<table>
<thead>
<tr>
<th>Title</th>
<th>The relationship between antral follicle count in a bovine ovary and developmental competence of in vitro-grown oocytes derived from early antral follicles</th>
</tr>
</thead>
<tbody>
<tr>
<td>Author(s)</td>
<td>Nagai, Katsuhisa; Yanagawa, Yojiro; Katagiri, Seiji; Nagano, Masashi</td>
</tr>
<tr>
<td>Citation</td>
<td>Biomedical Research, 37(1): 63-71</td>
</tr>
<tr>
<td>Issue Date</td>
<td>2016-02-25</td>
</tr>
<tr>
<td>Doc URL</td>
<td><a href="http://hdl.handle.net/2115/60734">http://hdl.handle.net/2115/60734</a></td>
</tr>
<tr>
<td>Type</td>
<td>article</td>
</tr>
</tbody>
</table>
The relationship between antral follicle count in a bovine ovary and developmental competence of in vitro-grown oocytes derived from early antral follicles

Katsuhisa Nagai, Yojiro Yanagawa, Seiji Katagiri, and Masashi Nagano

Laboratory of Theriogenology, Department of Veterinary Clinical Sciences, Graduate School of Veterinary Medicine, Hokkaido University, Sapporo 060-0818, Japan

(Received 4 December 2015; and accepted 19 December 2015)

ABSTRACT

To clarify the relationship between ovarian reserve and the developmental competence of bovine oocytes, oocyte-granulosa complexes (OGCs) collected from early antral follicles (≤1 mm) in ovaries with high (≥25) and low (<25) antral follicle counts (AFCs) were used. OGCs derived from different AFC groups were cultured for growth followed by maturation, fertilization and blastocyst formation. Viability of OGCs during growth culture was similar between groups; however, OGCs in the high-AFC group had a larger number of granulosa cells than the low-AFC group at 12 days of growth. The proportion of matured oocytes in the high-AFC group was higher than that in the low-AFC group. Mitochondrial activity of oocytes before maturation in the high-AFC group was higher than that in the low-AFC group; however, accumulation of reactive oxygen species was similar between groups. Cleavage rate in the high-AFC group tended to be higher than that in the low-AFC group, although blastocyst development was similar between groups. In conclusion, oocytes derived from ovaries with high AFC have higher maturational ability and fertilizability than those from low AFC. The difference may be caused by high proliferation of granulosa cells from ovaries with high AFC.

Major functions of ovaries are producing fertilizable oocytes having developmental competence that result in successive conception and secreting sex steroid hormones that induce the estrous cycle and sustain pregnancy. Ovarian reserve is defined as potential ability of these functions (38), and is proposed as a factor of the fertility for mammalian female animals including humans (6) and cattle (22). The number of small antral follicles (antral follicle count; AFC) in a pair of ovaries detected by ultrasonography is considered as an indicator of ovarian reserve (7), because it correlates with the number of primordial follicles (23) and is stable in individual animals (1, 10).

During the long lifetime of women, ovarian reserve decreases as aging. Therefore, the relationship between oocyte quality and ovarian reserve in aged women is well investigated, and it is reported that aged women with small ovarian reserve had low quality oocytes (12, 32). In addition, the time of onset of menopause is closely correlated with ovarian reserve; small reserve resulted in earlier menopause (11). The size of the ovarian reserve varies between individuals of the same species or strain, but the relationship between oocyte quality and small ovarian reserve of young women is not known. Therefore, we need to investigate the relationship between oocyte quality and ovarian reserve of young animals.

It is reported that dairy cattle can be used as a novel experimental model for women reproduction; because cattle are single-ovulating species that have
two or three well characterized FSH-induced waves
of growth and atresia of antral follicles during their
reproductive cycles (16) similar to women (2). Cows
with low AFC, estimating small ovarian reserve, tend
to have low fertility; for example, a long open peri-
dium (27), low steroidogenic capability (25) and poor
responsiveness to superstimulatory treatments (21)
even though they were in the same age class. In the
previous study (28), we used the high- and low-
AFC cows (6.9 ± 1.8 and 6.6 ± 2.6 years old, respec-
tively) and showed that oocytes collected by ovum
pick-up from the high-AFC cows had higher fertil-
izability than those from the low-AFC cows at 3 to
4 days interval of ovum pick-up. However, only in
the high-AFC cows, the quality of oocytes was im-
paired by extending interval of ovum pick-up to 7
days, possibly due to the earlier degradation of fol-
licles compared to the low-AFC cows. From these
results we have speculated that the growth rate of
follicles is different between the high- and low-AFC
cows, and the difference affects on the quality of
oocytes.

Dairy cows are commonly culled before ending
their fertile life, and we can collect their ovaries at
an abattoir. Ministry of Agriculture, Forestry and
Fisheries in Japan reported that mean parity of culled
Holstein cows was 3.5 in 2012. From the mean par-
ity, the age at culling was estimated around 7 years
old. It means that we can investigate the relation-
ship between oocyte quality and ovarian reserve of
young animals. However, it is difficult to use the
conventional in vitro maturation, fertilization and
culture system for clarifying the relationship be-
 tween ovarian reserve and oocyte quality. Because
oocytes derived from antral follicles of 2–8 mm in
diameter, which are commonly used in the conven-
tional culture system, are already at various stage of
degradation (14, 30) and have various maturational
and developmental competences depending on their
degradation stages (13, 26, 29). For investigating
oocyte quality correctly, we should use oocytes de-
 rived from follicles before degradation. Recently, we
reported that in vitro-grown bovine oocytes derived
from early antral follicles, just before recruitment to
follicular waves (0.5–1 mm in diameter), achieved
good development to blastocysts corresponding to
the development of in vitro-grown oocytes (19, 20).

In the present study, to investigate the relation-
ship between ovarian reserve estimated with AFC
and oocyte quality, we cultured bovine oocyte-gran-
ulosa complexes (OGCs) collected from early antral
follicles in ovaries with high and low AFCs. During
culture for growth, we examined viabilities of OGCs
and the proliferation of granulosa cells. Nuclear mat-
uration, mitochondrial activity and the accumulation
of reactive oxygen species (ROS) in oocytes after
growth culture were also evaluated. Moreover, the
embryonic development of oocytes derived from dif-
f erent AFC ovaries were investigated after in vitro
fertilization.

MATERIALS AND METHODS

Chemicals. All the chemicals used for this study
were purchased from Sigma-Aldrich (St. Louis, MO,
USA), unless otherwise stated.

Collection of early antral follicles and growth cul-
ture of oocytes. Ovaries from Holstein cows obtained
at a local abattoir were kept at 20°C and were trans-
ported to the laboratory within 6 to 10 h after collec-
tion. After 3 washes in physiological saline, sliced
ovarian cortex tissues (<1 mm in thickness) were
dissected from the sliced ovarian tissues using a surgical blade (No. 20). OGCs were
isolated from follicles using a pair of fine forceps
and those with normal appearance were individually
cultured in 96-well culture plates (Falcon 353872;
Becton Dickinson, Franklin Lakes, NJ, USA) with
a growth medium for 12 days at 39°C in humidified air with 5% CO2. The growth medium
consisted of HEPES-buffered TCM-199 (Invitrogen, Grand Island, NY, USA)
supplemented with 0.1% polyvinyl alcohol, 25 mM
HEPES, 10 mM sodium bicarbonate, and 50 mg/mL
gentamicin sulfate (isolation medium, pH 7.4, at
37°C), as described by Huang et al. (19). Under a
stereomicroscope, early antral follicles (0.5–1.0 mm
in diameter) were dissected from the sliced ovarian
tissues using a surgical blade (No. 20). OGCs were
isolated from follicles using a pair of fine forceps
and those with normal appearance were individually
cultured in 96-well culture plates (Falcon 353872;
Becton Dickinson, Franklin Lakes, NJ, USA) with
200 μL of the growth medium for 12 days at 39°C
in humidified air with 5% CO2. The growth medium
consisted of HEPES-buffered TCM-199 (Invitrogen)
supplemented with 0.91 mM sodium pyruvate, 1 μg/
ML estradiol-17β (E2), 5% fetal calf serum (FCS;
Invitrogen), 4 mM hypoxanthine, 4% polyvinylpyr-
rrolidone (MW 360,000), and 50 μg/mL gentamicin
sulfate. At the onset of growth culture, OGCs were
photographed under an inverted microscope (CK 40;
Olympus, Tokyo, Japan) attached with a CCD cam-
era (Moticam 2000; Shimadzu Rika Corporation,
Tokyo, Japan), and viability of OGCs was deter-
dined by their morphologies as previously reported
(19). During growth culture, half (100 μL) of the
growth medium was replaced with the same amount
of fresh medium every 4 days.

Evaluation of granulosa cell proliferation during
growth culture. The OGCs with morphologically
normal appearance at 8 and 12 days of growth cul-
tured in a well was measured by a cell counter (Luna FL Dual Fluorescence Cell Counter; Logos Biosystems, Virginia, USA). Briefly, after removing cumulus-oocyte complexes and 175 μL of culture media from a well, 100 μL of Dulbecco’s phosphate buffered saline without calcium and magnesium (DPBS(−); Nissui Pharmaceutical, Tokyo, Japan) supplemented with 0.125% trypsin and 0.05% EDTA was added to a well. After 10 min of trypsinization and pipetting several times, 25 μL of FCS was added to stop the digestion. Then 18 μL of cell suspensions was mixed with 2 μL of acridine orange/propidium iodide cell viability kit (F23001; Logos Biosystems), and the numbers of live and dead granulosa cells were counted.

**Prematurational and maturational culture of in vitro-grown oocytes.** Before maturational culture, the cumulus-oocyte complexes recovered from morphologically normal OGCs were cultured individually in a well in microwell plates (Mini Trays 163118; NUNC, Roskilde, Denmark) that contained 6 mL of prematuration medium as described previously (20). The prematuration medium consisted of HEPES buffered TCM-199 supplemented with 0.2 mM sodium pyruvate, 2 × 10−6 units/mL FSH (from porcine pituitary), 0.5 mM 3-isobutyl-1-methylxanthine (IBMX), 1 μg/mL E2, 10% FCS, and 50 μg/mL gentamicin sulfate at 39°C in humidified air with 5% CO2 for 10 h. Some of cumulus-oocyte complexes were subsequently cultured for another 22 h at 39°C in humidified air with 5% CO2 for maturation. The maturational medium consisted of HEPES buffered TCM-199 supplemented with 0.2 mM sodium pyruvate, 0.02 units/mL FSH, 1 μg/mL E2, 10% FCS, and 50 μg/mL gentamicin sulfate, and each cumulus-oocyte complex was cultured individually in a well in microwell plates (31).

**Evaluation of oocyte nuclear status before and after maturation.** Before maturational culture, the cumulus-oocyte complexes before and after maturational culture were denuded from cumulus cells by vortexing. Denuded oocytes were incubated for 15 min in the dark at 37°C in DPBS(−) supplemented with 10 μM 2′,7’-dichlorodihydrofluorescein diacetate (DCHFDA), 1 μg/mL Hoechst 33342 and 10% FCS. Oocytes at germinal vesicle stage before maturational culture and oocytes progressed to metaphase II stage after maturational culture were subjected to the evaluation of ROS accumulation. Membrane potentials of mitochondria in oocytes were calculated as the ratio of fluorescent intensity of activated mitochondria, expressed as red fluorescence of JC-1 staining, to less activated mitochondria, expressed as green fluorescence of JC-1 staining (Δψm, red/green fluorescent intensity).

Oocytes before and after maturational culture were denuded from cumulus cells by vortexing. Denuded oocytes were incubated for 15 min in the dark at 37°C in DPBS(−) supplemented with 10 μM 2′,7’-dichlorodihydrofluorescein diacetate (DCHFDA), 1 μg/mL Hoechst 33342 and 10% FCS. Oocytes at germinal vesicle stage before maturational culture and at metaphase II stage after maturational culture were subjected to the evaluation of ROS accumulation, and images of them were acquired using a digital fluorescent microscope. The mean fluorescent intensity of the images was calculated using analysis software, and ROS accumulation in oocytes was defined as the mean fluorescent intensity of oocytes.

**In vitro fertilization and culture of oocytes.** After maturational culture, some oocytes were co-cultured with spermatozoa (5 × 10³ cells/mL) in 100-μL droplets (5–13 oocytes/droplet) of fertilization medium covered with mineral oil in a humidified atmosphere at 39°C with 5% CO2, 5% O2, and 90% N2 for 18 h (29). Fertilization medium was modified Brackett and Oliphant isotonic medium (5) supplemented

---

**In vitro fertilization and culture of oocytes.** After maturational culture, some oocytes were co-cultured with spermatozoa (5 × 10³ cells/mL) in 100-μL droplets (5–13 oocytes/droplet) of fertilization medium covered with mineral oil in a humidified atmosphere at 39°C with 5% CO2, 5% O2, and 90% N2 for 18 h (29). Fertilization medium was modified Brackett and Oliphant isotonic medium (5) supplemented
with 2.5 mM theophylline and 3 mg/mL fatty acid-free bovine serum albumin (BSA). After the thawing of frozen semen from a Holstein bull, motile sperm were separated using a Percoll (GE Healthcare, Pittsburgh, PA, USA) gradient (45% and 90%). Then inseminated oocytes (presumptive zygotes) were cultured as previously described (37). Briefly, after co-incubation with spermatozoa, presumptive zygotes were freed from cumulus cells by vortexing and washing three times in culture medium. Cumulus-free zygotes were cultured in 30-μL droplets (5–26 zygotes/droplet) of culture medium covered with mineral oil in a humidified atmosphere at 39°C with 5% CO₂, 5% O₂, and 90% N₂ for 6 days. The culture medium consisted of modified synthetic oviduct fluid containing 1 mM glutamine, 12 essential amino acids for basal medium Eagle, 7 nonessential amino acids for minimum essential medium, 10 mg/mL insulin, and 5 mM glycine, 5 mM taurine, 1 mM glucose, and 3 mg/mL fatty acid-free BSA. Cleavage and blastocyst rates were determined after 2 days (approximately 30 h) and 6 days (approximately 150 h) of culture, respectively. Total cell numbers in a blastocyst obtained after 6 days of culture were counted using an air-drying method (36).

Experimental group. AFC in an ovary was determined by the number of antral follicles of ≥2 mm in diameter. Usually, AFC means the number of follicle in pair of ovaries, but we could not collect the pair of ovaries from a cow at the abattoir. For adapting the criteria of AFC (high: ≥25, intermediate: 16-24, and low: ≤15) described by Ireland et al. (22), we compared the AFC in an ovary detected by ultrasonography to the AFC detected by the direct inspection. In the preliminary study, the number of follicles in an ovary detected by direct inspection (15.5 ± 2.4, n = 4) after ovariectomizing was approximately double compared to the number of follicles detected by ultrasonography (7.0 ± 1.4, n = 4). Cushman et al. (10) reported positive correlation between the number of antral follicles in one ovary and that in the contralateral ovary. Therefore, we allocated the ovary having more than 25 follicles of ≥2 mm in diameter as the high-AFC group and others as the low-AFC group. Then 189 and 342 ovaries were used as the high- and low-AFC groups, respectively.

Experimental design. During OGC collection, the number of early antral follicles and the number of collected OGCs in the high- and low-AFC groups were counted. OGCs having normal appearance were also counted and submitted to growth culture. The viability of cultured OGCs was examined every 4 days. In the present study, some of OGCs were used for the evaluation of proliferation of granulosa cells, and nuclear maturation, mitochondrial activity and ROS accumulation of oocytes.

Statistical analysis. The numbers of follicles and OGCs in high- and low-AFC ovaries were compared by Student’s t-test. The viability of OGCs during growth culture was compared by Tukey-Kramer’s HSD. The number of granulosa cells at 8 and 12 days of growth culture, mitochondrial activity and ROS accumulation in oocytes were compared by Student’s t-test. The data of nuclear status was analyzed by chi-square test. Percentage data of cleavage and blastocyst rates were subjected to an arcsine square-root transformation, and analyzed by Student’s t-test. Cell numbers in blastocysts were also analyzed by Student’s t-test. All analyses were performed using software (JMP Pro 10.6; SAS Institute Inc., Cary, NC, USA). Values were considered significantly different at P < 0.05.

RESULTS

As shown in Table 1, the mean number of early antral follicles dissected from one ovary in the high-AFC group was larger than that from the low-AFC groups (P < 0.01). The mean numbers of OGCs collected and OGCs having normal appearance in the high-group were larger than those in the low-AFC group. However, the proportion of OGCs having normal appearance based on total OGCs was similar between the high- and low-AFC groups (66.0 ± 0.1% and 67.4 ± 0.2%, respectively).

As shown in Fig. 1, the proportions of live OGCs in both groups were similar and decreased until 8 days of growth culture. As shown in Table 2, there was no difference in the mean number of granulosa cells in a well between the high- and low-AFC groups at 8 days of growth culture; however, the proportion of live cells in the high-AFC group was higher than that in the low-AFC group (P < 0.05). Numbers of granulosa cells in both groups increased from 8 to 12 days of growth culture. Although the high-AFC group had larger number of cells than the low-AFC group at 12 days of growth culture (P < 0.05), the proportions of live cells were similar between groups.

As shown in Table 3, the proportions of oocytes at germinal vesicle stage before maturational culture were similar between the high- and low-AFC groups. However, the proportion of oocytes at metaphase II
Relationship between ovarian reserve and oocyte quality 67

stage after maturational culture in the high-AFC group was higher than that in the low-AFC group ($P < 0.05$).

As shown in Fig. 2, before maturational culture (at germinal vesicle stage), the mitochondrial activity of oocytes in the high-AFC group was higher than that in the low-AFC group ($P < 0.05$). After maturational culture (at metaphase II stage), the mitochondrial activities of oocytes in both groups increased compared to those before maturational culture ($P < 0.05$) and were similar between the high- and low-AFC groups. As shown in Fig. 3, before and after maturational culture, ROS accumulations of oocytes in both AFC groups were similar; however, ROS accumulation decreased after maturational culture compared to before maturational culture in both groups ($P < 0.05$).

The results of embryonic development of oocytes were described in Table 4. Cleavage rate in the high-AFC group tended to be higher than that in the low-AFC group ($P = 0.09$). There were no significant differences in both blastocyst rates based on inseminated and cleaved oocytes, although values of standard deviation were large. The numbers of cells in blastocysts in both groups were similar between groups.

**DISCUSSION**

In the present study, the high-AFC group had larger number of early antral follicles ($\leq 1$ mm) in an ovary than the low-AFC group, and the proportion of oocytes having normal appearance was similar between groups. Our results correspond to the previous report that AFC in an ovary was positively correlated to number of follicles at various stages, and the proportion of healthy follicles was similar between the high- and low-AFC groups (23). However, the number of granulosa cells at 12 days of growth culture in the high-AFC group was larger than those in the low-AFC group. The high viability of granulosa cells at 8 days of growth culture in the high-AFC group might contribute to larger number of granulosa cells at 12 days of growth culture. These results seem to indicate that the function of granulosa cells is different depending on their origin of ovaries with different AFC. In the present study, oocytes in the high-AFC group also showed higher competence of nuclear maturation than that in the low-AFC group. Granulosa cells regulate the meiotic resumption of oocytes through gap junction (35). It suggests that granulosa cells derived from the high-AFC cows are able to regulate the functions of meiosis more properly. Scheetz *et al.* (34) reported that the cultured granulosa cells derived from the low-AFC cows showed lower secretion of estradiol, which is essential for granulosa cell proliferation (17). In the present study, we added the much concentration (1 μg/
However, Scheetz et al. (34) also reported that the granulosa cells derived from the low-AFC cows secreted more progesterone than that from the high-AFC cows. For developing follicles healthily, the ratio of estradiol to progesterone is very important (3). In further study, we should examine the function of mL) of estradiol to growth medium compared to that in the bovine dominant follicle of 10.5 mm in diameter (around 700 ng/mL) (3). It means that the estradiol is enough for proliferation of granulosa cells. However, Scheetz et al. (34) also reported that the granulosa cells derived from the low-AFC cows secreted more progesterone than that from the high-AFC cows. For developing follicles healthily, the ratio of estradiol to progesterone is very important (3). In further study, we should examine the function of

### Table 2 Relationship between AFC, granulosa cell proliferation and survivability during in vitro growth culture

<table>
<thead>
<tr>
<th>Duration of growth culture</th>
<th>AFC group</th>
<th>No. of oocytes (replicates)</th>
<th>Number of granulosa cells ($\times 10^3$)</th>
<th>Proportion (%) of live cells</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>total</td>
<td>live</td>
</tr>
<tr>
<td>8</td>
<td>High</td>
<td>54 (5)</td>
<td>37.0 ± 22.9</td>
<td>33.1 ± 22.4</td>
</tr>
<tr>
<td></td>
<td>Low</td>
<td>45 (5)</td>
<td>32.5 ± 23.7</td>
<td>28.0 ± 23.7</td>
</tr>
<tr>
<td>12</td>
<td>High</td>
<td>78 (9)</td>
<td>54.5 ± 32.1$^a$</td>
<td>39.3 ± 32.3$^a$</td>
</tr>
<tr>
<td></td>
<td>Low</td>
<td>72 (9)</td>
<td>37.9 ± 20.7$^b$</td>
<td>24.9 ± 16.0$^b$</td>
</tr>
</tbody>
</table>

Values are mean ± SD.

$^a, b$ Different superscripts indicate a significant difference ($P < 0.05$).

### Table 3 Nuclear status of oocytes derived from ovaries with high and low AFCs before and after in vitro maturation

<table>
<thead>
<tr>
<th>In vitro maturation</th>
<th>AFC group</th>
<th>No. of oocytes (replicates)</th>
<th>Proportion (%) of oocytes at</th>
<th>GV</th>
<th>M I</th>
<th>A I / T I</th>
<th>M II</th>
</tr>
</thead>
<tbody>
<tr>
<td>Before</td>
<td>High</td>
<td>70 (5)</td>
<td>90.0</td>
<td>8.6</td>
<td>2.3</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Low</td>
<td>65 (5)</td>
<td>83.1</td>
<td>17.0</td>
<td>0</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>After</td>
<td>High</td>
<td>157 (9)</td>
<td>0</td>
<td>15.9</td>
<td>1.9</td>
<td>82.1$^*$</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Low</td>
<td>118 (9)</td>
<td>0</td>
<td>22.9</td>
<td>7.6</td>
<td>69.5$^b$</td>
<td></td>
</tr>
</tbody>
</table>

GV: germinal vesicle, M I: metaphase I, A I / T I: anaphase I or telophase I, M II: metaphase II.

$^a, b$ Different superscripts indicate a significant difference ($P < 0.05$).
steroidogenesis of granulosa cells derived from different AFC cows. Another possibility of the cause of difference in the proliferation of granulosa cells is the difference of oocyte function between groups. Differentiation and proliferation of granulosa cells are regulated by oocyte secreted factors (OSFs) such as GDF-9 or BMP-15 (15). Therefore, the secretion of OSFs from oocytes may be different between the high- and low-AFC groups. In the previous study, Scheetz et al. (34) cultured only granulosa cells and examined the function of granulosa cells. For the correct evaluation of granulosa cell function of different AFC groups, the expression levels of OSFs in oocytes should be examined by co-culture with oocyte and granulosa cells.

Also the mitochondrial activity in oocytes before maturational culture was higher in the high-AFC group than in low-AFC group. Mitochondrial activity is one of indicators of cytoplasmic maturation of oocytes (4). Huang et al. (19, 20) reported that in vitro-grown oocytes acquired high mitochondrial activity after 10-h prematurational culture, and showed higher maturation and development competences compared to oocytes without prematurational culture. Jeseta et al. (24) reported that the kinetics of mitochondrial activity and adenosine triphosphate (ATP) production during maturational culture were influenced by the atretic grade of granulosa cells. This indicates the importance of the correlation between oocyte and granulosa cells. Low mitochondrial activity of oocytes may be affected by lower number and also lower function of granulosa cells in the low-AFC group. In the ROS accumulation, there was no difference between the high- and low-AFC groups by the measurement of mean fluorescent intensity in ooplasm; nevertheless there was difference in the mitochondrial activity before maturational culture. ROS in ooplasm is considered to be a byproduct of electron transport chain activity in mitochondria when producing ATP (33). Our results suggest that the activity of anti-oxidant enzyme in the high-AFC group is higher than that in the low-AFC group. Expression patterns of mRNA coding anti-oxidant enzymes were reported to change in according to days from follicular emergence (39), follicular size and their status (9) in cattle. It was also reported that mRNA expression level of glutathione peroxidase between dominant and subordinate follicles was different and that glutathione peroxidase could prevent cell apoptosis from oxidative stress during high steroidogenic period (18). Thus, evaluation of anti-oxidant enzyme expression in oocytes and granulosa cells derived from different AFC cows is necessary.

In conclusion, oocyte-granulosa complexes derived from early antral follicles in ovaries with high AFC have higher maturational ability, and it is assumed

<table>
<thead>
<tr>
<th>AFC group</th>
<th>No. of oocytes (replicates)</th>
<th>Cleavage rate (n)</th>
<th>Blastocyst rate (%) based on inseminated oocytes</th>
<th>Cell number in blastocysts (n)</th>
</tr>
</thead>
<tbody>
<tr>
<td>High</td>
<td>127 (9)</td>
<td>65.3 ± 11.7</td>
<td>11.4 ± 10.0</td>
<td>19.9 ± 20.3</td>
</tr>
<tr>
<td>Low</td>
<td>80 (8)</td>
<td>45.0 ± 30.6</td>
<td>8.4 ± 15.9</td>
<td>13.7 ± 26.1</td>
</tr>
</tbody>
</table>

Values are mean ± SD.
to have higher fertilizability than those from ovaries with low AFC. Higher proliferation of granulosa cells during growth culture and higher mitochondrial activity in ooplasm before maturational culture may support both nuclear and cytoplasmic maturation of in vitro-grown oocytes from ovaries with high AFC. In further study, we need to investigate factors and mechanisms of proliferation of granulosa cells and activation of mitochondria in oocytes derived from different AFC ovaries.

Acknowledgements

This study was supported by a Grant-in-Aid for Scientific Research from the Japan Society for the Promotion of Science to M. Nagano (No. 25450441).

REFERENCES


