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Author(s)	味岡,拓
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博士論文

Distribution of glycerol dialkyl glycerol tetraethers in Lake Biwa basin and the reconstruction of lake water pH during the last 300,000 years 琵琶湖集水域系における GDGT の分布と 過去 30 万年間の湖水 pH の復元

Division of Earth System Science, Graduate School of Environmental Science, Hokkaido University 北海道大学大学院環境科学院地球圈科学専攻

Taku Ajioka

味岡 拓

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Abstract

We investigated glycerol dialkyl glycerol tetraether (GDGT) distributions in soils and river/lake sediments in the Lake Biwa basin of central Japan, in order to understand their source and fate in a terrestrial environment. GDGTs analyzed in 16 soil profiles exhibited significant depth variation at each site. Branched GDGTs were generally most abundant in the surface litter layer (O layer) and decreased with depth, while the maximum concentration of crenarcheol appeared in the upper soil layer (A layer), above the maximum concentration of the other isoprenoid GDGTs. This finding is consistent with different microbial communities contribution to these GDGT pools the subsurface The relationship between methylation index of branched sequences. soil tetraethers/cyclization ratio of branched tetraethers (MBT'/CBT) and pH/MAAT in Lake Biwa basin soils was consistent with those of a previous report for global soils. The GDGT distributions in surface sediments from the lake differed from those of soils and river sediments in the watershed, suggesting that the GDGTs were produced in the lake water rather than supplied from the land. The CBT for lake sediments from different water depths corresponded to water pH values. We thus infer that the CBT for lake sediments may serve as proxy for lake water pH and can be applied to paleoenvironmental reconstruction.

Based on these results, we generated a 280,000-year record of water pH and temperature in Lake Biwa, central Japan, by analysing the methylation index (MBT') and cyclisation ratio (CBT) of branched tetraethers in sediments from piston and borehole cores to understand the responses of precipitation and air temperature in central Japan to the East Asian monsoon variability on the orbital timescale. Because water pH in Lake Biwa is determined by phosphorus and alkali cation inputs driven by precipitation and temperature, the record of water pH should indicate changes in precipitation and temperature in central Japan. Comparison with pollen assemblage in a Lake Biwa core suggests that lake water pH was determined by summer temperature in low eccentricity period before 55 ka, while it was determined by summer precipitation in high eccentricity period after 55 ka. From 130 to 55 ka, variation in lake pH (summer precipitation) was lagged behind that in summer temperature by several thousand years. This perspective is consistent with the conclusions of previous studies (Igarashi and Oba, 2006; Yamamoto, 2009) that the temperature variation preceded the precipitation variation in central Japan.

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Chapter 1

Introduction

1.1. Long-term variation in past East Asian monsoon

The East Asian monsoon governs the climate of East Asia (Wang et al., 2003), and East Asian monsoon variability on orbital timescales has been the topic of many studies, which have revealed that it has responded to precession; however, the timing of monsoon variability continues to be debated. Kutzbach (1981) hypothesised that the Asian monsoon responds to insolation changes at low latitudes, which are regulated by precession. According to this hypothesis, the summer monsoon is maximal when Northern Hemisphere summer insolation is maximal in the precession cycle. Indeed, oxygen isotope records from cave stalagmites in China have demonstrated that summer monsoon variability was pronounced at the precession cycle and maximal at July-August precession (e.g., Wang et al., 2001, 2008; Yuan et al., 2004; Dykoski et al., 2005). However, some proxy records are not consistent with this hypothesis. Clemens and Prell (2003) reported that Indian summer monsoon variability showed both precession and obliquity cycles and was maximal at the November perihelion on the precession band. The pollen record in the north-western Pacific off of central Japan shows that the East Asian monsoon has been strongest at the October-November perihelion in precession cycles (Heusser and Morley, 1985; Igarashi and Oba, 2006). Thus, the conclusions have varied according to the proxy record used.

1.2. Proxies used for paleoclimate reconstruction in Lake Biwa sediments

Lake sediments provide a good, widely available palaeoclimate archive. Proxies applicable to lake sediments include pollen and diatom fossils, δD of long-chain *n*-alkanes, lignin, biogenic opal, and pigments, among others. In Lake Biwa, sediment cores have been investigated using pollen fossils (Miyoshi et al., 1999; Nakagawa et al., 2008), lignin (Ishiwatari and Uzaki, 1987; Ishiwatari et al., 2009; Ohira et al., 2013), diatom frustules (Kuwae et al., 2004), biogenic opal (Xiao et al., 1997), pigments (Ishiwatari et al., 2009), and δD of long-chain *n*-alkanes (Seki et al., 2012). Although paleoclimate information is available from these proxy records, more records generated by other new proxies are necessary to better understand the response of the East Asian monsoon to orbital forcing.

1.3. GDGTs and its applicability for paleoclimatology

Glycerol dialkyl glycerol tetraethers (GDGTs) in natural environments include isoprenoid and branched GDGTs (Appendix 1; Nishihara and Koga, 1987; Sinninghe Damsté et al., 2000). Isoprenoid GDGTs derive from Archaea (De Rosa and Gambacorta, 1988), while the biological sources of branched GDGTs include *Acidobacteria* (Sinninghe Damsté et al., 2000, 2011; Weijers et al., 2006a). These compounds in sediments are used for paleoenvironmental reconstruction in marine and terrestrial realms (Schouten et al., 2002; Weijers et al., 2007a).

Isoprenoid GDGTs have been found in marine sediments (e.g. Schouten et al., 2000), marine suspended particles (e.g. Schouten et al., 2001), marine settling particles (e.g. Wuchter et al., 2006), lake sediments (e.g. Powers et al., 2004) and soils (e.g. Weijers et al., 2006b). Crenarchaeol, an isoprenoid GDGT with one cyclohexane and 4 cyclopentane rings, is thought to derive from Group 1 Crenarchaeota, a phylum of

Archaea (Sinninghe Damsté et al., 2002). The Group 1 Crenarchaeota was subsequently reclassified into a new phylum, Thaumarchaeota (Brochier-Armanet et al., 2008; Spang et al., 2010). Schouten et al. (2002) proposed a paleotemperature proxy, TEX_{86} (tetraether index of tetraethers consisting of 86 carbons), based on the empirical relationship between the relative abundance of isoprenoid GDGTs in seafloor sediments and sea surface temperature. TEX₈₆ has also been applied to lake sediments (Powers et al., 2004, 2010). The relative abundance of isoprenoid GDGTs is controlled not only by temperature but also by a contribution from anaerobic methane oxidizing Archaea. Zhang et al. (2011) proposed the methane index (MI) as an indicator of the overprint of the isoprenoid GDGT pool by lipid contributed by archaea mediating the anaerobic oxidation of methane. In soils, amoA (encoding a subunit of the key enzyme ammonia monooxygenase) gene abundance of Thaumarchaeota is correlated with crenarchaeol concentration (Leininger et al., 2006), suggesting that ammonia-oxidizing archaea play an important role in contributing the archaeal lipids in soils. The presence of isoprenoid GDGT in soils was reported (Weijers et al., 2006b), but little is known about the detailed distribution of isoprenoid GDGTs in terrestrial environments.

Branched GDGTs were first identified in peat (Sinninghe Damsté et al., 2000) and subsequently found in soil (Weijers et al., 2007a), and marine (Hopmans et al., 2004) and lake sediments (e.g. Sinninghe Damsté et al., 2009; Tierney and Russell, 2009). The stereochemistry of the glycerol moieties of branched GDGTs suggests that they derive from Bacteria rather than Archaea (Weijers et al., 2006a). Their carbon isotope signature suggests a heterotrophic origin (Pancost and Sinninghe Damsté, 2003). Branched GDGT I was recently detected in an *Acidobacterium* (Sinninghe Damsté et al., 2011). Hopmans et al. (2004) found branched GDGTs in marine sediments and proposed the BIT (branched and isoprenoid tetraether) index as an indicator of the contribution of soil organic matter (OM) to marine environments, because soils contain abundant branched GDGTs. Weijers et al. (2007a) reported that the relative abundance of branched GDGTs in soils reflects soil pH and mean annual air temperature (MAAT). The cyclization ratio of br tetraethers (CBT) correlated with soil pH and the methylation index of br tetraethers (MBT) correlated with both soil pH and MAAT. Based on these empirical relationships, Weijers et al. (2007b) proposed the MBT/CBT paleotemperature index, which they applied to marine sediments from the Congo River fan and obtained reasonable results (Weijers et al., 2007b). Recently, Peterse et al. (2012) improved the relationship among branched GDGT distribution, soil pH and MAAT with additional dataset, and suggested MBT' instead of MBT. Niemann et al. (2012) applied MBT/CBT index to sediment core in a high Alpine lake in southern Switzerland where GDGTs in catchment soils contribute to, and reasonably reconstructed MAAT. However, the MAAT estimated from the application of MBT/CBT to a marine core from offshore from Norway was much higher than the true value. The higher estimate was interpreted to reflect summer temperature (Rueda et al., 2009). Sinninghe Damsté et al. (2008) also found that the MBT/CBT index afforded higher MAAT for soils from Mt. Kilimanjaro and attributed it to local bias. Tierney and Russell (2009) found that the branched GDGTs in lake sediments differed from those in soils, and suggested in situ production of branched GDGTs in lake water as the cause. The same phenomenon has been reported for a New Zealand lake (Zink et al., 2010), a Scottish lake (Tyler et al., 2010) and a Tibetan salt lake (Wang et al., 2012). Tierney et al. (2010) noted that the correlation between MBT/CBT from sediments and MAAT for 46 lakes in East Africa differed from that of the global soil set. The branched GDGTs in suspended particulate matter from 23 American lakes exhibited a similar regression equation to that for East African lakes (Schoon et al., 2013). These studies demonstrated that the MBT/CBT is useful for paleotemperature estimates for lake sediments, but it is not clear whether or not the branched GDGTs produced in lake water respond to pH and temperature differently from those in soils.

1.4. Aims of this study

I have investigated the branched and isoprenoid GDGTs in soils and river and lake sediments from the Lake Biwa basin to clarify (i) spatial and depth variation in branched and isoprenoid GDGTs in soils, (ii) the source of GDGTs in river and lake sediments and (iii) the response of MBT and CBT to pH and temperature in lake environments. Based on these results, we examined branched GDGTs in sediments from borehole BIW08-B and piston core BIW07-6 in Lake Biwa, central Japan, to reconstruct lake water pH and temperature during the last 280,000 years. We then evaluated the variability of the East Asian summer and winter monsoons based on estimated summer precipitation and winter lake water temperature.

Chapter 2

Materials and methods

2.1. Environmental setting of Lake Biwa basin

Lake Biwa is located at an altitude of 84 m in central Japan and is surrounded by mountains ca. 1,000 m high. It is the largest lake in Japan, with an area of 674 km² and a watershed area of 3,850 km² (Fig. 1). More than 118 rivers flow into it and the Seta River discharges from it. The climate of the area is affected by the East Asian monsoon (Yoshino, 1965, 1978); summer monsoon brings warm and humid conditions, whereas the winter monsoon brings snowfall in the northern part of the area and dryness in the southern part.

The MAAT is 14.7 °C at Hikone Meteorological Observatory (87.3 m) and 6.3 °C at Mount Ibuki Weather Station (1,375.8 m), respectively (Japan Meteorological Agency, available at http://www.jma.go.jp/jma/index.html). Water temperature and pH data were obtained from the Lake Biwa Environmental Research Institute (http://www.pref.shiga.lg.jp/biwako/koai/hakusyo/index.html).

2.2. Samples

2.2.1. Soils

Soil samples (0–30 cm, 5 cm interval) were collected in August/September 2009 at a total of 25 sites between 86 and 1,149 m above sea level; 5 sites were from Mt. Ibuki, 12 the Ane River drainage area and 8 from the Oku-Biwako area (Table 1 and Fig. 1). Soil samples were kept at 0 °C during sampling and freeze-dried in the laboratory. The MAAT at each sampling site was calculated from its altitude via a lapse rate model developed from the relationship between MAAT and the altitude at the nearby observation sites (4 sites). The average root mean square error of this lapse rate model is 0.22 °C.

2.2.2. River sediments

Four river sediment samples were taken from two sites in the Ane River with a shovel in August/September 2009 (Fig. 1). Site 1 was the sandbank in the river, and samples 1-1 and 1-2 were composed mainly of coarse and fine sand, respectively (Table 2). At site 2, sediments were sampled from a stagnant backwater in the river. Samples 2-1 and 2-2 comprised black fine sand and homogeneous black silt, respectively.

2.2.3. Lake Biwa surface sediments

Lake surface sediments were retrieved with a gravity corer and an Ekman-Birge sampler from June–October 1997 (Murase and Sakamoto, 2000; Table 3). Seven surface sediments were taken from the offshore transect Echi River estuary at intervals of ca. 2–3 km. One additional sample was from the Ane River estuary (Fig. 1). Along the Echi River estuary transect, sediments at sites 671 and 716 were medium sand and silt, respectively, and all other sediments were clay (Table 3). Water temperature and pH were obtained from the Lake Biwa Environmental Research Institute (http://www.pref.shiga.lg.jp/biwako/koai/hakusyo/index.html).

2.2.4. Cores BIW07-6 and BIW08-B

Piston core BIW07-6 (18.42 m long) was taken from the central part of Lake Biwa (35°13′59.02″N, 136°02′51.89″E; water depth of 55 m) in 2007 (Fig. 1; Takemura et al.,

2010). The sediments in the core consisted of homogenous dark-grey silty clay (Fig. 2). The age–depth model for core BIW07-6 was generated from six well-dated widespread ash layers and 13 accelerator mass spectrometry (AMS) ¹⁴C age of plant wood fossils analyzed at three AMS facilities during the past decade; the Center for Chronological Research at Nagoya University (Laboratory code; NUTA2, Nakamura et al., 2000, 2004), the W.M. Keck Carbon Cycle Accelerator Mass Spectrometry Laboratory (UCIAMS, Southon et al., 2004a, b) and the Korean Institute of Geoscience and Mineral Resources (KIGAM, Hong et al., 2010) (Table 4,). The ¹⁴C ages were converted to calendar ages using the IntCal13 dataset (Reimer et al., 2013).The average sedimentation rate was found to be 0.4 m/ky (Fig. 3). The sediment was stored at 4°C for 0.5 years. Then, 137 samples (1-cm thick) were collected from 0.30 m (0.6 ka) to 18.29 m (46 ka) and immediately freeze-dried. The average sampling interval was ~0.3 ky.

Borehole core BIW08-B (100.3-m long) was collected from its site (35°13'41.15"N, 136°03'21.19"E; water depth of 53) in 2008 (Fig. 1). The sediments consisted of dark-grey massive silty clay from 0 to 89 m, sandy silt containing abundant sand and plant debris from 89 to 99 m, and dark-grey massive silty clay from 99 to 100.3 m (Fig. 2; Takemura et al., 2010). The age–depth model for core BIW08-B was generated from 17 dated widespread ash layers, two AMS ¹⁴C age of plant wood fossils analyzed at the Korean Institute of Geoscience and Mineral Resources (KIGAM, Hong et al., 2010) (Table 5), and nine dated Anhysteretic Remanet Magnetization (ARM) events. The ¹⁴C age were converted to calendar ages using the IntCal13 dataset (Reimer et al., 2013). The ARM events were correlated with those detected in core BIW07-6 (Hayashida et al., 2007). The average sedimentation rate was found to be 0.3 m/ky (Fig. 3). The sediment

was stored at 4°C for 0.5 years. Then, 152 samples (2.5-cm thick) were collected from 13 m (43 ka) to 88 m (280 ka), and the samples were immediately freeze-dried. The average sampling interval was ~1.9 ky.

2.3. Analytical methods

2.3.1. pH measurement

Soil and river sediment samples were freeze-dried and sieved at 2 mm. The pH values of the soils and river sediments were measured in a soil/water mixture of 1/2.5 (w/w) with a pH meter (pH BOY-P2, Shindengen Electronic MFG. CO., LTD, Japan).

2.3.2. Lipid extraction and separation

Lipids were extracted (3×) from a freeze-dried sample using a DIONEX ASE-200 at 100 °C and 1000 psi for 10 min with 11 ml CH₂Cl₂/CH₃OH (6:4 v/v) and concentrated. The extract was separated into four fractions using column chromatography (SiO₂ with 5% distilled water; column of 5.5 mm inner diameter and 45 mm length): F1 (hydrocarbons) with 3 ml *n*-hexane; F2 (aromatic hydrocarbons) with 3 ml hexane/toluene (3:1 v/v); F3 (ketones) with 4 ml toluene; F4 (polar compounds) with 3 ml toluene/CH₃OH (3:1 v/v).

2.3.3. GDGT analysis

An aliquot of F4 was dissolved in hexane/propan-2-ol (99:1 v/v) and filtered. The GDGTs were analyzed using high performance liquid chromatography-mass spectrometry (HPLC-MS) with a Shimadzu SIL-20AD system connected to a Bruker Daltonics micrOTOF-HS time-of-flight mass spectrometer. A Prevail Cyano column

 $(2.1 \times 150 \text{ mm}, 3 \mu\text{m}; \text{Alltech})$ at 30 °C was used, following the methods set out by Hopmans et al. (2000) and Schouten et al. (2007). The conditions were: flow rate 0.2 ml/min, isocratic with 99% hexane and 1% propan-2-ol (5 min) followed by a linear gradient to 1.8% propan-2-ol over 45 min. Detection was achieved using atmospheric pressure chemical ionization (APCI) MS in positive ion mode. The spectrometer was run in full scan mode (m/z 500–1,500). Compounds were assigned from comparison of mass spectra and retention times with GDGT standards (from the main phospholipids of Thermoplasma acidophilum via acid hydrolysis) and values from a previous study (Hopmans et al., 2000). Quantification was achieved by integrating the summed peak areas in the $(M+H)^+$ and $(M+H+1)^+$ chromatograms and comparing them with the peak area from an internal standard C_{46} glycerol trialkyl glycerol tetraether (GTGT; Patwardhan and Thompson, 1999) in the (M+H)⁺ ion chromatogram, according to the method set out by Huguet et al. (2006). The correction value for the ionization efficiency between GDGTs and the internal standard was obtained by comparing the peak areas from T. acidophilum-derived mixed GDGTs with that from C₄₆ GTGT. The standard deviation of a replicate analysis was 3.0% of the concentration for each compound.

2.3.4. Calculations of GDGT proxies

TEX₈₆ was calculated from the concentrations of GDGT-1, GDGT-2 and GDGT-3, plus a regioisomer of crenarchaeol using the following equation (Eq. 1; Schouten et al., 2002):

 $TEX_{86} = ([GDGT-2] + [GDGT-3] + [crenarchaeol regioisomer])/([GDGT-1] +$

Temperature values were calculated according to the following equations based on core top calibrations of global marine sediments (Eq. 2; Kim et al., 2010) and lake sediments (Eq. 3; Powers et al., 2010):

$$T = 81.5 \times TEX_{86} - 26.6 \tag{Eq. 2}$$

$$T = 55.2 \times TEX_{86} - 14.0$$
 (Eq. 3)

where T = temperature (°C). The analytical accuracy was 0.45 °C in our laboratory. The BIT index (Eq. 4) was calculated as per Hopmans et al. (2004):

BIT = ([GDGT I] + [GDGT II] + [GDGT III])/([GDGT I] + [GDGT II] + [GDGT III] + [GDGT II]) + [GDGT II] + [GDGT II

The methane index (MI; Eq. 5) was calculated as per Zhang et al. (2011):

MI = ([GDGT-1] + [GDGT-2] + [GDGT-3])/([GDGT-1] + [GDGT-2] + [GDGT-3] +[crenarchaeol] + [crenarchaeol regioisomer]) (Eq. 5)

CBT (Eq. 6) and MBT' (Eq. 7) were calculated as per Weijers et al. (2007a) and Peterse et al. (2012):

$$CBT = -\log([GDGT Ib] + [GDGT IIb])/([GDGT I] + [GDGT II])$$
(Eq. 6)

$$MBT' = ([GDGT I] + [GDGT Ib] + [GDGT Ic])/([GDGT I] + [GDGT Ib] + [GDGT Ic] + [GDGT II] + [GDGT IIb] + [GDGT IIc] + [GDGT III])$$
(Eq. 7)

The pH (Eq. 8) and MAAT (Eq. 9) were calculated according to the following based on a global soil dataset (Peterse et al., 2012):

$$pH = 7.90 - 1.97 \times CBT$$
 (Eq. 8)

$$MAAT = 0.81 - 5.67 \times CBT + 31.0 + MBT'$$
(Eq. 9)

The pH and MAAT were also calculated according to the following equations based on an African lake sediment dataset (Tierney et al., 2010; Eqs. 10 and 11):

$$pH = 10.32 - 3.03 \times CBT$$
 (Eq. 10)

$$MAAT = 11.84 - 9.32 \times CBT + 32.54 \times MBT'$$
(Eq. 11)

The analytical accuracy of CBT and MBT' was 0.015 and 0.006, respectively, in this study.

Chapter 3

Branched and isoprenoid glycerol dialkyl glycerol tetraethers in soils and lake/river sediments in Lake Biwa basin and implications for MBT/CBT proxies

Source of GDGTs in river and lake sediments were determined by comparing the distribution of GDGTs in soils, river and lake sediments to clarify spatial and depth variation in branched and isoprenoid GDGT.

3.1. Results

3.1.1. Soils

3.1.1.1. Soil type

A typical soil sequence (Table 1) from the surface to subsurface included an O layer in the top with litter, an A layer, consisting of dark brown soil (brown forest soil), and a B layer, consisting of light brownish mineral soil (brown forest soil). The O layer (max. thickness 5 cm) developed in the forest sites where old trees (mainly cedar) were present, whereas it was absent from the forest sites where young trees were present. The boundary between the A and B layers was clear, and the depth of the boundary varied from 5–30 cm (Table 1).

3.1.1.2. Soil pH

The pH ranged from 3.3–8.0 in the surface soils (0–5 cm) (Fig. 4a). The pH in the O layer was variable, with a minimum in the A layer and increasing toward the B layer (Fig. 5a).

3.1.1.3. GDGTs in surface soils

The concentration of total isoprenoid GDGTs and total branched GDGTs ranged from 0.0088–4.3 and from 0.76–40 μ g/g, respectively (Fig. 4b and 4c). The branched GDGTs contributed 69.1–99.7% to the total GDGTs.

The distribution of isoprenoid GDGTs revealed that the average relative abundances of GDGT-0, GDGT-1 to -3 and crenarchaeol in soils were 24, 28 and 45%, respectively. TEX₈₆ varied from 0.18–0.89 (Fig. 4d), which corresponds to temperatures from -12 to 46 °C based on the marine core-top calibration of Kim et al. (2010). MI ranged from 0.20–0.87 (Fig. 4e). Among the branched GDGTs, GDGT I was most abundant (avg. 63%), followed by GDGT II (23%), GDGT Ib (5%), GDGT III (4%) and GDGT IIb (3%). BIT ranged from 0.75–1.00 and exceeded 0.9 at most sites (Fig. 4f). CBT ranged from 0.28–2.77 (Fig. 4g) and MBT' from 0.19–0.89 (Fig. 4h).

3.1.1.4. Depth variation of GDGTs

The depth profiles of the GDGT concentrations varied by site (Fig. 6). A typical sequence appeared at site 6 (Fig. 5a and 5b); here, the branched GDGTs were most abundant in the surface O layer and decreased with depth. Isoprenoid GDGTs increased with depth and decreased after the maximum. The maximum concentration of crenarchaeol (5–10 cm) occurred at shallower depth (Fig. 7a) than the maximum concentration of other isoprenoid GDGTs (15–20 cm). This typical profile was partially displayed at the other sites. The upper half was observed at sites 5, 7, 10 and 15, the lower half at sites 3, 9, 12, 18, 19 and 20, and the middle part at sites 4, 11, 22, 25 and 26 (Fig. 6).

The values of TEX₈₆ ranged from 0.1-0.8 (Fig. 5b). It was highly variable in the top

layer (0–5 cm) but converged to values between 0.6 (22 °C) and 0.8 (39 °C) below this depth (Fig. 5b). MI values ranged from 0.20–1.00 and increased with depth (Fig. 5c). The BIT index ranged from 0.79–1.00 (Fig. 5d); it was > 0.9 for most samples and did not change significantly with depth. At site 12, exceptionally, it increased with depth. CBT values decreased with depth (Fig. 5e), the reverse of the increase in soil pH (Fig. 5a and 5e). Maximum MBT' values were observed around the O/A boundary and decreased below this depth (Fig. 5f).

3.1.2. River sediments

The measured pH of Ane River sediments ranged from 5.8–7.2, higher than the values for the surface soils (Fig. 4a). in the same catchment area (3.6–6.6).

The concentration of isoprenoid GDGTs ranged from 0.015–0.91 μ g/g (Fig. 4b). The total concentration was higher in finer sediments than in coarse sediments (Fig. 4b). The average relative abundance of crenarchaeol, GDGT-0, GDGT-1 and GDGT-2 was 43%, 31%, 10% and 10%, respectively. TEX₈₆ values ranged from 0.47–0.57 (Fig. 4d), corresponding to a temperature range of 16–21 °C based on the marine core top calibration of Kim et al. (2010). MI values ranged from 0.32–0.42 (Fig. 4e).

The concentration of branched GDGTs ranged from 0.052–6.8 μ g/g and the values were higher in fine sediments than in coarse sediments (Fig. 4c). The average relative abundances of GDGT I, GDGT II, and GDGT III were 33%, 37% and 17%, respectively. BIT values ranged from 0.86–0.94 and were higher for finer sediments (Fig. 4f), as were CBT values (0.50–1.19; Fig. 4g); these values were within the range for the surface soil values in the same catchment (Fig. 4g). MBT' values ranged from 0.38–0.40 (Fig. 4h) and were lower than those in the soils in the same catchment (0.61–0.87).

3.1.3. Lake sediments

The total concentration of isoprenoid GDGTs ranged from 0.064–2.2 µg/g and was higher in finer sediments (Fig. 8a). Among isoprenoid GDGTs, GDGT-0 was the most abundant (38–74%) and increased toward the coast (Fig. 8c). Crenarchaeol was the second most abundant (18–52%) and increased toward offshore sites. TEX₈₆ values ranged from 0.32–0.52 and corresponded to a temperature ranges of 0–16 °C and 6–16 °C based on marine (Kim et al., 2010) and lake core-top (Powers et al., 2010) calibrations, respectively (Fig. 8e). MI values ranged from 0.13–0.44 and were higher for shallower sites (Fig. 8f). The total concentration of branched GDGTs ranged from 0.30–4.4 µg/g and was higher in finer sediments (Fig. 8b). The relative abundance of GDGT I, GDGT II, and GDGT III was 14–37%, 27–34% and 13–34%, respectively, followed by GDGT IIb (ca. 12%) and GDGT Ib (7%). The others were < 5% (Fig. 8d). BIT values ranged from 0.79–0.95 and were lower in deeper sites (Fig. 8h). MBT' values ranged from 0.23–0.45 and were higher in coarser sediments (Fig. 8i).

3.2. Discussion

3.2.1. Depth variation of GDGTs in soils

The maximum concentration of branched GDGTs appeared in the O layer and decreased with depth (Fig. 6). Active decomposition of OM occurred in litter layer, because heterotrophs obtain energy from the decomposition of plant remains. The producers of branched GDGTs are heterotrophs (Pancost and Sinninghe Damsté, 2003), and they produce more branched GDGTs in the O layer. The relative abundance of

branched GDGTs changed with depth. Compounds Ib and IIb increased with depth and other compounds with one or two pentane rings also increased (as seen at site 6 in Fig. 7b). This finding suggests that branched GDGTs in deeper soils were not a remnant of those produced in surface layers, but were produced in situ in the subsurface soil.

In contrast to branched GDGTs, the crenarchaeol concentration was highest in the upper part of the A layer and the concentration of other isoprenoid GDGTs was maximal around the lower part of the A layer or the upper part of the B layer (Fig. 6). Crenarchaeol is assumed to be specific to Thaumarchaeota (Sinninghe Damsté et al., 2002; Schouten et al., 2013). Microbial studies have shown that members of group 1 Crenarchaeota (Thaumarchaeota) are the dominant archaeal taxon in soils (Bates et al., 2011) and can be found at deeper soil depths (Hansel et al., 2008; Hartmann et al., 2009), and the ratio of Archaea: Bacteria increases with soil depth (Kemnitz et al., 2007). Thaumarchaeota include autotrophic nitrifiers that obtain energy via ammonia oxidation (Könneke et al., 2005; Park et al., 2010; Jung et al., 2011; Kim et al., 2012) and possibly heterotrophs (Biddle et al., 2006; Lloyd et al., 2013). We did not measure the ammonia concentration in brown forest soils, but it is generally high around the O/A boundary (Ohta and Kumada, 1978). Abundant ammonia would support the production of Thaumarchaeota in the upper A layer. Other isoprenoid GDGTs, i.e. GDGT-0 to -3 are not only produced by Thaumarchaeota but also by some Euryarchaeota. In particular, anaerobic methane-oxidizing Archaea, ANME-1, produce GDGT-0 to GDGT-3 (e.g. Pancost et al., 2001; Wakeham et al., 2003; Blumenberg et al., 2004; Liu et al., 2011; Schouten et al., 2013). The increase in GDGT-0 to -3 around the A/B boundary suggests a contribution from Euryarchaeota. The function of the Euryarchaeota in subsurface soils is unclear. Further microbial studies are needed to clarify the behavior of

Euryarchaeota in subsurface soil sequences.

3.2.2. Relationship between MBT'/CBT and soil pH/MAAT

CBT values decreased with depth, corresponding to an increase in soil pH (Figs. 5a and 5e). A negative correlation was observed between CBT and soil pH. This tendency has been reported by Weijers et al. (2007a). CBT in 0–5 cm surface soils had the following relationship with soil pH (Eq. 12):

$$CBT = -0.4805 \times pH + 3.7979 \ (r^2 \ 0.69, \ n = 25)$$
(Eq. 12)

CBT in all samples had the following relationship:

$$CBT = -0.4748 \times pH + 3.5828 (r^2 \ 0.51, n = 81)$$
(Eq. 13)

These relationships between CBT and soil pH are consistent with those of the global soil sample set published by Peterse et al. (2012).

Maximum MBT' values appeared in the O and A layers (Fig. 5f). No significant relationship was observed between MBT' and MAAT, because the range of MAAT (6.5 °C) in our sample set was not large enough to evaluate the correlation with MBT'. Instead, we found a significant correlation between MBT' and soil pH (r^2 0.72, n = 81). The relationship between MBT'/CBT and MAAT is expressed as:

$$MBT' = -0.133 \times pH + 0.0154 \times MAAT + 1.19 (r^2 0.77, n = 25)$$
(Eq. 14)

This equation may be converted to:

$$MBT' = 0.2186 \times CBT + 0.0379 \times MAAT - 0.0487 (r^2 \ 0.68, n = 25)$$
(Eq. 15)

This regression equation is similar to that published by Peterse et al. (2012). The depth variation in MBT'/CBT-based temperature increased with depth by a maximum of 6.5 °C (Fig. 5g). Recently, Huguet et al. (2013) reported that tree roots and their surfaces contain branched GDGTs that had different MBT/CBT values from those of the surrounding soils. Because we sieved the soil samples to remove roots, it is less likely that the GDGTs in roots affected changes in MBT'/CBT in our samples. The downward increase in MBT'/CBT-based temperature is thus likely to reflect the addition of newly produced branched GDGTs in subsurface soils. Generally, pH increases and free oxygen decreases with depth in soil sequences. This change in microenvironment results in a microbial community gradient, producing branched GDGTs with different composition. As a result, MBT' and CBT are altered in the burial process and the estimated pH and MAAT values are biased.

3.2.3. Sources of GDGTs in river and Lake Biwa surface sediments

On the ternary plot for branched GDGTs, crenarchaeol, and GDGT-0, most soils contain more branched GDGTs than river and lake sediments, although some soils have a similar composition to river and lake sediments (Fig. 9a). In contrast, the ternary diagram for crenarchaeol, GDGT-0 and GDGT-1 to -3 distinguishes soils and river sediments from lake sediments (Fig. 9b). Soils and river sediments are richer in

GDGT-1 to -3 than lake sediments (Fig. 9b). The preferential degradation or fractionation of GDGTs during transportation cannot be excluded for the interpretation of this phenomenon, but we suggest that difference in the source organisms are more likely to explain it. The GDGT composition of soils and river sediments suggests two distinct archaeal sources: Euryarchaeota with GDGT-0 and GDGT-1 to -3, but no crenarchaeol (Source E1 in Fig. 9b), and Thaumarchaeota with abundant crenarchaeol (Source T in Fig. 9b). The GDGT composition of lake sediments adds one more archaeal source: lacustrine Euryarchaeota with abundant GDGT-0 (Source E2 in Fig. 9b). Shallower lake sediments are richer in GDGT-0, suggesting that the contribution from source E2 is more important than in shallower sediments.

The CBT- MBT' plot indicates that most soils have higher MBT' than river and lake sediments (Fig. 10). The soils from the Mt. Ibuki area are exceptional and have a similar composition to lake and river sediments (Fig. 10). The limestone area of Mt. Ibuki is so small that the GDGTs produced in the area would not contribute significantly to the lake sediments. Thus, the difference in branched GDGTs between soils and the surface sediments from Lake Biwa suggests that most of the branched GDGTs in the lake sediments were not derived from soils, but were produced within the lake. However, the relatively higher MBT' values in coastal sediments suggest that they might be affected by a contribution from soil GDGTs to some extent. It is still not clear whether branched GDGTs were produced in the lake water or surface sediments. Further studies on the intact GDGTs are necessary to understand the production of branched GDGTs in Lake Biwa.

3.2.4. MBT'/CBT index for lake sediments

Lake Biwa is a monomictic lake, its water stratifies from spring to autumn and mixes in winter, which is controlled by air temperature. The modern measured temperature of surface water in Lake Biwa has an annual average of 16.7° C and is lowest (~7.5°C) in winter and highest in (~27.9°C) in summer, whereas the temperature of bottom water at a depth of 80 m ranges from 7.0 to 7.7°C with an annual average of ~7.3°C (Fig. 9b; off Imadzu: $35^{\circ}23.68'$ N, $136^{\circ}07.95'$ E; data from the Lake Biwa Environmental Research Institute, http://www.pref.shiga.lg.jp/biwako/koai/hakusyo/index.html). The measured pH value in the surface water of Lake Biwa has an annual average of 8.1 and is lowest (~7.6) in winter and highest (~8.9) in summer, whereas the pH of the bottom water at a depth of 80 m ranges from 7.2 to 7.7 with an average value of ~7.4 (Fig. 11a; off Imadzu: $35^{\circ}23.68'$ N, $136^{\circ}07.95'$ E; data from the Lake Biwa Environmental Research Institute, http://www.pref.shiga.lg.jp/biwako/koai/hakusyo/index.html).

In Lake Biwa, its water pH depends on the geology of the drainage basin, evaporation, the photosynthesis of phytoplankton and submerged plants, the respiration of organisms, and the decomposition of OM by microbes (Wetzel, 2001). In volcanic regions, lake water receives strong mineral acids, particularly sulphuric acid, which decreases pH to less than 4 (Wetzel, 2001). There is, however, no active volcano in the Lake Biwa watershed. Thus, this factor is not important for pH of Lake Biwa water. In contrast, Ca²⁺ supplied from limestone increases lake water pH (Wetzel, 2001). In the Lake Biwa drainage basin, limestone is exposed only in the Mt. Ibuki area, and its contribution toward controlling lake water pH should be minor. Evaporation of lake water increases water pH (Wetzel, 2001), but pollen records in Lake Biwa cores suggest moist environments throughout the last 430 ky (Miyoshi et al., 1999), so evaporation

has not been a factor controlling lake water pH. Consumption of CO₂ by the photosynthesis of phytoplankton and submerged plants increases lake water pH (e.g., Talling, 1976). On the other hand, regeneration of CO₂ by the respiration of organisms and degradation of OM by microbes decreases lake water pH (Wetzel, 2001). Actually, a tripling of the concentration of chlorophyll from winter to spring in Lake Biwa increases lake water pH by 0.85 (data from the Lake Biwa Environmental Research Institute, http://www.pref.shiga.lg.jp/biwako/koai/hakusyo/index.html). These observations suggest that photosynthesis in the lake water is the major factor controlling water pH in Lake Biwa.

Temperature and pH were calculated by applying the MBT'/CBT index to Lake Biwa surface sediments using Biwa (this study; Eqs. 12 and 14) and global soil (Peterse et al., 2012) and lake (Tierney et al., 2010) calibrations. The estimated pH values in clay sediments were 7.2, 7.3 and 9.3 according to Biwa soil, global soil calibrations and global lake calibrations, respectively (Fig. 11a). The estimated temperature in clay sediments were 5.6 °C, 6.3 °C and 16.2 °C according to Biwa soil, global soil calibrations and global lake calibrations, respectively (Fig. 11b). The pH and temperature estimated using Lake Biwa and global soil calibrations were closer to the measured winter pH and temperature, whereas those estimated using the global lake calibration were very different from the measured pH and temperature. Furthermore, the reconstructed water pH and temperature estimated using soil calibrations have gradients with water depth and correspond to those of bottom water (Fig. 11). Together, these findings suggest that the MBT'/CBT index in Lake Biwa sediments reflect water pH and temperature, and soil calibration is applicable to their reconstruction.

3.3. Conclusions

The GDGTs in soils showed depth variations in branched and isoprenoid GDGT concentrations with depth. Branched GDGTs were abundant in the top layer and decreased with depth, while the maximum concentration of isoprenoid GDGTs occurred in subsurface soil. These findings suggest that different microbial communities developed in the subsurface soil sequence.

The relationship between MBT'/CBT and pH/MAAT in Lake Biwa basin soils was consistent with those for global soils (Peterse et al., 2012). The MBT'/CBT values in Lake Biwa sediments differed from those in soils from its watershed, suggesting that the branched GDGTs in the lake sediments did not originate from soil GDGTs but were produced in the lake environment. The CBT and MBT' values in surface sediments in the lake corresponded to lake water pH and temperature. The MBT'/CBT index may thus be useful for the estimation of paleo-pH and paleotemperature in Lake Biwa.

Chapter 4

Water pH and temperature in Lake Biwa from MBT'/CBT indices during the last 280,000 years

Based on the results of Chapter 3, paleo-pH and paleotemperature of the lake water were estimated by analyzing GDGTs in the cores BIW07-6 and BIW08-B (Fig. 1).

4.1. Results

The concentrations of isoprenoid and branched GDGTs varied between 0.01 and 2.55 and between 0.05 and 12.58 µg/g, respectively (Fig. 12). CBT-based pH ranged from 6.0 to 7.6 (Fig. 12). The measured pH value in the surface water of Lake Biwa has an annual average of 8.1 and is lowest (~7.6) in winter and highest (~8.8) in summer, whereas the pH of the bottom water at a depth of 59 m ranges from 7.2 to 7.7 with an average value of ~7.5 (off Minami-Hira: 35°11'39"N, 135°59'39"E; data from the Lake Biwa Environmental Research Institute, http://www.pref.shiga.lg.jp/biwako/koai/hakusyo/index.html). On the other hand, the measured pH of surface soils in the drainage basin ranged from 3.3 to 8.0 with an average of 5.0 (Fig. 4a), which are lower than those in lake water. The CBT-based pH values of the core sediments are lower than those in the present lake water. CBT-based pH was maximal at 279, 249, 210, 160, 133, 117, 78, 45, 25 and 6 ka and minimal at 264, 220, 178, 149, 123, 95, 52, 29 and 17 ka. MBT'/CBT-based temperature ranged from 4 to 11°C and was lower during glacials and higher during interglacials (Fig. 12). The modern measured temperature of surface water in Lake Biwa has an annual average of 16.9°C and is lowest (~7.6°C) in winter and highest in (~27.9°C) in summer, whereas

the temperature of bottom water at a depth of 59 m ranges from 7.2 to 8.2°C with an annual average of ~7.7°C (off Minami-Hira; data from the Lake Biwa Environmental Research Institute, http://www.pref.shiga.lg.jp/biwako/koai/hakusyo/index.html).

The BIT index ranged from 0.83 to 0.99 (Fig. 12). BIT values exceeded 0.95 in most intervals, but BIT values lower than 0.9 were found at 0–6, 29, 30, 114, 210 and 240 ka (Fig. 12).

4.2. Discussion

4.2.1. Factor controlling water pH in Lake Biwa

Described in section 3.2.4., photosynthesis in the lake water is the major factor controlling water pH. In Lake Biwa, photosynthesis is mainly controlled by phosphorus concentration in the water (Ishida et al., 1982; Tezuka, 1985). The anthropogenic eutrophication of Lake Biwa induced high primary production, resulting in the increase of the pH of the lake water by more than 1 from 1960s to 1970s (Nakayama, 1981). Phosphorus concentration in the lake is determined by the inflow of phosphorus from the catchment soils, which is govern by precipitation in the watershed (Kunimatsu, 1993). At the present day, the East Asian summer monsoon brings most of annual precipitation to the study area, exceptionally at high elevations in the northern part where snowfall brought by the East Asian winter monsoon is relatively important (http://www.jma.go.jp/jma/index.html). Therefore, summer precipitation in the water pH of the lake. Phosphorus concentration may also be governed by air temperature because the dissolution of silicate depends on temperature in chemical weathering process (White and Blum, 1995). Thus, both higher precipitation and higher temperature potentially

increase the inflow of phosphorus to the lake, enhancing primary production, and thus increases the lake water pH. Increases in precipitation and temperature intensify chemical weathering. The chemical weathering increases the inflows of Ca+ and phosphorus to the lake, both increasing lake water pH.

It is not clear whether branched GDGTs are produced in the water column or the sediment surface in Lake Biwa (this study; Ajioka et al., 2014). If branched GDGTs are produced in the water column, the main production depth of branched GDGTs is also unclear. Very recently, Buckles et al. (2014) found high abundance of intact branched GDGTs in the suspended particulates in the oxygen deficient bottom water in Lake Challa and suggested that oxygen deficient environment ($O_2 < 2 \text{ mg/L}$) is a favorable condition for branched GDGT production. In Lake Biwa, such a condition ($O_2 < 2 \text{ mg/L}$) exists the bottom water above sediment surface from autumn to early winter (http://www.pref.shiga.lg.jp/biwako/koai/hakusyo/index.html). It is thus likely that branched GDGTs were produced mainly in the bottom water in Lake Biwa. This is just a speculation by analogy with the case of Lake Calla. Evidence is necessary to test this speculation from the original data of suspended and sinking particles in Lake Biwa,

4.2.2. Changes in lake water pH during the last 280 ky

CBT-based pH during the last 280 ky showed a glacial-interglacial pattern with higher pH in interglacial periods (Fig. 12). Variation pattern in CBT-based pH was consistent with that of *Cryptomeria* pollen abundance and lagged behind that of the Tp value in cores BIW95-4 and Takashima-oki BT in Lake Biwa from 130 ka to 55 ka (Fig. 13; Hayashi et al., 2010a, b). Igarashi and Oba (2006) assumed that the abundance of *Cryptomeria* pollen as a proxy of summer precipitation because the natural distribution

of *Cryptomeria* corresponds to the area of precipitation higher than 2000 mm per year. The correspondence between CBT-based pH and *Cryptomeria* pollen abundance indicates that both proxy records are robust records of precipitation.

The pollen temperature index, T_p (= 100 x $T_w/[T_c + T_w]$, where T_w is the sum of temperate taxa and T_c is the sum of subalpine taxa) is used as a proxy of summer air temperature (Igarashi and Oba, 2006). Igarashi and Oba (2006) reported that variation in the Tp value preceded the abundance of *Cryptomeria* pollen in core MD01-2421 off central Japan in the western North Pacific, indicating that the variation of summer air temperature preceded that of summer precipitation by several thousand years. Yamamoto (2009) interpreted that this time lag was caused the latitudinal shift of the Baiu Front (early summer rain front). Our new record indicates that CBT-based pH was lagged behind the Tp record, implying that summer precipitation was lagged behind summer air temperature, as was already pointed by pollen studies (Igarashi and Oba, 2006; Hayashi et al., 2010a, b).

On the other hands, variation pattern in CBT-based pH was not consistent with that of *Cryptomeria* pollen abundance but with Tp value in core BIW95-4/BT from 55 ka to the present (Fig. 13; Hayashi et al., 2010a, b). This correspondence suggests that the lake water pH was determined by summer temperature rather than summer precipitation in this period. Nakagawa et al. (2008) suggested that the response of the East Asian monsoon to summer insolation was different between low and high eccentricity periods, and that the response was not clear in the low eccentricity (< 0.024) periods. The eccentricity was lower in the period from 55 ka to the present than the period from 130 to 55 ka. We speculate that higher amplitude variation in summer insolation in the latter period induced higher amplitude precipitation variation, which resulted in a condition
that precipitation controlled lake water pH. In contrast, lower amplitude variation in the former period induced lower amplitude precipitation variation. Instead, the other climatic forcings such as greenhouse gas forcing were relatively important, and temperature variation was a major factor controlling the variation in lake water pH.

The age uncertainity (2 σ values [95% confidence level]) of core BIW08B ranges from 5 to 11 ky in MIS 5 and 6 because of large error of radiogenic ages of tephras (Table 5). However, the pollen composition (Tp and *Cyptomeria* abundance) in core Lake BIW95-4/BT nearby the study site have a consistent variation with that in a marine core MD01-2421 from the offshore of central Japan in the western North Pacific. The age-depth model of core MD01-2421 was established by oxygen isotope stratigraphy using benthic foraminifera isotopes (Oba et al., 2006). Assumed synchronous vegetation change in central Japan, the correspondence of pollen assemblages between MD01-2421 and Lake Biwa cores assures that the uncertainty of the age-depth models of Lake Biwa cores in MIS 5 and 6 is smaller than that indicated by the dating error of each tephras, and the ages of sediments are precise enough to discuss variation on orbital timescales.

With caution of this limitation of age-depth model, comparison with Chinese stalagmite records reveals regional perspective on the East Asian summer monsoon variability. In the period from 55 ka to the present, the variation in CBT-based pH was synchronous with that of δ^{18} O in stalagmites in China (Fig. 13). This correspondence suggests that summer temperature in central Japan varied synchronously with the δ^{18} O of the precipitated water in China.

On the other hands, in the period from 143 to 55 ka, the variation in CBT-based pH was lagged behind that of the δ^{18} O in stalagmites in China by several thousand years

(Fig. 13; Wang et al., 2001, 2008). The mismatch between Japanese precipitation and Chinese stalagmite δ^{18} O records was already pointed out by Yamamoto (2009) and Clemens et al. (2010). There are two possible interpretations of this phenomenon. First, precipitation in Japan and China during early summer is principally determined by the position of the Baiu Front (an early summer rain front) that develops at the atmospheric boundary between warm moist air masses flowing from the south and cold air masses from the north (Yoshino, 1965). The latitudinal shift of the Baiu Front in response to orbital forcing may have resulted in the difference of response of precipitation between central Japan and China (Yamamoto, 2009). Second, synchronous variations in stalagmite δ^{18} O from Hulu Cave (Wang et al., 2001) and Sanbao Cave (Wang et al., 2008) in central China, and from Dongge Cave (Yuan et al., 2004) in southern China, suggest that stalagmite δ^{18} O is not a simple index of local precipitation (Yamamoto, 2009). The negative shift of stalagmite δ^{18} O from Chinese caves is also synchronous with the rapid rise of air temperature in central Japan at glacial terminations, suggesting that the negative shift of stalagmite δ^{18} O was partly related to temperature increase in air masses derived from the summer monsoon because heavy rains, which occur more frequently in warmer air mass, induce isotopically lighter water (Yamamoto, 2009). Clemens et al. (2010) showed a different idea that winter precipitation contributed to stalagmite δ^{18} O. This question is still open. Our new result confirms the mismatch between the precipitation records in central Japan and Chinese stalagmite $\delta^{18}O$ records. This is an interesting topic in the Asian monsoon study.

In this study, we assume that the branched GDGTs produced in the lake contributed to sedimentary branched GDGTs in the study site. However, the spike of high CBT (low estimated pH) at 13 ka is exceptionally affected by the contribution of soil GDGTs (Fig. 12). Ohira et al. (2013) reported a spike of abundant lignin at the same layer in core BIW08-B. Because high CBT and low CBT-based pH (3.3 to 8.0 with an average of 5.0; Fig. 4a) in the soils, the high CBT (low estimated pH) at 13 ka associated with high lignin abundance likely reflected the soil GDGT contribution.

4.2.3. Changes in MBT'/CBT -based winter lake water temperature during the last 280 ky

MBT'/CBT-based winter temperature showed a glacial-interglacial pattern (Fig. 12). The variation is consistent with the summer air temperature estimated from the pollen assemblage in Lake Biwa (Nakagawa et al., 2008). However, the MBT'/CBT-based temperatures (3 to 10°C) are more realistic than the pollen-based temperatures (–10 to 5°C; Nakagawa et al., 2008). The winter water temperature in Lake Biwa is controlled by winter cooling (data from the Lake Biwa Environmental Research Institute, http://www.pref.shiga.lg.jp/biwako/koai/hakusyo/index.html, Japan Meteorological Agency; available at http://www.jma.go.jp/jma/index.html) and reflects the intensity of the Asian winter monsoon. The rise of MBT-CBT' based winter lake water temperature ware lagged behind the rise of Tp-based summer air temperature by ~10 ky in Lake Biwa cores (Fig. 13). This suggests warmer summer and colder winter in MIS 5e, and the seasonal temperature variation was larger in this period.

4.2.4. Changes in microbial ecosystem of Lake Biwa during the last 280 ky

BIT index was higher than 0.8 throughout the core (Fig. 12). Because most branched GDGTs are produced in the lake (this study; Ajioka et al., 2014), this high BIT implies that high bacterial production in Lake Biwa. The BIT record indicates negative spikes (< 0.9) at 6–0 ka, 114, 210, and 240 ka, which correspond to the maximal peak of crenarchaeol concentration (Fig. 12). Because crenarchaeol is specific to Thaumarchaeota (Sinninghe Damsté et al., 2002), the production of Thaumarchaota was enhanced in those periods. Although the reason why Thanumarchaotal production was enhanced in the specific periods is not clear, changes in nutrient condition is a likely candidate to cause such an ecological change.

4.3. Conclusions

We analysed the MBT'/CBT indices in sediment cores retrieved from Lake Biwa to reconstruct changes in summer precipitation and temperature and winter temperature in central Japan during the last 280 ky. Comparison with pollen assemblage in a Lake Biwa core suggests that lake water pH was determined by summer temperature in low eccentricity period, while it was determined by summer precipitation in high eccentricity period. From 130 to 55 ka, variation in lake pH (summer precipitation) was delayed behind that in summer temperature by several thousand years. Thaumarchaeotal production was enhanced in specific periods in interglacials.

Chapter 5

Summary

In this study, branched GDGTs in sediments from borehole BIW08-B and piston core BIW07-6 in Lake Biwa, central Japan, was investigated to reconstruct lake water pH and temperature during the last 280,000 years.

The GDGTs in soils showed depth variations in branched and isoprenoid GDGT concentrations with depth. Branched GDGTs were abundant in the top layer and decreased with depth, while the maximum concentration of isoprenoid GDGTs occurred in subsurface soil. These findings suggest that different microbial communities developed in the subsurface soil sequence.

The relationship between MBT'/CBT and pH/MAAT in Lake Biwa basin soils was consistent with those for global soils (Peterse et al., 2012). The MBT'/CBT values in Lake Biwa sediments differed from those in soils from its watershed, suggesting that the branched GDGTs in the lake sediments did not originate from soil GDGTs but were produced in the lake environment. The CBT and MBT' values in surface sediments in the lake corresponded to lake water pH and temperature. The MBT'/CBT index may thus be useful for the estimation of paleo-pH and paleotemperature in Lake Biwa.

MBT'/CBT indices were applied to sediment cores retrieved from Lake Biwa to in order to reconstruct changes in summer precipitation and temperature and winter temperature in central Japan during the last 280 ky. Comparison with pollen assemblage in a Lake Biwa core suggests that lake water pH was determined by summer temperature in low eccentricity period, while it was determined by summer precipitation in high eccentricity period. From 130 to 55 ka, variation in lake pH (summer precipitation) was delayed behind summer temperature by several thousand years. Thaumarchaeotal production was enhanced in specific periods in interglacials.

Although CBT has been used for estimating soil pH, this study revealed that the CBT in lake sediments reflects the pH of lake water. Using the CBT as a proxy of lake water pH, changes in lake water pH during the last 280,000 years was successfully reconstructed. The soil pH may have responded summer precipitation and temperature , both are sensitive to summer monsoon variability.

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Table 1								
locations,	soil types, veget	ation, lithology of bas	sement rocks and	l mean annual	air temper	atures of soil samples.		
Site No.	Latitude (N)	Longitude (E)	Altitude (m)	Depth (cm)	Soil type	Vegetation	Lithology of basement rock ^b	MAAT ^c (°C)
1	35°23.56'	136°13.27'	86	0-5	0	Cropland	Sand	14.1
3	35°30.93'	136°22.23'	590	0-5	0	Japanese cedar	Granite	11.1
				5-20	Α			
				20-30	В			
4	35°30.40'	136°22.34'	714	0-5	0	Japanese cedar	Granite	10.3
				5-30	А			
5	35°30.38'	136°22.45'	773	0-5	0	Japanese cedar	Granite	9.9
				5-20	Α			
				20-30	В			
6	35°31.19'	136°22.65'	652	0-5	0	Japanese cedar	Granite	10.7
				5-15	А			
				15-30	В			
7	35°31.05'	136°22.17'	588	0-5	0	Japanese cedar and bamboo	Granite	11.1
				5-30	А			
8	35°31.02'	136°22.16'	579	0-5	А	Rice paddy	Granite	11.1
9	35°29.82'	136°21.61'	534	0-15	А	Mixed trees and grass	Granite	11.4
				15-30	В			
10	35°29.00'	136°21.05'	440	0-5	0	Pine and shruh	Shale	12.0
10	55 20.00	100 21:00		5-15	B			12.0
11	35°28 28'	136°20 97'	344	0-5	0	Iananese cedar	Shale	12.6
	35 20.20	150 20.57	511	5-25			Shale	12.0
12	35°26 00'	136°21 57'	283	0-20	A	Japanese cedar and hamboo	Shale	12.0
12	35°25 55'	136°25 01'	11/9	0-20	B	Grasses	Limestone	7.6
14	35°25.35	136°25.04'	086	0-5	0	Grasses	Limestone	9.6
14	35°25.13	136°25'22'	980	0-5	0	Japapasa cadar	Limestone	8.0
15	35°24.25'	136°25.40'	925	0-20	A	Grassas	Sandstone	9.0
10	25°22 95'	126926 12	217	0-5	A	Jananasa aadar	Chart slate and conditions	12.2
17	33 22.83	150 20.15	217	0-3	A	Japanese cedar	Chert, state and sandstone	15.5
18	35°24.36'	136°19.81'	133	0-5	A	Japanese cedar	Tuff	13.8
				5-10	B			
19	35°30.83'	136°09.21'	107	0-15	A	Japanese cedar and bamboo	Sandstone and shale	14.0
20	35°30.72'	136°09.36'	127	0-5	0	Japanese cedar	Shale	13.9
				5-20	В			
21	35°30.41'	136°09.61'	212	0-5	В	Mixed trees	Sandstone and shale	13.4
22	35°30.35'	136°09.19'	196	0-5	0	Japanese cedar	Shale	13.5
				5-20	Α	-		_
23	35°28.97'	136°07.47'	89	0-5	0	Grasses	Greenstone	14.1
24	35°27.29'	136°08.99'	302	0-5	0	Maple	Mudstone	12.8
25	35°28.60'	136°08.55'	297	0-15	A	Broad leaf trees	Greenstone	12.8
				15-25	В			
26	35°29.17'	136°09.00'	360	0-5	0	Japanese cedar	Sandstone	12.5
				5-20	А			
Soil type;	O, A and B mea	an litter layer, organi	c material-rich da	ark brown soil	layers and	organic material-poor yellowish	h brown soil layers, respectively.	
Lithology	refers to Isomi	(1956), Kurimoto et	al. (1999), Saito a	and Sawada (2	2000) and N	Vakae et al. (2001).		
Mean and	nual air temperat	ures were calculated	d by the linearly i	nterpolation or	n altitude be	etween temperatures at two ne	arby stations.	

* These soils are artificially disturbed.

Table 2							
Locations,	lithology, measure	d sediment pH and	mean annual ai	r temperature	s of riverine sedime	nts.	
Site No.	Latitude (N)	Longitude (E)	Altitude (m)	Depth (cm)	Lithology	Sediment pH	MAAT (°C)
1-1	35°23.54'	136°13.31'	84	0-5	Coarse sand	6.9	14.1
1-2			84	0-5	Fine sand	5.8	14.1
2-1	35°30.95'	136°22.04'	541	0-1	Fine sand	7.2	11.4
2-2			541	0-1	Silt	7.0	11.4

Table 3									
Locations,									
Site No.	Latitude (N)	Longitude (E)	Water depth (m)	Depth (cm)	Lithology	Water pH ^a	Water temperature (°C) ^a		
521	35°23.16'	136°12.00'	50.0	0-2	Clay	7.5	7.9		
671	35°13.08'	135°58.14'	26.2	0-2	Medium sand	7.7	9.8		
676	35°13.14'	135°59.10'	79.2	0-2	Clay	7.4	7.4		
686	35°13.08'	136°00.84'	70.0	0-2	Clay	7.4	7.5		
701	35°13.14'	136°02.70'	55.1	0-2	Clay	7.5	7.8		
706	35°13.14'	136°04.74'	28.0	0-2	Clay	7.6	9.4		
716	35°13.08'	136°06.24'	13.3	0-2	Silt	7.9	13.9		
721	35°13.32'	136°08.10'	9.8	0-2	Clay	8.1	15.4		
$\frac{1}{2}$ We the set of the set									

^a Water pH and temperatures were measured at near station off Imadzu (N35°23.68', E136°07.95').

Age controls of core bi wor-o										
Depth	Course	14 C			Mod	Model age				
(m)	Source	(BP, 1σ)			(cal B	(cal BP, 2σ)				
0.980	KgP				3150 ±	12	[1]			
1.115	SOh	3650	\pm	30	4086 -	3887	[2]			
1.450	K-Ah				7350 ±	80	[3]			
2.501	U-Oki				10286 ±	45	[4]			
3.510	14 C (wood)	11855	±	40	13844 -	13486	[5]			
3.700	14 C (wood)	12400	±	45	14940 -	14110	[6]			
4.010	14 C (wood)	12865	±	45	15880 -	15000	[5]			
5.380	14 C (wood)	14510	±	80	17940 -	17235	[5]			
5.630	Sakate				19484 ±	112	[4]			
6.130	14 C (wood)	18050	±	90	22005 -	21555	[5]			
6.620	14 C (wood)	19010	±	100	23275 -	22310	[5]			
7.160	14 C (wood)	20440	±	110	24815 -	23955	[5]			
8.530	14 C (wood)	23760	±	170	29160 -	28033	[5]			
9.870	AT				30009 ±	189	[4]			
11.560	14 C (wood)	27950	±	280	32965 -	31470	[6]			
12.030	14 C (wood)	28760	±	320	34495 -	32265	[6]			
15.360	14 C (wood)	36530	±	820	42970 -	40090	[5]			
17.660	14 C (wood)	41200	±	1500	48642 -	42855	[6]			
18.340	14 C (wood)	42570	±	1270	49190 -	44295	[6]			

Table 4Age controls of core BIW07-6

[1] Tani et al. (2013); [2] Fukuoka and Matsui (2002); [3] Kitagawa and van der Plicht (1998); [4] Staff et al. (2012); [5] Kitagawa et al. (2010); [6] Kitagawa et al. (in preparation).

Table	5
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Age controls of core BIW08-B

Depth	0		Model age					
(m)	Source	(B	P, 1c	5)	(cal]	Remarks		
0.00	Core-top				-57	±	0	[1]
1.12	¹⁴ C (wood)	1920	±	60	1868	±	160	[1]
2.09	SOh	3650	±	30	4086	-	3887	[2]
2.18	14 C (wood)	4720	±	110	5408	±	324	[1]
2.45	K-Ah				7350	±	80	[3]
3.28	U-Oki				10286	±	45	[4]
4.34	ARM-1 event				13837	±	103	[5][6]
4.97	ARM-2 event				16586	±	155	[5][6]
5.63	Sakate				19484	±	112	[4]
6.04	ARM-3 event				23181	±	277	[5][6]
7.61	DSs (Plinian)				29247	±	270	[1]
7.85	AT				30009	±	189	[4]
8.82	ARM-4 event				31078	±	442	[5][6]
9.84	ARM-6 event				32720	±	803	[5][6]
10.58	ARM-7 event				34529	±	770	[5][6]
12.58	ARM-8 event				40594	±	1425	[5][6]
13.32	SI				42080	±	1250	[1]
13.69	ARM-9 event				42619	±	1329	[5][6]
14.83	ARM-10 event				44154	±	2028	[5][6]
23.58	DNP				82922	±	10758	[6]
23.66	DMP2				83039	±	10788	[6]
26.81	KTZ				96715	±	7080	[6]
27.73	AsoABCD				104263	±	5352	[6]
28.41	Unnamed tephra				108098	±	5505	[6]
34.81	Unnamed tephra				114925	±	5544	[6]
38.99	Unnamed tephra				120837	±	7675	[6]
40.59	Aso-3b				123619	±	9005	[7]
44.67	Aso2				142975	±	9059	[7]
83.91	Aso1				266000	\pm	28000	[7]

[1] Kitagawa et al. (in preparation); [2] Fukuoka and Matsui (2002); [3] Kitagawa and van der Plicht (1998); [4] Staff et al. (2012); [5] Hayashida et al. (2007); [6] Takemura et al. (in preparation); [7] Matsumoto et al. (1991).



Fig. 1. Map showing the sampling sites.



Fig. 2. Lithologic columns of (a) core BIW07-6 and (b) borehole core BIW08-B (Takemura et al., 2010).



Fig. 3. Age–depth models of cores BIW07-6 and BIW08-B. The red, green and blue dots show volcanic ashes, ¹⁴C of plant debris and total organic carbon, and ARM events, respectively (see Tables 4 and 5). Error bars indicate the 95% confidence intervals (2σ).



Fig. 4. Measured pH (a), concentration of total isoprenoid GDGTs (b) and branched GDGTs (c), TEX₈₆ (d), methane index (e), BIT (f), CBT (g) and MBT' (h) for soils, river sediments and Lake Biwa surface sediments. For river and Lake Biwa surface sediments, the red, purple, blue and sky blue dots indicate coarse, medium, fine sand and silt samples, respectively. The pH values for Lake Biwa surface sediments represent the observed mean annual water pH.



Fig. 5. Depth variation in measured soil pH (a), TEX_{86} (b), methane index (c), BIT (d), CBT (e) and MBT' (f), and estimated temperature based on MBT'/CBT (g) according to the calibration of Peterse et al. (2012).



Fig. 6. Depth variation in the concentration of total branched GDGTs, GDGT-0, GDGT-1 to -3 and crenarchaeol.



Fig. 7. Vertical variation in concentration of total branched GDGTs and isoprenoid GDGTs (a) and relative abundances of branched GDGTs (b) at site 6. Relative abundances of GDGT Ib, Ic, IIb, IIc, III, IIIb and IIIc are multiplied by 10.



Fig. 8. Total iso (a) and br (b) GDGT concentrations, distributions of iso (c) and br (d) GDGTs, TEX_{86} (e), methane index (f), BIT (g), CBT (h), MBT' (i) and water depth (j) for Lake Biwa surface sediments. Light blue shading indicates sand or silt.



Fig. 9. Ternary diagrams indicating relative abundance of crenarchaeol, branched GDGTs and GDGT-0 (a) and GDGT-1 to -3, crenarchaeol and GDGT-0 (b) in soils, river sediments and Lake Biwa surface sediments.



Fig. 10. Plots of CBT vs. MBT' for soils, river sediments and Lake Biwa surface sediments.


Fig. 11. Reconstructed water pH (a) and MBT'/CBT-based temperature (b) in Lake Biwa surface clays. The temperature based on lake calibration was obtained from MBT/CBT rather than MBT'/CBT.



Fig. 12. Variations in the concentrations of total isoprenoid and branched GDGTs and crenarchaeol, CBT-based pH, MBT'/CBT-based temperature and BIT in cores BIW07-6 and BIW08-B during the last 280,000 years. Red, blue and green bars indicate age controls with 95% confidence intervals of volcanic ashes, the ¹⁴C of plant debris and total organic carbon, and ARM events, respectively.



Fig. 13. Variations in CBT-based pH and MBT'/CBT-based temperature from cores BIW07-6 and BIW08-B (this study), Tp and *Cryptomeria* (%) from cores BIW95 and Takashima-oki BT in Lake Biwa (Hayashi et al., 2010a and b), the δ^{18} O of stalagmites in Sanbao and Hulu caves in central China (Wang et al., 2001, 2008), and eccentricity during the last 150,000 years. Red, blue and green bars indicate age controls with 95% confidence intervals of volcanic ashes, the ¹⁴C of plant debris and total organic carbon, and ARM events, respectively. The names of tephras that appeared commonly in BIW07-6/BIW08-B and BIW95-4/BT are shown in the panel.



Appendix 1. Structures of isoprenoid and branched GDGTs.

Site	Denth	GDGT 0	GDGT-	Crenarchaeol	Isoprenoid	Branched			Maggurad	CBT	MBT'/CBT
No	(cm)	(ng/g)	1 to 3	(ng/g)	GDGTs	GDGTs	CBT	MBT'	nH	-based	-based
110.	(em)	(118/5)	(ng/g)	(118/8)	(µg/g)	(µg/g)			pm	pН	temperature (°C)
1-3	0-5	23	64	180	0.29	0.76	0.43	0.61	6.6	7.05	17.41
2	o -		27		0.10		0.41	0.04		2.1.5	12.01
3	0-5	46	37	21	0.10	11.24	2.41	0.86	4.4	3.15	13.91
	5-10	111	106	1	0.22	10.11	2.03	0.88	4.4	3.89	16.47
	10-15	370	639	1	1.01	6.32	1.55	0.86	4.3	4.85	18.80
	15-20	479	914	1	1.39	4.26	1.37	0.83	4.5	5.21	18.80
	20-30	216	449	4	0.67	1.04	1.16	0.82	4.5	5.61	19.67
	0.5	1.61		201	0.50	1 < < 0	1 40	0.00		1.00	15.10
4	0-5	161	232	291	0.70	16.60	1.48	0.80	4.5	4.98	17.13
	5-10	215	3/1	170	0.78	12.11	1.26	0.83	4.9	5.42	19.55
	10-15	361	584	16/	1.13	8.30	1.15	0.79	4.8	5.64	18.74
	15-20	480	740	117	1.36	2.96	1.00	0.75	4.9	5.93	18.38
	20-25	1245	2673	112	4.05	3.31	1.04	0.77	4.8	5.85	18.89
	25-30	1002	2415	84	3.52	2.97	1.00	0.78	5.0	5.93	19.35
~	0.5	100	1.40	22	0.00	22.04	0.17	0.07	2.6	2.62	15.50
5	0-5	128	143	22	0.29	22.96	2.17	0.87	3.6	3.62	15.52
	5-10	104	110	56	0.27	11.24	2.05	0.87	4.2	3.86	16.08
	10-15	329	403	88	0.82	15.19	1.80	0.86	4.3	4.35	17.37
	15-20	668	801	68	1.54	11.53	1.65	0.82	4.5	4.66	16.94
	20-25	921	1109	63	2.10	7.14	1.53	0.79	4.6	4.89	16.57
	25-30	1100	1424	70	2.61	5.26	1.46	0.80	4.8	5.02	17.26
_	~ -				0.40	10.00					
6	0-5	160	221	223	0.60	18.89	2.11	0.85	4.1	3.74	15.16
	5-10	154	238	397	0.80	10.55	1.85	0.85	4.3	4.26	16.80
	10-15	329	465	386	1.19	7.43	1.40	0.83	4.5	5.14	18.73
	15-20	370	498	143	1.02	2.37	1.26	0.81	4.5	5.42	18.75
	20-25	172	291	42	0.51	1.08	1.14	0.81	4.6	5.66	19.32
	25-30	120	273	16	0.41	0.48	1.05	0.78	4.6	5.82	19.09
7	0-5	111	77	12	0.20	15.33	1.38	0.78	4.7	5.18	17.12
	5-10	100	93	5	0.20	7.70	1.27	0.79	5.1	5.41	18.22
	10-15	188	218	6	0.41	5.11	1.17	0.79	5.1	5.60	18.63
	15-20	257	323	17	0.60	3.99	1.09	0.79	5.0	5.76	19.10
	20-25	366	441	26	0.83	2.94	1.05	0.78	4.9	5.84	19.07
	25-30	272	304	26	0.60	3.38	1.12	0.78	5.0	5.70	18.81
8	0-5	280	155	211	0.66	3.84	1.02	0.70	5.9	5.90	16.88
9	0-5	46	90	150	0.31	3.99	0.82	0.68	5.2	6.29	17.30
	5-10	70	108	79	0.27	2.15	0.82	0.72	5.3	6.28	18.54
	10-15	69	95	44	0.21	1.02	0.83	0.71	5.1	6.26	18.05
	15-20	35	47	25	0.11	0.51	0.85	0.69	5.1	6.23	17.28
	20-25	22	25	15	0.06	0.32	0.87	0.66	5.0	6.18	16.38
	25-30	22	24	15	0.06	0.31	0.83	0.65	5.0	6.27	16.13
10	0-5	9	0	0	0.01	2.95	2.77	0.84	4.1	2.45	11.22
	5-10	14	9	0	0.02	1.86	2.98	0.78	4.5	2.03	8.12
	10-15	18	13	0	0.03	1.35		0.78	4.5		
11	0-5	132	61	37	0.23	39.53	1.40	0.76	4.1	5.14	16.32
	5-10	92	192	32	0.32	10.86	1.32	0.78	4.3	5.31	17.55
	10-15	172	361	37	0.57	3.54	1.07	0.79	4.5	5.79	19.13
	15-20	140	272	35	0.45	7.93	1.36	0.79	4.4	5.22	17.50
	20-25	211	313	32	0.56	4.93	1.26	0.78	4.6	5.42	17.71
12	0-5	949	944	2264	4.34	9.71	1.03	0.64	4.7	5.86	14.88
	5-10	911	721	1698	3.48	8.76	0.92	0.64	4.5	6.10	15.35
	10-15	557	417	885	1.94	5.78	0.85	0.60	5.0	6.22	14.72
	15-20	377	250	302	0.96	3.91	0.76	0.54	5.8	6.40	13.34
13	0-5	44	99	206	0.37	3.23	0.28	0.30	8.0	7.36	8.50
14	0-5	0	0	64	0.06	2.68	0.72	0.19	7.3	6.49	2.57
									(Continu	e to the n	ext page)

Appendix 2. GDGT concentrations, proxy data and estimated pH and temperature for soils.

Site No.	Depth (cm)	GDGT-0 (ng/g)	GDGT- 1 to 3 (ng/g)	Crenarchaeol (µg/g)	Isoprenoid GDGTs (µg/g)	Branched GDGTs (μg/g)	CBT	MBT'	Measured pH	CBT -based pH	MBT'/CBT -based temperature (°C)
15	0-5	100	82	79	0.26	22.47	1.78	0.78	4.4	4.40	15.00
	5-10	150	224	28	0.40	12.98	1.45	0.81	4.1	5.04	17.65
	10-15	237	359	106	0.70	20.92	1.53	0.83	4.4	4.89	17.90
	15-20	186	286	134	0.61	26.70	1.57	0.85	4.3	4.80	18.21
16	0-5	6	8	25	0.04	0.88	2.15	0.68	4.6	3.67	9.83
17	0-5	95	211	405	0.72	3.52	0.49	0.37	6.1	6.93	9.42
18	0-5	41	114	298	0.48	5.78	0.49	0.65	6.1	6.93	18.09
	5-10	12	27	74	0.12	1.49	0.77	0.74	5.0	6.38	19.40
19	0-5	30	43	151	0.24	12.36	1.08	0.71	5.4	5.78	16.69
	5-10	21	33	37	0.10	2.74	0.99	0.75	4.8	5.95	18.47
	10-15	17	29	25	0.08	1.52	0.94	0.72	4.9	6.05	17.78
20	0-5	116	160	414	0.72	20.22	0.98	0.58	5.9	5.97	13.32
	5-10	20	37	76	0.14	2.68	0.86	0.72	5.2	6.21	18.23
	10-15	20	37	30	0.09	0.97	0.84	0.76	5.2	6.25	19.71
	15-20	16	31	12	0.06	0.52	0.73	0.74	5.3	6.47	19.77
21	0-5	89	182	402	0.70	7.88	0.81	0.58	6.4	6.30	14.25
22	0-5	160	283	541	1.02	6.72	1.54	0.83	4.3	4.86	17.86
	5-10	87	172	464	0.74	4.39	1.34	0.85	4.5	5.26	19.49
	10-15	157	354	183	0.71	2.97	1.28	0.84	4.7	5.38	19.50
	15-20	111	264	66	0.45	2.28	1.26	0.86	4.8	5.42	20.37
23	0-5	25	41	166	0.23	9.09	0.79	0.74	6.0	6.35	19.14
24	0-5	13	0	15	0.03	4.89	1.45	0.77	4.7	5.04	16.42
25	0-5	77	145	267	0.51	4.82	1.62	0.79	4.4	4.71	16.17
	5-10	121	294	157	0.59	4.05	1.49	0.80	4.5	4.96	17.05
	10-15	109	281	79	0.48	2.65	1.42	0.79	4.8	5.09	17.27
	15-20	103	246	88	0.45	3.46	1.44	0.80	4.4	5.06	17.42
	20-25	77	192	47	0.32	2.11	1.37	0.81	4.6	5.19	18.17
26	0-5	439	654	910	2.06	20.62	1.95	0.89	3.3	4.06	17.44
	5-10	480	799	884	2.22	18.56	1.85	0.88	3.6	4.26	17.50
	10-15	461	674	531	1.70	13.20	1.79	0.86	3.8	4.36	17.35
	15-20	657	945	264	1.89	6.61	1.56	0.85	3.9	4.84	18.28

Appendix 3. GDGT concentrations, proxy data and estimated pH and temperature for river sediments.

Site No.	Depth (cm)	GDGT-0 (ng/g)	GDGT- 1 to 3 (ng/g)	Crenarchaeol (ng/g)	Isoprenoid GDGTs (µg/g)	Branched GDGTs (μg/g)	CBT	MBT'	Measured pH	CBT -based pH	MBT'/CBT -based temperature (°C)
1-1	0-5	4	5	6	0.02	0.05	0.50	0.39	6.9	6.92	9.98
1-2	0-5	39	28	44	0.11	0.51	0.67	0.41	5.8	6.57	9.64
2-1	0-1	31	24	50	0.10	0.53	1.13	0.39	7.2	5.67	6.33
2-2	0-1	301	195	406	0.91	6.76	1.19	0.39	7.0	5.56	6.01

Appendix 4-1. GDGT concentrations, proxy data for lake surface sediments.

Site No.	Water Depth (cm)	GDGT-0 (ng/g)	GDGT- 1 to 3 (ng/g)	Crenarchaeol (ng/g)	Isoprenoid GDGTs (µg/g)	Branched GDGTs (µg/g)	BIT	MI	TEX ₈₆	TEX ₈₆ ^L	TEX ₈₆ ^H
671	26.2	30	7	27	0.06	0.30	0.90	0.21	0.52	-0.42	-0.28
676	79.2	1223	127	881	2.24	4.41	0.79	0.13	0.39	-0.63	-0.41
686	70.0	574	143	798	1.52	4.31	0.80	0.15	0.32	-0.69	-0.49
701	55.1	762	79	509	1.35	4.33	0.86	0.13	0.44	-0.46	-0.36
706	28.0	350	50	185	0.59	2.96	0.92	0.21	0.42	-0.50	-0.38
716	13.3	122	48	59	0.23	1.46	0.95	0.44	0.37	-0.63	-0.43
721	9.8	475	46	116	0.64	3.47	0.95	0.28	0.46	-0.41	-0.34
521	50.0	347	38	134	0.52	1.71	0.91	0.22	0.50	-0.53	-0.30

Appendix 4-2. GDGT proxy data and estimated pH and temperature for lake surface sediments.

Site	Water				CBT-based pH		MBT'/CBT-based temperature (°C)				
No.	Depth (cm)	CBT	MBT'	Ajioka et al. (2014)	Peterse et al. (2012)	Tierney et al. (2010)	Ajioka et al. (2014)	Peterse et al. (2012)	Tierney et al. (2010)		
671	26.2	0.52	0.33	6.82	6.87	8.74	6.87	7.94	17.3		
676	79.2	0.36	0.25	7.16	7.20	9.24	5.78	6.49	16.4		
686	70.0	0.36	0.24	7.16	7.19	9.23	5.50	6.16	16.0		
701	55.1	0.35	0.24	7.17	7.21	9.26	5.52	6.18	16.1		
706	28.0	0.31	0.27	7.25	7.28	9.37	6.58	7.38	17.5		
716	13.3	0.71	0.45	6.42	6.49	8.16	9.17	10.86	19.9		
721	9.8	0.28	0.35	7.32	7.34	9.46	8.87	10.04	20.4		
521	50.0	0.46	0.28	6.95	7.00	8.93	6.05	6.91	16.5		

Appendix 5. GDGT concentrations, proxy data and estimated pH and temperature for BIW07-6.

Sample	Top	Mid	Age	Crenarchaeol	Isoprenoid	Branched				CBT	MBT'/CBT
No.	depth	depth	(ka)	(µg/g)	GDGTs	GDGTs	CBT	MBT'	BIT	-based	-based
30	(m)	(m)	0.000	0.18	$(\mu g/g)$	$(\mu g/g)$	0.40	0.28	0.86	рн 7.07	6 41
38	0.30	0.295	0.909	0.18	0.30	1.40	0.40	0.28	0.80	7.07	0.41 6.26
36 46	0.36	0.455	1.432	0.22	0.45	1.74	0.37	0.27	0.85	7.14	6.58
55	0.55	0.545	1.727	0.13	0.26	1.14	0.34	0.30	0.87	7.20	7.23
63	0.63	0.625	1.988	0.23	0.45	1.56	0.29	0.27	0.83	7.29	6.72
71	0.71	0.705	2.250	0.18	0.35	1.40	0.29	0.27	0.85	7.30	6.66
80	0.80	0.795	2.545	0.16	0.35	1.39	0.30	0.27	0.86	7.28	6.65
88	0.88	0.875	2.807	0.19	0.44	1.78	0.28	0.26	0.87	7.31	6.42
96	0.96	0.955	3.068	0.29	0.65	2.29	0.26	0.25	0.85	7.36	6.50
105	1.05	1.045	3.547	0.17	0.39	1.59	0.28	0.26	0.87	7.32	6.51
113	1.13	1.125	4.075	0.18	0.51	2.03	0.28	0.28	0.89	7.32	7.06
121	1.21	1.205	4.881	0.19	0.49	1.94	0.27	0.26	0.88	7.34	6.53
130	1.30	1.295	5.788 6.504	0.22	0.05	2.15	0.17	0.27	0.87	7.30	7.50
136	1.50	1.375	0.394	0.17	0.32	1.23	0.24	0.27	0.90	7.39	6.94
140	1.40	1.435	7.504	0.07	0.31	1.00	0.22	0.20	0.95	7.44	7 17
163	1.63	1.625	7.839	0.06	0.28	1.83	0.20	0.26	0.95	7.50	6.92
171	1.71	1.705	8.062	0.07	0.31	1.90	0.24	0.25	0.95	7.40	6.56
191	1.91	1.905	8.621	0.05	0.23	1.55	0.27	0.26	0.96	7.35	6.52
199	1.99	1.985	8.845	0.07	0.32	2.04	0.26	0.26	0.95	7.37	6.71
205	2.05	2.045	9.012	0.06	0.25	1.77	0.24	0.26	0.96	7.40	6.73
213	2.13	2.125	9.236	0.06	0.30	1.98	0.21	0.25	0.96	7.46	6.65
221	2.21	2.205	9.459	0.06	0.26	1.85	0.28	0.26	0.96	7.31	6.51
230	2.30	2.295	9.711	0.05	0.18	1.26	0.33	0.27	0.95	7.22	6.59
238	2.38	2.375	9.934	0.04	0.16	1.37	0.29	0.27	0.96	7.30	6.79
246	2.46	2.455	10.158	0.05	0.24	1.47	0.29	0.28	0.95	7.30	7.10
255	2.55	2.545	10.435	0.05	0.20	1.00	0.31	0.26	0.96	7.20	0.35
203	2.03	2.025	10.704	0.00	0.29	2.00	0.24	0.25	0.90	7.41	0.02 6.60
271	2.71	2.765	11 141	0.05	0.24	1.77	0.24	0.25	0.96	7.41	6.26
288	2.84	2.835	11.410	0.03	0.15	1.35	0.34	0.27	0.97	7.19	6.31
296	2.92	2.915	11.680	0.02	0.12	1.29	0.41	0.26	0.98	7.05	5.74
305	3.01	3.005	11.983	0.03	0.18	1.35	0.34	0.27	0.97	7.20	6.44
313	3.09	3.085	12.252	0.03	0.18	1.32	0.34	0.24	0.97	7.20	5.72
321	3.17	3.165	12.521	0.03	0.14	1.14	0.32	0.26	0.97	7.23	6.33
330	3.26	3.255	12.824	0.03	0.19	1.38	0.37	0.30	0.97	7.14	7.07
338	3.34	3.335	13.093	0.06	0.53	4.48	0.75	0.45	0.98	6.34	8.88
346	3.42	3.415	13.362	0.03	0.19	1.52	0.39	0.29	0.97	7.09	6.64
355	3.51	3.505	13.666	0.03	0.16	1.45	0.37	0.29	0.97	7.12	6.85
363	3.59	3.585	13.966	0.02	0.13	1.23	0.40	0.29	0.97	/.0/	6.65
3/1	3.07 3.76	3.005	14.207	0.02	0.09	0.90	0.46	0.30	0.98	0.95 6 70	0.00
388	3.70	3.835	14.382	0.02	0.17	0.83	0.34 0.41	0.33	0.98	7.05	6.15
396	3.92	3 915	15.078	0.01	0.13	1.22	0.41 0.52	0.27	0.90	6.81	6.28
405	4.01	4.005	15.355	0.02	0.21	1.37	0.45	0.28	0.97	6.98	6.09
413	4.09	4.085	15.497	0.02	0.12	1.07	0.45	0.27	0.97	6.97	5.91
421	4.17	4.165	15.632	0.02	0.11	0.92	0.40	0.27	0.98	7.07	6.06
430	4.26	4.255	15.784	0.02	0.12	1.03	0.47	0.27	0.97	6.92	5.62
438	4.34	4.335	15.919	0.02	0.09	0.76	0.48	0.31	0.97	6.90	6.59
446	4.42	4.415	16.054	0.01	0.04	0.54	0.60	0.27	0.97	6.66	5.04
455	4.51	4.505	16.206	0.01	0.07	0.64	0.60	0.30	0.97	6.66	5.63
463	4.59	4.585	16.342	0.02	0.11	1.05	0.60	0.30	0.98	6.66	5.81
4/1	4.67	4.665	16.4//	0.01	0.06	0.58	0.66	0.34	0.97	6.52	6.39
480	4.75	4.745	16.012	0.02	0.15	1.40	0.90	0.45	0.99	6.03 6.78	1.87
400	4.65	4.825	16.882	0.02	0.11	0.75	0.54	0.52	0.97	0.78 6.87	0.02 5.34
505	5.00	4.995	17.034	0.01	0.05	0.99	0.43	0.29	0.98	7.00	6.55
513	5.08	5.075	17.169	0.01	0.03	0.44	0.48	0.25	0.97	6.91	5.23
521	5.16	5.155	17.304	0.01	0.04	0.42	0.32	0.24	0.97	7.25	5.69
530	5.25	5.245	17.456	0.01	0.05	0.45	0.37	0.24	0.96	7.14	5.62
538	5.33	5.325	17.591	0.01	0.04	0.40	0.39	0.29	0.98	7.09	6.59
546	5.41	5.405	17.859	0.01	0.04	0.46	0.50	0.26	0.97	6.86	5.31
555	5.50	5.495	18.509	0.02	0.05	0.56	0.47	0.24	0.97	6.92	4.88
563	5.58	5.575	19.086	0.02	0.08	0.49	0.51	0.30	0.95	6.85	6.39
571	5.66	5.655	19.603	0.01	0.06	0.64	0.40	0.25	0.97	7.07	5.64
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1411 13.85 13.845 37.258 0.02 0.07 0.86 0.34 0.25 0.97 7.19 1436 14.10 14.095 37.866 0.03 0.14 1.28 0.38 0.26 0.97 7.19	5.91
1436 1410 14095 37866 003 014 128 038 026 007 712	5.88
1.50 11.10 11.075 57.000 0.05 0.17 1.20 0.50 0.20 0.77 7.12	6.10
1461 14.33 14.325 38.425 0.02 0.09 0.97 0.33 0.25 0.97 7.22	5.86
1486 14.58 14.575 39.033 0.02 0.12 1.20 0.25 0.26 0.97 7.38	6.62
1511 14.83 14.825 39.641 0.02 0.10 0.99 0.30 0.24 0.97 7.29	5.99
1536 15.08 15.075 40.250 0.02 0.09 0.91 0.25 0.24 0.97 7.38	6.06
1561 15.33 15.325 40.858 0.02 0.09 0.93 0.28 0.25 0.97 7.32	6.37
1586 15.55 15.545 41.221 0.03 0.12 0.92 0.28 0.25 0.96 7.32	6.29
1011 15.80 15.795 41.600 0.02 0.08 0.73 0.27 0.24 0.97 7.34 1636 16.05 16.045 41.978 0.02 0.00 0.68 0.22 0.25 0.07 7.45	6.02 6.50
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	6 79
1686 16.54 16.535 42.721 0.02 0.07 0.74 0.30 0.25 0.97 7.51	6.15
1711 16.79 16.785 43.100 0.02 0.11 0.87 0.27 0.24 0.96 7.34	6.11
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Sample No.	Top depth (m)	Mid depth (m)	Age (ka)	Crenarchaeol (µg/g)	Isoprenoid GDGTs (µg/g)	Branched GDGTs (µg/g)	CBT	MBT'	BIT	CBT -based pH	MBT'/CBT -based temperature (°C)
1736	17.04	17.035	43.479	0.03	0.13	1.21	0.25	0.24	0.97	7.38	6.17
1761	17.29	17.285	43.858	0.04	0.16	1.37	0.18	0.23	0.96	7.52	6.37
1786	17.54	17.535	44.237	0.03	0.14	1.44	0.18	0.24	0.97	7.53	6.48
1811	17.79	17.785	44.787	0.04	0.18	1.50	0.28	0.23	0.96	7.32	5.78
1836	18.04	18.035	45.495	0.03	0.16	1.38	0.19	0.25	0.96	7.50	6.77
1861	18.29	18.285	46.204	0.03	0.14	1.31	0.30	0.27	0.97	7.28	6.71

Appendix 6. GDGT concentrations, proxy data and estimated pH and temperature for BIW08-B.

Sample	Top	Mid	Age	Crenarchaeol	Isoprenoid	Branched				CBT	MBT'/CBT
No	depth	depth	Age (ka)	(ug/g)	GDGTs	GDGTs	CBT	MBT'	BIT	-based	-based
INO.	(m)	(m)	(Kd)	(µg/g)	$(\mu g/g)$	$(\mu g/g)$				pН	temperature (°C)
8-18	13.495	13.5075	42.353	0.04	0.21	2.01	0.29	0.26	0.97	7.29	6.58
8-1 28	13.995	14.0075	43.047	0.04	0.18	1.61	0.41	0.26	0.97	7.05	5.87
8-2 12	14.480	14.4925	43.700	0.05	0.27	2.21	0.32	0.26	0.97	7.23	6.33
8-2 30	14.930	14.9425	44.652	0.07	0.32	2.76	0.29	0.26	0.97	7.30	6.56
9-17	15.495	15.5075	47.156	0.06	0.30	2.72	0.29	0.28	0.97	7.30	6.90
9-1 27	15.995	16.0075	49.371	0.03	0.17	1.58	0.52	0.31	0.98	6.82	6.43
9-2 18	16.490	16.5025	51.564	0.03	0.18	1.96	0.62	0.35	0.98	6.61	6.86
9-2 37	16.965	16.9775	53.669	0.04	0.17	1.95	0.52	0.32	0.98	6.83	6.78
10-1 16	17.490	17.5025	55,995	0.03	0.16	1.93	0.55	0.34	0.98	6.75	7.06
10-1 36	17 990	18 0025	58 210	0.04	0.24	2.84	0.61	0.38	0.98	6 64	7 69
10-2.19	18 475	18 4875	60 359	0.06	0.27	2.92	0.48	0.30	0.98	6.90	6.75
10-2 38	18.950	18 9625	62 464	0.09	0.54	7.53	0.44	0.35	0.98	6.99	8.08
11-1.6	10.250	19 5025	64 856	0.05	0.34	3 29	0.37	0.33	0.90	7.14	7 37
11 1 26	10.000	20.0025	67.071	0.05	0.37	3.05	0.37	0.34	0.90	6.06	7.57
11-120	20.450	20.0025	60 110	0.05	0.32	3.05 4.01	0.45	0.34	0.98	0.90	7.01
11-2 10	20.450	20.4025	71 325	0.08	0.49	2.04	0.37	0.32	0.97	7.14	7.00
11-2 30	20.950	20.9025	71.323	0.04	0.20	2.04	0.30	0.29	0.97	7.20	7.50
12-115	21.970	21.9823	73.844	0.08	0.57	5.11	0.55	0.51	0.97	7.24	7.39
12-2 14	22.450	22.4625	//.9/1	0.11	0.58	4.00	0.27	0.31	0.96	7.34	8.00
12-2 34	22.950	22.9625	80.186	0.12	0.55	4.54	0.30	0.34	0.96	7.29	8.42
13-1 19	23.985	23.9975	84.504	0.07	0.33	2.95	0.32	0.32	0.97	7.24	7.92
13-214	24.465	24.4775	86.588	0.08	0.50	3.58	0.35	0.30	0.97	7.19	7.33
13-2 34	24.965	24.9775	88.759	0.07	0.33	3.85	0.38	0.30	0.97	7.10	6.97
14-1 2	25.510	25.5225	91.125	0.07	0.33	2.96	0.35	0.29	0.97	7.18	7.01
14-1 22	26.010	26.0225	93.296	0.08	0.64	3.49	0.38	0.33	0.97	7.12	7.71
14-2 14	26.470	26.4825	95.293	0.09	0.81	5.49	0.42	0.32	0.98	7.03	7.38
14-2 34	26.970	26.9825	98.130	0.14	1.25	7.11	0.35	0.31	0.97	7.18	7.50
15-1 10	27.495	27.5075	102.438	0.12	0.62	4.19	0.41	0.31	0.96	7.06	7.12
15-1 30	27.995	28.0075	105.828	0.15	0.77	4.57	0.31	0.30	0.96	7.25	7.37
15-2 15	28.475	28.4875	108.181	0.11	0.92	3.63	0.24	0.30	0.96	7.40	7.93
16-1 14	29.475	29.4875	109.247	0.08	0.41	2.22	0.32	0.32	0.95	7.23	7.94
16-1 34	29.975	29.9875	109.781	0.11	0.70	5.17	0.32	0.28	0.97	7.24	6.89
16-2 17	30.470	30.4825	110.309	0.09	0.61	5.36	0.41	0.33	0.98	7.05	7.60
16-2 37	30.970	30.9825	110.842	0.21	0.93	7.80	0.39	0.33	0.96	7.08	7.73
17-1 10	31.480	31.4925	111.386	0.24	0.80	5.76	0.39	0.32	0.95	7.09	7.46
17-1 30	31.980	31.9925	111.920	0.37	1.30	8.54	0.39	0.31	0.94	7.09	7.13
17-2 15	32,460	32,4725	112.432	0.42	1.29	7.45	0.44	0.33	0.93	7.00	7.39
17-2 35	32.960	32.9725	112.965	0.55	1.74	8.69	0.35	0.29	0.92	7.18	7.02
18-1.9	33,480	33.4925	113,520	0.47	1.79	6.45	0.33	0.31	0.91	7.21	7.55
18-1 29	33 980	33 9925	114 053	0.61	1.62	6 54	0.30	0.31	0.88	7 29	7 77
18-2.14	34 450	34 4625	114 554	0.72	2.44	7.83	0.29	0.28	0.88	7 31	7.15
18-2 34	34 950	34 9625	115 141	0.41	1.69	6.49	0.26	0.20	0.00	7.36	7.40
10 2 34	35 570	35 5825	116.018	0.36	2.16	7 39	0.20	0.30	0.92	7.30	7.40
19-1 31	36.070	36.0825	116 725	0.56	1.02	5 71	0.27	0.30	0.95	7.31	8.09
19-2.14	36.470	36.4825	117 291	0.10	1.02	5.90	0.24	0.32	0.90	7.33	8 58
10 2 34	36.970	36 0825	117.008	0.14	1.00	5.70	0.24	0.33	0.97	7.40	8.12
20 1 14	37.480	37 4025	118 710	0.10	0.52	1 13	0.27	0.32	0.97	7.35	8.12
20-1 14	27.020	27.0025	110.719	0.09	0.52	4.15	0.29	0.33	0.97	7.30	8.25
20-1 34	37.960 28.460	37.9923	119.420	0.17	0.92	2 27	0.29	0.32	0.90	7.29	8.15
20-2 19	28.0400	28 0725	120.103	0.07	0.37	2.57	0.34	0.37	0.97	7.19	0.90 8 5 0
20-2 39	20,400	30.9723	120.812	0.09	0.40	5.50	0.39	0.30	0.97	7.10	8.30
21-1 19	39.490	39.5025	121.728	0.14	0.90	9.19	0.39	0.33	0.98	7.10	7.04
21-210	39.990	40.0025	122.597	0.11	0.67	7.00	0.60	0.31	0.98	0.05	6.06
21-2 35	40.465	40.4775	123.423	0.12	0.61	7.82	0.75	0.36	0.98	6.33	6.39
22-2.2	40.990	41.0025	125.576	0.09	0.50	5.93	0.52	0.33	0.98	6.82	6.90
22-2.22	41.490	41.5025	127.948	0.06	0.21	2.01	0.56	0.32	0.96	6.74	6.57
23 21	42.040	42.0525	130.557	0.08	0.41	5.02	0.52	0.28	0.98	6.82	5.76
24 4	42.485	42.4975	132.668	0.09	0.55	4.75	0.43	0.30	0.98	7.02	6.76
24 24	42.985	42.9975	135.040	0.05	0.23	2.79	0.47	0.30	0.98	6.92	6.43
25 7	43.490	43.5025	137.436	0.08	0.33	3.83	0.44	0.30	0.97	6.99	6.67
25 27	43.990	44.0025	139.808	0.06	0.29	3.97	0.55	0.34	0.98	6.77	7.17
26 7	44.500	44.5125	142.228	0.05	0.21	2.72	0.47	0.30	0.98	6.92	6.47
26 27	45.000	45.0125	144.049	0.06	0.26	3.52	0.49	0.33	0.98	6.89	7.10
27 15	45.490	45.5025	145.585	0.06	0.26	3.45	0.48	0.31	0.98	6.91	6.72
27 34	45.965	45.9775	147.074	0.07	0.25	2.12	0.57	0.32	0.96	6.71	6.47
28 17	46.480	46.4925	148.689	0.05	0.22	2.45	0.58	0.35	0.98	6.69	7.07
29 4	46.980	46.9925	150.256	0.04	0.24	2.81	0.46	0.31	0.98	6.96	6.91

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Sample	Top	Mid	Age	Crenarchaeol	Isoprenoid	Branched	CDT	MDT'	DIT	CBT	MBT'/CBT
No.	(m)	(m)	(ka)	$(\mu g/g)$	GDG1s (µg/g)	GDG1s (µg/g)	CBI	MBT	BII	-based pH	-based temperature (°C)
29 24	47.480	47.4925	151.824	0.04	0.16	1.46	0.57	0.31	0.97	6.72	6.09
30 4	47.995	48.0075	153.439	0.05	0.19	2.06	0.48	0.33	0.97	6.92	7.13
30 24	48.495	48.5075	155.006	0.04	0.18	1.89	0.55	0.35	0.97	6.76	7.28
31 12	48.995 49.495	49.0075	150.574	0.08	0.26	2.18	0.54	0.30	0.97	6.79 7.02	6.09
32 15	49.990	50.0025	159.693	0.11	0.48	4.49	0.33	0.28	0.97	7.21	6.65
32 34	50.465	50.4775	161.183	0.14	0.67	5.04	0.35	0.28	0.96	7.18	6.70
33 19	50.980	50.9925	162.797	0.12	0.44	5.03	0.37	0.27	0.97	7.14	6.27
34 4	51.485	51.4975	164.381	0.10	0.35	3.65	0.47	0.31	0.97	6.92	6.61
34 23	51.985	51.9975	165.948	0.14	0.53	4.85	0.38	0.30	0.96	7.11	6.93
36 4	52.480	52.4925	167.500	0.07	0.26	2.76	0.49	0.31	0.97	6.89	6.63
30 24 37 4	52.980 53.465	52.9925 53.4775	109.008	0.08	0.51	5.24 4.41	0.47	0.30	0.97	6.94 6.90	0.03 6.59
37 24	53 965	53 9775	172.156	0.11	0.31	2.57	0.48	0.31	0.97	6.90 6.69	6.88
38 2	54.480	54.4925	173.770	0.08	0.30	2.97	0.48	0.29	0.97	6.90	6.26
38 22	54.980	54.9925	175.338	0.09	0.45	4.68	0.58	0.35	0.98	6.69	7.20
39 4	55.485	55.4975	176.921	0.09	0.42	4.23	0.58	0.34	0.97	6.70	6.82
39 24	55.985	55.9975	178.489	0.09	0.34	3.14	0.73	0.33	0.97	6.38	5.80
40 2	56.490	56.5025	180.072	0.11	0.54	6.63	0.47	0.33	0.98	6.93	7.32
40 22	56.990	57.0025	181.640	0.09	0.49	4.08	0.45	0.31	0.97	6.98 6.05	6.94 6.65
41 5 41 25	57.46U 57.960	57 9725	103.113	0.08	0.41	4.22 5.52	0.40 0.46	0.30	0.98 0.98	0.95	0.00 7 10
42.4	58.475	58.4875	186.296	0.15	0.63	6.05	0.42	0.30	0.97	7.04	6.86
42 24	58.975	58.9875	187.863	0.19	0.93	7.47	0.43	0.30	0.97	7.01	6.86
43 4	59.500	59.5125	189.509	0.07	0.29	2.83	0.44	0.31	0.97	6.98	6.87
43 24	60.000	60.0125	191.077	0.05	0.21	1.99	0.43	0.28	0.97	7.02	6.31
44 4	60.510	60.5225	192.676	0.17	0.91	5.94	0.30	0.27	0.96	7.28	6.71
44 24	61.010	61.0225	194.243	0.16	0.72	4.91	0.29	0.30	0.96	7.30	7.57
45 4	61.455	61.4675	195.638	0.21	0.89	6.01	0.26	0.30	0.95	7.36	7.63
45 24 46 4	62 505	62 5175	197.200	0.24	1.15	0.45	0.54	0.27	0.95	7.20 6.90	6.35 6.94
46.24	63.005	63.0175	200.498	0.29	1.10	9.14	0.34	0.32	0.96	7.19	7.40
47 4	63.485	63.4975	202.003	0.34	1.36	9.82	0.34	0.29	0.95	7.19	6.97
47 24	63.985	63.9975	203.570	0.38	1.28	7.26	0.31	0.31	0.93	7.26	7.65
48 2	64.475	64.4875	205.107	0.48	1.53	7.59	0.36	0.32	0.92	7.16	7.78
48 22	64.975	64.9875	206.674	0.85	2.55	9.04	0.28	0.31	0.88	7.31	7.80
49 4	65.435	65.4475	208.116	0.77	2.52	7.92	0.28	0.31	0.87	7.31	7.76
49 24 50 4	65.935	65.94/5	209.684	0.30	1.16	5.41	0.26	0.32	0.93	7.36	8.13
50 4 50 24	66 975	66 9875	211.577	0.17	0.78	0.14	0.34 0.41	0.32	0.90	7.19	6.73
514	67.475	67.4875	212.545	0.15	0.51	4.48	0.50	0.2)	0.96	6.87	6.59
51 24	67.975	67.9875	216.080	0.13	0.49	4.04	0.52	0.36	0.96	6.82	7.76
52 5	68.470	68.4825	217.632	0.13	0.46	5.04	0.46	0.34	0.97	6.95	7.70
52 25	68.970	68.9825	219.199	0.14	0.54	5.51	0.56	0.33	0.97	6.75	6.91
53 5	69.510	69.5225	220.892	0.20	0.93	7.82	0.40	0.33	0.97	7.08	7.64
53 25	70.010	70.0225	222.460	0.22	0.94	7.83	0.38	0.33	0.96	7.11 6.07	7.91 6 01
54 5 54 75	70.500	70.5125	223.990 225 561	0.15	0.82	1.58	0.45	0.31	0.97 0.96	0.97 7 02	0.91
55.6	71.495	71.5075	227.116	0.12	0.53	4.79	0.46	0.34	0.97	6.95	7.73
55 26	71.955	71.9675	228.558	0.35	1.65	12.32	0.37	0.32	0.96	7.13	7.68
56 6	72.525	72.5375	230.345	0.42	2.00	12.58	0.34	0.33	0.95	7.19	7.90
56 26	73.025	73.0375	231.913	0.43	1.75	9.78	0.35	0.32	0.94	7.17	7.63
57 6	73.525	73.5375	233.480	0.43	1.48	7.74	0.32	0.33	0.93	7.24	8.14
57 26	74.025	74.0375	235.048	0.42	1.48	6.49	0.30	0.32	0.91	7.27	8.06
586 5894	75.005	75 0175	230.353	0.6/	2.03	8.15	0.26	0.32	0.89	1.36 7.25	8.1/ 0 20
50 20 59 6	75 515	75 5275	230.120 239.719	0.55	1.54 2.06	8.03	0.20	0.34	0.00	7.55 7.44	0.00 9.16
59 26	76.015	76.0275	241.287	0.31	0.98	4.93	0.22	0.36	0.09	7.31	9.13
60 6	76.505	76.5175	242.823	0.22	1.00	5.83	0.29	0.36	0.95	7.29	9.08
60 26	77.005	77.0175	244.391	0.23	1.07	6.69	0.27	0.36	0.95	7.33	9.07
61 2	77.475	77.4875	245.864	0.19	0.94	7.40	0.37	0.33	0.96	7.14	7.95
61 22	77.975	77.9875	247.432	0.16	0.76	8.01	0.34	0.31	0.97	7.20	7.51
62 2	78.405	78.4175	248.780	0.30	1.32	10.24	0.21	0.30	0.96	7.47	8.03
62 22	78.905	78.9175	250.348	0.18	1.08	6.87 7.75	0.32	0.29	0.96	7.24	7.17
02 39 64 3	19.33U 70 600	19.3423 70.6125	231.08U 252 527	0.21	1.30	1.13 0.16	0.29	0.33	0.90	7.51 7.16	8.29 7 50
0 4 J	77.000	17.0123	232.321	0.51	1.57	2.10	0.50	0.51	Continu	e to the n	(i) (ext page)
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	Top	Mid			Isoprenoid	Branched				CBT	MBT'/CBT
Sample	denth	denth	Age	Crenarchaeol	GDGTs	GDGTs	CBT	MRT'	BIT	-based	_based
No.	(m)	(m)	(ka)	$(\mu g/g)$	(µg/g)	(µø/ø)	СБТ	MIDT	DII	nH	temperature (°C)
6/ 13	70.815	79 8275	253 201	0.00	0.02	0.18	0.36	0.34	0 00	7.16	8 26
64 22	20.065	P0.0275	252.084	0.00	0.02	0.10	0.30	0.34	0.55	7.10	0.20 7 77
04 23	80.003	80.0773	255.964	0.27	1.52	0.15	0.20	0.30	0.95	7.57	7.77
65 3	80.480	80.4925	255.285	0.23	1.28	9.16	0.26	0.30	0.96	1.37	/.69
65 23	80.980	80.9925	256.853	0.18	0.77	6.34	0.28	0.33	0.96	7.32	8.45
65 31	81.180	81.1925	257.480	0.07	0.37	3.04	0.31	0.34	0.97	7.26	8.53
66 3	81.455	81.4675	258.342	0.14	0.76	4.76	0.36	0.32	0.96	7.16	7.79
66 23	81.955	81.9675	259.910	0.11	0.44	4.38	0.49	0.35	0.97	6.88	7.72
67 2	82.415	82.4275	261.352	0.12	0.45	4.78	0.44	0.33	0.97	6.98	7.35
67 22	82.915	82.9275	262.920	0.04	0.19	1.86	0.63	0.36	0.97	6.60	7.11
67 38	83.315	83.3275	264.174	0.03	0.18	1.68	0.76	0.40	0.98	6.32	7.43
68 27	84.050	84.0625	266.478	0.17	0.78	6.45	0.44	0.30	0.97	7.00	6.73
69 5	84.450	84.4625	267.732	0.19	0.69	6.40	0.51	0.32	0.96	6.85	6.93
69 25	84.950	84.9625	269.300	0.20	0.82	6.75	0.44	0.31	0.96	6.99	6.88
70 4	85.510	85.5225	271.055	0.31	1.17	9.55	0.44	0.33	0.96	6.99	7.40
70 24	86.010	86.0225	272.623	0.23	0.83	6.13	0.41	0.33	0.95	7.05	7.71
71 2	86.425	86.4375	273.924	0.18	0.74	6.37	0.41	0.30	0.96	7.04	6.80
718	86.575	86.5875	274.394	0.03	0.13	1.07	0.39	0.32	0.97	7.10	7.38
71 22	86.925	86.9375	275.492	0.35	1.86	10.25	0.35	0.33	0.95	7.17	7.84
72 2	87.425	87.4375	277.059	0.33	1.16	7.07	0.33	0.35	0.94	7.21	8.54
72 22	87.925	87.9375	278.627	0.31	1.02	6.03	0.33	0.38	0.93	7.23	9.36
73 9	88.370	88.3825	280.022	0.37	1.47	8.62	0.34	0.39	0.94	7.20	9.65