Protective Efficacy of Neutralizing Monoclonal Antibodies in a Nonhuman Primate Model of Ebola Hemorrhagic Fever

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Abstract

Ebola virus (EBOV) is the causative agent of severe hemorrhagic fever in primates, with human case fatality rates up to 90%. Today, there is neither a licensed vaccine nor a treatment available for Ebola hemorrhagic fever (EHF). Single monoclonal antibodies (MAbs) specific for Zaire ebolavirus (ZEBOV) have been successfully used in passive immunization experiments in rodent models, but have failed to protect nonhuman primates from lethal disease. In this study, we used two clones of human-mouse chimeric MAbs (ch133 and ch226) with strong neutralizing activity against ZEBOV and evaluated their protective potential in a rhesus macaque model of EHF. Reduced viral loads and partial protection were observed in animals given MAbs ch133 and ch226 combined intravenously at 24 hours before and 24 and 72 hours after challenge. MAbs circulated in the blood of a surviving animal until virus-induced IgG responses were detected. In contrast, serum MAb concentrations decreased to undetectable levels at terminal stages of disease in animals that succumbed to infection, indicating substantial consumption of these antibodies due to virus replication. Accordingly, the rapid decrease of serum MAbs was clearly associated with increased viremia in non-survivors. Our results indicate that EBOV neutralizing antibodies, particularly in combination with other therapeutic strategies, might be beneficial in reducing viral loads and prolonging disease progression during EHF.

Introduction

Ebola virus (EBOV) has a non-segmented, single strand negative-sense RNA genome and, together with Marburg virus, constitutes the family Filoviridae [1]. EBOV causes severe hemorrhagic fever in humans and nonhuman primates (NHPs) with the highest human case fatality rates among hemorrhagic fever viruses. Currently, there is neither an effective prophylaxis nor treatment available for Ebola hemorrhagic fever (EHF). While all Marburg virus isolates currently belong to a single virus species, multiple EBOV species have been described [1,2]. *Zaire ebolavirus* (ZEBOV), first identified in 1976, is the most virulent species with case fatality rates in humans approaching 90% and almost 100% lethality in experimental macaque models [1], the current gold standard animal model among several established ZEBOV disease models [3].

The EBOV transmembrane glycoprotein (GP) is responsible for both receptor binding and fusion of the virus envelope with the host cell membrane [4,5], and is the only known target for neutralizing antibodies against this virus. The presence of EBOV-neutralizing antibodies was confirmed in the sera of convalescent patients and experimentally infected NHPs [6,7]. The protective efficacy of passive immunization with hyperimmune sera or purified polyclonal antibodies was evaluated using rodent models and shown to be effective in mice and guinea pigs, whereas evidence of protective efficacy in primates, including humans, remains elusive [6,7,8]. In contrast, we have shown that certain GP-specific antibodies enhance filovirus infection *in vitro*, a
Infectivity of ZEBOV in Vero E6 cells in a dose-dependent neutralization test [20]. Both MAbs significantly reduced the infection of ZEBOV in Vero E6 cells in a dose-dependent manner (Figure 2), whereas the negative control MAbs (ch61) did not. The 50% inhibitory concentrations of ch133 and ch226 were 1.6 and 2.1 μg/ml, respectively. These values were similar to those of the original mouse MAbs (3.2 and 0.8 μg/ml, respectively) [19], indicating that genetic modification of these MAbs did not significantly affect their ability to neutralize ZEBOV in vitro.

**Results**

**MAbs ch133 and ch226 neutralize ZEBOV in vitro**

In a previous study, we have identified different amino acid residues important for the neutralizing activity of the two mouse MAbs, ZGP133/16.3 and ZGP226/8.1, using a surrogate virus system [15]. All escape mutants selected in the presence of ZGP133/3.16 contained a single amino acid substitution at position 549 in the ZEBOV GP. In contrast, for ZGP226/8.1 three different escape mutants were isolated containing amino acid substitution at position 134, 194, or 199 in the ZEBOV GP, suggesting that this antibody recognizes a different conformational epitope. Mapping of these epitopes, together with that of KZ52, on the 3-D structure of the ZEBOV GP molecule indicates that these MAbs likely bind to different epitopes, although the ZGP133 epitope may partially overlap with that of KZ52 (Figure 1).

To evaluate the protective efficacy in nonhuman primates, we converted ZGP133/16.3 and ZGP226/8.1 into the human-mouse chimeric MAbs ch133 and ch226. MAb ch61 specific for the influenza virus hemagglutinin (HA) was generated as a control antibody. The neutralizing activities of chimeric MAbs ch133 and ch226 were analyzed in vitro by performing a focus reduction neutralization test [20]. Both MAbs significantly reduced the infectivity of ZEBOV in Vero E6 cells in a dose-dependent manner.
(25 mg of each MAb; 50 mg total) 24 hours before and 24 and 72 hours after challenge with a lethal dose of ZEBOV, strain Kikwit ($10^3$ plaque-forming units). A control animal (CTRL) was identically challenged and treated at the same time points with MAb ch61 by the same route and dose. Animals CTRL and EBO1 developed fulminant EHF with viremia levels exceeding $10^4$ TCID$_{50}$ equivalents/ml prior to day 8 and had to be euthanized on days 7 and 8, respectively (Figure 3A). This is a normal disease progression for rhesus macaques infected with a lethal dose of ZEBOV. Animal EBO2 showed a delayed onset of clinical signs and prolonged time to death with viremia levels still below $10^3$ TCID$_{50}$/ml on day 8 (Figure 3B), although it had to be euthanized with characteristic signs of EHF on day 11. Furthermore, virus titers in liver, spleen, and adrenal gland were more than 1 log higher in the control animal (CTRL) compared to EBO2 (Table 1), again showing the delayed disease progression in this animal. Animal EBO3 was protected from clinical disease and survived. This animal had only very low level viremia detected by qRT-PCR on day 8 (Figure 3A); however, virus isolation was negative (Figure 3B). In addition, the survivor EBO3 showed no significant ZEBOV-specific changes in blood chemistry or hematology throughout the study; its liver enzyme levels (i.e. alanine aminotransferase (ALT)), as well as platelet counts, were always within the normal range (Figures 3C and 3D). To exclude viral escape under neutralizing pressure, we sequenced viral RNA isolated from blood collected on days 8 (EBO1) and 11 (EBO2). No mutation was found in the GP genes, indicating that virus escape did not occur as previously described for both MAbs in vitro [15].

Serum antibody levels in treated nonhuman primates

MAb concentrations in the serum samples collected throughout the experiment were monitored using enzyme-linked immunosorbent assay (ELISA). MAb concentrations (ch133 and ch226) on days 1–5 after challenge were maintained above 75 μg/ml, but drastically decreased to almost undetectable levels in animals EBO1 and EBO2 on day 8 (Figure 4A). The rapid decrease in serum MAb concentrations was timely associated with increased

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**Figure 3. Hallmark laboratory parameters after ZEBOV challenge.** Viral RNAs in the blood samples were detected as described in Materials and Methods (A). Virus titers in the blood samples collected 8 days after challenge were determined as TCID$_{50}$ in Vero E6 cells (B). For CTRL titers for the sample collected on day 7 are shown. Platelet counts (C) were determined from whole blood samples; alanine aminotransferase levels (ALT) (D) were determined from serum at the indicated time points.

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viremia in these non-survivors (Figures 3A and 4A). Notably, higher levels of MAb concentrations were detected in the serum of the surviving animal (EBO3), remaining above 50 μg/ml until day 11 before IgG concentrations steadily increased (Figure 4A). This increase is due to a challenge virus-induced IgG response against ZEBOV proteins leading to recovery from infection as demonstrated by the increase of nucleoprotein (NP)-specific IgGs (Figure 4B). The concentration of MAb ch61 in the serum of CTRL remained above 150 μg/ml until day 7 indicating no consumption of neutralizing antibodies despite the increase of nucleoprotein (NP)-specific IgGs (Figure 4B). The concentration of MAb ch61 in the serum of CTRL remained above 150 μg/ml until day 7 indicating no consumption of neutralizing antibodies. The concentration of MAb ch61 in the serum of EBO3 ch133+ch226, surviving animals during the prophylactic treatment study. Antibodies specific to GP (A), NP (B), and influenza virus HA (C) were detected by ELISA, as described in Materials and Methods. Levels of anti-HA antibody were measured in the animal given only the control MAb (anti-HA, ch61). This passive transfer study, serum MAb concentrations dropped remarkably at the terminal stage of the disease, indicating substantial consumption of these antibodies in the blood. Since both MAbs used in this study do not bind sGP dimers [15], this observation is likely due to uncontrolled virus replication leading to production of large quantities of the shed trimeric glycoprotein functioning as antibody decoy in the blood. Therefore, it seems that high levels of plasma antibodies are required to suppress virus replication until host immune responses are sufficiently induced. This could be achieved through additional injections (i.e. day 5 and day 7) or through higher MAb concentrations in the initial injections. Complete neutralization of ZEBOV using ch133 and ch226 in vitro was achieved at concentrations of greater than 40 μg/ml (Figure 2). The antibody dose used in this NHP study, which gave approximately 50–100 μg/ml blood at each treatment, did not significantly differ from the in vitro situation, but the half-life time of the MAb was less than 4 days. This again indicates that more than three injections (every 2–3 day) may have been beneficial. One possible reason for relatively short half-life of our chimeric MAbs might be reduced stability due to desialylation during the prolonged protein expression process in CHO cell culture. In general, sialic acids likely influence the solubility, thermal stability, and resistance to protease attack of various glycoproteins. Thus, it seems possible that the stability of MAbs may be improved by enhancing sialylation during expression in CHO cells [26]. It has been demonstrated that EBOV utilizes multiple cellular pathways for entry into host cells [27]. Direct inhibition of GP attachment to cell surface or endosomal receptor(s) and blocking fusion of viral and host membranes are likely to be key mechanisms of neutralization. Preventing cathepsin cleavage is another formal possibility but remains controversial [28,29]. The GP gene of EBOV has two overlapping reading frames expressing the full-length transmembrane GP and two non-structural soluble GPs (sGP and ssGP), which are secreted from ZEBOV-infected cells [21–24]. Due to its high abundance in the blood of infected humans, it has been proposed that sGP facilitates virus spread by deactivating anti-GP antibodies. Indeed, it has been shown that sGP can reduce the neutralizing activities of anti-GP serum in vitro [23]. A similar function was also suggested for the shed GP ectodomain resulting from GP cleavage on the cell or virus surface [25]. In this passive transfer study, serum MAb concentrations dropped remarkably at the terminal stage of the disease, indicating substantial consumption of these antibodies in the blood. Since both MAbs used in this study do not bind sGP dimers [15], this observation is likely due to uncontrolled virus replication leading to production of large quantities of the shed trimeric glycoprotein functioning as antibody decoy in the blood. Therefore, it seems that high levels of plasma antibodies are required to suppress virus replication until host immune responses are sufficiently induced. This could be achieved through additional injections (i.e. day 5 and day 7) or through higher MAb concentrations in the initial injections. Complete neutralization of ZEBOV using ch133 and ch226 in vitro was achieved at concentrations of greater than 40 μg/ml (Figure 2). 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**Discussion**

Passive transfer of antibodies leads to either complete inhibition of virus replication, resulting in sterile immunity, or incomplete protection in which the virus replicates at a reduced level, allowing the host to mount virus-induced immune responses resulting in virus clearance. In this study, passive transfer of a mixture of two neutralizing MAbs conferred partial protection in rhesus macaques against lethal ZEBOV challenge with 1 of 3 animals surviving and another one showing delayed disease progression. Our data demonstrate that animals can be protected even when primary challenge virus replication occurs and that “sterile immunity” is not necessarily required for protection from lethal ZEBOV infection.
single human MAb KZ52 did not protect rhesus macaques from lethal ZEBOV challenge given a higher dose of MAb (50 mg/kg, two times) [18] than that used in this NHP study (approximately 10–13 mg/kg, 3 times). Thus, the combination of two MAbs as done in this study seems to have improved treatment efficacy at lower antibody doses. Efficacy might be even higher with a cocktail of three or more MAbs, in particular if they target distinct epitopes and thus have independent mechanisms of action.

Previous studies on viral vector-based EBOV vaccine have suggested that the induction of cellular immune responses is also an important protective mechanism for EBOV infection [30,31]. Since dysfunction of the immune system is critical for the pathogenesis of EHF in humans and NHPs [6,8,32], strategies need to be developed improving the immune functions (both humoral and cellular) disrupted during EHF. Thus, combined treatment with neutralizing MAbs and immune-modulating compounds should be evaluated in a future NHP study.

Materials and Methods

Challenge virus

ZEBOV (strain Kikwit) (kindly provided by the Centers for Disease Control and Prevention, Atlanta, Georgia, USA) was propagated in commonly used African green monkey kidney Vero E6 cells (kindly provided by Dr. R. Baric, University of North Carolina, Chapel Hill, NC, USA). The supernatants were cleared of cell debris, aliquoted, and stored in liquid nitrogen until used. All infectious work with ZEBOV was performed in the biosafety level 4 (BSL-4) laboratories at the Integrated Research Facility in the Rocky Mountain Laboratories (RML), Division of Intramural Research, National Institute of Allergy and Infectious Diseases (NIAID), National Institutes of Health (NIH), Hamilton, Montana, USA.

Monoclonal antibodies

Total RNA was extracted from mouse hybridoma cells producing mouse MAb ZGP133/3.16 or ZGP226/8.1, both of which were shown previously to neutralize ZEBOV [15,19]. The variable heavy- and light-chain regions were amplified by RT-PCR with primers designed for these antibodies. The PCR products were subcloned into the pBR322-based plasmid, heavy- and light-chain (IgG1) construction vectors (pDN11-g1 and pCB-k, respectively), and the light chain cassette was transferred from pCB-k into the heavy-chain expression vector pDN11-g1 (Figure 5). The resulting plasmids (pDN11-kg1) expressing human-mouse chimeric MAbs ch133 and ch226 were designated DN11-ch133kg1 and DN11-ch226kg1, respectively. Stable cell lines expressing recombinant MAbs ch133 and ch226 were obtained by transfection of CHO DG44 cells (Invitrogen) with DN11-ch133kg1 and DN11-ch226kg1, respectively. Forty-eight hours after transfection, cells were seeded at a density of \(1 \times 10^3\) cells/0.1 ml/well onto 96-well microculture plates in IS-CHO CD medium (Irvine Scientific) containing G418 (400 \(\mu g/ml\)). Two weeks after transfection, proliferating clones were isolated and...
transferred to 24-well plates. Culture supernatants were collected and analyzed for production of each MAb (ch133 and ch226) by ELISA. For each MAb, the clone showing the highest expression level was propagated further. A chimeric MAb (ch61) specific for influenza virus hemagglutinin [strain A/Viet Nam/1194/2004 (H5N1)] was generated as a control MAb using the same methodology. MAb-expressing cell clones were maintained in IS-CHO CD medium and the recombinant MAbbs ch133, ch226, and ch61 were purified from culture supernatants using rProtein A Sepharose Fast Flow (GE Healthcare) and Endospecy ES-50M (Seikagaku Biobusiness Corporation). MAb purity (90%<) and endotoxin levels (<1.0 EU/ml) were confirmed by performing SDS-PAGE and an EndoTrap red test (Profos AG), respectively.

Neutralization assay
Vero E6 cells were seeded in a 48-well plate to generate a confluent monolayer on the day of infection. MAb dilutions were prepared in DMEM supplemented with 2% fetal bovine serum and 25 µl were incubated with 200 focus forming units of ZEBOV in a total volume of 50 µl. After 30 min at 37°C the media was removed from cells, the virus-serum mixture was added and incubated for 60 min at 37°C. Then the mixture was removed from the cells and 0.5 ml of a 1.2% carboxymethyl cellulose (CMC)/MEM (Life Technologies) solution was added per well. Following incubation for 4 days at 37°C the plates were fixed with 10% neutral buffered formalin and removed from BSL4 laboratories according to standard operating procedures. Subsequently, the cells were permeabilized and foci were stained with a rabbit anti-VP40 antibody (kindly provided by Dr. Y. Kawaoka, University of Wisconsin, Madison, WI) followed by a FITC-rabbit anti-VP40 antibody (kindly provided by Dr. Y. Kawaoka, University of Wisconsin, Madison, WI) and endotoxin levels (<1.0 EU/ml) were confirmed by performing SDS-PAGE and an EndoTrap red test (Profos AG), respectively.

Passive immunization and protection experiments
In the prophylactic treatment study, rhesus macaques (male adults, 4.0–5.2 kg) were given a mixture of MAb ch133 and ch226 (25 mg each/animal; total of 50 mg per animal) (n = 3) or ch61 (50 mg/animal) (n = 1) intravenously 24 hours prior to infection (day –1) to challenge with an intramuscular injection of 10^3 plaque-forming units of ZEBOV. The same amounts of antibodies were administered again using the same route 24 and 72 hours after challenge. Blood samples were collected throughout the study (on days 1, 0, 1, 3, 5, 8, 11, 16, 21, 27, and 31), and used to determine virus titers and antibody concentrations. Animals were monitored daily for clinical signs (fever, posture, respiration, feces/urine, food intake, recumbence, attitude, and skin turgor) using a FITC-labeled secondary antibody (Sigma). Foci were counted using a fluorescent microscope (Carl Zeiss Microimaging LLC).

Enzyme-linked immunosorbent assay (ELISA)
The filovirus GP-based ELISA was performed as described previously [34]. Briefly, ELISA plates (Nunc MaxiSorp) were coated with purified soluble ZEBOV GP lacking transmembrane domain (100 ng/50 µl/well), followed by blocking with 3% skim milk (200 µl/well). Serial dilutions of NHP serum samples and purified antibodies (ch133, ch226, and ch61) of known concentrations were prepared, added to the ELISA plates and incubated for 1 hour at room temperature. Bound antibodies were visualized by adding a secondary peroxidase-conjugated goat anti-human IgG Fcγ fragment antibody (Jackson ImmunoResearch) and 3,3',5,5'-tetramethylbenzidine (Sigma). The addition of 1 M phosphoric acid stopped the reaction and the optical density (OD) at 450 nm was measured. Antibody concentrations in the NHP serum samples were determined based on the OD values obtained for the standard curves from purified ch133, ch226, and ch61. For ch61, A/Viet Nam/1194/2004 (H5N1) virus particles treated with 0.5% Triton X-100 were used as the ELISA antigen. ZEBOV NP-specific antibodies in the serum samples (1:10000 dilution) were detected performing ELISA using a recombinant NP antigen [35] and peroxidase-conjugated goat anti-monkey IgG γ chain antibody (Rockland).

Virus detection
Total RNA was isolated from whole blood samples using the QIamp viral Mini RNA kit (Qiagen). All quantitative real-time RT-PCRs were performed by employing the QIAquick 1-step Rotorgene kit (Qiagen) and ZEBOV-specific primers and probes based on the nucleoprotein sequence (NP; bp 2661–2721, GenBank accession number AF086833). We performed virus titration by TCID<sub>50</sub> in Vero E6 cells from the blood and selected tissue samples. Briefly, 10-fold serial dilutions of the blood and tissue homogenates were prepared and used to infect Vero E6 cells. Cells were monitored for cytopathic effects (CPE) and the TCID<sub>50</sub> was calculated for each sample employing the Reed and Muench method [36].

Animal ethics statement
Healthy, adult rhesus macaques (Macaca mulatta) were handled in the Rocky Mountain Laboratories (RML) Animal BSL-2 and BSL-4 containment space. Research was conducted in compliance with the Animal Welfare Act and other federal statutes and regulations relating to animals and experiments involving animals, and adhered principles stated in the Guide for the Care and Use of Laboratory Animals, National Research Council, 1996. The facility where this research was conducted (RML) is fully accredited by the Association for the Assessment and Accreditation of Laboratory Animal Care International and has an approved OLAW Assurance #A4149-01. Research was conducted under a protocol approved by the IACUC. All steps were taken to ameliorate the welfare and to avoid the suffering of the animals in accordance with the “Weatherall report for the use of non-human primates” recommendations. Animals were housed in adjoining individual primate cages allowing social interactions, under controlled conditions of humidity, temperature and light (12-hour light/12-hour dark cycles). Food and water were available ad libitum. Animals were monitored (pre- and post-infection) and fed commercial monkey chow, treats and fruit twice daily by trained personnel. Environmental enrichment consisted of commercial toys. All procedures were conducted by trained personnel under the supervision of veterinarians and all invasive clinical procedures were performed while animals were anesthetized. Early endpoint criteria, as specified by the IACUC approved...
score parameters, were used to determine when animals should be humanely euthanized.

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Author Contributions

Conceived and designed the experiments: AT HF. Performed the experiments: AM RY HM. Ishijima MY VM MK FF DB HF AT. Analyzed the data: AM RY M. Igarashi EN AT. Contributed reagents/materials/analysis tools: YS M. Igarashi MS. Wrote the paper: AM RY EN HF AT.

References