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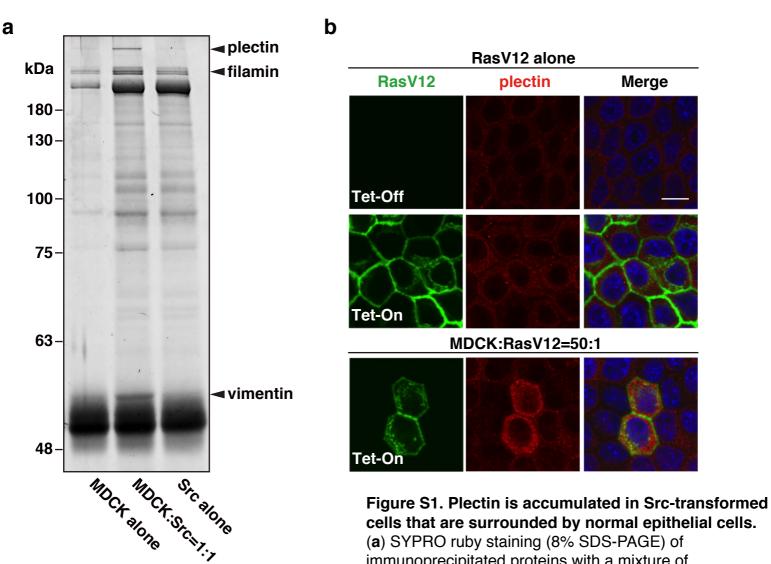
Supplementary Information

Plectin is a novel regulator for apical extrusion of

RasV12-transformed cells

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Supplementary Figure S1-6



immunoprecipitated proteins with a mixture of anti-phospho-tyrosine antibodies. Cells were cultured C under three different conditions: i) normal MDCK cells MDCK:cSrc=50:1 alone, ii) 1:1 mix culture of normal and Src-transformed **cSrc** plectin Merge MDCK cells, and iii) Src-transformed MDCK cells alone. Cell lysates were collected after 16 h incubation with the temperature shift, followed by immunoprecipitation. The XY arrowheads indicate the band for plectin, filamin, and vimentin. (b) Immunofluorescence images of plectin in RasV12-transformed MDCK cells that are cultured alone or co-cultured with normal MDCK cells in the presence or XZ absence of tetracycline. MDCK-pTR GFP-RasV12 cells were mixed with normal MDCK cells or cultured alone on cSrc alone collagen gels. Cells were fixed after 16 h incubation with tetracycline and stained with anti-plectin antibody (red) and Hoechst (blue). Scale bar, 10 µm. (c) Immunofluorescence XY images of plectin in the mix culture of normal and Src-transformed MDCK cells. MDCK-pTR GFP-cSrcY527F cells were mixed with normal MDCK cells or cultured alone on collagen gels. Cells were fixed after 16 h incubation with tetracycline and stained with anti-plectin antibody (red) and XZ Hoechst (blue). Scale bar, 10 µm.

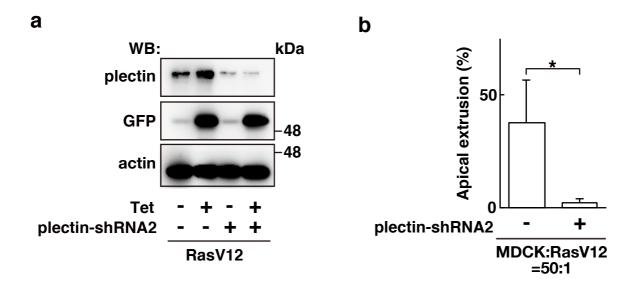


Figure S2. Plectin plays a positive role in apical extrusion of RasV12-transformed cells. (a) Establishment of MDCK-pTR GFP-RasV12 cells stably expressing plectin-shRNA2. Expression of GFP-RasV12 was induced with tetracycline treatment, followed by western blotting with the indicated antibodies. (b) Quantification of the effect of plectin-knockdown on apical extrusion of RasV12 cells. Data are mean \pm SD from seven and three independent experiments. *P < 0.05; n=100-290 cells for each experimental condition.

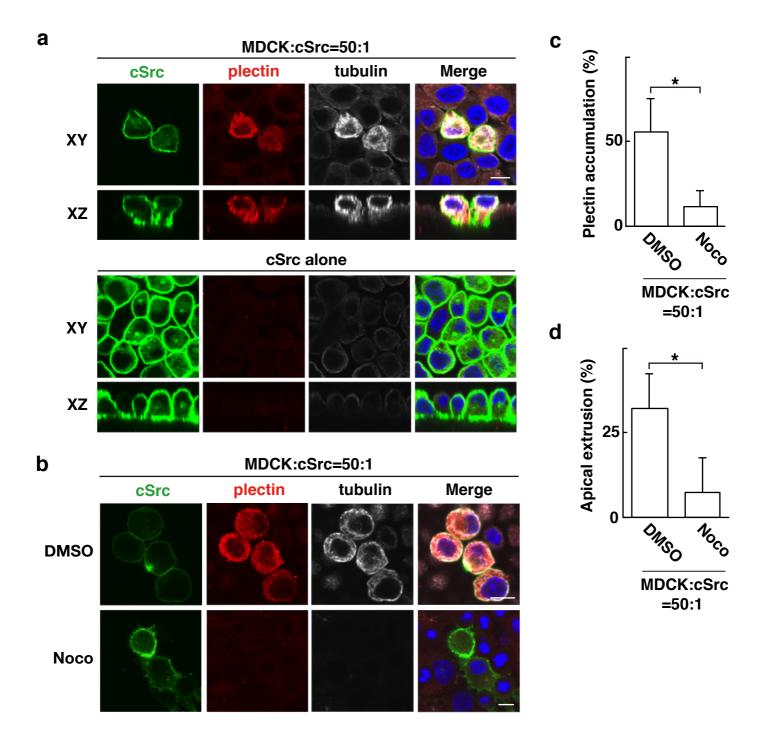


Figure S3. Microtubules and plectin co-regulate their accumulation and promote apical extrusion of Src-transformed cells. (a) Accumulation of plectin and tubulin in Src-transformed cells surrounded by normal epithelial cells. MDCK-pTR GFP-cSrcY527F cells were mixed with normal MDCK cells or cultured alone on collagen gels. Cells were fixed after 16 h incubation with tetracycline and stained with anti-plectin (red) and anti-tubulin (white) antibodies and Hoechst (blue). (b,c) Effect of nocodazole on accumulation of plectin and tubulin. (c) Data are mean \pm SD from three independent experiments. *P<0.05; n=150-180 cells for each experimental condition. (a,b) Scale bars, 10 µm. (d) Effect of nocodazole on apical extrusion of Src-transformed cells. Data are mean \pm SD from three independent experiments. *P<0.05; n=60-120 cells for each experimental condition.

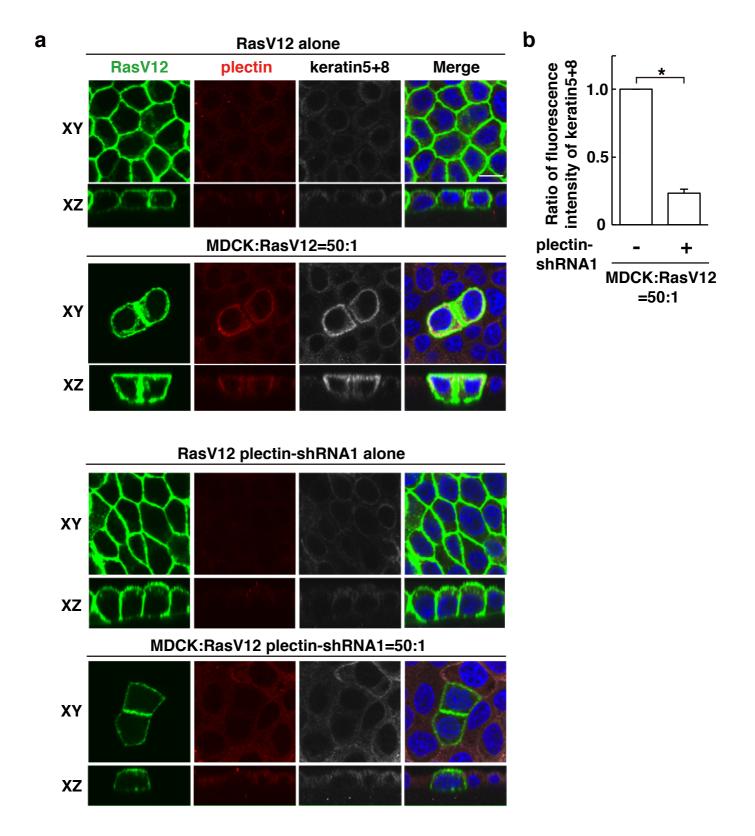
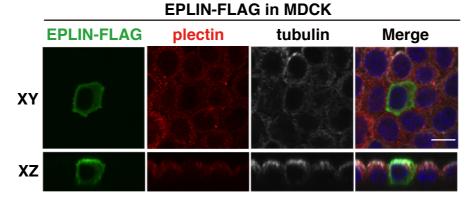


Figure S4. Plectin regulates accumulation of keratin intermediate filaments in RasV12-transformed cells surrounded by normal epithelial cells. (a) Accumulation of plectin and keratin5+8 in RasV12-transformed cells surrounded by normal epithelial cells. MDCK-pTR GFP-RasV12 cells or MDCK-pTR GFP-RasV12 plectin-shRNA1 cells were mixed with normal MDCK cells or cultured alone on collagen gels. Cells were fixed after 16 h incubation with tetracycline and stained with anti-plectin (red) and anti-keratin5+8 (white) antibodies and Hoechst (blue). (b) Effect of plectin-knockdown on accumulation of keratin5+8. Data are mean ± SD from three independent experiments. *P<0.001; n=30 cells for each experimental condition. Values are expressed as a ratio relative to plectin-shRNA1 (-).



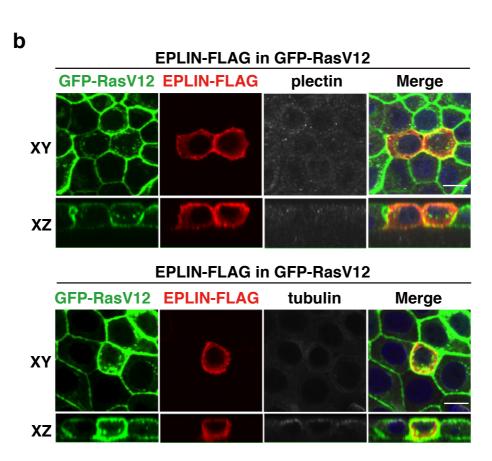


Figure S5. Overexpression of EPLIN alone is not sufficient to cause accumulation of plectin or microtubules. EPLIN-FLAG was transiently expressed within a monolayer of MDCK cells (a) or MDCK-pTR GFP-RasV12 cells (b) in a mosaic manner. Cells were fixed after 24 h of transfection and stained with anti-FLAG, anti-plectin, and/or anti-tubulin antibodies and Hoechst. Scale bars, 10 μm.

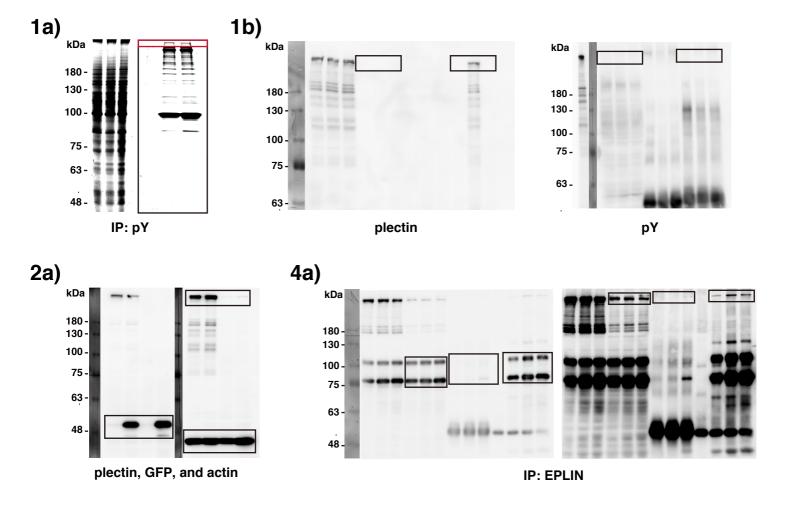


Figure S6. Full-length gels and blots