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博士学位論文

Studies on the multisensory integration in cricket's
auditory system

(コオロギ聴覚系における多感覚統合に関する研究)

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学位論文内容の要旨

博士の専攻分野の名称 博士（生命科学） 氏名 染谷 真琴

学位論文題名

Studies on the multisensory integration in cricket's auditory system

(コオロギ聴覚系における多感覚統合に関する研究)

動物は異なる感覚器官で受容された複数のモダリティの感覚刺激の情報を中枢神経系で統合する。この過程は多感覚統合と呼ばれ、それによる神経活動の変化は物体の知覚や刺激によって誘導される行動の修飾をもたらす。多感覚統合は捕食者の検出感度の向上や、より正確な位置や距離の把握に有効であり、生存のための回避行動にも利益をもたらすと推測される。昆虫の中枢神経系には捕食者の接近に関連した刺激を検出するいくつかのニューロンが同定されているが、これらのニューロンが複数モダリティの感覚入力を統合しうるのかは不明であった。本学位論文では、捕食者であるコウモリが発する超音波エコーに応答するコオロギの聴覚ニューロン AN2 を対象として、やはり捕食者の接近を検知する気流感覚器である尾葉からの機械感覚入力と前肢鼓膜器官からの聴覚入力の多感覚統合に着目し、この問題に取り組んだ。

第一章では、AN2 を含む聴覚ニューロンが尾葉からの機械感覚入力を受けるかを検証した。尾葉は空気流の変化によって捕食者の接近を検出し、コオロギに逃避行動を引き起こす。まず、頸部腹側縦連合からの細胞外記録により、音刺激および気流刺激に応答する上行性神経活動を検証し、次に細胞内記録法により前胸神経節内における多感覚ニューロンを探索した。その結果、すでに聴覚ニューロンとして同定されている AN2, ON1 を含むいくつかの前胸介在ニューロンが気流刺激に対してスパイク応答を示した。気流応答は尾葉を切除すると消失したことから、鼓膜器官で受容された聴覚入力と尾葉で受容された気流感覚入力が、前胸神経節内の感覚情報処理の初期段階において統合されることが示唆された。

第二章では、聴覚および気流感覚入力が AN2 のスパイク発火に与える影響を検証した。音と気流刺激の同時提示刺激に対する応答はそれぞれの刺激の単独

提示に対する応答より有意に大きく、多感覚統合によって応答が増大されることがわかった。しかし、同時提示に対する応答は、基本的に単独提示に対する応答の和にほぼ等しく、AN2は音と気流入力を線形に加算していた。単独提示応答の線形和よりも大きな加算は、スパイク閾値に近い弱い強度の刺激を同時提示した場合にのみ観察された。したがって、AN2の多感覚統合における非線形加算は、興奮性シナプス後電位（EPSP）からスパイクへ変換する閾値特性を反映したものであると考えられる。

AN2は音源の位置、音圧および音の長さといった情報を高頻度のスパイク発火（バースト発火）で符号化することが報告されている。音と気流の同時提示は、単独提示よりも高い頻度でバースト発火を誘発したことから、多感覚統合は音情報表現へも影響することが示唆された。しかし、単独提示した音刺激の強度を増大した場合にも同様にバースト発火は増大し、同程度の大きさのスパイク活動に含まれるバースト数は単独提示と同時提示に違いは見られなかった。したがって、バースト発火の増加は単にスパイク応答の増大によるもので、多感覚統合特異的なものではないと考えられる。したがって、AN2は刺激のモダリティによらず興奮性入力を統合しており、その統合過程は一般的なシナプス入力加算のメカニズムによるものであると推測された。

第三章では、AN2におけるバースト発火をもたらす神経基盤を理解する手がかりとして、感覚刺激のモダリティ、およびバースト発火と閾値以下の膜電位応答の関連を検証した。細胞内記録したAN2の膜電位応答からスパイク成分を取り除き、閾値下の興奮性シナプス後電位（EPSP）を推定した。第二章においてほとんどのバースト発火は一発目の活動電位に続いて生じていたため、この初発スパイク後のEPSPに注目したところ、同時提示刺激に対する応答は単独提示刺激に対する応答よりも脱分極がより持続していた。この期間の脱分極の高とその持続時間は、いずれもバースト発火の発生および個々のバースト発火の大きさと相関を示したが、その関係性に刺激モダリティによる違いはみられなかった。以上の結果から、閾値以下の膜電位変化はバーストの発生およびその大きさを決定する重要な要因であるが、それに対する影響は入力強度の増大によるものであり多感覚統合特異的ではないことがわかった。

AN2はコオロギの捕食者であるコウモリが発する超音波を検出し、そのバー

スト発火は飛行中のコオロギに音源と反対方向に飛行する逃避行動を引き起こすことから、捕食者検出ニューロンとして働くと考えられている。本研究によって、AN2 が尾葉への気流刺激に対しても興奮性の応答を示し、鼓膜からの聴覚入力と線形に加算したスパイク出力をすることによってバースト発生を増大させることがわかった。本研究結果は、感覚情報処理の初期に単一ニューロン内で複数のモダリティの感覚入力統合され、逃避行動に利用されている可能性を示すものである。

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General Discussion

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Acknowledgment

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1 **General Introduction**

2 The central nervous systems of animals integrate various modalities of
3 signals detected by distinct sensory organs in a process known as
4 multisensory integration (Pouget et al., 2002; Fetsch et al., 2013).
5 Multisensory integration modulates the neural activity, which leads to
6 improvement of sensory perception and enhancement of the behavioral
7 responses to cross-modal stimuli compared to those of unimodal stimuli (Stein
8 et al., 1988; Alais and Burr, 2004; Gu et al., 2008; McMeniman et al., 2014).
9 For example, Human accurately perceives an object by integrating the visual
10 and tactile signals (Ernst and Banks, 2002).

11 Thus, multisensory integration offers some advantages in detection of
12 predators and in successful escaping because the attack of predator gives rise
13 to multiple sensory cues. Several sensory interneurons in insects have been
14 identified as detectors of predators' signals, and their activity elicits
15 avoidance behaviors. For example, the visual interneurons, such as LPLC2 in
16 fruit flies and DCMD in locusts are sensitive to looming motion stimuli that
17 are perceived as an approaching predator. If the spiking responses of these
18 neurons exceed a certain threshold, escape behavior is triggered (Fotowat and
19 Gabbiani, 2007; Herberholz and Marquart, 2012; Klapoetke et al., 2017). The
20 several identified auditory interneurons in crickets, moths, and mantids are
21 also sensitive to ultrasound, such as the echolocation call of bats that prey on
22 these insects, and activity of these neurons evoked by these stimuli elicits

23 defensive steering in flight (Hoy et al., 1989; Pollack, 2015). However, it
24 remains unknown whether these key neurons are sensitive to multiple
25 sensory cues relevant to a predator's attack. If these neurons have
26 multimodal sensitivity, how are their spiking outputs altered by
27 simultaneous sensory inputs of different modalities? To address these
28 questions, which are main concerns of my dissertation, I testes multimodal
29 sensitivity in prothoracic auditory neurons in the crickets, and focused on the
30 auditory neuron 2 (AN2) that was identified as one of the multimodal neurons
31 sensitive to both ultrasound and airflow.

32 **Chapter I**

33 **Identification of Multimodal Neurons of Crickets**

34 1.1 Introduction

35 Crickets process information on aero dynamics with two distinct sensory
36 systems, cercal and auditory systems, which have the distinctly different
37 sensory organs and frequency ranges of aero dynamics (Fig. 1-1A). Both
38 sensory systems provide crucial signals of the predator attack and trigger the
39 escape behavior.

40 The cerci are abdominal appendages covered with 500-750 filiform hairs
41 that sense surrounding air particle displacement (Palka and Olberg, 1977;
42 Miller et al., 2011). Information about airflow dynamics is processed by a local
43 circuit within the terminal abdominal ganglion and transmitted by several
44 ascending giant interneurons (GIs) (Fig. 1-1B) (Miller et al., 1991; Theunissen
45 and Miller, 1991). The airflow signals detected by cerci are considered to be
46 perceived as a sign of a predator's attack for crickets standing on the ground,
47 because short air puff stimuli elicit running or jumping escapes (Tauber and
48 Camhi, 1995; Oe and Ogawa, 2013; Sato et al., 2017).

49 On the other hand, sound with the higher frequency than 3 kHz
50 (Imaizumi and Pollack, 1999) is detected by auditory receptor neurons of
51 tympanal organ located at tibia of the front legs. The auditory receptor
52 neurons project their afferent axons into the prothoracic ganglion (Hennig,
53 1988; Imaizumi and Pollack, 1999, 2005). Several prothoracic neurons
54 directly receive excitatory synaptic inputs from auditory receptor afferents
55 (Hennig, 1988). Sound information is conveyed by several kinds of ascending

56 projection neurons to brain (Hennig, 1988). The acoustic signals mediated by
57 the ascending auditory neurons elicit escape steering during the flight (Nolen
58 and Hoy, 1984; Schildberger and Hörner, 1988; Hedwig, 2006; Marsat and
59 Pollack, 2012).

60 The cercal GIs project large long axon to brain and have axon collaterals
61 arborizing into each segmental ganglion including the prothoracic ganglion
62 (Hirota et al., 1993), suggesting the possibility that prothoracic auditory
63 neurons are also sensitive to the airflow via the cercal system. Actually, it has
64 been revealed that some prothoracic auditory neurons respond to wind
65 stimuli (Atkins and Pollack, 1987). However, the wind sensitivity of auditory
66 neurons related to avoidance behaviors has not been tested.

67 In this chapter, I explored whether the auditory neurons, which play key
68 roles in the sensory coding relevant to biologically important events, receive
69 the airflow inputs from cerci without the neural pathways through
70 protocerebrum.

71 1.2 Materials and Methods

72 *Animals*

73 Adult female crickets (*Gryllus bimaculatus*) were used for the experiments.

74 The crickets were bred and housed on a 12-h light/dark cycle at a constant
75 temperature of 28°C.

76

77 *Extracellular recording*

78 For preparation of the extracellular recording, crickets were pinned dorsal
79 side up on a silicone platform after removing the middle and hind legs. The
80 connective nerve cords between the prothoracic and subesophageal ganglia
81 were exposed by removing the gut and surrounding fat and were surgically
82 cut to block the descending signals from the cephalic ganglia. Ascending
83 spikes were extracellularly recorded using glass suction electrodes filled with
84 isotonic saline placed proximally on the cut end of the nerve cord (Fig. 1-1A).

85 For recording all ascending signals as multiple spike units, the whole nerve
86 cord was suctioned by a glass micro-pipettes, while a part of ascending
87 activities was recorded from a piece of split nerve cord for the single-unit
88 recording. Signals were digitized at 20 kHz with a Powerlab 4/30 analog-
89 digital converter (ADInstruments) and filtered with a 150–3,000 Hz band
90 pass implemented in Chart version 7 software (ADInstruments).

91 Recorded spikes were sorted using a commercially available software
92 (Spike Taro, Chinou Jouhou Shisutemu), which uses a sorting algorithm

93 based on the following two criteria: peak detection to separate individual
94 units from a compound spike, and classification based on the correlation
95 between separated units in terms of spike waveform and height. This
96 algorithm has been used to analyze extracellular recordings of insect
97 receptors (Kidokoro-Kobayashi et al., 2012). Spikes with heights in the range
98 of $\pm 35\%$ and waveforms with a correlation coefficient of >0.90 were
99 considered to be generated by a single neuron. Units that fired more than
100 once on average in a trial were selected for analysis. In the extracellular
101 recording of the whole nerve cord, 134 units were sorted from 4 recordings.

102

103 *Stimulation for extracellular recording experiments*

104 The airflow stimulus consisted of short puffs of nitrogen gas from a nozzle
105 connected to a PV820 pneumatic picopump (World Precision Instruments,
106 Sarasota, FL, USA). Eight nozzles were arranged around the cerci on the
107 same horizontal plane with their ends positioned 45° apart at a distance of 40
108 mm from the cerci. The velocity of airflow was 1.12 m/s as measured at the
109 cerci. The duration of the airflow stimulus was 200 ms.

110 For sound stimulation, a 10-kHz pure tone was generated by RpvdsEx
111 software (Tacker Davis Technologies, Alachua, FL, USA) and presented
112 through 1.5 inch full-range sealed loudspeaker (MM-SPS2, Sanwa Supply,
113 Okayama, Japan). Eight speakers were arranged around the cerci on the
114 same horizontal plane with their ends positioned 45° apart. The duration of

115 sound pulses was 200 ms with symmetrical rise and fall times of 2 ms. The
116 sound intensity was 70 dB SPL in the recording from whole nerve cord and
117 76 dB SPL in the recording from a split nerve cord.

118 Single airflow or sound stimulus was provided independently from
119 various directions for multi-unit recording or from ipsilateral side to the nerve
120 cord for single-unit recording. To confirm whether the airflow was not
121 detected by the tympanum, responses to airflow and sound stimuli were
122 collected again after both frontal legs were ablated or both tympanums were
123 gummed up by adhesion bond.

124

125 *Intracellular recording*

126 For preparation of the intracellular recording, the wings and hind legs were
127 removed and the crickets were positioned ventral side-up on a block of clay
128 and restrained with small metal clamps. The prothoracic ganglion was
129 exposed and stabilized on a stainless-steel platform using a silver wire ring.
130 To eliminate descending signals from the cephalic ganglion, the connective
131 nerve cords between the prothoracic and subesophageal ganglia were cut. In
132 the ablation experiment, both cerci were removed to confirm that the auditory
133 neurons received the sensory signals from the cerci.

134 Glass microelectrodes (40-60 M Ω) were filled with 8% Lucifer yellow CH
135 lithium salt (Sigma-Aldrich, RRID:SCR_008988) in 200-mM LiCl or 2-mM
136 Oregon Green BAPTA-1 hexapotassium salt in 200-mM potassium acetate.

137 Lucifer yellow was used for most of the recordings and both kinds of the
138 electrode solution have no influence on the spontaneous activities and neural
139 responses. Glass microelectrode was inserted into dendrites or axons of
140 auditory neurons and membrane potentials of recorded neurons were
141 measured with current clamp mode. Electrophysiological signals were
142 digitized at 20 kHz with a Powerlab 4S analog-to-digital converter
143 (ADInstruments) and recorded with Chart version 7 software
144 (ADInstruments). To load the fluorescent dye, a hyperpolarizing current of -1
145 to -2 nA was injected during recording. To identify the recorded neuron, the
146 prothoracic ganglion was dissected after recording, fixed in 4%
147 paraformaldehyde in 200-mM phosphate buffer over night, dehydrated for 15
148 min each in a series of 50, 70, 90, 100, and 100% ethanol, and cleared in
149 methyl-salicylate.

150

151 *Stimulation in the intracellular recordings*

152 The airflow stimulus consisted of short puffs of nitrogen gas from a nozzle
153 connected to an IM-300 microinjector (Narishige, Japan). For stimulation, the
154 nozzle ends were positioned 15 mm away from the cerci on either side of the
155 cricket. The duration of the airflow stimulus was 200 ms. The velocity of
156 airflow was from 0.17 to 0.43 m/s (see each figure legend). For finding the
157 auditory neurons, 5- or 15-kHz pure tone was generated by R PvdsEx software

158 (Tacker Davis Technologies) and presented through an ATH-CM707
159 headphone speaker (audio-technica, Tokyo, Japan). A pair of speakers were
160 positioned 25 mm away from the tympanum on either side. The duration of
161 sound pulses was 30 ms with symmetrical rise and fall times of 2 ms. The
162 sound intensity was 60 dB SPL.

163

164 *Data analysis*

165 Spike traces were binned at 20-ms resolution through all figures in this
166 chapter. To analyze subthreshold voltage traces, raw traces of the recordings
167 were filtered by a median filter with a 5-ms window (Shi et al., 2017). Then,
168 the resting level of membrane potential was subtracted from the filtered
169 potentials to indicate the stimulus-evoked postsynaptic potentials. The
170 latency of EPSP was defined as the duration from the stimulus onset to the
171 points when the filtered potential exceeded 3 mV, which corresponded to 2 SD
172 (2.89 mV) of the spontaneous fluctuation in the resting potentials of AN2 and
173 omega neuron 1, from the resting potential.

174 MATLAB software (MathWorks, RRID:SCR_001622) was used for data
175 processing and analysis, and R programming software v3.4.0 (R Core Team
176 2013) was used for statistical analysis. To avoid pseudoreplication, the data
177 obtained from the same samples were averaged for each individual. All values
178 are reported as mean \pm standard error of the mean. A p -value of < 0.05 was
179 considered to be statistically significant.

180 **1.3 Results**

181 **1.3.1 Exploring multimodal neurons by extracellular recordings**

182 First, I examined existence of ascending multimodal neurons in the thoracic
183 or abdominal ganglia that responded to the acoustic and airflow stimuli. The
184 spiking responses of the ascending neurons to the sound stimulus applied to
185 tympanums and the airflow stimuli applied to cerci were extracellularly
186 recorded from the distal cut end of a nerve cord between the subesophageal
187 and prothoracic ganglia (Fig. 1-2A, B). Individual spikes were sorted into
188 multiple units based on their wave shapes of spikes (see Methods). I found
189 multi-modal units which showed firing responses to both sound and airflow
190 stimuli (Fig. 1-3A, B). These multimodal units accounted for 46% of all sorted
191 units (Fig. 1-3C). Thirty percent of units were sensitive to only airflow stimuli,
192 but no units responded to only sound stimuli. The multimodal units showed
193 remarkable increase in spiking activity in responses to the both stimuli (Fig.
194 1-3D). The auditory responses completely disappeared after the tympanums
195 were removed (Fig. 1-3D). After the tympanum removal, 96% of them showed
196 no significant increase in spiking responses to sound (unpaired t-test). These
197 results indicate that a part of ascending neurons is sensitive to both airflow
198 and sound stimuli.

199 However, the multi-modal units might contain multiple cellular
200 activities derived from auditory-specific and cercal-specific neurons or
201 activity of a single cell might split into multiple units. To confirm the

202 existence of multimodal neurons, single unit recording was performed from a
203 split nerve cord (Fig. 1-4A). In the single unit recording, the multimodal spike
204 unit was recorded, which showed spiking responses to both sound and airflow
205 stimuli (Left 2 panels in Fig. 1-4B, C). After both tympanums were covered
206 with adhesion bond, the units showed no response to the sound while it still
207 responded to the airflow. This result also supported the existence of ascending
208 multimodal neurons in the thoracic or abdominal ganglia.

209

210 **1.3.2 Multimodal neurons in the prothoracic ganglion**

211 To identify the multimodal neurons observed with extracellular recordings, I
212 performed intracellular recording in the prothoracic ganglion to monitor the
213 membrane potential responses to auditory and airflow stimuli and to stain
214 the recorded neuron with fluorescent dye. I found that ascending neuron
215 identified as AN2 and local interneuron identified as omega neuron 1 (ON1),
216 both of which have been identified as the auditory ascending neuron (Wohlers
217 and Huber, 1982), were sensitive to the airflow stimuli to cerci (Fig. 1-5A, B).
218 Their responses to the airflow were significantly larger in the firing rate than
219 the spontaneous activities (Fig. 1-5C). In addition to AN2 and ON1, omega
220 neurons 2 (ON2), T-shape neuron 1 (TN1) and descending neuron 1 (DN1)
221 also showed distinct spiking responses to airflow stimuli. However, their
222 airflow-evoked responses were no significance statistically because of small
223 size of samples. In contrast, the auditory ascending neuron 1 (AN1) showed

224 no response to the airflow stimuli (Fig. 1-6A, B), of which firing rate and
225 membrane potential during the airflow stimulation were not different from
226 their spontaneous activity (Fig. 1-6C, D). My results demonstrated that
227 certain types of auditory neurons including ascending, descending, and local
228 interneurons have bi-modal sensitivity.

229

230 **1.3.3 Response property of the auditory neurons to airflow stimulation**

231 Next, I examined the response properties of AN2 and ON1 to airflow stimuli
232 (Fig. 1-7A). In both AN2 and ON1, the airflow stimulus evoked transient
233 firing (Fig. 1-7B). The latency from stimulus onset to the first spike in AN2
234 was shorter than that in ON1 although this time difference was not
235 significant (Fig. 1-7C). I observed no difference in their firing rate in the
236 transient spiking responses (Fig. 1-7D) and during the later phase following
237 the transient response (Fig. 1-7E).

238 There was significant difference in the time course of the subthreshold
239 membrane potential changes between AN2 and ON1 (Fig. 1-8A). Both AN2
240 and ON1 showed transient depolarization of their membrane potential in
241 response to airflow stimuli. In AN2, the depolarized membrane potential
242 quickly returned to the resting level. In contrast, ON1's membrane potential
243 depolarization was sustained even after the stimulus termination. The
244 latency of EPSP of AN2 was shorter than that of ON1 (Fig. 1-8B). There was
245 no difference in EPSP amplitude during the transient phase (Fig. 1-8C),

246 whereas the EPSP during the later phase following the transient
247 depolarization was larger in ON1 than AN2 (Fig. 1-8D). Therefore, although
248 both AN2 and ON1 show transient depolarization in response to airflow
249 stimuli, ON1's EPSP was longer lasting and evoked sustained firing. This
250 difference in time course of the responses suggests that different neural
251 pathways convey the airflow signals to the ascending and local interneurons.

252 ON1 shows directional selectivity for the auditory stimulus and plays a
253 prominent role in producing the directionality of AN1 and AN2 (Selverston et
254 al., 1985; Horseman and Huber, 1994). Thus, I investigated the directional
255 selectivity for airflow stimulus in ON1. ON1 tended to generate more spikes
256 in response to the airflow stimuli from ipsilateral to its soma than to the
257 contralateral stimuli (Fig. 1-9A, B). There was no difference in the latency
258 from stimulus onset to the first spike between the stimulus directions (Fig. 1-
259 9C). Since ON1 prefers the ipsilateral stimulus to its soma in its auditory
260 response (Wohlert and Huber, 1978), the directional preference in the airflow
261 response coincided with that for auditory stimuli.

262 Finally, I examined whether the cercal system mediated the airflow-evoked
263 excitatory synaptic inputs to AN2 and ON1. As shown in Fig. 1-7A and 1-8A,
264 AN2 and ON1 showed remarkable EPSPs and spiking responses to airflow
265 stimuli. In the individuals of which both cerci were ablated, both AN2 and
266 ON1 showed no change in their membrane potential in response to the airflow
267 stimuli (Fig. 1-10), meaning that the mechanosensory signals to AN2 and

268 ON1 are mediated by the cercal system.

269 1.4 Discussion

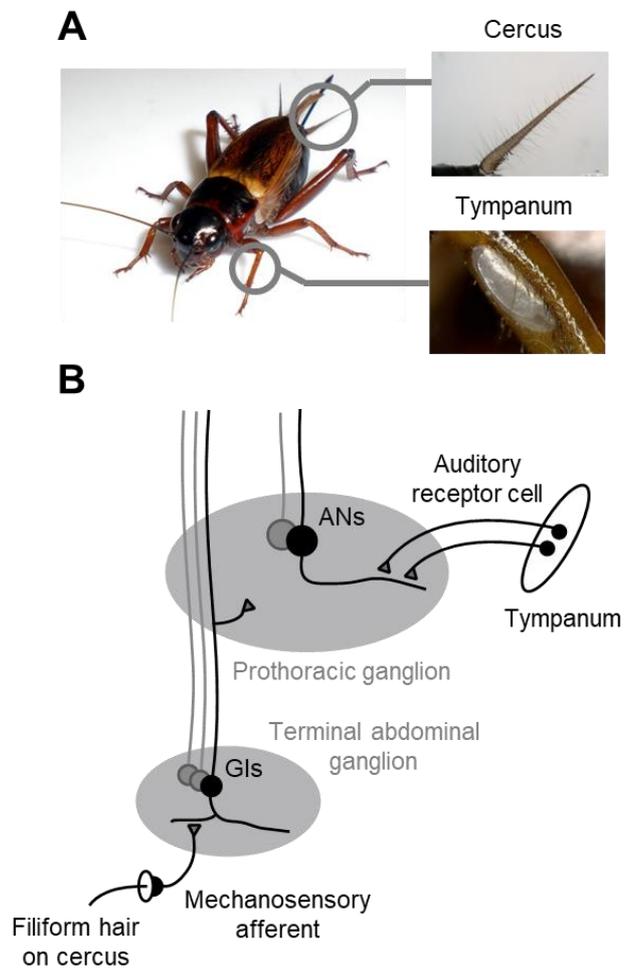
270 In this study, I demonstrated that certain types of prothoracic auditory
271 neurons including AN2 were sensitive to mechanosensory stimuli received via
272 the cercal organ. On the other hand, AN1 considered to be a key neuron to
273 elicit positive phonotaxis (Schildberger and Hörner, 1988) was insensitive to
274 the airflow. AN1 responds to calling songs of male crickets, of which carrier
275 frequency is about 5 kHz (Horseman and Huber, 1994), and induces oriented
276 walking behavior in female attracted by the male's song (Schildberger and
277 Hörner, 1988). In contrast, AN2 responds to ultrasound like echolocation call
278 of bats that prey on the crickets. Thus, our results suggest that the airflow
279 signals would influence the processing of auditory information relevant not
280 to mating but to predators. Further, I revealed that ON1 was also sensitive
281 to mechanosensory stimuli. This local interneuron inhibits activities of the
282 contralateral ON1, AN1 and AN2, resulting in the enhancement of the
283 directional selectivity in AN1 and AN2 (Selverston et al., 1985; Horseman
284 and Huber, 1994). The airflow-driven activity of ON1 might modulate the
285 directional selectivity of AN1 and AN2.

286 The interneurons AN2, ON1, ON2, TN1 and DN1, which were examined
287 in this study, have been identified as auditory neurons in the prothoracic
288 ganglion (Wohlers and Huber, 1982). Some previous studies have reported
289 that these auditory interneurons are also affected by other modalities of
290 sensory inputs. For example, AN2 and ON1 receive inhibitory inputs from

291 mechanoreceptors within the subgenual organ of legs which detects
292 mechanical vibration (Wiese, 1981; Kühne et al., 1984). TN1 and DN1
293 responds to wind stimuli (Atkins and Pollack, 1987). In addition to these
294 interneurons, certain T-shaped neurons, which extend both ascending and
295 descending axons, and descending neurons are sensitive to sound, wind and
296 light stimuli (Atkins and Pollack, 1987). These findings indicate that the
297 auditory information is not processed by modality-specific pathways. Rather,
298 the sensory systems within the prothoracic ganglion process multiple sensory
299 modalities. Notably, AN2 directly receive excitatory inputs from receptor
300 neurons in the tympanum (Hennig, 1988). Considering this previous finding
301 and my results together, inputs from multiple sensory modalities are
302 integrated in the early stage of sensory processing.

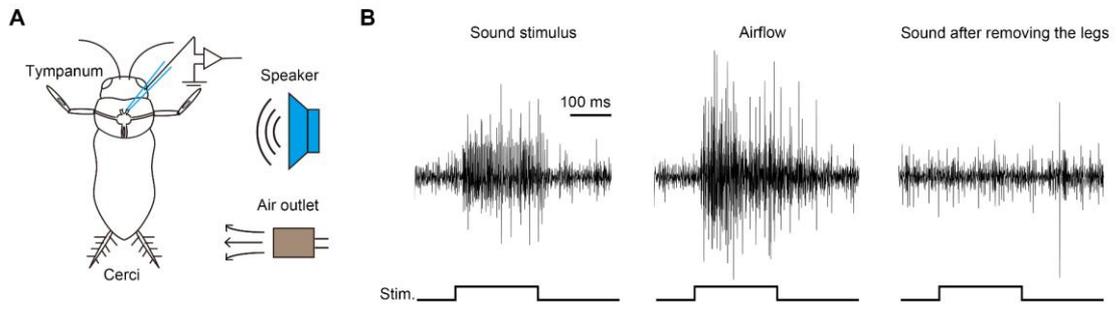
303 I demonstrated that the airflow inputs to AN2 and ON1 was mediated
304 by the cercal system. Neural pathways from the cerci to wind sensitive
305 auditory interneurons remain unclear. However, the wind-sensitive giant
306 interneurons (GIs), which project their ascending axons to the brain, arborize
307 their axon-collaterals into the thoracic ganglion (Hirota et al., 1993). Most of
308 the GIs show directional selectivity and prefer airflow from ipsilateral side to
309 their axons (Jacobs et al., 2008). ON1 also had directional preference for the
310 airflow from ipsilateral side to its soma, which suggests that GIs with their
311 axons ipsilateral to ON1's cell body provide airflow inputs to ON1.

312 Fig. 1-1.



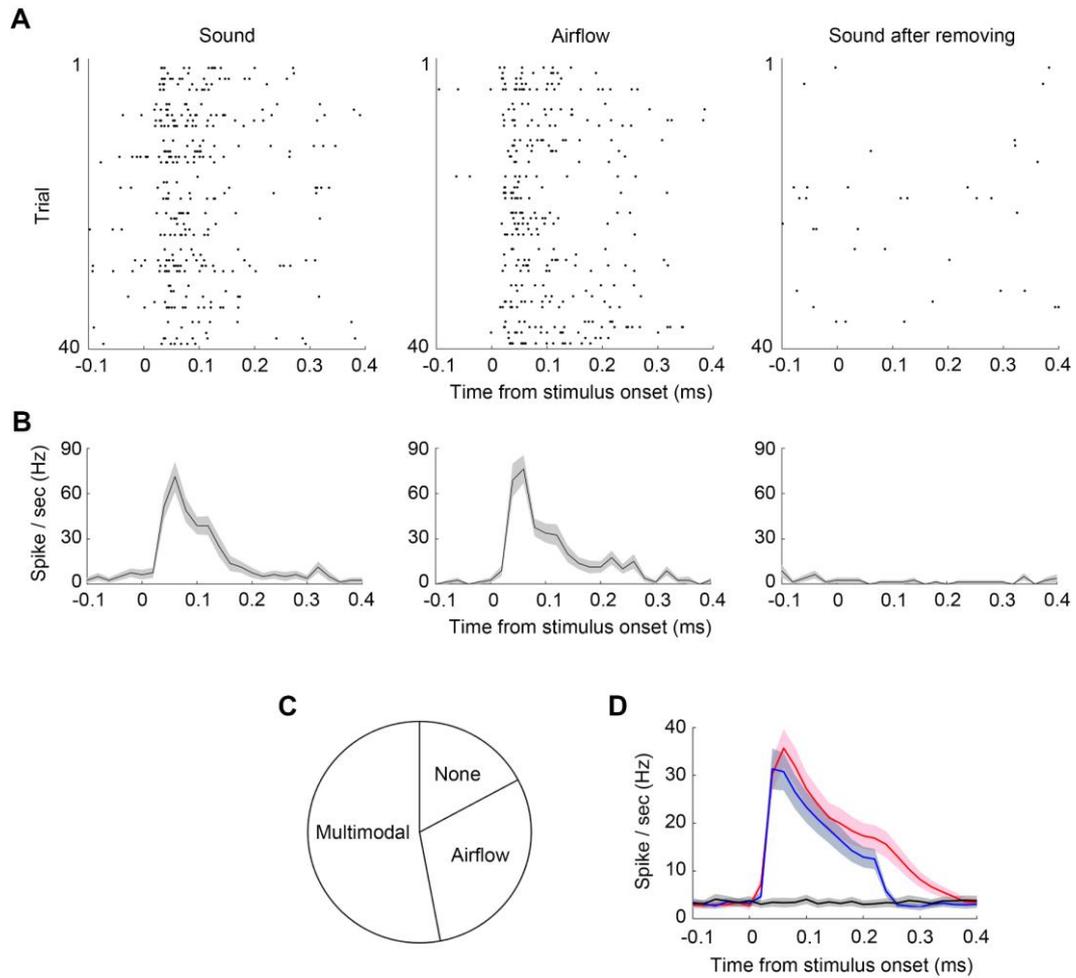
313 Figure 1-1. Auditory and cercal sensory systems of crickets. A) Two sensory
314 organs, tympanum and cercus. B) Diagram indicating the auditory and
315 cercal-sensory pathways. GIs, wind-sensitive giant interneurons; ANs,
316 auditory ascending neurons.

317 Figure 1-2



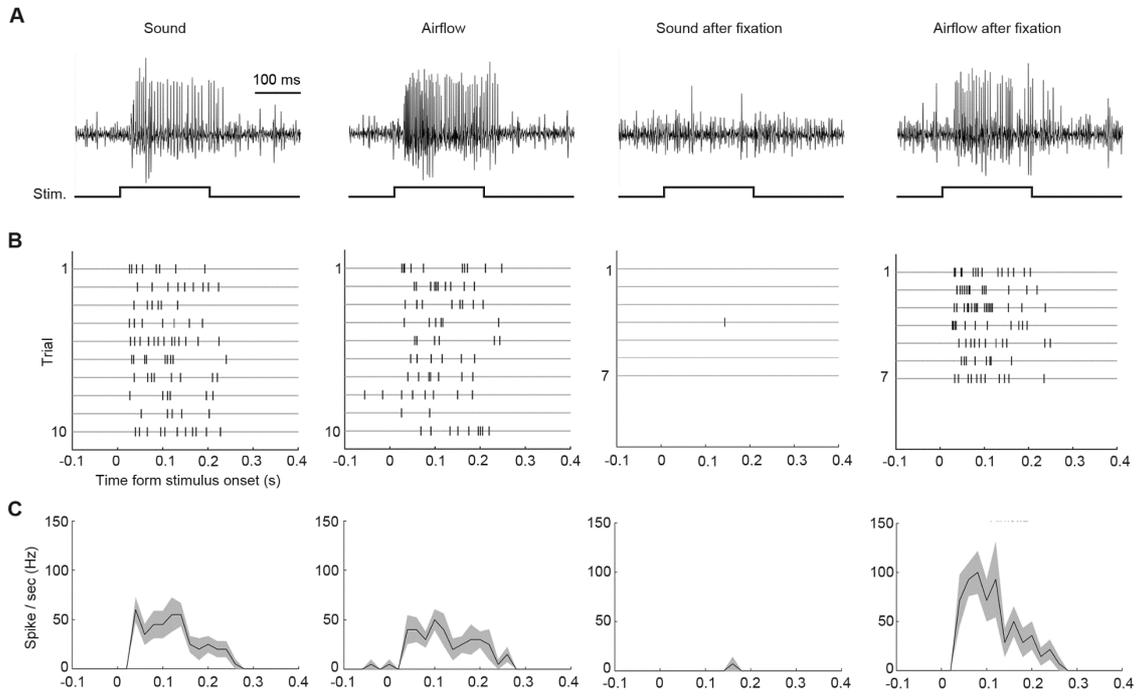
318

319 Figure 1-2. Extracellular recordings of ascending neurons. A) Experimental
320 setup for extracellular recording and stimulation. B) Typical ascending
321 activities evoked by sound (left panel) and airflow (middle). Right panel shows
322 sound-evoked response after both frontal legs were removed. Lower
323 rectangular traces indicate the stimulation.



326 Figure 1-3. Ascending spike units responding to airflow and sound stimuli. A)
327 Raster plot of a typical multimodal unit sensitive to sound and airflow. Each
328 dot indicates individual spike for sound (left), airflow (middle) and sound
329 stimulations after the frontal legs were removed (right). B) Time course of
330 firing rates of the unit shown in A. Stimulation in each column corresponds
331 to that in Fig. 1-3A. C) Fraction of multimodal, airflow-specific, and non-
332 responsive spike units in all recorded units. Each unit was labeled whether
333 its firing rate during the stimulation was significantly different from
334 spontaneous firing rate (paired t-test and Holm-Bonferroni correction). D)
335 Time course of mean firing rate of all multimodal units for sound (blue),
336 airflow (red), and sound stimulation after leg removal (black).

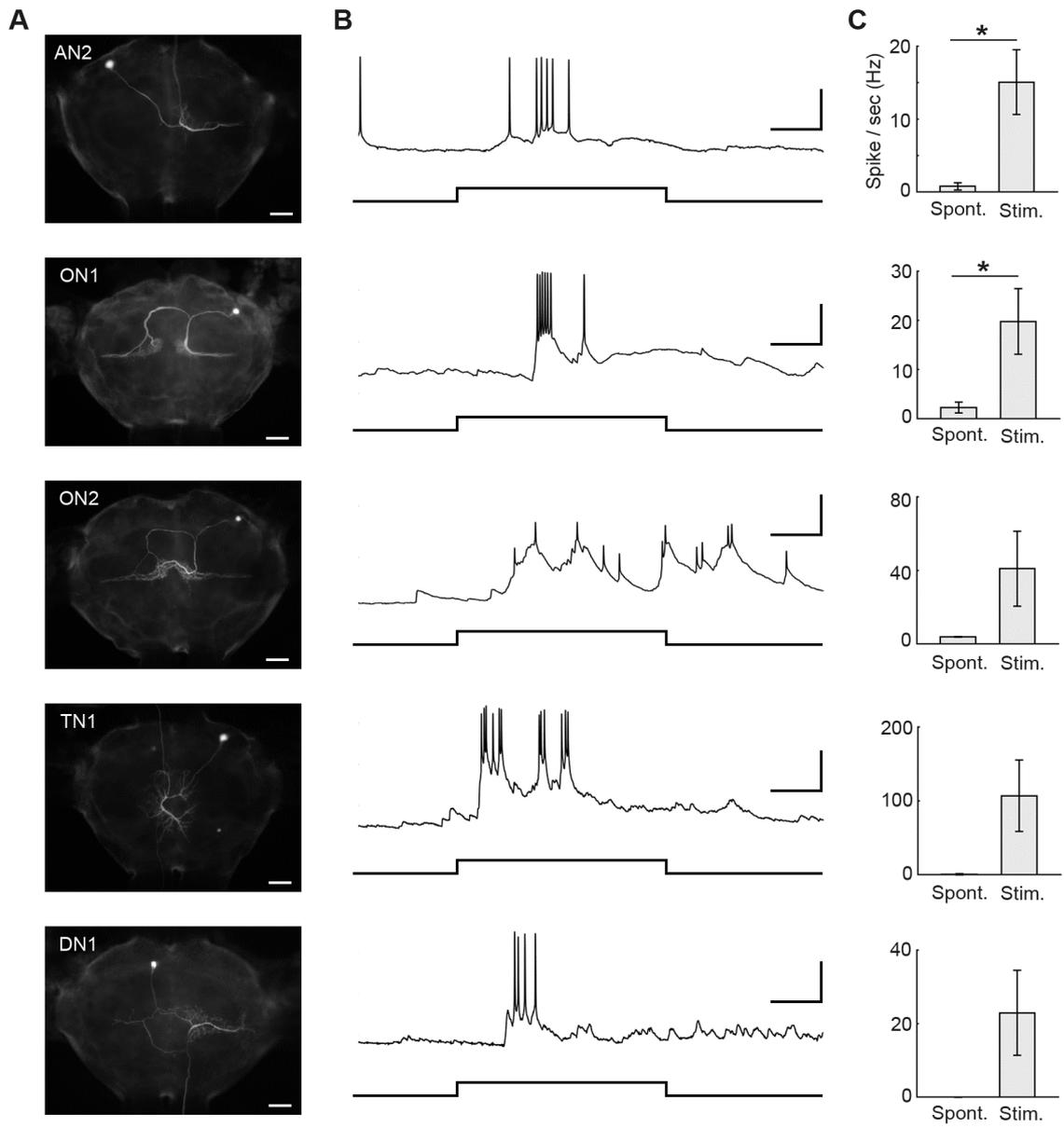
337 Figure 1-4



338

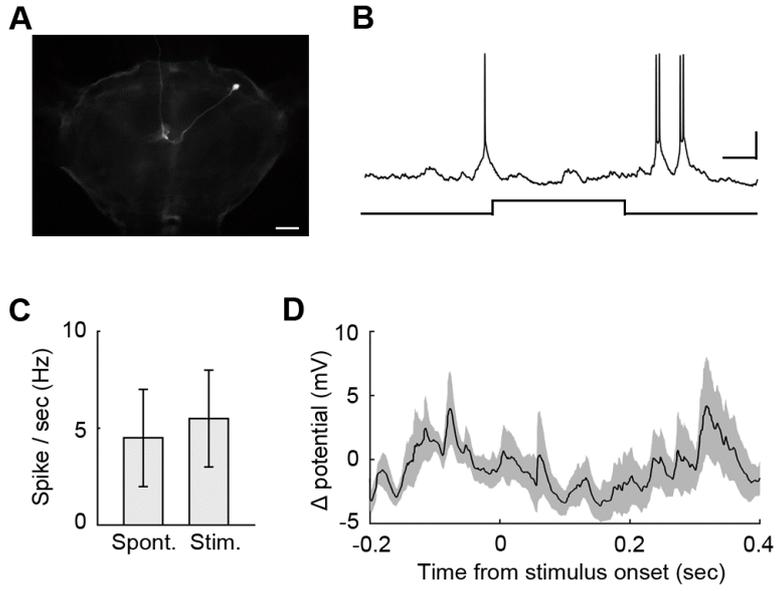
339 Figure 1-4. Single-unit recording of ascending neurons from the split nerve
340 cord. A) Raw traces of typical ascending activities for sound and airflow
341 stimulations in naïve (left two panels) and auditory-blocked (right two panels)
342 crickets in which both tympanums were gummed up. B), C) Raster plots (B)
343 and time course of the firing rate (C) of typical multimodal unit shown in Fig.
344 1-4B. Stimulation for each column corresponds to that in Fig. 1-4A.

345 Figure 1-5



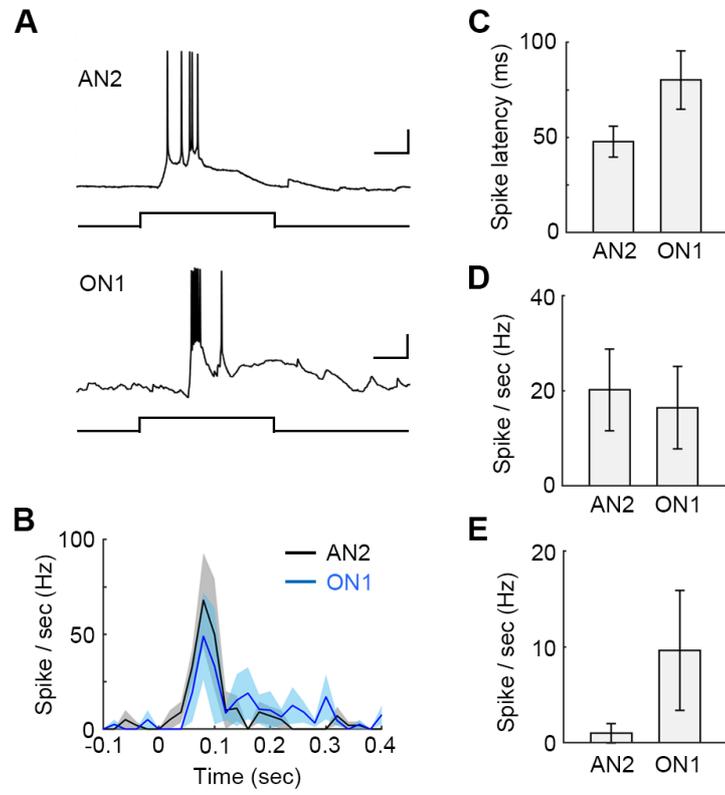
346 Figure 1-5. Airflow-evoked membrane-potential responses in identified
347 auditory neurons. A) Fluorescent images of auditory neuron AN2, ON1, ON2,
348 TN1 and DN1 recorded intracellularly. Scale bars indicate 100 μm . B) Typical
349 traces of membrane potential during the airflow stimulation. Lower
350 rectangular traces indicate the airflow stimulation. Vertical and horizontal
351 scale bars indicate 20 mV and 50 ms, respectively. C) Mean firing rate during
352 the spontaneous activity (Spont) and airflow stimulation (Stim), each of
353 which was measured as mean value for 200 ms before or during the airflow
354 stimulation. Sample sizes were 11 AN2s, 8 ON1s, 2 ON2s, 3 TN1s and 3 DN1s
355 (paired t-test, $p_{AN2} = 0.06 \times 10^{-1}$, $p_{ON1} = 0.04$, $p_{TN1} = 0.16$ and $p_{DN1} = 0.18$).
356 The cell type in each row corresponds to that in Figure 1-5A. Velocity of
357 airflow stimulus was from 0.17 to 0.43 m/s.

358 Figure 1-6



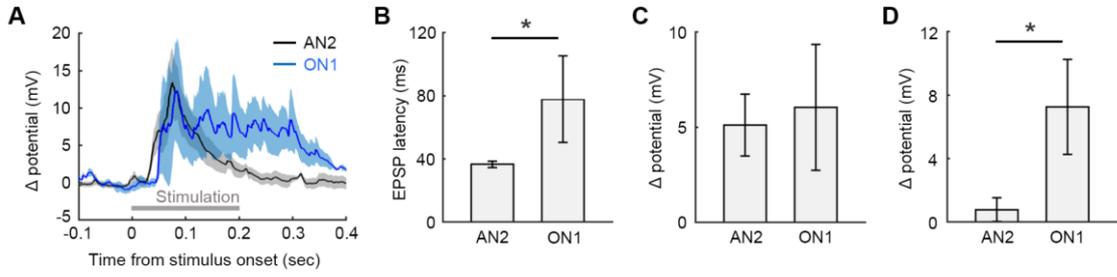
359

360 Figure 1-6. Neural response of AN1 to airflow stimulus. A) Fluorescent image
361 of AN1. Scale bar indicates 100 μm . B) Typical trace of membrane potential
362 in AN1 for airflow stimulation. Vertical and horizontal scale bars indicate 10
363 mV and 50 ms, respectively. C) Firing rate during spontaneous activity
364 (Spont) and stimulation (Stim) ($n = 2$ AN1s). D) Subthreshold change in the
365 membrane potential in AN1. After action potentials were removed using a
366 median filter with 5-ms window, the difference in the filtered membrane
367 potential from the average of resting potentials before stimulation was
368 calculated. Velocity of airflow stimulus was 0.43 m/s, and stimuli were
369 delivered from contralateral side to AN1's soma.



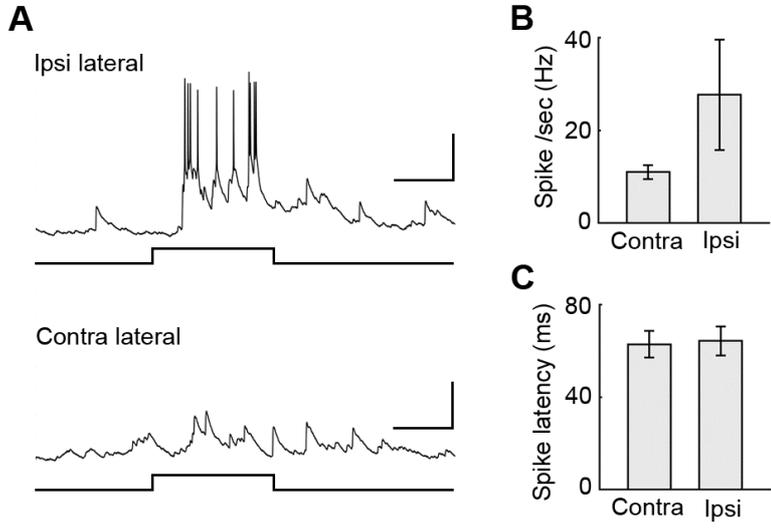
371 Figure 1-7. Firing-response properties of AN2 and ON1 for airflow
372 stimulation. A) Typical traces of membrane potentials in AN2 and ON1.
373 Vertical and horizontal scale bars indicate 10 mV and 50 ms, respectively. B)
374 Time course of firing rate in AN2 and ON1 ($n = 5$ neurons respectively
375 through Fig. 1-7). C) Latency from stimulus onset to the evoked first spike
376 (Wilcoxon rank sum test, $p = 0.10$). D) Firing rates in the earlier phase within
377 200 ms after the stimulus onset (unpaired t -test, $p = 0.77$). E) Firing rates in
378 the later phase which was time period from 200 to 300 ms after the stimulus
379 onset (unpaired t -test, $p = 0.24$). Velocity of airflow stimulus was 0.18 m/s.
380 Stimuli were delivered from ipsilateral side to ON1's soma and from
381 contralateral side to AN2's soma.

382 Figure 1-8



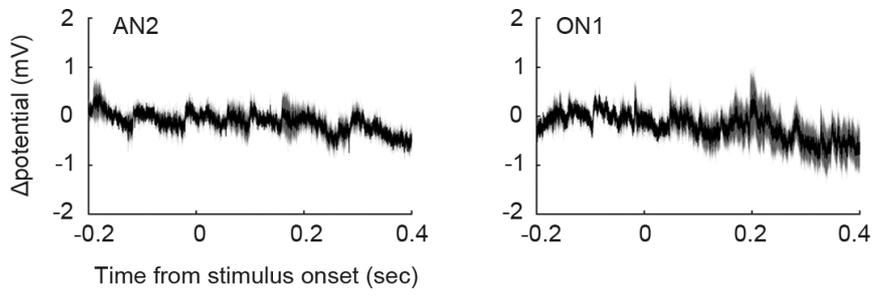
383

384 Figure 1-8. Subthreshold membrane-potential response properties of AN2
385 and ON1 for airflow stimulation. A) Time courses of subthreshold activity in
386 AN2 (black; $n = 5$ neurons) and ON1 (blue; $n = 3$ neurons). Gray line indicates
387 the airflow stimulation. B) Latency of EPSPs evoked by airflow stimulus
388 (sample size was the same as shown in A through B-D; Wilcoxon rank sum
389 test, $p = 0.04$). C) Mean postsynaptic potentials in the earlier phase within
390 200 ms after the stimulus onset (unpaired t-test, $p = 0.78$). D) Mean
391 potentials in the later phase which was time period from 200 to 300 ms after
392 the stimulus onset (unpaired t-test, $p = 0.04$). Velocity and direction of
393 stimulus are the same as shown in Fig. 1-7.



398 Figure 1-9. Directional selectivity to airflow stimulation in ON1. A) Typical
399 traces of membrane potentials in ON1 during airflow stimuli from ipsil-
400 (upper) or contralateral (lower) side to its soma. B) Mean firing rates in
401 responses to ipsil- (Ipsi) or contralateral (Contra) stimulation ($n = 3$ neurons;
402 paired t -test, $p = 0.29$). C) Latency from the stimulus onset to the first spike
403 for ipsil- or contralateral stimulation ($n = 3$ neurons; Wilcoxon signed-rank
404 test, $p = 0.75$). Velocity of airflow stimulus was 0.18 m/s.

405 Figure 1-10



406

407 Figure 1-10. Airflow-evoked responses in AN2 and ON1 without cerci. Time
408 courses of difference in the raw membrane potentials in AN2 and ON1 for
409 airflow stimulation after the both cerci were ablated. Velocity of airflow
410 stimulus was 0.17 m/s.

411 **Chapter II**

412 **Multisensory Integration in an Auditory Neuron AN2**

413 **2.1 Introduction**

414 In the previous chapter, I demonstrated that AN2 was sensitive to airflow
415 stimuli to cerci. AN2 is one of the primary auditory interneurons that receives
416 direct synaptic inputs within the prothoracic ganglion from auditory receptor
417 neurons of the tympanal organ located in the tibia of the front legs (Wohlers
418 and Huber, 1982; Hennig, 1988). This neuron is sensitive to high-frequency
419 sounds (Wohlers and Huber, 1978; Popov and Markovich, 1982), and its high-
420 frequency firing responses can trigger an avoidance steering response in
421 flying crickets (Nolen and Hoy, 1986). Therefore, AN2 is considered to be a
422 neuron that detects the presence of bats. The location of the sound source and
423 its intensity and duration are encoded by ‘burst firing’ of AN2 with short
424 inter-spike intervals, the behavioral functions of which have been well
425 studied (Marsat and Pollack, 2005, 2006, 2010). My previous finding of the
426 airflow-sensitivity of AN2 suggests that AN2 could detect other modality of
427 sensory clue of a bat’s attack, such as strong turbulent airflow caused by the
428 bat’s wingbeats.

429 It has been reported that the simultaneous presentation of different
430 modal stimuli, termed as cross-modal stimulation, elicits enhancement or
431 depression of spiking responses comparing to those responses to each
432 component stimulus (Meredith and Stein, 1983). Furthermore, enhancement
433 in cross-modal stimulation results in nonlinear (supra- or sub-linear) or linear
434 summation of unimodal responses (Stein and Stanford, 2008). In this chapter,

435 I examined multisensory integration in the spiking outputs of AN2 in
436 response to the cross-modal stimulus combining the sound and airflow.

437 In addition, cluster of spikes in short intervals, called bursts, plays an
438 important role in the sensory signaling by AN2. The burst firing of AN2 can
439 improve the detection of sound stimuli and the representation of sound-source
440 location (Marsat and Pollack, 2006). The fine temporal structure of the burst
441 firing represents additional stimulus-related information such as intensity
442 and duration of sound (Marsat and Pollack, 2010). Thus, I also examined
443 effect of the multisensory integration on burst firings in AN2.

444 **2.2 Materials and Methods**

445 See the method in Chapter I about *Animals* and *Intracellular recordings*.
446 Throughout the experiment in this chapter, 4 or 8% Lucifer yellow CH lithium
447 salt in 200-mM LiCl was used for intracellular recordings.

448

449 *Stimulation*

450 See Methods in Chapter I about the apparatuses for airflow and sound
451 stimulations. The velocity of airflow was 0.17 m/s at the position of cerci and
452 the duration was 200 ms. For sound stimulation, only 15-kHz pure tone was
453 used.

454 For unimodal stimulation of the cerci, single air puffs were used (Fig. 2-
455 1). Different manners of stimulation were used for experiments with
456 repetitive stimulation or various auditory intensities. In the repetitive-
457 stimulation experiments that were performed to test multisensory
458 integration, 9 sound pulses at 60 dB were given with 500-ms inter-stimulus
459 interval, and airflow was provided coincident with the 4th sound pulse. To
460 analyze multisensory effects, I compared the responses to the 3rd sound pulse
461 with those to the combined stimuli. All responses to sound pulses (1st-3rd and
462 5th-9th) were used for comparing burst firing with non-burst firing. In the
463 experiments with varying auditory intensities that were performed to test the
464 effects of input amplitude, single sound pulses of 50, 60, 70, 80, or 90 dB were
465 applied for the unimodal stimulation, and a single air puff combined with

466 various intensities of sound pulses was applied for the cross-modal
467 stimulation. All stimuli were delivered contralateral to the soma of the
468 recorded AN2. For the cross-modal stimuli with both types of stimulation,
469 differences in the stimulus onset between the sound pulse and airflow were <
470 15 ms, and the onset of the sound stimulus was defined as the stimulus onset
471 for cross-modal stimuli.

472

473 *Data analysis*

474 In the repetitive stimulation experiments, neural responses to sound pulses,
475 airflow, and combined stimuli were recorded from 8 neurons in different
476 crickets. In the various intensities experiments, the responses to sound pulses
477 were recorded from 4 neurons in different crickets, and those to airflow and
478 combined stimuli were obtained from 3 out of these 4 neurons. In addition to
479 these experiments, responses to airflow stimuli were also recorded from 5
480 neurons. In the cerci ablation experiments, neural responses to airflow
481 stimuli were recorded from 2 neurons in 2 crickets. Spikes were detected
482 using custom-written MATLAB routines (R2016a, MathWorks,
483 RRID:SCR_001622). Raw traces of membrane potential were filtered with a
484 150-Hz high-pass filter, and spike peaks were detected based on the filtered
485 traces using the findpeaks function in MATLAB. To analyze the time course
486 of the firing rate, spike trains were binned at a size of 1 ms and convolved
487 with a moving-average filter with a 20-ms window. Summation of the

488 responses (r_{sum}) to the uni-modal stimulus was defined as

489
$$r_{sum} = r_{sound} + r_{airflow}$$

490 where r_{sound} and $r_{airflow}$ indicate the firing rate in response to sound or
491 airflow stimulus, respectively. Linearity of the multisensory integration was
492 defined as

493
$$Linearity = \frac{r_{cross}}{r_{sum}}$$

494 where r_{cross} indicates the firing rate in the response evoked by the cross-
495 modal stimulus. Histogram of inter-spike intervals (ISI) was fitted to a kernel
496 probability distribution using normal kernel smoothing with a 0.3-ms
497 bandwidth. To test the ISI distribution results from a Poisson process, the ISI
498 histogram was also fitted to a single exponential curve. A burst was defined
499 as a cluster of spikes separated by short intervals (< 6.5 ms; Marsat and
500 Pollack 2006). The first and last spike within a burst were defined as the
501 onset and offset of the burst, respectively. Size of the burst was defined as a
502 product of the burst duration and the number of spikes within the burst
503 (Marsat and Pollack, 2010).

504 2.3 Results

505 2.3.1 Multisensory enhancement in AN2 spiking activity

506 To examine the multisensory effects of simultaneous inputs with different
507 modalities on the spiking activities of AN2, I compared neural responses to
508 single air puffs, the 3rd sound pulses, and cross-modal stimulation combining
509 an air puff with the 4th sound pulse (Fig. 2-2A). Cross-modal stimuli elicited
510 significantly larger numbers of spikes than unimodal stimuli did, but the
511 amplitudes of the responses to the cross-modal stimuli were similar to the
512 arithmetic sum of the responses to the 3rd sound pulse and air puff that were
513 separately presented (Fig. 2-2A, B). The linearity of the responses to cross-
514 modal stimuli was examined using various intensities of sound combined with
515 airflow, the velocity of which was constant (Fig. 2-2C). Higher intensities of
516 sound stimuli elicited greater spiking responses in both uni- and bimodal
517 stimulus conditions, but the responses to cross-modal stimuli were close to
518 the arithmetic sum of the responses to airflow and to sound pulses regardless
519 of the sound intensity. To verify the effects of the sound intensity on the
520 linearity of the summation in the cross-modal responses, I examined the
521 relationship between the linearity of summation and firing rates in the
522 summed responses to unimodal stimuli (Fig. 2-2D). Linearity depended on
523 the magnitude of the summed responses, and supra-linear summation was
524 observed only in small responses in which the firing rate was lower than 10
525 Hz. This suggests that AN2 linearly and additively integrates auditory and

526 mechanosensory input, and supra-linear summation occurred in the
527 integration of small sensory inputs evoked by stimuli around the threshold.

528 The linearity of summation in the cross-modal responses could depend
529 on the stimulus intensity (Meredith and Stein, 1986; Stein and Stanford,
530 2008) but also on the latency from stimulus onset (Rowland and Stein, 2007;
531 Rowland et al., 2007). Thus, I examined the temporal profile of the
532 summation linearity. The cross-modal responses were larger in their firing
533 rate than sound-evoked responses (Fig. 2-3A) and the arithmetic sum of
534 unimodal responses (Fig. 2-3B) during the period of 30–60 ms after stimulus
535 onset. However, there was no significance in the firing rates in this period
536 between the cross-modal and summed responses ($n = 8$ neurons, paired t-test,
537 $p = 0.07$, the firing rates were 85.05 ± 10.44 Hz for cross-modal and $66.61 \pm$
538 13.00 Hz for the summed responses, respectively). In addition, even if I
539 focused on this period, the supra-linear summation occurred only in the small
540 cross-modal responses (Fig. 2-3C). The supra-linear summation would simply
541 result from the fact that summed inputs of the subthreshold unimodal stimuli
542 were larger than the spike threshold and evoked action potentials. Our
543 results suggest that AN2 linearly sums auditory and mechanosensory inputs
544 and that supra-linearity cross-modal summation would arise from the spike
545 threshold.

546

547 **2.3.2 Multisensory enhancement in burst firing of AN2**

548 In previous studies (Marsat and Pollack, 2006, 2007), the responses of AN2
549 to 30-kHz long-duration tones with Gaussian amplitude envelopes showed a
550 characteristic distribution of inter-spike intervals (ISI), with a peak at short
551 intervals and broad tail of longer intervals. Based on this ISI distribution, the
552 spike trains with intervals shorter than 6.5 ms are classified into ‘bursts,’
553 which are used for encoding precise ultrasound information (Marsat and
554 Pollack 2006, 2010). In the current study, ISI distribution of the multiple
555 spikes that were evoked in 58% of responses (170/294 in 17 individuals) to 15-
556 kHz tone pulses at 60 dB showed similar features, which included a peak at
557 short intervals and a peaky but broad tail at longer intervals (Fig. 2-4A). In
558 addition, the bimodal peaks in ISI distribution in our data could be also
559 separated at 6.5 ms intervals. To confirm that the peak at the shorter
560 intervals indicated the distribution of the burst firing, I fitted a single
561 exponential curve to the whole ISI histogram. If the peaks surpass this curve,
562 these activities were considered to be burst firings different from the ISI
563 distribution expected by Poisson process (Bastian and Nguyenkim, 2001;
564 Marsat and Pollack, 2012). The distinct peak at the intervals shorter than 6.5
565 ms was higher than the exponential curve, indicating that the 15-kHz tone
566 pulse stimulus enabled AN2 to generate the burst firings, which could be
567 separated from the isolated spikes using the same criteria as in previous
568 studies.

569 I examined effects of cross-modal stimuli on generation of burst firing.

570 Burst firing responses were more frequently evoked by cross-modal stimuli
571 than by unimodal stimuli (Fig. 2-5A). To clarify whether this enhancement of
572 burst firing activity was specific to multisensory integration or simply
573 resulted from increased firing rates due to linear summation, I evaluated the
574 effects of response magnitude on the probability of burst occurrence among
575 cross- and unimodal responses. The larger the spiking responses were, the
576 higher the probability of burst occurrence was regardless of the sensory
577 modality (Fig. 2-5B). Further, there was little difference among the uni- and
578 cross-modal responses in the dynamic ranges of the curves, that indicated a
579 relationship between the response magnitude and the bursting probability.
580 This means that multisensory integration does not specifically affect the
581 generation of burst firing. Rather, more frequent burst firing would result
582 simply from an increase in spiking responses via linear summation of
583 auditory and mechanosensory inputs.

584 I also focused on temporal aspects of burst generation (Fig. 2-5C). Most
585 of the bursts occurred following the first action potentials evoked by the
586 auditory and cross-modal stimuli. In contrast, the bursts evoked by the
587 airflow stimuli followed several isolated spikes to appear in the middle of
588 spike trains. Taken together, these results illustrate that AN2 integrates
589 sound and airflow inputs and enhances the response magnitude, resulting in
590 burst firing activity at the beginning of responses. However, the bursts were
591 also enhanced by stronger unimodal stimuli, and no difference in the timing

592 of burst generation was observed between the auditory and cross-modal
593 responses.

594

595 **2.3.3 Effect of temporal coincidence of the stimuli on the burst firing**

596 In multisensory integration, the neural responses to the cross-modal
597 stimulus were enhanced when the stimuli were presented at the same time
598 and/or provided from the same location (Holmes and Spence, 2005). This
599 spatio-temporal coincidence detection is considered as a mechanism to
600 precisely detect a single event or object presenting multiple sensory cues. To
601 examine whether AN2 responses to the cross-modal stimulus depended on
602 the timing of stimulus presentation, the sound and airflow stimuli were
603 provided in various intervals between them (Fig. 2-6A). The number of spikes
604 evoked by stimulus did not depend on the temporal difference in the stimulus
605 onset (Fig. 2-6B). However, the cross-modal stimuli elicited a single burst
606 when the airflow and sound stimuli were presented at approximately the
607 same time (Fig. 2-6C). When the stimulus onsets separated over 40 ms, AN2
608 generated multiple bursts. To focus on the temporal structure of burst, I
609 measured the duration and size, which is product of number of spikes within
610 burst and duration of burst and is positively correlated with magnitude of the
611 behavioral response (Marsat and Pollack, 2010). Figures 2-6D and 2-6E show
612 the relationships between the time lag between sound and airflow stimuli and
613 the maximum duration or maximum size of the burst, both of which were

614 measured from the largest burst in the response if multiple bursts were
615 evoked. Both duration and size became larger as the difference of stimulus
616 interval was shorter (Fig. 2-6D, E). Not only the maximum duration and size
617 of the individual burst, but the summation of size in all bursts elicited by a
618 single stimulation also depended on the difference of stimulus onset (Fig. 2-
619 6F, G). These results suggest that AN2 represents the coincidence of the
620 stimulus onsets by using the temporal structure of burst.

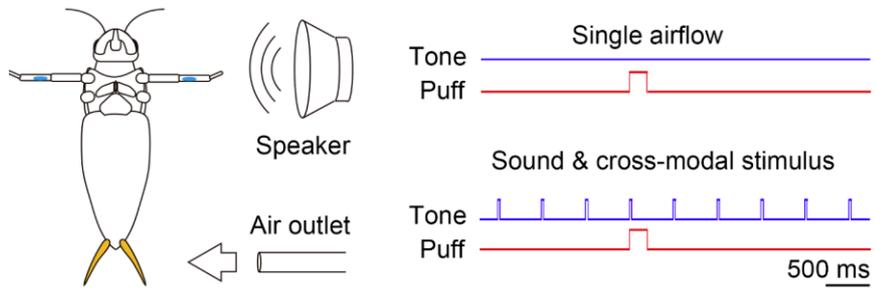
621 2.4 Discussion

622 I demonstrated that the cross-modal stimulation enhanced the magnitude of
623 firing activity. Further, simply put, the spike outputs evoked in AN2 by cross-
624 modal stimuli were equal to linear summation of those evoked by the
625 respective unimodal stimuli. Supra-linear summation in spiking activity was
626 observed only when the unimodal stimuli were as weak as the threshold for
627 generating spikes, suggesting that supra-linear summation would result from
628 the non-linear transduction from graded potentials to action potentials by
629 threshold properties of voltage-gated channels (Holmes and Spence, 2005).
630 Our result that supra-linear summation was observed only around spike
631 threshold is consistent with that in recent studies of the MSTd (dorsal medial
632 superior temporal) area of macaques (Morgan et al. 2008) and the superior
633 colliculus of cats (Perrault et al., 2003, 2005; Stanford, 2005). Because the
634 supra-linearity of the spike threshold is based on intrinsic active membrane
635 properties, the supra-linear summation of spike responses of AN2 would not
636 be specific to multisensory integration.

637 As well as the mean firing rate of the response, the burst firing was also
638 enhanced by the cross-modal stimulation. Considering that AN2 uses bursts
639 to encode the sound information such as intensity, duration, and location
640 (Marsat and Pollack, 2006, 2010), airflow stimulus would have some impacts
641 on acoustic representation in AN2. Although it is unclear the role of cercal
642 signals on the acoustic representation in AN2, multisensory integration may

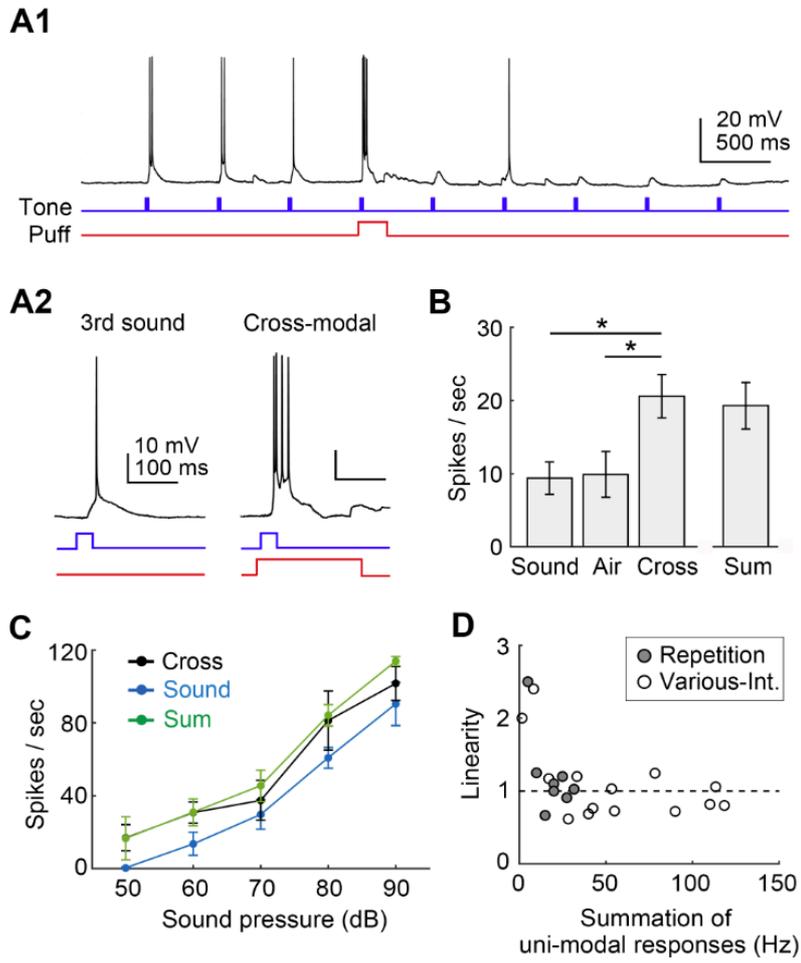
643 improve neural representation of the component stimulus (Ibrahim et al.,
644 2016; Atilgan et al., 2018).

645 Figure 2-1



646 Figure 2-1. Experimental setup (left) and stimulation protocol of repetitive-
647 stimulation experiments (right). Single short durations of airflow were
648 delivered separately (upper). Nine tone pulses were repetitively applied and
649 single airflow stimuli were combined with the 4th pulse (lower).

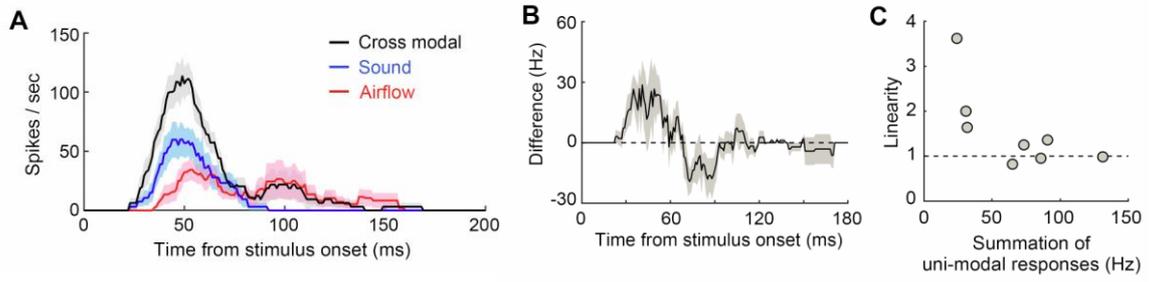
650 Figure 2-2



652 Figure 2-2. Multisensory enhancement of AN2 firing activity. A) Typical
653 responses to sound and cross-modal stimuli in the repetitive stimulation
654 experiments (A1), and the enlarged views of the responses to the 3rd sound
655 pulse and cross-modal stimulus combining the 4th sound pulse with airflow
656 (A2). B) Firing rates of responses to 3rd sound, airflow, and cross-modal
657 stimuli for 200 ms after stimulus onset. Cross-modal stimuli caused
658 significant increases in the firing rate than unimodal stimuli ($n = 8$ neurons,
659 one-way repeated measures ANOVA, $p = 0.02$; paired t-tests and Holm-
660 Bonferroni correction, cross vs. sound, $p = 0.03$; cross vs. airflow, $p =$
661 0.09×10^{-2} ; sound vs. air, $p = 0.91$). Right bar indicates the arithmetic sum
662 (Sum) of the firing responses to sound and airflow. There was no significant
663 difference in the firing rate between the sum of the unimodal responses and
664 the cross-modal responses ($n = 8$ neurons, paired t-test, $p = 0.39$). C) Sound
665 intensity-dependent curves of the responses to the sound (blue, $n = 4$ neurons),
666 cross-modal stimuli (black, $n = 3$ neurons excluding the point at 80 dB [$n = 2$
667 neurons]) and those predicted from the arithmetic sum of the unimodal
668 responses (green, $n = 3$ neurons). For cross-modal stimulation, the constant
669 velocity (0.17 m/s) airflow was combined with sound at various intensities (50
670 to 90 dB). D) Relationship between linearity of the cross-modal response and
671 the firing rate in summation of the unimodal responses to sound and airflow.
672 Filled circles indicate the mean value of data measured from repetitive
673 stimulation experiments for each individual ($n = 8$ neurons). Open circles

674 indicate the data of the response measured from various intensities of
675 stimulation experiments (n = 14 data points from 3 neurons). 'Linearity = 1'
676 shown by a dashed line indicates that the cross-modal response was equal to
677 the linear summation of the unimodal responses. 'Linearity > 1' or '< 1'
678 indicates supra- or sub-linear summation, respectively.

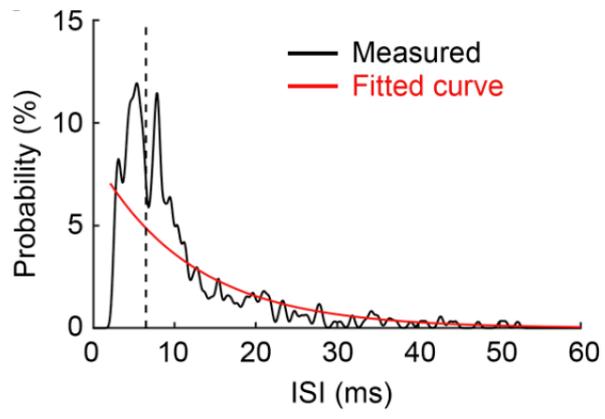
679 Figure 2-3



680

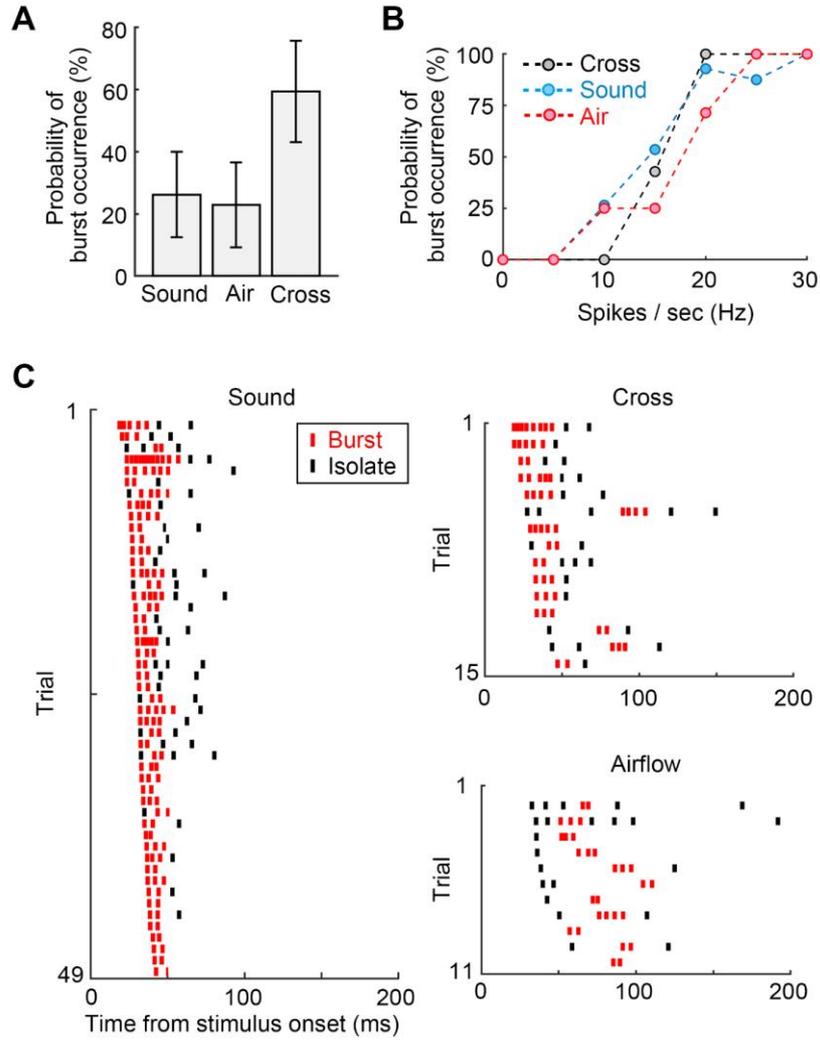
681 Figure 2-3. Dependency of linearity on latency from stimulus onset. A)
682 Variation in the mean firing rate over time in response to cross-modal (black),
683 sound (blue), and airflow (red) stimuli in the repetitive stimulation
684 experiments (n = 8 neurons). B) Difference in firing rate between cross-modal
685 responses and the summation of auditory and cercal responses (n = 8 neurons).
686 C) Relationship between linearity of the cross-modal response and the firing
687 rates in the summation of unimodal responses during the period of 10–60 ms
688 after the stimulus onset, which corresponds to the period of 30–60 ms shown
689 in F.

690 Figure 2-4

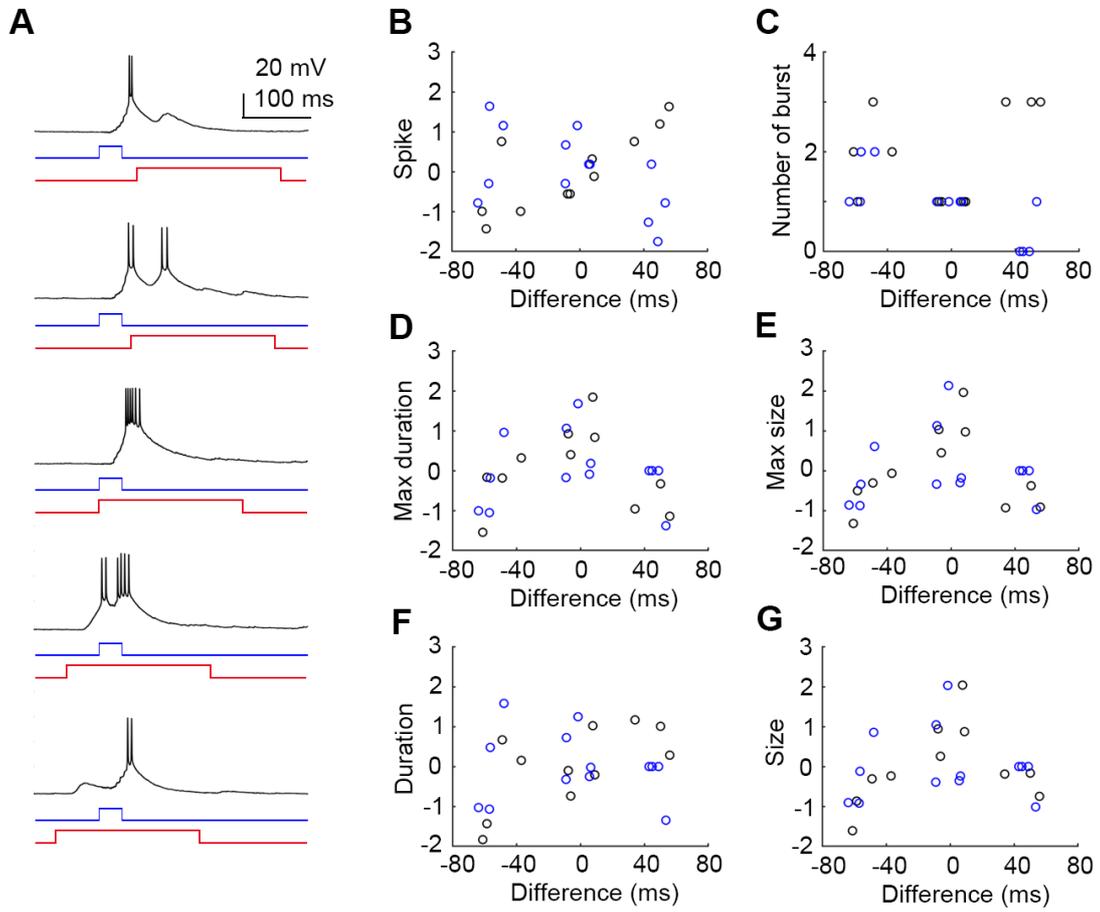


691 Figure 2-4. Distribution of ISIs measured from all spiking responses to 15-
692 kHz sound pulses of 60 dB (black; n = 424 ISIs from 16 neurons). A single
693 exponential curve was fitted to the whole distribution (red; $f(x|\mu) = \frac{1}{\mu} e^{-\frac{x}{\mu}}$,
694 $\mu = 11.87$). Dashed line indicates 6.5 ms ISI that was used for criterion of the
695 burst.

696 Figure 2-5



697 Figure 2-5. Multisensory enhancement of bursting. A) Probability of burst
698 occurrence in the unimodal and cross-modal responses in the repetitive-
699 stimulation experiments ($n = 8$ neurons, Friedman test, $p = 0.04$; Wilcoxon
700 signed-rank test, cross vs. sound, $p = 0.13$; cross vs. air, $p = 0.06$; air vs.
701 sound, $p = 1.00$). B) Dependency of the probability of burst generation on the
702 firing rate in uni- and cross-modal responses. The data of both uni- and cross-
703 modal responses were obtained from all samples in the repetitive stimulation
704 and various intensities stimulation experiments ($n = 11$ neurons for sound
705 and cross-modal) and the single airflow stimulation ($n = 16$ neurons for
706 airflow). The firing rates were measured for 200 ms after the stimulus onset.
707 C) Timing of burst firing in response to sound (left), cross-modal (upper right),
708 and airflow (lower right) stimuli. Red and black lines indicate bursting and
709 isolated spikes for individual trials, respectively. Each trial was sorted by the
710 latency of the first spike. The same samples as in C were used.



713 Figure 2-6. Dependency of the burst structures on temporal coincidence of
714 stimulus. A) Traces of membrane potentials in AN2 in response to the
715 combination of sound (blue) and airflow (red) stimuli with various inter-
716 stimulus intervals. Positive value means that the onset of sound precedes the
717 onset of airflow. B–F) Relationships between the time lag between the sound
718 and airflow stimulus onset and firing rates normalized as Z-score (B), number
719 of bursts (C), maximum burst duration (D), or maximum burst size (E). The
720 values of the burst structure in (E) and (E) were measured from the largest
721 burst in the response if multiple bursts were evoked. F, G) Summation of
722 duration (F) and size (G) of all evoked bursts. Color indicates the data
723 recording from different individuals through Fig. 2-6 from B to G. The max.
724 duration and size of the burst were normalized as Z-score through D to G.

725 **Chapter III**

726 **Neural Mechanisms Underlying Multisensory Enhancement in AN2**

727 **3.1 Introduction**

728 In the previous chapter, I demonstrated the effect of multisensory integration
729 on the spiking outputs of AN2. The cross-modal stimulus combining the sound
730 pulse and airflow enhanced spiking activities including bursting activity. And,
731 the linearity of summation of the multisensory integration depended on the
732 magnitude of the evoked responses. As discussed in the previous chapter, it
733 was possible that supra-linear summation would result from the non-linear
734 transduction from graded potentials to action potentials. To reveal the
735 cellular mechanisms relevant to multisensory enhancement in AN2, I focused
736 on sub-threshold changes in the membrane potential evoked by uni- and
737 cross-modal stimuli. Also, I examined the modulation of spike threshold that
738 affects spike outputs. In several sensory neurons, fast membrane
739 depolarization lowers the spike threshold, which is known as a dynamic
740 threshold (Azouz and Gray, 2003; Wilent, 2005; Jeanne and Wilson, 2015).
741 The dynamic threshold helps neurons to detect a coincidence of inputs (Azouz
742 and Gray, 2003; Jeanne and Wilson, 2015).

743 In this chapter, I investigated the difference of subthreshold activity
744 among modalities, and between bursting and isolated spikes. And I showed
745 the modulation of spike threshold by temporal pattern of action potentials.

746

747 **3.2 Materials and Methods**

748 Data, which was obtained by the experiments shown in Chapter II, was used
749 for the analysis. See Methods in Chapter II about *Animals, Intracellular*
750 *recordings* and *Stimulation*.

751

752 *Data analysis*

753 A burst was defined as a cluster of spikes separated by short intervals (< 6.5
754 ms; Marsat and Pollack, 2006). This criterion was applicable to my data as
755 shown by figure 2-4. The first and last spike within a burst were defined as
756 the start and end of the burst, respectively. To analyze subthreshold voltage
757 traces, raw traces of the recordings were filtered by a median filter with a 5-
758 ms window (Shi et al., 2017). To focus on the time variation of EPSPs, the
759 extracted post-synaptic potentials were normalized by the potentials at the
760 peak time of the first spike.

761 I adopted the following procedure to detect spike thresholds (Jeanne and
762 Wilson, 2015). After filtering using a moving-average filter with 150- μ m
763 window, the spike threshold was defined as the membrane potential
764 immediately preceding a spike that was detected as the inflection point in the
765 plot of membrane potential and the slope of the potential, dV/dt . The
766 inflection point was defined as the point where the d^2V/dt^2 fell below the 10%
767 maximum of d^2V/dt^2 in the stereotypical spike trajectory. The rate of
768 depolarization was calculated as the slope of the filtered voltage trace for 1.5

769 ms before the spike threshold. The inter-spike interval (*ISI*) of the n^{th} spike
770 within a spike train was calculated from the interval between the $(n - 1)^{th}$
771 and n^{th} spike.

772 MATLAB software (MathWorks, RRID:SCR_001622) was used for data
773 processing and computation. All values are reported as mean \pm standard error
774 of the mean. A p -value of < 0.05 was considered to be statistically significant.

775 **3.3 Results**

776 **3.3.1 Cross-modal stimulation maintains depolarization following initial**
777 **spiking**

778 First, I compared evoked EPSPs in response to cross- and unimodal stimuli.
779 It was technically difficult to suppress the spike generation of AN2 by
780 injecting minus current through a sharp electrode because of cell size and
781 geometry. Instead, the EPSPs were extracted from traces of membrane
782 potential changes using a 5-ms median filter (Fig. 3-1A). For EPSPs evoked
783 by airflow, the membrane potential was transiently depolarized with longer
784 latency and decreased for 150 to 200 ms after stimulus onset even though
785 airflow was continuously applied (Fig. 3-1B). Cross-modal stimuli evoked
786 larger EPSPs than either unimodal stimulus types did (Fig. 3-1B), but there
787 was no significant difference in the peak value of EPSPs among the stimulus
788 modalities (peak potentials for cross-modal, sound, and airflow stimulation
789 were 16.04 ± 2.11 , 13.10 ± 2.28 , and 12.78 ± 1.60 mV, respectively; $n = 8$
790 neurons, one-way repeated measures ANOVA, $p = 0.31$). In contrast, the
791 EPSPs evoked by cross-modal stimuli were significantly larger in their mean
792 during the transient elevation (0-150 ms after the stimulus onset) than those
793 evoked by the sound and airflow stimuli (Fig. 3-1C). However, EPSP evoked
794 by the cross-modal stimuli were much smaller in both peak and mean of
795 potential changes than the arithmetic sum of the EPSPs evoked by sound and
796 airflow stimuli, implying that the subthreshold potentials were sub-linearly

797 summed in AN2.

798 As shown in figure 2-5C, the cross-modal stimuli enhanced spiking
799 activity to evoke burst firing more frequently at the beginning of responses
800 compared to unimodal stimuli, suggesting differences in depolarization
801 following the first spikes in response to cross-modal versus unimodal stimuli.
802 To examine this possibility, I aligned the EPSP traces with the peak of their
803 first spikes and found that EPSPs evoked by cross-modal stimuli lasted longer
804 and had more sustained depolarization compared to EPSPs evoked by sound
805 or airflow alone (Fig. 3-2A). To quantify this, I measured the average value of
806 normalized changes in membrane potential over 6.5 ms following the first
807 spike (Fig. 3-2B). This corresponded to a border line of ISIs for separating
808 bursts and isolated spikes and probably affected the burst firing and offset of
809 the sustained peak (Fig. 3-2C), defined as the time required EPSPs to
810 decrease from 100% to 90% of the normalized potential at the first spike
811 (shown by the dashed line in Fig. 3-2A). Cross-modal stimuli evoked slightly
812 larger depolarizations after the first spike compared to unimodal stimuli, but
813 this difference was not significant (Fig. 3-2B). In contrast, the offset of peak
814 for cross-modal stimuli was significantly delayed compared to those for
815 unimodal stimuli (Fig. 3-2C). These results demonstrate that cross-modal
816 stimuli evoke longer-lasting EPSPs than unimodal stimuli.

817 The followings two factors could be considered as causes resulting in the
818 elongation of EPSP following the first spikes: the first was the time lag in the

819 response latency to the sound and airflow, the other was mechanical
820 fluctuation of the stimulus onsets of the airflow. To test the first possibility, I
821 measured the latency of the first spikes for each type of unimodal stimuli. The
822 latency of the airflow-evoked spike was significantly longer than sound- and
823 cross-modal stimulus-evoked spikes (Fig. 3-3A), meaning that the airflow
824 responses were delayed to the auditory response. From this quantification of
825 the response latencies, the time shift between the sound-evoked and airflow-
826 evoked responses was 24.10 ± 15.81 ms ($n = 6$ neurons), which was longer
827 than the offset of peak in EPSP evoked by the cross-modal stimuli ($13.88 \pm$
828 3.61 ms shown in 4F, $n = 8$ neurons), but no significance (Wilcoxon rank sum
829 test, $p = 0.57$). In addition, the variance of the peak offsets in the cross-modal
830 responses was much smaller than that of the time shift between the auditory-
831 and airflow-evoked first spikes (104.31 ms for the peak offset and 1500.18 ms
832 for the time lag, respectively). These facts suggest that the long-lasting peak
833 of the cross-modal responses would be partly resulted from the time difference
834 of auditory and cercal inputs, but this would not be all cause for the EPSP
835 elongation. In regard to the second possibility, I found no relationship
836 between the time lag between the sound and airflow stimulus onsets and the
837 depolarization size (Fig. 3-3B) or peak offset (Fig. 3-3C) in the EPSP evoked
838 by cross-modal stimuli, indicating that mechanical lag of the unimodal
839 stimuli had no effects on the EPSP shape.

840

841 **3.3.2 Subthreshold activity accounts for burst firing in response to cross-**
842 **modal stimuli in AN2.**

843 Our results revealed differences in the time course of depolarization following
844 the first spike in response to cross-modal and unimodal stimuli. However, it
845 was unclear whether differences in EPSPs accounted for differences in burst
846 firing. To investigate this hypothesis, I compared EPSPs accompanied by
847 burst firing and by non-burst firing (isolated spikes). In responses to
848 unimodal (sound) and cross-modal stimuli, both of which evoked bursts at the
849 beginning of their responses (Fig. 2-5C), EPSPs accompanying bursts were
850 larger than those accompanying isolated spikes (Fig. 3-4A1, A2). As shown in
851 Fig. 3-2A, subthreshold potentials were normalized to the voltage at the base
852 of the first spike and aligned with the peak time of the first spike (Fig. 3-4B1,
853 B2), and the mean potentials over 6.5 ms following the first spike as well as
854 the peak offset were measured. There were significant differences in mean
855 potential changes between burst firing and isolated spikes evoked by
856 unimodal and cross-modal stimuli (Fig. 3-4C). The peak offset for burst firing
857 was also significantly delayed compared to that for isolated spikes in response
858 to the unimodal and cross-modal stimuli (Fig. 3-4D). These results suggest
859 that longer-lasting depolarization following the first spike accounted for the
860 generation of burst firing regardless of sensory modality.

861 I further tested whether these parameters of the subthreshold potential
862 are also correlated with the temporal size of bursts. Both the mean potential

863 change after the first spike and the peak offset were significantly correlated
864 with burst duration. The correlation coefficient between the peak offset and
865 the burst duration was higher (0.71) than that between the mean potential
866 change and burst duration (0.33) (Fig. 3-5A, B). However, distribution of the
867 plots for auditory and cross-modal stimuli overlapped each other. These
868 results indicated that the size and persistence of the depolarization after
869 individual spikes were key factors in determining burst generation, and that
870 the persistence of depolarization could affect the duration of the burst firing.
871 Further, both the size and persistence of EPSPs were enhanced not only by
872 multisensory integration but also by higher stimulus intensity regardless of
873 sensory modality.

874

875 **3.3.3 Inter-spike interval modulates spike threshold but modality don't.**

876 Spike output depends on the spike threshold in addition to the magnitude of
877 EPSPs. It has been reported that in some sensory systems such as *Drosophila*
878 olfactory neurons and mammalian visual cortical neurons, rapid membrane
879 depolarization decreases the spike threshold, creating a dynamic threshold
880 (Azouz and Gray, 2000, 2003; Jeanne and Wilson, 2015). To examine whether
881 cross-modal stimuli modulates the spike threshold in AN2, I compared spike
882 thresholds between the cross-modal and uni-modal stimulus conditions. I
883 found that the spike threshold was dynamically altered, especially during
884 burst firing (Fig. 3-6A, B); however, there was no difference in first spike

885 thresholds among the different stimulus modalities (Fig. 3-6C). Next, I
886 examined which factors are related to the spike threshold. The rate of
887 depolarization immediately preceding individual spikes was poorly correlated
888 with the spike threshold (Fig. 3-6D). In contrast, inter-spike interval (ISI)
889 was negatively correlated with the spike threshold regardless of the
890 stimulation modality (Fig. 3-6E). In summary, spike threshold was
891 dynamically altered in accordance with ISI in AN2, whereas the sensory
892 modality had no effect on the spike threshold.

893

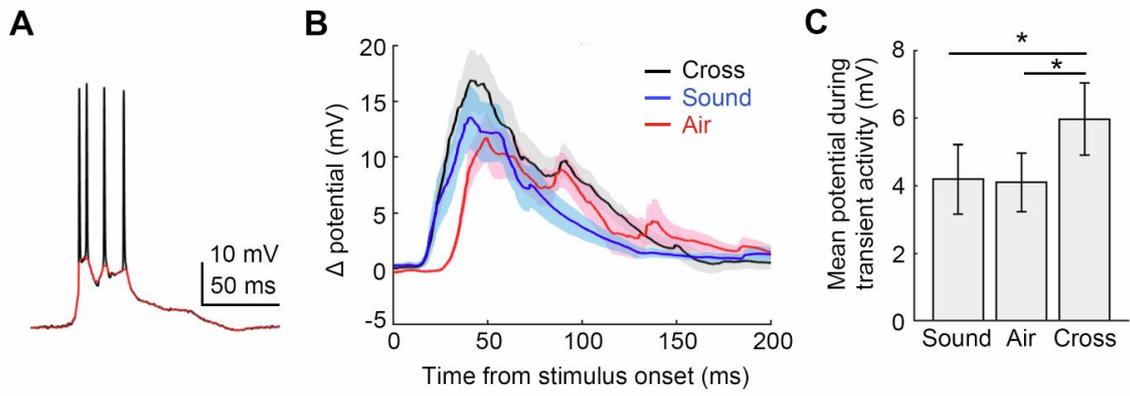
894 3.4 Discussion

895 I demonstrated that bimodal stimuli combining sound and airflow evoked
896 linearly or supra-linearly summed spike outputs. In contrast, different
897 modalities of sensory inputs were sub-linearly summed as reflected by EPSPs.
898 Many studies on linearity in synaptic responses to cross-modal stimulation
899 have revealed various types of computation including supra-linear, sub-linear,
900 and linear summations. In neurons in the optic tectum of *Xenopus*,
901 multimodal inputs are supra-linearly summed in the evoked EPSPs that
902 result from NMDA receptor activation (Truszkowski et al., 2017). The linear
903 summation of EPSPs is observed in the superior colliculus of rats (Skaliora et
904 al., 2004), and sub-linear summation has been reported in the lamprey optic
905 tectum and mouse neocortex (Olcese et al., 2013; Kardamakis et al., 2016).
906 Thus, linearity of summation of EPSPs evoked by multimodal stimulation
907 varies among the brain regions and animals. In previous studies, linear
908 summation was observed as changes in membrane potential of a few
909 millivolts from resting potentials (Skaliora et al. 2004), whereas sub-linear
910 summation has been observed when multimodal stimuli evoke larger EPSPs
911 close to the spike threshold (Olcese et al. 2013). In the present study, the
912 analyzed EPSPs that were extracted from spiking responses were ‘over-
913 threshold’ depolarizations by over 10 mV. Therefore, sub-linear summation in
914 the synaptic potentials of AN2 might result from active properties of the cell
915 membrane, which are mediated by voltage-gated cation channels.

916 In addition, I found the elongation of EPSP evoked by cross-modal
917 stimuli that might result from time difference of auditory- and cercal-sensory
918 inputs. AN2 receives direct excitatory inputs from the auditory receptor
919 afferents (Hennig 1988) whereas cercal sensory inputs are mediated by at
920 least 2 synapses that include synapses from mechanoreceptor afferents to
921 cercal projection neurons such as GIs and that from those projection neurons
922 to AN2. It is possible that the cercal sensory pathway may involve in other
923 prothoracic interneurons. AN2 might apply the time difference of
924 multisensory inputs to elongate the compound EPSP generating the burst
925 firings.

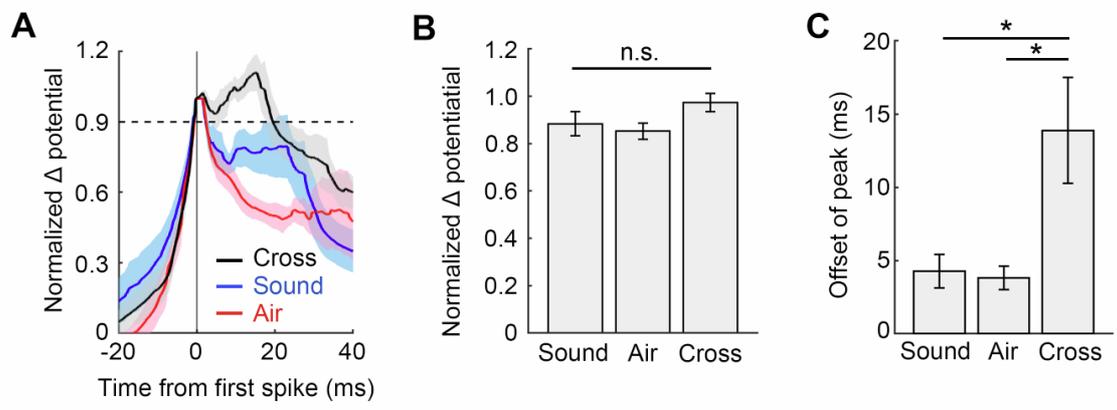
926 The modulation of spike threshold has been observed in some sensory
927 neurons (Azouz and Gray, 2003; Wilent, 2005). In these neurons, the rate of
928 depolarization modulates the spike threshold. In AN2, the rate of
929 depolarization and sensory modality did not affect the spike threshold
930 whereas the spike threshold was modulated by the inter-spike interval. This
931 intrinsic property might prevent the generation of inappropriate bursts.

932 Figure 3-1



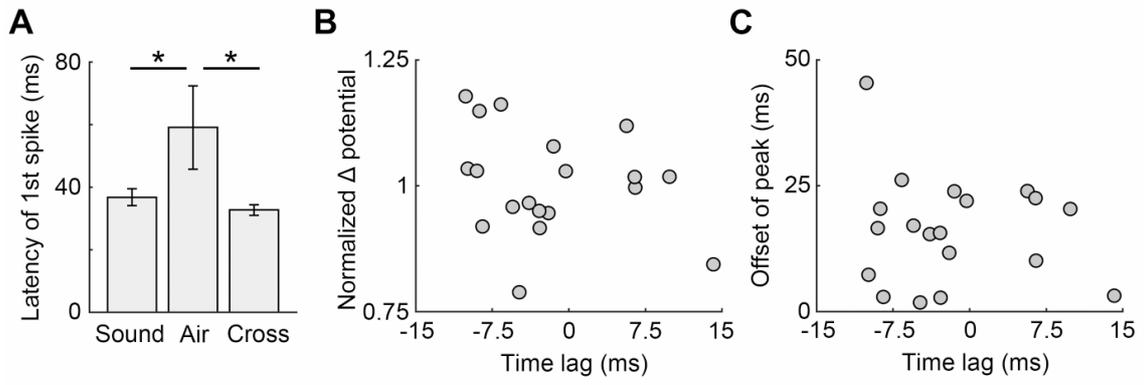
933 Figure 3-1. Excitatory post-synaptic potentials (EPSPs) evoked by cross-
934 modal and unimodal stimuli. A) Extracted PSPs. Action potentials were
935 removed from the raw trace of membrane potential (black trace) using a 5-ms
936 median filter to estimate the graded potential of PSPs (red trace). B) Time
937 variation in the membrane potential changes in EPSPs evoked by cross-modal
938 (black), sound (blue), and airflow (red) stimuli in repetitive stimulation
939 experiments (n = 8 neurons). C) Mean changes of membrane potential in
940 EPSPs for 150 ms after the stimulus onset (n = 8 neurons, Friedman test,
941 $p = 0.01$; Wilcoxon signed-rank test with Holm-Bonferroni correction, cross vs.
942 sound, $p = 0.47 \times 10^{-1}$; cross vs. air, $p = 0.02$; sound vs. air, $p = 0.84$).

943 Figure 3-2

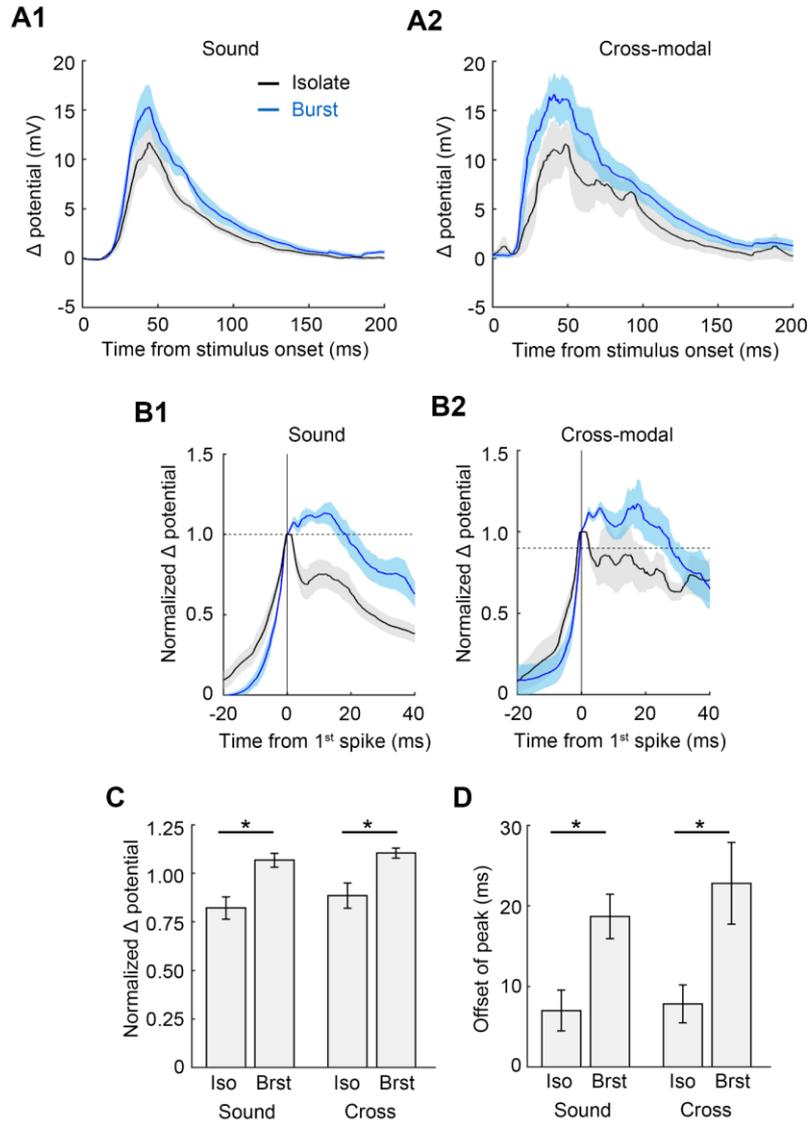


944 Figure 3-2. EPSPs relevant to first spike for stimulus. A) Time variation in
945 graded potential changes in EPSPs following the first spike. EPSP traces
946 were temporally aligned to peak time of the first spike and normalized in
947 magnitude by a value of the graded potentials at that moment (n = 8, 7, and
948 7 neurons for cross-modal, sound, and airflow, respectively). The dashed line
949 indicates 90% of the EPSP at the time of the first spike (0 ms). E, F) Mean
950 values of the normalized changes in membrane potential over 6.5 ms
951 following the first spike (E; n = 8, 7, and 7 neurons for cross-modal, sound,
952 and airflow, respectively. Kruskal-Wallis test, $p = 0.14$) and the offset of peak
953 EPSPs (F; n = 8, 7, and 7 neurons for cross-modal, sound, and airflow,
954 respectively, One-way factorial ANOVA, $p = 0.01$; unpaired t-test with Holm-
955 Bonferroni correction, cross vs. sound, $p = 0.02$; cross vs. air, $p = 0.02$; sound
956 vs. air, $p = 0.47$) evoked by different modalities of stimuli. The offset of peak
957 was measured as the time when EPSPs were > 90% of the normalized EPSP
958 indicated by the broken line in D.

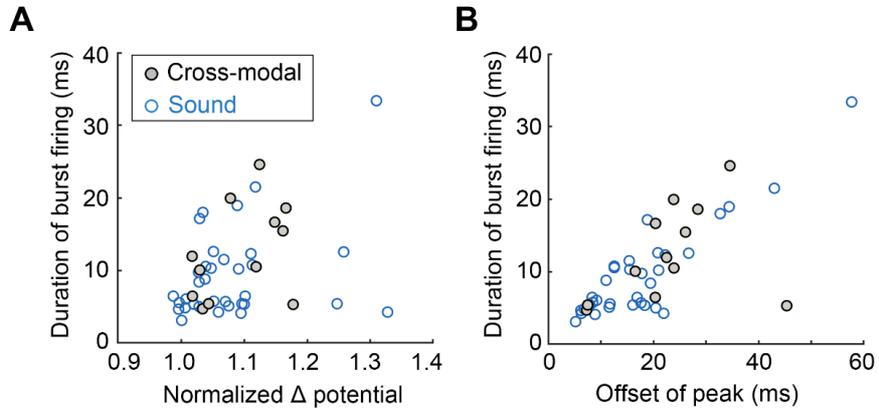
959 Figure 3-3.



960 Figure 3-3. Effects of the time difference in response latency and in stimulus
961 onset on EPSP's properties. A) Latency of the first spikes evoked by different
962 types of stimulation from the stimulus onset. The spike latency of the airflow
963 response was significantly longer than those of sound and cross-modal
964 responses (n = 7, 7, and 8 neurons for sound, airflow and cross-modal
965 stimulations, respectively, Kruskal-Wallis test, $p = 0.04 \times 10^{-1}$; Wilcoxon
966 rank sum test with Holm-Bonferroni correction, sound vs. air, $p = 0.04$; cross
967 vs. air, $p = 0.04 \times 10^{-1}$; sound vs. cross, $p = 0.28$). B, C) Relationships
968 between the time lag between the sound and airflow stimulus onset and the
969 normalize membrane potentials within 6.5 ms after the 1st spike peak (B) or
970 the offset of peak (C) in EPSPs evoked by cross-modal stimuli (n = 19
971 responses from 8 neurons, Spearman's rank correlations, $r = -0.32$, $p = 0.18$
972 for B, $r = -0.04$, $p = 0.89$ for C). Negative value of the time lag indicates
973 that the airflow preceded the combined sound stimulus onset.

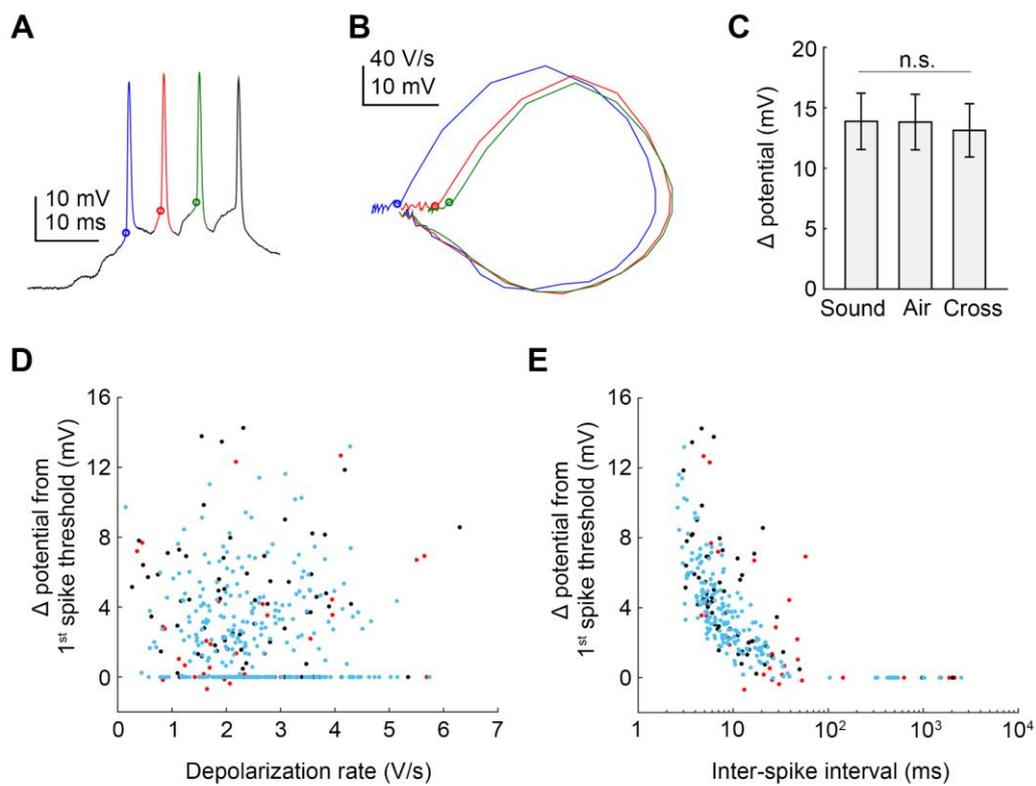


975 Figure 3-4. Excitatory post-synaptic potential (EPSP) characteristics
976 affecting burst firing. A) Extracted EPSPs accompanied by isolated spikes
977 (black) or burst firing (blue) in response to sound (A1) and cross-modal (A2)
978 stimuli. All data used for Figure 3-4 were obtained from the responses to uni-
979 and cross-modal stimuli using 60-dB sound in both the repetitive stimulation
980 and various intensities of stimulation experiments. Sample sizes used for A
981 to D were $n = 8$ and 5 neurons for isolated spikes and bursts evoked by sound
982 stimuli, and $n = 9$ and 7 neurons for isolated spikes and bursts evoked by
983 cross-modal stimulation, respectively. B) Time variation of the graded
984 potential changes in EPSPs following the first spike evoked by sound (B1) or
985 cross-modal (B2) stimuli. Colors of lines correspond to those in A. C, D) Mean
986 values of the normalized changes in membrane potential for 6.5 ms after the
987 first spike (C; Wilcoxon rank sum test, isolated vs. burst for sound stimuli,
988 $p = 0.03 \times 10^{-1}$; isolated vs. burst for cross-modal stimuli, $p = 0.05 \times 10^{-1}$)
989 and of the offset of peak in EPSPs (D; unpaired t-test, isolated vs. burst for
990 sound stimuli, $p = 0.01$; isolated vs. burst for cross-modal stimuli, $p = 0.01$)
991 accompanied by isolated spikes or bursts in response to sound and cross-
992 modal stimuli.
993



995 Figure 3-5. EPSP characteristics relevant to structure of burst. A, B)
996 Relationship between the duration of burst firing and the mean of normalized
997 potential changes (A; $n = 46$ burst responses from 10 neurons, Spearman's
998 rank correlation, $r = 0.33$, $p = 0.02$) or the offset of peak EPSPs (B; sample
999 size was the same as in A, Spearman's rank correlation, $r = 0.71$; $p =$
1000 0.03×10^{-6}). Blue open circles represent data for sound stimuli and gray filled
1001 circles represent data for cross-modal stimuli.

1002 Figure 3-6



1003 Figure 3-6. Modulation of the spike threshold in AN2. A) Trace of a typical
1004 firing response and estimated spike threshold potentials. Blue, red, and green
1005 circles indicate the threshold for the first, second, and third spikes,
1006 respectively. B) Phase portraits (dV/dt versus V) for the 3 spikes shown in A.
1007 Colors of traces correspond to those in A. C) Thresholds of the first spikes
1008 evoked by different modalities of stimuli ($n = 7, 7$ and 8 neurons; One-way
1009 factorial ANOVA, $p = 0.81$). Delta potentials of spike thresholds were the
1010 differences between the threshold potential and resting membrane potential
1011 before stimuli. D, E) Relationship between spike threshold and the preceding
1012 depolarization rate (D; Spearman's rank correlation, $r_{cross} = -0.20$, $r_{sound} =$
1013 -0.07 , and $r_{airflow} = 0.05$; $p_{cross} = 0.08$, $p_{sound} = 0.22$, and $p_{airflow} = 0.73$) or
1014 inter-spike interval (E; Spearman's rank correlation, $r_{cross} = -0.84$, $r_{sound} =$
1015 -0.92 , and $r_{airflow} = -0.68$; $p_{cross} = 1.59 \times 10^{-21}$, $p_{sound} = 1.98 \times 10^{-119}$, and
1016 $p_{airflow} = 4.00 \times 10^{-7}$). Sample size shown in D was the same as shown in E
1017 ($n = 293, 44$ and 78 spike thresholds from $8, 7$ and 8 neurons in sound, air
1018 and cross-modal stimulations respectively). The depolarization rate was
1019 measured as the slope of the change in membrane potential for 1.5 ms prior
1020 to each spike-threshold point. Delta (Δ) potential from first spike threshold
1021 was measured as the difference between the threshold potential and the first
1022 spike threshold. Colors of dots in D and E indicate data for cross-modal (black),
1023 sound (blue), and airflow (red) stimuli.

1024

1025 **General Discussion**

1026 Multisensory integration is fundamental computation observed in various
1027 sensory systems and different brain regions (Meredith and Stein, 1986;
1028 Ghazanfar and Schroeder, 2006; Deeg et al., 2009; Zahar et al., 2009;
1029 Kardamakis et al., 2016). Multisensory integration is performed in not only
1030 higher processing stage for association but also earlier stages close to sensory
1031 surface such as the primary auditory cortex of macaque and the primary
1032 visual cortex of mouse (Schroeder and Foxe, 2005; Ghazanfar and
1033 Chandrasekaran, 2007; Lakatos et al., 2007; Ibrahim et al., 2016). In
1034 *Drosophila* larvae, certain sensory neurons directly receive sensory inputs
1035 from receptor cells of different modalities (Ohyama et al., 2015). The
1036 prothoracic auditory circuit of crickets is also such a sensory system to
1037 integrate various modalities in the early stage as discussed in Chapter I.
1038 These findings imply that multisensory integration is fundamental
1039 mechanisms in early stage of sensory processing.

1040 Finally, I refer to the behavioral relevance of multisensory integration
1041 in AN2. AN2 is considered to play a crucial role in the detection of predatory
1042 bats, because it is sensitive to ultrasound-like echolocation calls of bats, and
1043 its firing activity is necessary and sufficient to trigger avoidance steering in
1044 the opposite direction of the sound source (Nolen and Hoy, 1984, 1986).
1045 Notably, bursting of AN2 correlates with abdominal movement in the
1046 avoidance steering behavior (Marsat and Pollack 2006, 2010). I demonstrated

1047 that AN2 generates bursting in response to 15-kHz tone pulses of which the
1048 carrier frequency was lower than that used in previous studies (Marsat and
1049 Pollack 2006, 2007, 2010), and that cross-modal stimuli combining the tone
1050 pulse with airflow enhanced the generation of bursting. AN2 represents
1051 directionality, intensity, and duration of sound stimuli with its bursting
1052 (Marsat and Pollack, 2006, 2010). Our results, therefore, suggest that
1053 multisensory integration in AN2 may enable quicker and more precise
1054 identification of alert signals via simultaneous detection of the searching echo
1055 and air flow caused by flying bats.

1056 The avoidance steering driven by the bursts of the AN2 are observed
1057 only in response to ultrasound during flight. So, it remains unclear whether
1058 flying crickets use wind stimuli as a sign of a bat's attack. Several types of
1059 insects, such as mantids and cockroaches, detect the predator's attack by
1060 their cercal organs and execute avoidance steering during flight (Ganihar et
1061 al., 1994; Triplehorn and Yager, 2006). It is possible that the wind sensitivity
1062 of AN2 might be useful for successful escape via multisensory integration
1063 with auditory inputs. On the other hand, crickets standing on the ground
1064 exhibit escape behaviors such as running or jumping in response to wind
1065 stimuli (Tauber and Camhi, 1995; Oe and Ogawa, 2013; Sato et al., 2017).
1066 AN2 is also involved in the phonotaxis via detection of conspecific calls
1067 (Schildberger and Hörner, 1988), meaning that AN2 could respond to
1068 ultrasound and also to airflow in standing crickets. Recently, I reported that

1069 10- or 15-kHz pure tones that will activate AN2 modulate wind-elicited escape
1070 behavior in crickets (Fukutomi et al., 2015; Fukutomi and Ogawa, 2017).
1071 Taken together, the findings in this study suggest that AN2 is an excellent
1072 model for understanding the neuroethological significance of multisensory
1073 integration.

1074 **Reference**

- 1075 Alais D, Burr D (2004) The ventriloquist effect results from near-optimal
1076 bimodal integration. *Curr Biol* 14:257–262.
- 1077 Atilgan H, Town SM, Wood KC, Jones GP, Maddox RK, Lee AKC, Bizley JK
1078 (2018) Integration of visual information in auditory cortex promotes
1079 auditory scene analysis through multisensory binding. *Neuron* 97:640–
1080 655.
- 1081 Atkins G, Pollack GS (1987) Response properties of prothoracic,
1082 interganglionic, sound-activated interneurons in the cricket *Teleogryllus*
1083 *oceanicus*. *J Comp Physiol A* 161:681–693.
- 1084 Azouz R, Gray CM (2000) Dynamic spike threshold reveals a mechanism for
1085 synaptic coincidence detection in cortical neurons in vivo. *Proc Natl Acad*
1086 *Sci* 97:8110–8115.
- 1087 Azouz R, Gray CM (2003) Adaptive coincidence detection and dynamic gain
1088 control in visual cortical neurons in vivo. *Neuron* 37:513–523.
- 1089 Bastian J, Nguyenkim J (2001) Dendritic modulation of burst-like firing in
1090 sensory neurons. *J Neurophysiol* 85:10–22.
- 1091 Deeg KE, Sears IB, Aizenman CD (2009) Development of multisensory
1092 convergence in the xenopus optic tectum. *J Neurophysiol* 102:3392–3404.
- 1093 Ernst MO, Banks MS (2002) Humans integrate visual and haptic information
1094 in a statistically optimal fashion. *Nature* 415:429.
- 1095 Fetsch CR, DeAngelis GC, Angelaki DE (2013) Bridging the gap between

1096 theories of sensory cue integration and the physiology of multisensory
1097 neurons. *Nat Rev Neurosci* 14:429–442.

1098 Fotowat H, Gabbiani F (2007) Relationship between the phases of sensory
1099 and motor activity during a looming-evoked multistage escape behavior.
1100 *J Neurosci* 27:10047–10059.

1101 Fukutomi M, Ogawa H (2017) Crickets alter wind-elicited escape strategies
1102 depending on acoustic context. *Sci Rep* 7:15158.

1103 Fukutomi M, Someya M, Ogawa H (2015) Auditory modulation of wind-
1104 elicited walking behavior in the cricket *Gryllus bimaculatus*. *J Exp Biol*
1105 218:3968–3977.

1106 Ganihar D, Libersat F, Wendler G, Camhi JM (1994) Wind-evoked evasive
1107 responses in flying cockroaches. *J Comp Physiol A* 175:49–65.

1108 Ghazanfar AA, Chandrasekaran CF (2007) Paving the way forward:
1109 integrating the senses through phase-resetting of cortical oscillations.
1110 *Neuron* 53:162–164.

1111 Ghazanfar A, Schroeder C (2006) Is neocortex essentially multisensory?
1112 *Trends Cogn Sci* 10:278–285.

1113 Gu Y, Angelaki DE, DeAngelis GC (2008) Neural correlates of multisensory
1114 cue integration in macaque MSTd. *Nat Neurosci* 11:1201–1210.

1115 Hennig RM (1988) Ascending auditory interneurons in the
1116 cricket *Teleogryllus commodus* (Walker): comparative physiology and
1117 direct connections with afferents. *J Comp Physiol A* 163:135–143.

- 1118 Herberholz J, Marquart G (2012) Decision making and behavioral choice
1119 during predator avoidance. *Front Neurosci* 6:125.
- 1120 Hirota K, Yuji S, Yoshichika B, Tsuneo Y, others (1993) Distinction in
1121 morphology and behavioral role between dorsal and ventral groups of
1122 cricket giant interneurons. *Zoolog Sci* 10:705–709.
- 1123 Holmes NP, Spence C (2005) Multisensory Integration: Space, time and
1124 superadditivity. *Curr Biol* 15:762–764.
- 1125 Horseman G, Huber F (1994) Sound localisation in crickets. *J Comp Physiol*
1126 *A* 175:389–398.
- 1127 Hoy R, Nolen T, Brodfuehrer P (1989) The neuroethology of acoustic startle
1128 and escape in flying insects. *J Exp Biol* 146:287–306.
- 1129 Ibrahim LA, Mesik L, Ji X, Fang Q, Li H, Li Y, Zingg B, Zhang LI, Tao HW
1130 (2016) Cross-modality sharpening of visual cortical processing through
1131 layer-1-mediated inhibition and disinhibition. *Neuron* 89:1031–1045.
- 1132 Imaizumi K, Pollack GS (1999) Neural coding of sound frequency by cricket
1133 auditory receptors. *J Neurosci* 19:1508–1516.
- 1134 Imaizumi K, Pollack GS (2005) Central projections of auditory receptor
1135 neurons of crickets. *J Comp Neurol* 493:439–447.
- 1136 Jacobs GA, Miller JP, Aldworth Z (2008) Computational mechanisms of
1137 mechanosensory processing in the cricket. *J Exp Biol* 211:1819–1828.
- 1138 Jeanne JM, Wilson RI (2015) Convergence, divergence, and reconvergence in
1139 a feedforward network improves neural speed and accuracy. *Neuron*

1140 88:1014–1026.

1141 Kardamakis AA, Pérez-Fernández J, Grillner S (2016) Spatiotemporal
1142 interplay between multisensory excitation and recruited inhibition in the
1143 lamprey optic tectum. *Elife* 5:e16472.

1144 Kidokoro-Kobayashi M, Iwakura M, Fujiwara-Tsujii N, Fujiwara S, Sakura
1145 M, Sakamoto H, Higashi S, Hefetz A, Ozaki M (2012) Chemical
1146 discrimination and aggressiveness via cuticular hydrocarbons in a
1147 supercolony-forming ant, *Formica yessensis*. *PLoS One* 7:e46840.

1148 Klapoetke NC, Nern A, Peek MY, Rogers EM, Breads P, Rubin GM, Reiser
1149 MB, Card GM (2017) Ultra-selective looming detection from radial
1150 motion opponency. *Nature* 551:237.

1151 Kühne R, Silver S, Lewis B (1984) Processing of vibratory and acoustic signals
1152 by ventral cord neurones in the cricket *Gryllus campestris*. *J Insect*
1153 *Physiol* 30:575–585.

1154 Lakatos P, Chen C-M, O’Connell MN, Mills A, Schroeder CE (2007) Neuronal
1155 oscillations and multisensory interaction in primary auditory cortex.
1156 *Neuron* 53:279–292.

1157 Marsat G, Pollack GS (2005) Effect of the temporal pattern of contralateral
1158 inhibition on sound localization cues. *J Neurosci* 25:6137–6144.

1159 Marsat G, Pollack GS (2006) A behavioral role for feature detection by
1160 sensory bursts. *J Neurosci* 26:10542–10547.

1161 Marsat G, Pollack GS (2007) Efficient inhibition of bursts by bursts in the

1162 auditory system of crickets. *J Comp Physiol A* 193:625–633.

1163 Marsat G, Pollack GS (2010) The structure and size of sensory bursts encode
1164 stimulus information but only size affects behavior. *J Comp Physiol A*
1165 196:315–320.

1166 Marsat G, Pollack GS (2012) Bursting neurons and ultrasound avoidance in
1167 crickets. *Front Neurosci* 6:95.

1168 McMeniman CJ, Corfas RA, Matthews BJ, Ritchie SA, Vosshall LB (2014)
1169 Multimodal integration of carbon dioxide and other sensory cues drives
1170 mosquito attraction to humans. *Cell* 156:1060–1071.

1171 Meredith MA, Stein BE (1986) Visual, auditory, and somatosensory
1172 convergence on cells in superior colliculus results in multisensory
1173 integration. *J Neurophysiol* 56:640–662.

1174 Meredith M, Stein B (1983) Interactions among converging sensory inputs in
1175 the superior colliculus. *Science* 221:389–391.

1176 Miller JP, Jacobs GA, Theunissen FE (1991) Representation of sensory
1177 information in the cricket cercal sensory system. I. Response properties
1178 of the primary interneurons. *J Neurophysiol* 66:1680–1689.

1179 Miller JP, Krueger S, Heys JJ, Gedeon T (2011) Quantitative characterization
1180 of the filiform mechanosensory hair array on the cricket cercus. *PLoS*
1181 *One* 6:e27873.

1182 Nolen TG, Hoy RR (1984) Initiation of behavior by single neurons: the role of
1183 behavioral context. *Science* 226:992–994.

1184 Nolen TG, Hoy RR (1986) Phonotaxis in flying crickets. I. Attraction to the
1185 calling song and avoidance of bat-like ultrasound are discrete behaviors.
1186 *J Comp Physiol A* 159:423–439.

1187 Oe M, Ogawa H (2013) Neural basis of stimulus-angle-dependent motor
1188 control of wind-elicited walking behavior in the cricket *Gryllus*
1189 *bimaculatus*. *PLoS One* 8:e80184.

1190 Ohyama T, Schneider-Mizell CM, Fetter RD, Aleman JV, Franconville R,
1191 Rivera-Alba M, Mensh BD, Branson KM, Simpson JH, Truman JW,
1192 Cardona A, Zlatic M (2015) A multilevel multimodal circuit enhances
1193 action selection in *Drosophila*. *Nature* 520:633.

1194 Olcese U, Iurilli G, Medini P (2013) Cellular and synaptic architecture of
1195 multisensory integration in the mouse neocortex. *Neuron* 79:579–593.

1196 Palka J, Olberg R (1977) The cercus-to-giant interneuron system of crickets.
1197 *J Comp Physiol* 119:301–317.

1198 Perrault TJ, Vaughan JW, Stein BE, Wallace MT (2003) Neuron-specific
1199 response characteristics predict the magnitude of multisensory
1200 integration. *J Neurophysiol* 90:4022–4026.

1201 Perrault TJ, Vaughan JW, Stein BE, Wallace MT (2005) Superior colliculus
1202 neurons use distinct operational modes in the integration of multisensory
1203 stimuli. *J Neurophysiol* 93:2575–2586.

1204 Pollack GS (2015) Neurobiology of acoustically mediated predator detection.
1205 *J Comp Physiol A* 201:99–109.

1206 Popov AV, Markovich AM (1982) Auditory interneurons in the prothoracic
1207 ganglion of the cricket, *Gryllus bimaculatus*. *J Comp Physiol A* 146:351–
1208 359.

1209 Pouget A, Deneve S, Duhamel JR (2002) A computational perspective on the
1210 neural basis of multisensory spatial representations. *Nat Rev Neurosci*
1211 3:741–747.

1212 Rowland BA, Quessy S, Stanford TR, Stein BE (2007) Multisensory
1213 integration shortens physiological response latencies. *J Neurosci*
1214 27:5879–5884.

1215 Rowland B, Stein B (2007) Multisensory integration produces an initial
1216 response enhancement. *Front Integr Neurosci* 1:4.

1217 Sato N, Shidara H, Ogawa H (2017) Post-molting development of wind-
1218 elicited escape behavior in the cricket. *J Insect Physiol* 103:36–46.

1219 Schildberger K, Hörner M (1988) The function of auditory neurons in cricket
1220 phonotaxis. *J Comp Physiol A* 163:621–631.

1221 Schroeder CE, Foxe J (2005) Multisensory contributions to low-level,
1222 “unisensory” processing. *Curr Opin Neurobiol* 15:454–458.

1223 Selverston AI, Kleindienst HU, Huber F (1985) Synaptic connectivity
1224 between cricket auditory interneurons as studied by selective
1225 photoinactivation. *J Neurosci* 5:1283–1292.

1226 Shi X, Barchini J, Ledesma HA, Koren D, Jin Y, Liu X, Wei W, Cang J (2017)
1227 Retinal origin of direction selectivity in the superior colliculus. *Nat*

1228 Neurosci 20:550–558.

1229 Skaliora I, Doubell TP, Holmes NP, Nodal FR, King AJ (2004) Functional
1230 topography of converging visual and auditory inputs to neurons in the
1231 rat superior colliculus. *J Neurophysiol* 92:2933–2946.

1232 Stanford TR (2005) Evaluating the operations underlying multisensory
1233 integration in the cat superior colliculus. *J Neurosci* 25:6499–6508.

1234 Stein BE, Scott Huneycutt W, Alex Meredith M (1988) Neurons and behavior:
1235 the same rules of multisensory integration apply. *Brain Res* 448:355–358.

1236 Stein BE, Stanford TR (2008) Multisensory integration: current issues from
1237 the perspective of the single neuron. *Nat Rev Neurosci* 9:255–266.

1238 Tauber E, Camhi J (1995) The wind-evoked escape behavior of the cricket
1239 *Gryllus bimaculatus*: integration of behavioral elements. *J Exp Biol*
1240 198:1895–1907.

1241 Theunissen FE, Miller JP (1991) Representation of sensory information in
1242 the cricket cercal sensory system. II. Information theoretic calculation of
1243 system accuracy and optimal tuning-curve widths of four primary
1244 interneurons. *J Neurophysiol* 66:1690–1703.

1245 Triplehorn JD, Yager DD (2006) Wind generated by an attacking bat:
1246 anemometric measurements and detection by the praying mantis cercal
1247 system. *J Exp Biol* 209:1430–1440.

1248 Truskowski TLS, Carrillo OA, Bleier J, Ramirez-Vizcarrondo CM, Felch DL,
1249 McQuillan M, Truskowski CP, Khakhalin AS, Aizenman CD (2017) A

1250 cellular mechanism for inverse effectiveness in multisensory integration.
1251 *Elife* 6:e25392.

1252 Wiese K (1981) Influence of vibration on cricket hearing: Interaction of low
1253 frequency vibration and acoustic stimuli in the omega neuron. *J Comp*
1254 *Physiol A* 143:135–142.

1255 Wilent WB (2005) Stimulus-dependent changes in spike threshold enhance
1256 feature selectivity in rat barrel cortex neurons. *J Neurosci* 25:2983–2991.

1257 Wohlers DW, Huber F (1978) Intracellular recording and staining of cricket
1258 auditory interneurons (*Gryllus campestris* L., *Gryllus bimaculatus*
1259 DeGeer). *J Comp Physiol A* 127:11–28.

1260 Wohlers DW, Huber F (1982) Processing of sound signals by six types of
1261 neurons in the prothoracic ganglion of the cricket, *Gryllus campestris* L.
1262 *J Comp Physiol* 146:161–173.

1263 Zahar Y, Reches A, Gutfreund Y (2009) Multisensory enhancement in the
1264 optic tectum of the barn owl: Spike count and spike timing. *J*
1265 *Neurophysiol* 101:2380–2394.

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1288 Someya M, Ogawa H (2017) Coincident multisensory inputs enhance burst

1289 activity via large and long-lasting EPSPs in insect auditory neuron, poster,

1290 Society for Neuroscience 47th Annual Meeting, Washington, DC, USA.

1291 Someya M, Ogawa H (2014) Multisensory integration of auditory and cercal

1292 sensory inputs by ascending projection neurons in the cricket, poster, The

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1294

1295 [Domestic]

1296 Someya M, Ogawa H (2017) Multisensory integration of auditory and

1297 mechanosensory inputs in identified projection neuron in the cricket, oral,

1298 The 40th Annual Meeting of the Japan Neuroscience Society, Yokohama,

1299 Japan.

1300 Someya M, Ogawa H (2016) Multisensory integration in early stage of sensory

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- 1303 Someya M, Ogawa H (2014) Multisensory ascending neurons responding to
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