Title

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Citation

Organic & biomolecular chemistry, 11(38): 6615-6622

Issue Date

2013-10-14

Doc URL

http://hdl.handle.net/2115/56729

Type

article (author version)

File Information

WoS_63009_Shuto.pdf
Design and Synthesis of the Stabilized Analogs of Belactosin A with the Unnatural 
\textit{cis}-Cyclopropane Structure

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Running Title: Stabilization of the \textit{cis}-Belactosin A Analogs

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Abstract

The belactosin A analog \(2a\), having the unnatural \(cis\)-cyclopropane structure instead of the \(trans\)-cyclopropane structure in belactosin A, is a much more potent proteasome inhibitor than belactosin A. However, its cell growth inhibitory effect is rather lower than that expected from its remarkable proteasome inhibitory effect, probably due to its instability under cellular conditions. We hypothesized that the instability of \(2a\) was due to its chemical and enzymatic hydrolysis of the strained \(\beta\)-lactone moiety. Thus, to increase the stability of \(2a\) by chemical modification, its analogs with a sterically more hindered \(\beta\)-lactone moiety and/or cyclopropyl strain-based conformational restriction were designed and synthesized, resulting in the identification of a stabilized analog \(6a\) as a proteasome inhibitor with cell growth inhibitory effects. Our findings suggest that the chemical and biological stability of \(2a\) is significantly affected by the steric hindrance around its \(\beta\)-lactone carbonyl moiety and the conformational flexibility of the molecule.

Introduction

The chemical and biological stability of small molecules depend on their chemical structures, and therefore it can be regulated by structural modifications.\(^1\) In the drug discovery process, compounds with insufficient stability often degrade rapidly \(in\) \(vivo\) and sometimes bind covalently to off-target molecules, resulting in the absence of the desired pharmacological effect, and even worse, producing an undesired toxic side-effect.\(^2\) The chemical and biological instability of compounds can be improved by changing the steric and/or electrostatic properties of the labile moiety. Furthermore, when the compound is unstable \(in\) \(vivo\) due to enzymatic degradation, it can be stabilized by changing of such structural features as molecular size, electrostatic property, hydrophobicity, and conformation to reduce the affinity for the degrading enzyme.

The ubiquitin-proteasome system is the major degradation pathway of intracellular proteins,\(^3\) which are involved in many physiologically important cellular processes, such as signal transduction,\(^4\) cell cycle progression,\(^5\) and unfolded protein response (UPR).\(^6\) Because inhibition of the proteasome causes cell cycle arrest to induce apoptosis, proteasome is an attractive target for the development of anti-cancer drugs.\(^7\) For example, a proteasome inhibitor bortezomib is clinically effective for the treatment of the multiple myeloma\(^8\) and mantle cell lymphoma.\(^9\)

Belactosin A is a proteasome inhibitor isolated from the \(Streptomyces\) sp. by Asai,\(^10\) which inhibits proteasome covalently by acylating the active site Thr residue \(via\) ring-cleavage of its strained \(\beta\)-lactone moiety.\(^11\) Because the binding site of belactosin derivatives differs from that of other proteasome inhibitors,\(^11\) belactosin A is an attractive potential lead for the development of novel proteasome inhibitors. In recent years, we have investigated the three-dimensional structure activity relationship (SAR) study of belactosin A and identified the unnatural \(cis\)-cyclopropane isomer \(1\) as a more potent proteasome inhibitor than belactosin A having the \(trans\)-cyclopropane structure.\(^13\) Furthermore, we investigated the SAR of \(1\) to result in identification of the optimized inhibitor \(2a\), which appeared to be as potent as the clinical drug bortezomib (Figure 2).\(^14\) Despite its remarkable proteasome inhibitory effect, however, its inhibitory effect on cell growth is not so strong, compared with other potent inhibitors such as bortezomib\(^15\) or carfilzomib,\(^16\) as summarized in Table 1. In our previous study, we investigated the stability of \(2b\), instead of \(2a\) due to its poor solubility in aqueous medium, and demonstrated that \(2b\) is gradually degraded in aqueous medium, while its half-life (\(t_{1/2} = 10\) h in pH 7.4 buffer)\(^14b\) is longer than that of other \(\beta\)-lactone-type proteasome inhibitors (omuralide, 13 min; salinosporamide A, 56 min).\(^17\) Furthermore, it was found that \(2b\) is significantly unstable under biological conditions (\(t_{1/2} = 2.3\) min in serum), which might be correlated with the relatively weak cell growth
inhibitory effect of 2a, because 2a can be as unstable as 2b. Thus, we planned to develop stabilized derivatives of 2a. Here we describe the design, synthesis, biological activities, and chemical and biological stability of the newly designed compounds.

Figure 1. Known proteasome inhibitors
Table 1. Inhibitory effect of 2a, bortezomib and carfilzomib on proteasome chymotrypsin-like (CT-L) activity and HCT116 cell growth

<table>
<thead>
<tr>
<th>compound</th>
<th>IC$_{50}$ [nM]</th>
<th>IC$_{50}$ ratio (cell growth/CT-L activity)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>proteasome (CT-L activity)</td>
<td>cell growth (HCT116)</td>
</tr>
<tr>
<td>2a</td>
<td>5.7</td>
<td>1820</td>
</tr>
<tr>
<td>bortezomib</td>
<td>4.5</td>
<td>5.0</td>
</tr>
<tr>
<td>carfilzomib</td>
<td>6.3</td>
<td>8.5</td>
</tr>
</tbody>
</table>

Figure 2: Previous SAR studies of belactosin A performed by our laboratory.

Figure 3: Stability of 2b in 0.1 M TEAA buffer (pH 7.4) and human AB serum at 37 °C

Results and Discussion

Design of compounds. The reactivity of the carbonyl group with nucleophiles is affected by the steric hindrance...
around its carbon atom, and we therefore designed compounds 3a-5a with various substituents at the \( \alpha \)-carbon of the \( \beta \)-lactone carbonyl group of 2a (Figure 4-a) to change the bulkiness at the position. The order of the steric hindrance around the \( \beta \)-lactone carbonyl group is thought to be 3a < 2a < 4a < 5a, as depicted in Figure 4-a.

On the other hand, because enzyme recognition can be influenced by the three-dimensional structure of the substrate, conformational restriction of 2a and its analogs might result in lowering the affinity for the degrading enzyme, and we therefore designed 6a as a conformationally restricted analog. The cis-oriented adjacent substituents on the cyclopropane ring are fixed in the eclipsed orientation, and accordingly, they exert significant mutual steric repulsion, which we previously termed “cyclopropylic strain”. Due to this characteristic structural feature, conformation of the substituents (Figure 5-a) on a cyclopropane ring can be restricted, and therefore, in compound 5a, conformer A (anti, the cyclopropane ring “down”/the side chain “up”) and B (syn, the cyclopropane ring “down”/the side chain “down”) would be preferable (Figure 5-b). Previously, we demonstrated that the bioactive conformation of the cis-cyclopropane belactosin derivatives seems to be syn. Therefore, we designed conformationally restricted analog 6a (Figure 4-b), whose conformation is restricted in the syn-form due to the significant steric repulsion between the introduced 1’R-methyl group and the cis-oriented amide group in its anti-form (Figure 5-b). Notably, this cyclopropylic strain-based conformational restriction can be achieved by the minimal structural change, i.e., only the introduction of a methyl group, allowing us to more rigorously investigate the relationship between the conformation and the stability.

The compounds 3a-6a were thought to be poorly soluble in aqueous medium, therefore we also planned to synthesize compounds 3b-6b, which are analogs of 3a-6a without the N-terminal Cbz group, to evaluate their stability under aqueous conditions instead of 3a-6a.

![Figure 4. Structure of newly designed compounds 3-6 and their parent compound 2. (a) Relative steric hindrance](image)
around the β-lactone carbonyl group is also shown. (b) The structure of conformationally restricted analog 6.

![Diagram showing conformational restriction](image)

**Figure 5.** The cyclopropylic strain-based conformational restriction. (a) General representation of the cyclopropylic strain. (b) Presumed stable conformation of 5 (syn/anti) and 6 (syn).

**Synthesis.** The target compounds 3a-6a would be obtained by condensation between the unit A or B and the unit C, D, or E. Although the synthesis of units A and B was described in our previous report, we needed to prepare the β-lactone units C-E (Scheme 1). In particular, in the synthesis of D and E, construction of the chiral all-carbon quaternary center adjacent to the β-lactone carbonyl group would be a key step.

**Scheme 1. Synthetic plan of 3a-6a**

![Scheme 1](image)

The β-lactone unit C was prepared as shown in Scheme 2, using a procedure similar to that for the preparation of the β-lactone unit in the total synthesis of belactosin A by Armstrong et al.21 4-Methylpentanoic acid 7 was condensed with (4R)-4-benzyl-2-oxazolidinone by the mixed anhydride method using LiCl as an additive22 to give 8, which was treated
with BrCH₂CO₂t-Bu/NaHMDS at -78 °C in THF to afford 9 stereoselectively. The oxazolidinone moiety of 9 was removed by hydrolysis with LiOH/H₂O₂ in aqueous THF to give 10. The α-position of the t-butyl ester in 10 was diastereoselectively chlorinated with CCl₄/LiHMDS in THF at -78 °C, which seemed to proceed through the Li-chelated seven-membered dianion transition state, followed by the ring-closing reaction under alkaline two-phase conditions to afford the β-lactone 11 (unit C, Pg = t-Bu).

Scheme 2. Synthesis of β-lactone units C and D

The synthesis of the unit D is also shown in Scheme 2. Starting from propionic acid 12, the β-lactone 16 was prepared according to the same procedure used for the synthesis of 11. Methanolysis of 16 yielded the ring-opened product 17, the substrate for the key reaction forming the asymmetric quaternary carbon center. Treatment of 17 with LiHMDS/3-bromo-2-methylpropene in THF at -78 °C to 0 °C afforded the desired alkylated product 18 as a single isomer. The reaction seemed to proceed through the Li-chelated six-membered transition state, in which the bulky t-butyl ester group prevents access of the electrophile from the upper side as shown in Scheme 2. Hydrogenation of 18 afforded 19, and subsequently its methyl ester moiety was selectively hydrolyzed with LiOH in aqueous THF, followed by ring-closing reaction with PyBOP to afford the β-lactone 20 (unit D, Pg = t-Bu). The relative stereochemistry of 20 was determined by NOE experiments (Figure 6-a).

The synthesis of the unit E is shown in Scheme 3. L-Isoleucine (21) was deaminated to afford 22, which was converted to the alcohol 25 according to the same procedure used for the synthesis of 17 described above. Next, we tried to construct the asymmetric quaternary carbon center by stereoselective methylation of 25 as in the synthesis of the unit D. Although the reaction was investigated under various conditions, it did not proceed at all. Because the bulky
(S)-sec-butyl side chain of 25 seems to lower the reactivity, we next examined the methylation reaction with the β-lactone as a substrate, which was prepared by removal of the t-butyl group of 24 with TFA. Thus, when 27 was treated with LDA/Mel in THF at -78 °C, the desired methylated product 28 was obtained as a diastereomeric mixture (dr 3:1), while the yield was low. The stereoselectivity of the reaction might be caused by steric repulsion due to the carboxy group as depicted in Scheme 3. The carboxy group of 28 was re-protected with a t-butyl group and subsequent methanolysis gave 26, which was obtained as a single isomer after silica gel column chromatography purification. The secondary alcohol moiety of 26 was oxidized with Dess-Martin periodinane and subsequent reduction of the resulting carbonyl group with (R)-2-methyl-CBS-1,3,2-oxazaborolidine resulted in complete inversion of its stereochemistry to give the corresponding epimer 29. Although we attempted to selectively hydrolyze the methyl ester moiety of 29, the desired mono-ester 30 was not obtained at all, even under S_N2 reaction conditions. Thus, we hydrolyzed both the methyl and t-butyl ester moieties of 29, and then the product was successively treated with TFAA and with benzyl alcohol, which gave the desired benzyl ester 31 exclusively. Finally, 31 was treated with PyBOP to yield the β-lactone 32 (unit E, Pg = Bn). The relative stereochemistry of 32 was determined by NOE experiments (Figure 6-b).
Scheme 3. Synthesis of β-lactone unit E

1) BrCH₂CO₂t-Bu
   NaHMDS, THF, -78 °C
2) LiOH, H₂O₂
   THF / H₂O, 0 °C to rt
3) LiHMDS, CCl₄
   THF, -78 °C
4) NaHCO₃ ether / H₂O
   4 steps 73%

2 steps 74%
The synthesized β-lactone units 11, 20 and 32 were deprotected and finally condensed with unit A or B to yield 3a-6a. Compounds 3b-6b were also synthesized by hydrogenolysis of 3a-6a (Scheme 4).

**Scheme 4. Synthesis of 3a-6a and 3b-6b**

Reagents and conditions: (a) TFA/CH2Cl2, -5 °C; (b) Pd/C, H2, THF, quant.; (c) TFA/CH2Cl2; (d) PivCl, Et3N, CH2Cl2, 0 °C to rt, 62% (3a, 2 steps from A), 100% (4a, 2 steps from A); (e) EDC·HCl, HOAt, Et3N, CH2Cl2, 0 °C, quant. (5a, 2 steps from A), 91% (6a, 2 steps from B); (f) Pd/C, H2, TFA/THF, 0 °C, quant. (3b-6b)
Chemical and Biological Stability of 2b-6b. We evaluated the chemical and biological stability of 2b-6b. The compounds were incubated in 0.1 M TEAA buffer (pH 7.4) or human AB serum at 37 °C, and the time courses were analyzed by HPLC to obtain the half-life (t1/2), the results of which are shown in Figure 6. In 0.1 M TEAA buffer, the order of their stability was 3b < 2b < 4b, 5b, 6b, which clearly corresponds to the order of the steric hindrance around their β-lactone carbonyl group (Figure 4-a), as we expected. Notably, 4b, 5b, and 6b which have a quaternary carbon adjacent to their β-lactone carbonyl group were quite stable and no degradation was observed under the conditions.

Similarly, in human AB serum, the order of their stability was 3b < 2b < 4b < 5b < 6b, where the relative stability of 2b-5b also depended on the steric hindrance around their β-lactone carbonyl group, while their half-life was remarkably short compared with those in 0.1 M TEAA buffer, suggesting that 2b-6b were degraded enzymatically in human AB serum. Furthermore, 6b, the conformationally restricted analog of 5b, was significantly more stable than 5b, of which the t1/2 was longer than 1 h. Therefore, the conformational restriction might result in lowering affinity for the degradation enzyme, as we hypothesized. This finding is an interesting example of the correlation between conformational flexibility and biological instability. Thus, we successfully identified 6a as a chemically and biologically stable analog of 2a.

<table>
<thead>
<tr>
<th>compound</th>
<th>t1/2 at 37 °C</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0.1 M TEAA buffer (pH 7.4) [h]</td>
</tr>
<tr>
<td>2b</td>
<td>10</td>
</tr>
<tr>
<td>3b</td>
<td>3.5</td>
</tr>
<tr>
<td>4b</td>
<td>ND</td>
</tr>
<tr>
<td>5b</td>
<td>ND</td>
</tr>
<tr>
<td>6b</td>
<td>ND</td>
</tr>
</tbody>
</table>

**Figure 7.** Chemical and biological stability of 2b-6b. (a) Time courses of 2b-6b in 0.1 M TEAA buffer (pH 7.4) at 37 °C analyzed by HPLC. (b) Time courses of 2b-6b in human AB serum at 37 °C analyzed by HPLC. (c) Calculated half-life of 2b-6b in each conditions. ND: not degraded

Pharmacological Effects of 6a. We investigated the inhibitory effect of the highly stable 6a on the CT-L activity of proteasome and HCT116 cell growth (Table 2). Notably, 6a (IC50 = 4.0 μM) showed a cell growth inhibitory effect
comparable to that of 2a (IC\textsubscript{50} = 1.8 \textmu M), despite its significantly lowered proteasome inhibitory activity (IC\textsubscript{50} = 1.3 \textmu M) compared with that of 2a (IC\textsubscript{50} = 0.0057 \textmu M). The IC\textsubscript{50} ratio (cell growth/CT-L activity) of 6a was 3.1, which is remarkably improved over that of 2a (319), and it was almost the same as those of bortezomib and carfilzomib (Table 1). These findings suggest that the lower cell growth inhibitory effect of 2a arises from its instability as we expected, and that structural optimization of 6a might lead to development of highly potent cell growth inhibitors.

### Table 2. Inhibitory Effect of 6a on Proteasome CT-L Activity and HCT116 Cell Growth

<table>
<thead>
<tr>
<th>compound</th>
<th>IC\textsubscript{50} [\textmu M]</th>
<th>IC\textsubscript{50} ratio (cell growth/CT-L activity)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2a</td>
<td>0.0057</td>
<td>1.8</td>
</tr>
<tr>
<td>6a</td>
<td>1.3</td>
<td>4.0</td>
</tr>
</tbody>
</table>

*Based on three experiments.

In summary, by chemical modification of 2a, we successfully developed a chemically and biologically stabilized analog 6a, in which the steric hindrance around the unstable \(\beta\)-lactone moiety and the cyclopropyllic strain-based conformational restriction would work together to stabilize the molecule. The cell growth inhibitory activity of 6a is comparable to its proteasome inhibitory activity, so that the structural optimization of 6a might result in highly potent cell growth inhibitors. The chemical and biological stability of 2a derivatives are well correlated to the steric hindrance around their \(\beta\)-lactone carbonyl group due to the bulkiness of their \(\alpha\)-carbon substituents. Furthermore, conformational restriction by the cyclopropyllic strain resulted in significant stabilization in human serum probably due to the decreased affinity for metabolic enzymes. The correlation between conformation and metabolic stability is not studied well, and this study presented an interesting example of their clear correlation.

### Abbreviations Used

Bn, benzyl; Boc, \(\text{-}\)-butoxycarbonyl; CBS, Corey-Bakshi-Shibata; Cbz, benzylxycarbonyl; CT-L, chymotrypsin-like; DMP, Dess-Martin periodinane; EDC, 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide; Et, ethyl; HOAt, 1-Hydroxy-7-azabenzotriazole; HPLC, high-pressure liquid chromatography; LDA, lithium diisopropylamide; LiHMDS, lithium hexamethyldisilazide; Me, methyl; Piv, pivaloyl; PyBOP, benzotriazol-1-yl-oxytritylpolidinophosphonium hexafluorophosphate; NaHMDS, sodium hexamethyldisilazide; NOE, nuclear Overhauser effect; SAR, structure activity relationship; TEAA, triethylammonium acetate; TFA, trifluoroacetic acid; TFAA, trifluoroacetic anhydride; THF, tetrahydrofuran; Thr, threonine; TMS, trimethylsilyl; UPR, unfolded protein response

### References


24. In the reaction, undesired diastereomer was not observed in the crude ¹H-NMR
27. In the reaction, undesired diastereomer was not observed in the crude ¹H-NMR
28. In this reaction, significant amount of trimethylsilylated product of 18 was obtained.
31. Absolute stereochemistry of 20 was determined based on the known specific optical rotation of compound 15.
36. The synthesis of the (2R,3S)-diastereomer of 24 was reported previously by Armstrong et al. using (R)-oxazolidinone instead of (S)-oxazolidinone (ref. 21). The [α]D and NMR data of 24 are different from those of the reported diastereomer. Thus, absolute configuration of 32 was determined as shown in Scheme 3.