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Efficient Synthesis of Photoreactive 2-Propoxyaniline Derivatives as Artificial Sweeteners

Yuta Murai, Takuma Yoshida, Lei Wang, Katuyoshi Masuda, Yasuyuki Hashidoko, Kenji Monde, Yasumaru Hatanaka, and Makoto Hashimoto

5-Nitro-2-propoxyaniline (1, Figure 1), named P-4000, has a sweetness intensity that is approximately 4000-times greater than that of sucrose and is well known as one of the strongest artificial sweeteners. On the other hand, it was once used as an artificial sweetener, but its use has been banned because of its possible toxicity. It is known that sweet taste is recognized by a sweet receptor, which is a heterodimeric receptor consisting of T1R2 and T1R3 subunits. However, the mechanism of sweetness and toxicity of 5-nitro-2-propoxyaniline has not yet been elucidated. Elucidation of the interactions between artificial sweeteners and their sweet receptor has attracted much interest and many researchers have studied the interactions by using NMR spectroscopy and X-ray crystallography. There have been only a few studies using homology modeling to determine the mechanism of action of 5-nitro-2-propoxyaniline for the gustatory receptor. However, satisfactory results have not been obtained, the detailed mechanism of action of 5-nitro-2-propoxyaniline for the receptor has not been elucidated. Identification of the biological mechanism as well as that of the target protein is a crucial point in the field of chemical biology. Photoaffinity labeling is one of the most powerful chemical tools to reveal interactions of bioactive compounds with biomolecules instead of X-ray crystallography and the solution-state NMR method, which are difficult to apply to a membrane protein such as the gustatory receptor. It is suitable for the analysis of biological interactions based on the affinity of biological active compounds for biomolecules. Therefore we have reported that synthesis of several photoaffinity labeling reagent for gustatory receptor.

Introduction:

5-Nitro-2-propoxyaniline (1, Figure 1), named P-4000, has a sweetness intensity that is approximately 4000-times greater than that of sucrose and is well known as one of the strongest artificial sweeteners. On the other hand, it was once used as an artificial sweetener, but its use has been banned because of its possible toxicity. It is known that sweet taste is recognized by a sweet receptor, which is a heterodimeric receptor consisting of T1R2 and T1R3 subunits. However, the mechanism of sweetness and toxicity of 5-nitro-2-propoxyaniline has not yet been elucidated. Elucidation of the interactions between artificial sweeteners and their sweet receptor has attracted much interest and many researchers have studied the interactions by using NMR spectroscopy and X-ray crystallography. There have been only a few studies using homology modeling to determine the mechanism of action of 5-nitro-2-propoxyaniline for the gustatory receptor. However, satisfactory results have not been obtained, the detailed mechanism of action of 5-nitro-2-propoxyaniline for the receptor has not been elucidated. Identification of the biological mechanism as well as that of the target protein is a crucial point in the field of chemical biology. Photoaffinity labeling is one of the most powerful chemical tools to reveal interactions of bioactive compounds with biomolecules instead of X-ray crystallography and the solution-state NMR method, which are difficult to apply to a membrane protein such as the gustatory receptor. It is suitable for the analysis of biological interactions based on the affinity of biological active compounds for biomolecules. Therefore we have reported that synthesis of several photoaffinity labeling reagent for gustatory receptor.

Abstract 5-Nitro-2-propoxyaniline is one of the strongest artificial sweeteners. However, little is known about the detailed relationship of the structure and biological activity between 5-nitro-2-propoxyaniline and its sweet receptor. Photoaffinity labeling is a useful method for revealing interactions of a small bioactive compound with molecules. Therefore, we synthesized photoreactive 2-propoxyaniline derivatives as useful tools for revealing the interactions by photoaffinity labeling.

Key words: photoaffinity labeling, chemical biology, photophore, artificial sweetener, sweet receptor

To the best of our knowledge, synthesis of photoreactive-2-propoxyaniline derivatives for photoaffinity labeling has not been reported yet. According to a previous report, even though the nitro group of 5-nitro-2-propoxyaniline was converted to photophores in this study.
selection of an appropriate photophore is important. In this paper, the synthesis of each photoreactive 2-propoxyaniline derivative with aryl azide, benzophenone and aryl diazirine and also the results of sweet-tasting effect assay for determining biological activities of photoreactive compounds are reported.

**Results and Discussion:**

The synthesis of 5-azide-2-propoxyaniline (6) started with commercially available 4-propoxybromobenzene (3) to synthesize (6) efficiently with a few steps. Because an azide group often cannot tolerate strong acidic condition or reductive condition, we decided to install an azide group in the final step. First, compound (3) was subjected to electrophilic nitration with nitric acid (fuming) in acetic anhydride to obtain compound (4) without ipso nitration. Subsequently, since H2-Pd/C reductive condition of the nitro group was simultaneously liable to reduce the bromo group, we chose sodium dithionite to selectively reduce the nitro group and afforded 5-bromo-2-propoxyaniline (5) in 78% yield. Finally, the bromo group in compound (5) was substituted with an azide group by sodium azide through CuI-catalyzed reaction to obtain compound (6).12 (Scheme 1)

We next tried to synthesize benzophenone derivative (10) from compound (4) as the common intermediate. Although the bromide of (4) was subjected to lithiation with n-BuLi followed by treatment with benzoylated reagents to produce a benzophenone moiety, the products were complicated. Consequently, we considered an alternative synthesis strategy for constructing compound (10), and it was prepared from commercially available N-(2-hydroxyphenyl)acetamide (7) according to similarly previous methods. Compound (7) was subjected to Williamson ether synthesis with 1-bromopropane to produce (8) in high yield. Subsequently, Friedel-Crafts benzoylation between compound (8) and benzoyl chloride with AlCl3 at room temperature for 3 h gave compound (9), and this was followed by deprotection of the acetyl group under an acidic condition to afford the benzophenone derivative (10) in 84% overall yield without cleaving the ether group by a three-step sequence. (Scheme 2)

For the (trifluoromethyl)phenyldiazirine moiety, compound (4) was not available for the starting material to install a trifluorocetyl group for the same reason in installing a benzophenone group. In addition, it was not possible to obtain a trifluorocetyl derivative from compound (7) or (8) by Friedel-Crafts acylation. Therefore, we selected alternative starting material (11), and it was subjected to demethylation by lithium chloride under a reflux condition to give (12), and this was followed by Williamson ether synthesis to produce (15). Compound (15) was able to be prepared in another way; commercially available 4-propoxybenzaldehyde was treated with CF3-TMS13 and worked up with 1 M HCl to obtain (14) in good yield. Compound (14) was oxidized with Dess-Martin periodinane under an acidic condition in excellent yield. (Scheme 3)

Nitrification of compound (15) with fuming HNO3 in acetic anhydride at 0 °C proceeded selectively (16). The trifluorocetyl moiety was converted to the (trifluoromethyl)diazirinyl moiety (20) using a general method. However, the diazirinyl moiety could not tolerate the condition in which the nitro group was reduced by Na2S2O4, and we therefore previously had the compound (19) reduced without breaking the diazirinyl moiety and obtained (21) in moderate yield. Subsequently, the diazirinyl moiety was converted to diazirinyl moiety (22)12 with activated MnO2 under a mild condition. (Scheme 4)

The activities of compound (6) and (22) were measured by a cell-based assay1, which determines the changes of intracellular calcium ion levels in the human sweet taste receptor hT1R2-hT1R3-expressing HEK293T cells by the fluorescent calcium indicator [fura-
2 acetoxyethyl ester] as described by Imada et al. Both the photoreactive 2-propoxyaniline, (6) and (22), exhibited almost the same sweetness activity as sucrose. On the other hand, compound (10) did not appreciably show the activity due to a bulky benzoyl group presumably. These results indicated that photoreactive 2-propoxyaniline and (22) have enough affinity with the sweet taste receptor to elucidate the binding site for the ligands in the sweet taste receptor.

Conclusion:
In summary, we have achieved for the first time a comprehensive synthesis of photoreactive (an arylazide, a benzophenone, and an arylidiazine) 2-propoxyanilines for investigation of the sweet activity as sucrose. On the other hand, compound photoreactive 2-propoxyaniline, (6) and (22), exhibited almost the same sweetness activity as sucrose. The reaction mixture was stirred for 1 d at 95 °C and the solution was removed. The residue was purified by silica column chromatography (CHCl3/n-hexane 1:1) to yield 6 (80.0 µg, 51%) as yellow oil.

\[ \text{H NMR (200 MHz, CDCl3): } \delta = 7.81 - 7.68 (m, 2H), 7.62 - 7.38 (m, 3H), 7.27 (s, 1H), 6.79 - 6.54 (m, 2H) \]

HRMS (ESI) m/z [M + H]⁺ calcd for C9H13BrN4O 194.0926, found 194.0929.

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Supporting Information
YES (this text will be updated with links prior to publication)

Primary Data
NO (this text will be deleted prior to publication)

References and Notes
CDCl₃: δ = -65.2. HRMS (ESI) m/z [M + H]+ calc for C₁₁H₁₃F₃N₃O 260.1011, found 260.1010.

(23) Cellular Responses with Cell-Based Assay
The hT1R2-hT1R3 and G16-gust44 cell lines (approx 80,000 cells) were treated in 96-well black-wall plates with a calcium indicator dye (FLIPR Calcium 4) for 1 hour at 37 °C. Fluorescence changes by excitation at 485 nm, emission at 525 nm, were monitored at 2 seconds intervals. The photoreactive compounds (6, 10, 22) and sucrose (as positive control) were added at 20 seconds, and scanning was continued for an additional 4 minutes.