Vanishing immunoglobulins: the formation of pauci-immune lesions in myeloperoxidase-antineutrophil cytoplasmic antibody-associated vasculitis

Running Title: Pathogenesis of pauci-immune lesion

Emika Futamata¹, Sakiko Masuda², Yuka Nishibata², Satoshi Tanaka³,
Utano Tomaru⁴, Akihiro Ishizu²

¹Undergraduate School of Health Sciences, Hokkaido University, Sapporo, Japan
²Faculty of Health Sciences, Hokkaido University, Sapporo, Japan
³Center for Cause of Death Investigation, Graduate School of Medicine, Hokkaido University, Sapporo, Japan
⁴Department of Pathology, Faculty of Medicine and Graduate School of Medicine, Hokkaido University, Sapporo, Japan

Correspondence: Akihiro Ishizu, Faculty of Health Sciences, Hokkaido University, Kita-12, Nishi-5, Kita-ku, Sapporo 0600812, Japan, Phone: +81-11-706-3385, Fax: +81-11-706-4916, E-mail: aishizu@med.hokudai.ac.jp
Dear editor,

Myeloperoxidase-antineutrophil cytoplasmic antibody (MPO-ANCA)-associated vasculitis is characterized by necrotizing crescentic glomerulonephritis and small vessel vasculitis with a presence of MPO-ANCA in the serum. Although the pathogenic role of MPO-ANCA in the formation of necrotizing lesions has been demonstrated, only little deposition of immunoglobulins can be detected in the affected tissues. This histological feature is what we call “pauci-immune.”

Neutrophil extracellular traps (NETs) are web-like DNA decorated with antimicrobial proteins, such as MPO, which are extruded from activated neutrophils [1]. NETs can trap microorganisms by the web-like DNA and kill them using the antimicrobial proteins scattered on the extracellular DNA. Although NET formation is regarded as an essential event in innate immunity, recent studies have noted the adverse effects of NETs on the hosts, e.g., thrombogenic potential and cytotoxicity to vascular endothelial cells [2].

The involvement of NETs in the pathogenesis of MPO-ANCA-associated vasculitis was firstly reported in 2009 [3]. The following studies have demonstrated the presence of NETs in crescentic glomeruli with pauci-immune necrotizing lesions [4]. In MPO-ANCA-associated vasculitis, neutrophils primed by inflammatory cytokines, such as TNF-α, express MPO on their surfaces, and MPO-ANCA can bridge the MPO and Fcγ receptors on the cell resulting in the formation of NETs [5].

In order to reveal the pathogenesis of pauci-immune lesions in MPO-ANCA-associated vasculitis, we determined that neutrophil elastase released from NET-forming neutrophils, which was activated by MPO-ANCA, could digest immunoglobulins. When the anti-MPO antibody (FITC-conjugated) was applied to
ethanol-fixed neutrophils, the antibody binding to the neutrophils was detected as ANCA steadily for 16 h (Figure 1a). On the contrary, the anti-MPO antibody (MPO-ANCA) induced NETs on TNF-α-primed neutrophils and then disappeared simultaneously. Since MPO has been shown to distribute on the DNA of NETs [1], it can be considered that the vanishing MPO-ANCA reflect a disappearance of the antibody rather than the antigen. Based on these findings and previous reports on the secretion of neutrophil elastase from NET-forming neutrophils [3], we speculated that neutrophil elastase could digest MPO-ANCA. As expected, time and dose-dependent digestions of the IgG by neutrophil elastase were demonstrated (Figure 1b). In addition, the MPO-ANCA, which was exposed to neutrophil elastase, could no longer bind to neutrophils (Figure 1c). The collective findings suggest that neutrophil elastase released from NET-forming neutrophils, which was activated by MPO-ANCA, could be involved in the pathogenesis of pauci-immune lesions, at least in part, in MPO-ANCA-associated vasculitis via digestion of MPO-ANCA.

Funding

This work was supported by a grant for Japan Research Committee of the Ministry of Health, Labour, and Welfare for Intractable Vasculitis.

Disclosure statement

The authors declare that no conflict of interest exists.
References


Figure 1. (a) Vanishing MPO-ANCA. Peripheral blood neutrophils from a healthy volunteer were seeded in slide chambers \((1 \times 10^6)\) ml, incubated for 30 min at 37 °C, and then fixed with 100% ethanol for 15 min at room temperature or exposed to 2 ng/ml of TNF-α for 15 min at 37 °C. After washing with PBS, the cells were allowed to react with 1 µg/ml of the FITC-conjugated anti-MPO antibody (Gene Tex, Irvine, CA) at 37 °C. After incubation for 15 min, 1 h, 4 h, and 16 h, the cells were washed with PBS and then mounted using the solution containing DAPI (Sigma-Aldrich, St. Louis, MO). Microphotographs (magnification, ×400) were taken under a fluorescent microscopy. (b) Time and dose-dependent digestions of the IgG by neutrophil elastase. Mouse IgG (1 µg) (BD Biosciences, Tokyo, Japan) was exposed to 20 µg of neutrophil elastase (abcam, Cambridge, UK) at 37 °C for 0 min, 15 min, 1 h, and 4 h. Alternatively, 1 µg of
mouse IgG was exposed to 0, 0.2, 2, and 20 µg of neutrophil elastase at 37 °C for 4 h. Thereafter, the samples were boiled for 5 min in the sample buffer followed by electrophoresis through 12% SDS polyacrylamide gel. The gels were stained with Coomassie Brilliant Blue. (c) Effect of neutrophil elastase exposure on MPO-ANCA. Peripheral blood neutrophils from a healthy volunteer were seeded in slide chambers (1 × 10⁶/ml), incubated for 30 min at 37 °C, and then fixed with 100% ethanol for 15 min at room temperature. After washing with PBS, the cells were allowed to react with 1 µg/ml of the FITC-conjugated anti-MPO antibody, which was treated without or with 20 µg/ml of neutrophil elastase at 37 °C for 4 h. After 1 h incubation at room temperature, the cells were washed with PBS and then mounted using the solution containing DAPI. Microphotographs (magnification, ×400) were taken under a fluorescent microscopy. All experiments were repeated more than twice. Since similar results were reproduced, the representative results were shown.