Title: Neuropathological and biochemical criteria to identify acquired Creutzfeldt-Jakob disease among presumed sporadic cases

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Running title: Criteria to identify acquired CJD
Abstract

As an experimental model of acquired Creutzfeldt-Jakob disease (CJD), we performed transmission studies of sporadic CJD using knock-in mice expressing human prion protein (PrP). In this model, the inoculation of the sporadic CJD strain V2 into animals homozygous for methionine at polymorphic codon 129 (129M/M) of the PRNP gene produced quite distinctive neuropathological and biochemical features, i.e., widespread kuru plaques and intermediate type abnormal PrP (PrPSc). Interestingly, this distinctive combination of molecular and pathological features has been, to date, observed in acquired CJD but not in sporadic CJD. Assuming that these distinctive phenotypic traits are specific for acquired CJD, we revisited the literature and found two cases showing widespread kuru plaques despite the 129M/M genotype, in a neurosurgeon and in a patient with a medical history of neurosurgery without dura mater grafting. By western blot analysis of brain homogenates, we revealed the intermediate type of PrPSc in both cases. Furthermore, transmission properties of brain extracts from these two cases were indistinguishable from those of a subgroup of dura mater graft-associated iatrogenic CJD caused by infection with the sporadic CJD strain V2. These data strongly suggest that the two atypical CJD cases, previously thought to represent sporadic CJD, very likely acquired the disease through exposure to prion-contaminated brain tissues. Thus, we propose that the distinctive combination of 129M/M genotype, kuru plaques, and intermediate type PrPSc, represents a reliable criterion for the identification of acquired CJD cases among presumed sporadic cases.

Key words: Creutzfeldt-Jakob disease, prions, prion diseases, PRNP, iatrogenic
INTRODUCTION

Prion diseases are lethal transmissible neurodegenerative diseases caused by an abnormal isoform of prion protein (PrP\textsuperscript{Sc}), a component of a proteinaceous infectious particle named prion, which is converted from the normal cellular isoform (PrP\textsuperscript{C}).\textsuperscript{1} The conformational conversion of PrP\textsuperscript{C} is believed to occur: (1) spontaneously in sporadic Creutzfeldt-Jakob disease (sCJD) and variably protease-sensitive prionopathy, (2) as a consequence of a pathogenic PRNP mutation in genetic CJD, Gerstmann-Sträussler-Scheinker syndrome, and fatal familial insomnia, (3) through an acquired prion infection in iatrogenic CJD, kuru, and variant CJD. Proven sources of iatrogenic CJD transmission include dura mater grafts, growth and gonadotrophic hormone parenteral administration, neurosurgical instruments, corneal grafts, and stereotactic intracranial electrodes,\textsuperscript{2} while kuru, the other acquired form of prion disease related to sCJD, likely originated from the transmission of prions through ritual cannibalism.\textsuperscript{3}

As an experimental model of acquired CJD, transmission studies using knock-in mice expressing human PrP\textsuperscript{C} have been extensively pursued over the last decade.\textsuperscript{4-15} These animal models have successfully clarified the pathogenesis of acquired CJD, particularly the relationship between PRNP polymorphism and disease susceptibility or disease phenotype.\textsuperscript{16} One of the most important findings in these animal models is the neuropathological and biochemical characteristics of acquired CJD that would help to distinguish acquired CJD cases from sporadic cases. In this review, we summarize recent advances provided by experimental transmission studies of sCJD prions with implication for the diagnostic approach aimed at identifying acquired CJD cases.

PHENOTYPIC HETEROGENEITY OF ANIMAL MODELS OF ACQUIRED CJD

To model acquired CJD, a range of sCJD isolates have been inoculated into knock-in mice expressing human PrP\textsuperscript{C} with each one of the three PRNP codon 129 genotypes (methionine homozygosity, M/M; valine homozygosity, V/V; or methionine/valine heterozygosity, M/V). Current classification of sCJD is largely based on the PRNP genotype at polymorphic codon 129 and the type of PrP\textsuperscript{Sc} (type 1 or type 2) accumulating in the brain, e.g., MM1, MM2, MV1,
MV2, VV1, or VV2. Types 1 and 2 are two isoforms of PrPSc which are distinguishable according to the size of the protease-resistant core (21 and 19 kDa, respectively). However, the molecular-phenotypic correlation based on these two molecular features is not complete. For example, the MM2 group comprises two clinicopathologically distinct subgroups, which include a cortical (MM2C) and a thalamic histotype (MM2T). Similarly, the MV2 group has also been divided into two distinct subtypes based on pathological criteria, which distinguish a cortical type (MV2C) and a kuru plaque type (MV2K). Finally, since both MM1 and MV1 and, MM2C and MV2C share the same clinicopathological features, they have been merged into single entities, i.e. the MM/MV1 subtype and the MM/MV2C subtype, respectively. In summary, six phenotypic subtypes are recognized in the current classification of sCJD, i.e., MM/MV1, MM/MV2C, MM2T, MV2K, VV1, or VV2. Interestingly, the transmission of each of these six sCJD subtypes to PrP-humanized knock-in mice, has led to the isolation and characterization of five sCJD prion strains with different transmission properties, namely M1 (sCJD-MM/MV1), M2C (sCJD-MM/MV2C), M2T (sCJD-MM2T), V1 (sCJD-VV1), or V2 (sCJD-MV2K and -VV2). Thus, five out of the six subtypes recognized by current sCJD classification seem to be associated with a specific strain of prions, the only exception being the MV 2K, which is indistinguishable from the VV2 subtype after transmission, and therefore also linked to the V2 strain. Disease phenotypes of the PrP-humanized knock-in mice inoculated with each sCJD prion strain are summarized in Table 1.

Among the various combinations of neuropathological and biochemical features observed in the inoculated mice, the most intriguing ones were those observed in 129M/M mice inoculated with the V2 strain. These animals showed widespread PrP amyloid plaques (kuru plaques) and a distinctive type of PrPSc with intermediate electrophoretic mobility (~20 kDa) between types 1 and 2, which has been designated as intermediate type (type i), accordingly. Since the combination of the 129M/M genotype, type i PrPSc, and widespread kuru plaques has not been observed in any subtypes of sCJD, these characteristic features likely result from the conformational adaptation of type 2 PrPSc with the 129V genotype (V2 PrPSc) to PrPc with the 129M genotype. Indeed, type i PrPSc and kuru plaques can also be observed in the 129M/V mice inoculated with the V2 PrPSc or in patients with sCJD-MV2K, which is considered to have originated from the spontaneous generation of V2 PrPSc.
PHENOTYPIC HETEROGENEITY OF ACQUIRED CJD PATIENTS

Among human CJD cases, the combination of 129M/M genotype, type i PrPSc, and kuru plaques has only been found in patients with the acquired form of the disease. Dura mater graft-associated CJD (dCJD), one of the two most prevalent subgroups of iatrogenic CJD, can be divided into two subtypes based on clinicopathological features, with the majority represented by a non-plaque-type caused by the M1 sCJD strain and the minority by a plaque-type caused by the V2 sCJD strain. While non-plaque-type dCJD patients show clinicopathological features identical to those affected by sCJD-MM/MV1, plaque-type dCJD patients show the combination of the 129M/M genotype, type i PrPSc, and kuru plaques. Kuru plaque formation combined with the 129M/M genotype has also been found in human growth hormone-associated CJD, the other most prevalent subgroup of iatrogenic CJD. Similar to dCJD, two neuropathologically distinct subtypes are also recognized in growth hormone-associated CJD subjects carrying the 129M/M genotype, based on the presence or absence of kuru plaques. Following Parchi’s classification, the size of protease-resistant PrPSc, among the relatively low number of cases examined to date, has been reported as type 1 in subjects carrying the 129M/M genotype, regardless of the presence of kuru plaques, and as type 2 in those carrying the 129M/V genotype. Since growth hormone-associated CJD cases and dCJD cases show similar divergent neuropathological phenotype, similar etiology has been suspected, i.e., contamination with different sCJD prion strain. Finally, widespread kuru plaques are also the neuropathological hallmark of kuru patients, especially in patients with the 129M/M or 129M/V genotype. Unfortunately, PrPSc typing of kuru brains has also been performed in a limited number of cases. Among them, the size of protease-resistant PrPSc has been reported as type 1 in a single 129M/M patient and as type 2 in a single 129M/V and three 129 V/V patients, respectively. Of note, transmission study using nonhuman primates indicated that kuru might have originated from the sCJD strain V2. Taken together, the data obtained from the phenotypic analyses of human cases and from experimental transmission studies, strongly suggest that the combination of 129M/M genotype, type i PrPSc, and widespread kuru plaques is a distinctive feature of acquired cases caused by an infection with
the sCJD strain V2.

IDENTIFICATION OF ACQUIRED CJD AMONG SPORADIC CASES

Assuming that the 129M/M genotype, type i PrP^Sc, and kuru plaques represent reliable markers of acquired CJD, we searched the literature for CJD cases with these distinctive features and found two of them, previously reported as sCJD, in a neurosurgeon and in a patient with a medical history of neurosurgery without dural grafting. In transmission experiments, these two atypical CJD cases showed the same properties as those of plaque-type dCJD (Table 2). Most significantly, at variance with most sCJD prions in which the codon 129 genotypic homology between inoculated animals and the inoculum strongly favors disease susceptibility, these two 129M/M cases were transmitted most efficiently into 129 V/V mice, despite the mismatched genotype. Moreover, like the sCJD strain V2 they induced the formation of PrP^Sc type 2 (V2 PrP^Sc) in the brain of these mice. Based on these findings, we concluded that the two atypical CJD cases, likewise plaque-type dCJD, actually represent acquired cases of CJD caused by the sCJD strain V2. In summary, the two atypical CJD cases and plaque-type dCJD cases showed the same transmission properties as those of the parental sCJD strain V2, with the highest susceptibility in 129 V/V mice and generation of V2 PrP^Sc, thus representing an example of the so-called “traceback phenomenon”.

CRITERIA TO IDENTIFY ACQUIRED CJD

Based on the findings described above, we have proposed neuropathological and biochemical criteria to identify acquired CJD cases caused by transmission of the sCJD strain V2 to the 129M/M individuals, denoted as acquired CJD-MMiK (129M/M genotype, type i PrP^Sc, and kuru plaques) (Fig. 1A). The distinction between type i PrP^Sc and type 1 PrP^Sc is rather difficult with conventional western blot analysis due to subtle differences in the electrophoretic mobility of the two fragments. This is presumably the reason why PrP^Sc type i has not been reported in acquired prion disease patients other than plaque-type dCJD. For their precise distinction, stringent conditions for protease treatment and high resolution gel electrophoresis systems such
as Bis-tris long gels or 10-20% gradient Tris-glycine long gels are needed.\textsuperscript{20} Alternatively, an analysis of the electrophoretic patterns of the carboxyl terminal PrP\textsuperscript{Sc} fragments using carboxyl terminus-directing antibodies is also quite useful to distinguish PrP\textsuperscript{Sc} type i from PrP\textsuperscript{Sc} type l.\textsuperscript{34}

Furthermore, the characteristic transmission properties also provide a sound basis for the differential diagnosis of the acquired CJD-MMiK as described above (Fig. 1B). To determine the transmission properties of a given CJD inoculum, the follicular dendritic cell assay following intraperitoneal inoculation into PrP-humanized knock-in mice\textsuperscript{4} can be a time-saving alternative to the standard intracerebral transmission that requires long incubation period until disease onset. In addition, \textit{in vitro} conversion assays using PrP\textsuperscript{C} carrying each of the codon 129 genotype as substrate, \textit{e.g.}, protein misfolding cyclic amplification,\textsuperscript{36,37} may also be useful to determine the seeded conversion activity and resulting PrP\textsuperscript{Sc} type.

Besides the neuropathological, biochemical, and transmission properties, clinical features of the acquired CJD-MMiK can also be distinctive as revealed by comparative studies of plaque-type and non-plaque-type dCJD patients.\textsuperscript{21,22} The distinctive clinical features of the acquired CJD-MMiK include gait disturbance as an initial symptom, slow progression of disease, and absence or late occurrence of periodic sharp-wave complexes on electroencephalogram.

**CONCLUDING REMARKS**

Animal models of acquired CJD have contributed greatly to the evaluation of the \textit{PRNP} genotypic effects on disease susceptibility or disease phenotype. The data obtained from animal experiments, combined with the phenotypic characterization of CJD patients have prompted the proposal of diagnostic criteria to identify acquired CJD among presumed sporadic cases as summarized in this review. However, this diagnostic approach is applicable only to a small portion of acquired CJD cases, \textit{i.e.}, the 129M/M individuals infected with the sCJD strain V2. Unfortunately, the other acquired CJD cases, \textit{i.e.}, the other combinations between host \textit{PRNP} genotype and prion strain, cannot be distinguished from sCJD by neuropathological and biochemical analyses, as revealed by the animal models. Nevertheless, further continuous surveillance of acquired CJD cases using the proposed diagnostic approach and the
identification of transmission routes in such cases may help to reduce the risk of iatrogenic CJD transmission in the future.

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224-233.


FIGURE LEGENDS

Fig. 1  Diagnostic criteria to identify acquired CJD. (A) The combination of the 129M/M genotype at the polymorphic codon 129 of the PRNP gene, intermediate type (type i) PrPSc with the intermediate electrophoretic mobility between PrPSc types 1 and 2, and widespread kuru plaques is a reliable criterion for identification of acquired CJD cases caused by infection with the sCJD strain V2, designated as acquired CJD-MMiK. (B) Transmission properties of acquired CJD-MMiK in PrP-humanized knock-in mice are also distinctive. Since the causative origin of acquired CJD-MMiK is the sCJD strain V2, acquired CJD-MMiK shows the same transmission properties as those of the sCJD strain V2. In particular, 129V/V mice show the highest susceptibility despite the mismatched codon 129 genotype, with PrPSc type 2 accumulation and plaque-like PrP deposition in their brain.
Table 1  Transmission properties of sCJD prions in PrP-humanized knock-in mice

<table>
<thead>
<tr>
<th>Inoculum</th>
<th>Mouse line</th>
<th>PrPSc type</th>
<th>PrP deposition†</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Prion strain</td>
<td>sCJD subgroup</td>
<td>129M/M</td>
<td>1</td>
<td>S</td>
</tr>
<tr>
<td>M1</td>
<td>MM/MV1</td>
<td>129M/V</td>
<td>1</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td></td>
<td>129V/V</td>
<td>1</td>
<td>S</td>
</tr>
<tr>
<td>M2C</td>
<td>MM/MV2C</td>
<td>129M/M</td>
<td>— ‡</td>
<td>—</td>
</tr>
<tr>
<td></td>
<td></td>
<td>129M/V</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td></td>
<td></td>
<td>129V/V</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>M2T</td>
<td>MM2T</td>
<td>129M/M</td>
<td>2</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td></td>
<td>129M/V</td>
<td>N.D.§</td>
<td>N.D.</td>
</tr>
<tr>
<td></td>
<td></td>
<td>129V/V</td>
<td>N.D.</td>
<td>N.D.</td>
</tr>
<tr>
<td>V1</td>
<td>VV1</td>
<td>129M/M</td>
<td>1</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td></td>
<td>129M/V</td>
<td>1</td>
<td>S</td>
</tr>
<tr>
<td></td>
<td></td>
<td>129V/V</td>
<td>1</td>
<td>S</td>
</tr>
<tr>
<td>V2</td>
<td>MV2K or VV2</td>
<td>129M/M</td>
<td>i†</td>
<td>K</td>
</tr>
<tr>
<td></td>
<td></td>
<td>129M/V</td>
<td>i+2‡</td>
<td>K</td>
</tr>
<tr>
<td></td>
<td></td>
<td>129V/V</td>
<td>2</td>
<td>P</td>
</tr>
</tbody>
</table>

† The patterns of PrP deposition in the mouse brain are classified into diffuse synaptic deposition (S), kuru plaques (K), or plaque-like deposition mainly observed in the white matter (P).
‡ The M2C sCJD strain was not transmissible to the PrP-humanized knock-in mice.
§ N.D., not done.
† Intermediate type that shows intermediate electrophoretic mobility (~20 kDa) between type 1 (21 kDa) and type 2 (19 kDa) PrPSc.
‡ A mixture of type i and type 2 PrPSc.
**Table 2**  Transmission properties of the atypical CJD cases or plaque-type dCJD cases in PrP-humanized knock-in mice

<table>
<thead>
<tr>
<th>Inoculum</th>
<th>Mouse line</th>
<th>PrP&lt;sup&gt;Sc&lt;/sup&gt; type</th>
<th>PrP deposition&lt;sup&gt;†&lt;/sup&gt;</th>
<th>References</th>
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<td>Atypical CJD</td>
<td>129M/M</td>
<td>i&lt;sup&gt;‡&lt;/sup&gt;</td>
<td>K</td>
<td>34</td>
</tr>
<tr>
<td></td>
<td>129M/V</td>
<td>i+2&lt;sup&gt;§&lt;/sup&gt;</td>
<td>K</td>
<td></td>
</tr>
<tr>
<td></td>
<td>129V/V</td>
<td>2</td>
<td>P</td>
<td></td>
</tr>
<tr>
<td>Plaque-type dCJD</td>
<td>129M/M</td>
<td>i</td>
<td>K</td>
<td>8,22,34</td>
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<tr>
<td></td>
<td>129M/V</td>
<td>i+2</td>
<td>K</td>
<td></td>
</tr>
<tr>
<td></td>
<td>129V/V</td>
<td>2</td>
<td>P</td>
<td></td>
</tr>
</tbody>
</table>

<sup>†</sup> The patterns of PrP deposition in the mouse brain are classified into kuru plaques (K) or plaque-like deposition mainly observed in the white matter (P).

<sup>‡</sup> Intermediate type that shows intermediate electrophoretic mobility (~20 kDa) between type 1 (21 kDa) and type 2 (19 kDa) PrP<sup>Sc</sup>.

<sup>§</sup> A mixture of type i and type 2 PrP<sup>Sc</sup>. 
Fig. 1

A

PRNP genotype: 129M/M

Codon 129

PrPSc type: intermediate type (type i)

Type 1  Type i

PrP deposition: kuru plaques

B

129MM  Type i PrPSc  Kuru plaques

129MV  Type i+2 PrPSc  Kuru plaques

129V/V  Type 2 PrPSc  Plaque-like deposition