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Rapid communication

Requirement for nuclear autoantigenic sperm protein mRNA expression in bovine preimplantation development

Running title: NASP gene knockdown in bovine embryos

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ABSTRACT

Nuclear autoantigenic sperm protein, NASP, is associated with DNA replication, cell proliferation, and cell cycle progression through its specific binding to histones. The aim of this study was to examine the roles of NASP in bovine preimplantation embryonic development.

Using NASP gene knockdown (KD), we confirmed the reduction of NASP mRNA expression during preimplantation development. NASP KD did not affect cleavage but significantly decreased development of embryos into the blastocyst stage. Furthermore, blastocyst hatching was significantly decreased in NASP KD embryos. Cell numbers in the inner cell mass of NASP KD blastocysts were also decreased compared to those of controls. These results suggest that NASP mRNA expression is required for preimplantation development into the blastocyst stage in cattle.

Key words: bovine embryo, NASP, knockdown.
INTRODUCTION

Following syngamy that is fusion of the male and female pronuclei after in vitro fertilization (IVF), the single-cell embryo, or zygote, undergoes a sequence of mitotic divisions. The process of cellular division without any accompanying increase in the volume of the cytoplasm is called cleavage, and the small cells produced by the series of early cleavage divisions are blastomeres. In mammals including cattle, preimplantation embryos are able to develop in culture media, and some can develop into the blastocyst stage. In cattle, when attempting the transfer of embryos produced by IVF to the uterus of a recipient cow, it is essential to have an in vitro culture (IVC) system for the effective production of blastocysts.

The cell number of a single bovine embryo drastically increases until the blastocyst stage during 8 d of IVC. Therefore, cleavage represents many rounds of mitosis with DNA replication. To assemble newly replicated DNA into chromatin, compensating active histone mRNA/protein synthesis occurs during S phase of the cell cycle, resulting in sufficient histone production. The nucleosome is assembled by wrapping a 146-bp DNA segment around a histone octamer that consists of two each of the core histones, H2A, H2B, H3, and H4. Repeating nucleosome units constitute chromatin. The transport of histones from the cytoplasm to the nucleus and their subsequent assembly into nucleosomes is mediated by a set of proteins that includes histone chaperones. Nuclear autoantigenic sperm protein, NASP, is one of these histone chaperones; it protects the H3–H4 tetramer from degradation in human cells (Cook et al. 2011), and transports linker histone H1, which is essential for normal development, into nuclei (Bustin et al. 2005; Alekseev et al. 2005).

NASP is associated with DNA replication, cell proliferation, and cell cycle progression, specifically binding to histones H1, H3, and H4, and affecting chromatin assembly (Finn et al. 2012). NASP was first characterized in rabbit testes, and is conserved in a wide range of species (Welch &
O’Rand 1990; Finn et al. 2012). NASP is involved in chromatin remodeling, which maintains the nucleosome spacing that is critical for the regulation of gene expression and for normal development. The essential role of NASP in early murine embryonic development is underscored by the report that NASP-/- null mice showed lethality at embryonic day 5.5 with implantation failure (Richardson et al. 2006).

In a previously reported study, we addressed the site-predominant expression of the NASP mRNA in the inner cell mass (ICM) of in vitro-derived blastocysts (Nagatomo et al. 2013). Little is known, however, as to whether NASP plays roles in preimplantation development in cattle. To elucidate the role of NASP during bovine preimplantation development, we used knockdown (KD) of NASP gene expression and evaluated the ability of NASP KD embryos to develop into the blastocyst stage.
MATERIALS AND METHODS

Preparation of bovine embryos by in vitro fertilization

Bovine oocyte retrieval, in vitro oocyte maturation, fertilization, and subsequent in vitro bovine embryo culture were performed according to the methods described in a previous paper (Nagatomo et al. 2013). Briefly, cumulus–oocyte complexes (COCs) collected from slaughterhouse-derived ovaries were matured by culturing in TCM-199 medium (Gibco, Grand Island, NY, USA) at 38.5°C in a humidified atmosphere of 5% CO₂ in air for 20–22 h. In vitro-matured oocytes were transferred to Brackett and Oliphant (BO) medium (Brackett & Oliphant 1975) containing 2.5 mM theophylline (Wako Pure Chemical Industries, Ltd., Osaka, Japan). Subsequently, frozen-thawed semen was centrifuged at 600 × g for 8 min in BO medium, and the spermatozoa were added to the COCs at a final concentration of 5 × 10⁶ cells/mL. After 18 h of incubation, presumptive zygotes were denuded and cultured in mSOFaa medium (Aono et al. 2013) at 38.5°C in a humidified atmosphere of 5% CO₂ in air for 8 d.

Microinjection of N4SP short hairpin RNA (shRNA) expression vectors

An RNA interference hairpin, with antisense/sense regions, a 12-bp stem loop (GCTTCCTGTCAC), and a 6-bp terminator element (TTTTTT), was designed to target nucleotides 878-898 of the N4SP mRNA (GenBank accession number NM_001038088.2). The sense- and antisense-strand oligonucleotides (Sigma-Aldrich Japan, Tokyo, Japan) were denatured at 95°C for 5 min, and then gradually cooled to 25°C for annealing. The double-strand DNA was ligated downstream of the U6 promoter in the pBAsi/mU6 Neo vector (Stratagene, La Jolla, CA, USA). The following sequence was used: 5’- GATCCGGAACACTACAGGAGAAAGTTAAGCCTTCCTGTCACTTAAACTTTTCTCTTG TAGTTCCCTTTTTATA -3’ (underlined and double-underlined portions indicate the stem-loop region
and terminator element, respectively). The shRNA expression vector (pBAsi/mU6/NASP-I) targeting
the \textit{NASP} mRNA (\textit{NASP} shRNA) was isolated using the EndoFree Plasmid ezFlow Mini Kit
(Biомiga, Inc., San Diego, CA, USA). Twelve h after the completion of insemination, the
synthesized \textit{NASP} shRNA (10 nG/\(\mu\)L) was then injected into cytoplasm of each bovine zygote by
using a FemtoJet® injection device (Eppendorf, Hamburg, Germany). After \textit{NASP} KD, \textit{NASP}
mRNA expression was confirmed by quantitative reverse transcription-PCR using the same
procedure as reported previously (Nagatomo \textit{et al.} 2013). The zygotes injected with \textit{NASP} shRNA
were cultured, to examine the effect of \textit{NASP} KD on subsequent embryonic development into the
blastocyst stage. Zygotes injected with the pBAsi/mU6 Neo vector without the \textit{NASP} shRNA insert
were used as a control in these experiments.

\textbf{Assessment of development to the blastocyst stage and the competence of hatching from the
zona pellucida}

 Rates of cleavage and blastocyst formation were assessed on day 2 (D2) and day 8 (D8) of IVC,
respectively. Embryos that developed into the blastocyst stage on D8 were allotted to experiments of
differential staining or cultured for a further 24 h to assess the number of blastocysts that showed
hatching from the zona pellucida.

\textbf{Differential staining of blastocysts}

 The differential staining of the ICM and trophectoderm (TE) within a blastocyst was performed
according to the method of Thouas \textit{et al.} (Thouas \textit{et al.} 2001). Briefly, the blastocysts were stained
with propidium iodide (Life Technologies, Carlsbad, CA, USA) as a 0.1 mg/mL solution in
phosphate-buffered saline containing 0.2\% (v/v) Triton X-100 at room temperature for
approximately 2 min, followed by staining with 25 \(\mu\)g/mL bisbenzimide (Sigma-Aldrich) in 99.5\%
ethanol at 4°C for 3 h. After washing with glycerol, the blastocysts were mounted on a glass slide, and observed with a fluorescence microscope (EVOS® Cell Imaging System, Advanced Microscopy Group, Mill Creek, WA, USA). The ICM and TE nuclei were differentially stained blue and pink, respectively.

Statistical analysis

Statistical analyses of all data were carried out by one-way analysis of variance and Fisher’s post-hoc least significant difference test using the Statview statistical-analysis software (Abacus Concepts, Inc., Berkeley, CA, USA). A value of $P < 0.05$ was considered significant.
RESULTS

Knockdown of NASP mRNA expression in bovine preimplantation embryos

To examine the role of NASP in early bovine embryogenesis, we first performed an RNA interference experiment by microinjecting NASP shRNA expression vectors into in vitro-derived zygotes. The relative expression level of the NASP mRNA was significantly decreased in NASP KD embryos during preimplantation development (at the 2–4-cell and blastocyst stages; \( P < 0.05 \)) (Fig. 1). Although there were no significant differences at the 8–16-cell and morula stages, the NASP mRNA tended to be decreased. During these stages, the NASP mRNA expression might be low in bovine embryos. The NASP KD embryos showed a lower rate of blastocyst formation compared to controls (43.2 ± 5.8% vs 28.4 ± 7.2%; \( P < 0.05 \)) (Table 1). However, KD of NASP did not affect the cleavage rate. Taken together, these results clearly show the importance of NASP mRNA expression for embryonic survival during preimplantation bovine embryonic development.

Effects of NASP knockdown on ICM cell number and subsequent hatching of blastocysts

Because we previously observed ICM-predominant expression of the NASP mRNA (Nagatomo et al. 2013), we further investigated the ratio of ICM cell number to total cell number in NASP KD blastocysts by the detergent procedure for differential staining of ICM and TE cells (Thouas et al. 2001). The proportion of ICM cells in blastocysts was significantly decreased in the NASP KD compared to the control (Fig. 2A and B). Next, we investigated the number of blastocysts showing hatching from the zona pellucida in NASP KD blastocysts. At 24 h after IVC for 8 d, the ratio of blastocysts showing hatching to total blastocysts was significantly decreased in the NASP KD compared to the control (Table 1). These findings clearly demonstrate the requirement of NASP gene expression for the normal formation of blastocysts in cattle.
DISCUSSION

The distinctiveness of bovine embryos is reflected in the lack of a precedent for the successful establishment of embryonic stem (ES) cells, trophoblast stem cells, and even induced pluripotent stem (iPS) cells, which might be due to insufficient genetic information on reliable marker genes in the ICM and TE cell lineages in cattle. We established a technique to determine which genes are expressed in a lineage-specific manner in the ICM and TE of bovine blastocysts, and reported 12 genes with novel site-predominant expression in bovine blastocysts. However, the biological functions of each gene have remained unknown (Nagatomo et al. 2013).

NASP was one of the genes showing ICM-predominant expression in bovine blastocysts, and encodes a histone chaperone protein that plays essential roles in mitosis. The mammalian NASP gene has a high degree of sequence similarity to a NASP homologue in Xenopus laevis, called N1/N2 (O'Rand et al. 1992). A significant relationship between NASP and carcinogenesis has been indicated in a variety of cancers, such as renal cell carcinoma, ovarian cancer, and prostate cancer (Alekseev et al. 2011; Chatterjee et al. 2006; Fang et al. 2015). In the cancer model, it is assumed that an alteration of NASP levels changes the amount of histone H3/H4 within a cell, resulting in impaired genome integrity, chromatin stability, and replication error (Cook et al. 2011); eventually, cells that show an abnormal level of NASP would be transformed into tumor cells.

Interestingly, pluripotent cells such as ES cells and iPS cells share a number of characteristics with cancer cells, including self-renewal, proliferation, and expression of stem-cell markers (Grskovic & Ramalho-Santos 2008). In the present study, the stable KD of NASP mRNA expression during bovine preimplantation development caused not only a decrease in blastocyst formation but also a reduction of ICM cell proportion. These results are consistent with the previous observation in murine research that blastocysts from the NASP-/- null mouse completely failed to
proliferate after outgrowth on a culture dish (Richardson et al. 2006). We previously found that the
NASP mRNA was predominantly expressed in the ICM of bovine blastocysts (Nagatomo et al. 2013),
suggesting that NASP plays an essential role in the normal proliferation of the ICM in bovine
blastocysts. Furthermore, the present study revealed that blastocyst hatching from the zona pellucida
was inhibited in NASP KD embryos. This might be due to the depletion of histones H3 and H4 being
required for acute DNA replication during embryo cleavage in NASP KD embryos. In fact, NASP
expression is high in ES cells and low in differentiated cells (Sun et al. 2008; Yocum et al. 2008).
Further studies are required to elucidate the function of NASP, and possibly other histone chaperones,
in lineage specification, maintenance, proliferation, and differentiation in bovine embryos during
preimplantation development.

Conclusions

Here, we have demonstrated that KD of the NASP gene disrupts development into the
blastocyst stage and subsequent hatching from the zona pellucida in bovine embryos. These findings
show that NASP mRNA expression is essential for normal bovine embryonic development. To the
best of our knowledge, this is the first report of NASP function in early embryos of cattle.
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FIGURE LEGENDS

Figure 1 Effect of NASP gene knockdown from the 2–4-cell stage to the blastocyst stage in bovine embryos. The NASP mRNA level was assessed by quantitative reverse transcription PCR. The results are shown as the mean ± SEM (error bars) of three replicate experiments, * P < 0.05. KD (knockdown): black. Control: white.

Figure 2 Effect of NASP gene knockdown on development of the inner cell mass. (A) The ratio of inner cell mass (ICM) cell number to total cell number in control and NASP knockdown (KD) blastocysts. The results are shown as the mean ± SEM (error bars), * P < 0.05. The number of embryos analyzed was 12 for the control and 10 for the NASP KD. (B) Representative blastocysts differentially stained by both bisbenzimide and propidium iodide in control and NASP KD embryos. Arrowheads indicate the ICM. Blue: nuclei in the ICM; pink: nuclei in the TE.
Table 1. Effect of NASP gene knockdown on the in vitro development of bovine embryos

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<th>No. of embryos examined</th>
<th>No. of embryos cleaved (% ± SEM)</th>
<th>No. of blastocyst (% ± SEM)</th>
<th>No. of hatched blastocyst (% ± SEM) (/blastocyst)</th>
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<td>control</td>
<td>74</td>
<td>59 (67.6 ± 4.0)</td>
<td>32 (43.2 ± 5.8)</td>
<td>7 (21.8 ± 2.9)</td>
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<td>KD</td>
<td>95</td>
<td>67 (70.5 ± 9.2)</td>
<td>27 (28.4 ± 7.2)</td>
<td>2 (7.4 ± 3.1)</td>
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* P < 0.05
Relative expression level (%)

2-4 cell  8-16 cell  Morula  Blastocyst

Figure 1
Figure 2

A

Proportion of ICM cells to total cells (%)

control

KD

B

control

KD

Figure 2