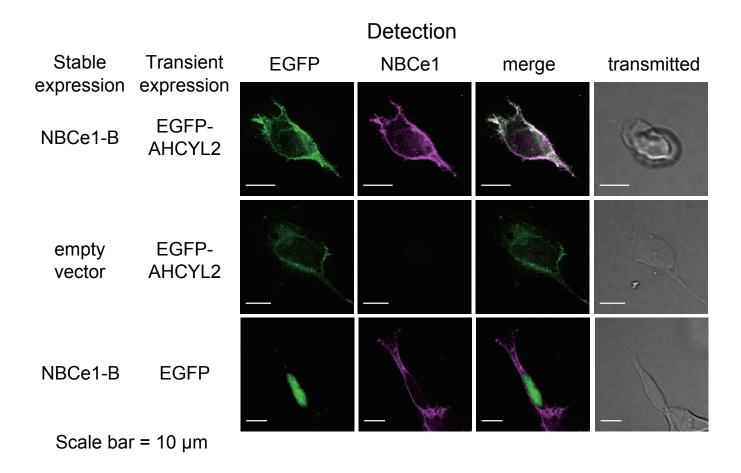
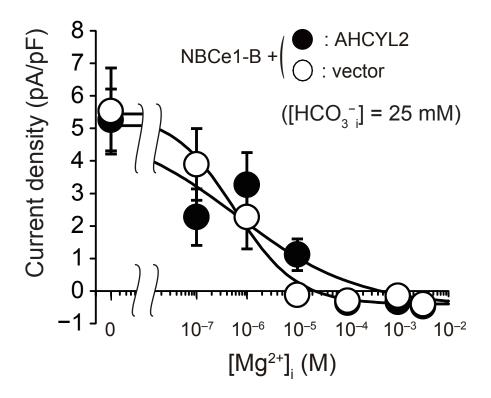
Title	AHCYL2 (long-IRBIT) as a potential regulator of the electrogenic Na+-HCO3- cotransporter NBCe1-B
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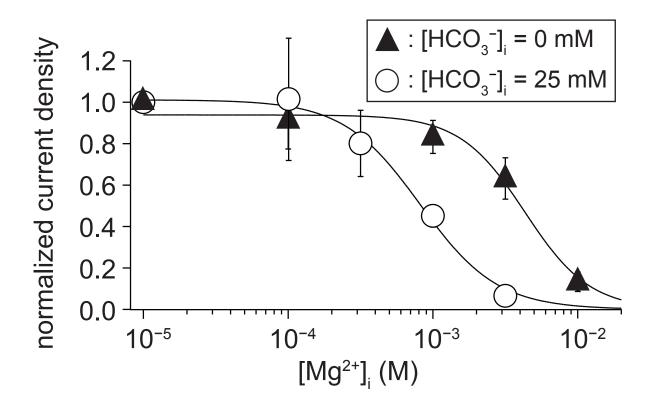




Supplemental Figure S1 Yamaguchi S. and Ishikawa T.



Supplemental Figure S2 Yamaguchi S. and Ishikawa T.



Supplemental Figure S3 Yamaguchi S. and Ishikawa T.

#### SUPPLEMENTAL FIGURE LEGEND

## **Supplemental Figure S1**

Co-localization of AHCYL2 and NBCe1-B exogenously expressed in HEK293 cells EGFP-tagged AHCYL2 or EGFP were transiently expressed in HEK293 cells stably transfected with NBCe1-B or mock transfected cells (empty vector). Cells were stained with mouse anti-NBCe1 antibody and Alexa 594 goat anti-mouse IgG antibody. AHCYL2 was detected by the fluorescence of EGFP (green), and NBCe1-B by fluorescence of Alexa 594 (magenta) by confocal microscopy. Cells were permeabilized with saponin to wash out cytosolic unbound proteins prior to fixation. Co-localization of AHCYL2 and NBCe1-B is indicated as merge and with white color. Scale bars, 10 μm.

### **Supplemental Figure S2**

AHCYL2 does not change  $Mg^{2+}_{i}$ -sensitivity of NBCe1-B currents stably transfected in HEK293 cells when recorded using pipette solutions with added 25 mM HCO<sub>3</sub><sup>-</sup> Dose-response curves for the inhibition of NBCe1-B currents by increasing free  $Mg^{2+}_{i}$  concentrations in the stably NBCe1-B-expressing cells, which were transiently transfected with AHCYL2 (solid circles). Shown are mean  $\pm$  S.E. (n = 8-11). The lines were fits to the Hill equation. The data for the stably NBCe1-B-expressing cells transiently transfected with empty vector alone (open circles) were taken from a recently published work (Yamaguchi & Ishikawa 2012) and are shown for comparison.

### **Supplemental Figure S3**

Mg<sup>2+</sup>i sensitivity of native NBCe1-B-like current in BPA cells under a nominally

# HCO<sub>3</sub> i-free condition

Dose-response curve for  $Mg^{2+}_{i}$ -inhibition of native NBCe1-B-like currents recorded from BPA cells using a nominally-HCO<sub>3</sub><sup>-</sup>-free pipette solution (filled triangles). NBCe1-B-like current densities at 0 mV were normalized to that at  $10^{-5}$  M free  $Mg^{2+}$  obtained from a BPA cell derived from the same bovine parotid gland. Shown are mean  $\pm$  S.E. (n = 3-11). The line is a fit to the Hill equation ( $K_i$  value:  $4.3 \times 10^{-3}$  M; Hill coefficient: 1.93). The data obtained under a  $HCO_3^-$ -loaded condition (open circles) were taken and adapted from Yamaguchi & Ishikawa (2008) and plotted for comparison. The current density values before normalization at  $10^{-5}$  M free  $Mg^{2+}$  with/without  $HCO_3^-$  were  $5.4 \pm 0.7$  and  $8.1 \pm 0.7$  pA/pF (n = 23 and 14), respectively.

#### MATERIALS AND METHODS FOR SUPPLEMENTAL FIGURES

## Immunofluorescence confocal microscopy

N-terminally EGFP-tagged AHCYL2 (pEGFPC1 vector) or EGFP alone was expressed in HEK293 cells stably transfected with bovine NBCe1-B or empty vector (pCIneo). The transfected cells were permeabilized in permeabilization buffer (80 mM PIPES, 1 mM MgCl<sub>2</sub>, 1 mM EGTA, and 4% polyethylene glycol, pH 7.2 with KOH) containing 0.1% saponin for 10 min on ice, and washed twice with ice-cold permeabilization buffer prior to fixation. The cells were fixed in PLP fixative (2% paraformaldehyde, 75 mM lysine, 37 mM Na phosphate, 10 mM Na periodate, pH 7.4) overnight at 4 °C, permeabilized in 1% SDS in PBS for 5 min, and blocked in image-iT signal enhancer (Life Technology) for 30 min and in PBS containing 5% normal goat serum and 0.2% BSA for 60 min. Cells were then stained with rabbit anti-NBCe1 antibody (1:1,000,

Chemicon). Following four times 5-min PBS washes, Alexa 594-conjugated goat anti-rabbit IgG (Life Technology) was applied for 1 hr at room temperature. Following four times 5-min PBS washes, the coverslips were mounted with ProLong Gold Antifade Reagent (Life Technology) and observed under IX-70 confocal fluorescence microscopy (Olympus) with a 60× oil-immersion objective.

### Whole-cell patch clamp experiment

Pipette solutions (pH = 7.4 with NMDG (N - methyl - D - glucamine)) contained (in mM): 10 BAPTA (1,2-Bis (2-aminophenoxy) ethane- N,N,N',N'- tetraacetic acid), 100 HEPES, 4 EDTA-2Na (ethylenediaminetetraacetic acid disodium salt), 2 NaHCO<sub>3</sub>, 23 cholineHCO<sub>3</sub>, 0-8 MgCl<sub>2</sub> (no MgCl<sub>2</sub> or appropriate amounts of MgCl<sub>2</sub> were added to yield  $Mg^{2+}$ -free or  $10^{-7}$  to  $10^{-2.5}$  M free  $Mg^{2+}$ , respectively), 0.044 - 0.202 CaCl<sub>2</sub> ( $10^{-9}$ M free Ca<sup>2+</sup>), 13-25 NMDG-glutamate, and 0-15 NMDG-Cl. Free Mg<sup>2+</sup> and Ca<sup>2+</sup> concentrations calculated using the program Maxchelator were (http://www.stanford.edu/~cpatton/maxc.html). The concentrations of NMDG-glutamate and NMDG-Cl were also varied to maintain the chloride concentration (14-16 mM Cl<sup>-</sup>). Pipette solutions with no added HCO<sub>3</sub> were the same as those described in the manuscript. The solutions containing HCO<sub>3</sub> were bubbled with 5% CO<sub>2</sub>/95% O<sub>2</sub>. The average series resistance of HEK293 cells or BPA cells, which was not electrically compensated, was  $23.1 \pm 0.6 \text{ M}\Omega$  (n = 60) or  $22.3 \pm 0.7 \text{ M}\Omega$  (n = 60) 45), respectively. The cell capacitance was  $18.3 \pm 1.0$  pF (n = 60) or  $32.6 \pm 1.3$  pF (n =45), respectively. Other experimental conditions were the same as those described in the manuscript.

# SUPPLEMENTAL REFERENCES

Yamaguchi S & Ishikawa T (2008). The electrogenic Na<sup>+</sup>-HCO<sub>3</sub><sup>-</sup> cotransporter NBCe1-B is regulated by intracellular Mg<sup>2+</sup>. *Biochem Biophys Res Commun* **376**, 100-104.

Yamaguchi S & Ishikawa T (2012). IRBIT reduces the apparent affinity for intracellular Mg<sup>2+</sup> in inhibition of the electrogenic Na<sup>+</sup>-HCO<sub>3</sub><sup>-</sup> cotransporter NBCe1-B. *Biochem Biophys Res Commun* **424**, 433-438.