



Title	Association between human exposure to heavy metals/metalloid and occurrences of respiratory diseases, lipid peroxidation and DNA damage in Kumasi, Ghana
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1 Association between human exposure to heavy metals/metalloid and occurrences of  
2 respiratory diseases, lipid peroxidation and DNA damage in Kumasi, Ghana

3

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22

23   **Abstract**

24   Heavy metals and metalloids contamination in soils, water, food and livers of wild rats have  
25   been studied in Kumasi, Ghana and despite the estimated risks to residents, there is no  
26   epidemiological study to ascertain these projections. In addition, the World Health  
27   Organization and International Agency for Research on Cancer have reported an increase in  
28   respiratory diseases and cancers, in Ghana. The study's purpose was therefore to explore  
29   the potential associations between metal exposure and occurrences of respiratory diseases,  
30   lipid peroxidation and/or DNA damage to different age groups and sexes in Kumasi.  
31   Human urine was collected from the general population in urban and control sites in  
32   Kumasi and nine metals were measured in each sample. Results showed that although Zn  
33   was the most abundant, total urinary As concentration was higher in 83% of samples  
34   compared to reference values. Urinary concentrations of metals, malondialdehyde (MDA)  
35   and 8-hydroxy-2-deoxy-guanosine (8-OHdG) were higher in urban sites compared to the  
36   control site. Based on the results obtained, there was no significant correlation between  
37   urinary metals and age. However, urinary Cd and MDA were highest in age groups 61-85  
38   and 3-20 years, respectively. Significantly higher levels of urinary Co, As and Cd were  
39   detected in female participants. The study revealed that exposure to As was significantly  
40   associated with increased odds of asthma (odds ratio (OR) = 2.76; CI: 1.11-6.83) and  
41   tachycardia (OR = 3.93; CI: 1.01-15.4). Significant association was observed between  
42   urinary metals and MDA and 8-OHdG indicating possibility of lipid peroxidation and/or  
43   DNA damage in Kumasi residents.

44   **Keywords:** Metals; Urine; Kumasi; Asthma; DNA damage

45     **Capsule:** Human exposure to metals/metalloid increased the occurrences of lipid  
46     peroxidation, DNA damage and was also associated with increased odds of asthma and  
47     tachycardia.

48

49     **1. Introduction**

50       Heavy metals and metalloids are among the most toxic substances (ATSDR 2015) and  
51       despite their natural abundance, they are formed mainly from human activities such as  
52       mining, smelting, combustion, tannery or fertilizer applications. Humans and animals could  
53       be exposed to metals via inhalation, consumption and/or dermal contact (Saoudi et al.  
54       2012). Despite the importance metals (iron, zinc, copper and manganese) play in  
55       maintaining normal physiological functions, excessive intake could result in health  
56       implications (Magge et al. 2013). In addition, exposure to cadmium, nickel, lead and  
57       arsenic could generate reactive oxygen species (ROS) leading to modifications of DNA and  
58       lipids (Stohs and Bagchi 1995). These modifications have been reported to contributes to  
59       the incidence of cancers and cardiovascular diseases (Shi et al. 2004). Indicators of DNA  
60       damage, oxidative stress and lipid peroxidation such as 8-hydroxy-2-deoxyguanosine (8-  
61       OHdG) and malondialdehyde (MDA) have widely been used to determine the health effects  
62       of human exposure to metals (Chen et al. 2005).

63       Besides DNA damage and lipid peroxidation, various epidemiological studies have also  
64       found and reported associations between heavy metal/metalloid (including arsenic,  
65       cadmium, copper, manganese, nickel, lead) exposure and the occurrence of respiratory

66 effects such as asthma, rhinitis, wheeze, bronchitis, and allergies (Gehring et al. 2015;  
67 Huang et al. 2016).

68 The growing rate of industrialization (including mining), and resulting increases in  
69 economic activity and population growth in Kumasi, Ghana, has led to increased pollution  
70 of the environment (Bortey-Sam et al. 2014). Studies of environmental contamination and  
71 possible health risks due to metal exposure via medicinal herbs (Nkansah et al. 2016a) ,  
72 geophagic white clay (Nkansah et al. 2016b), food (Nkansah et al. 2016c), dust (Nkansah et  
73 al. 2015), soils (Akoto et al. 2016; Akoto et al. 2017) and streams (Akoto et al. 2010)  
74 within Kumasi metropolis have been reported. Furthermore, levels of zinc, arsenic, copper  
75 and nickel in livers of wild rats sampled in Kumasi (Bortey-Sam et al. 2015) were higher  
76 compared to the levels in wild rats sampled around mining sites in Kabwe, Zambia  
77 (Nakayama et al. 2013). Despite these reports and estimated risks, there is no study to  
78 assess the impact of metal exposure to Kumasi residents.

79 In Ghana, there are an estimated 16,000 cancer cases annually and also an increase in  
80 occurrence of respiratory disease (GLOBACAN 2008; WHO 2011). In 2012, the estimated  
81 cancer incidence in Kumasi was 11.9 per 100,000 and was higher in females (15.7 per  
82 100,000) than males (7.3 per 100,000) (Laryea et al. 2014). Due to the high cancer  
83 incidence and respiratory symptoms in Kumasi (GLOBACAN 2008; Laryea et al. 2014;  
84 WHO 2011), and unavailability of research on the epidemiology and risks of metal  
85 exposure to residents, the objectives of this study were to: explore the potential associations  
86 between metal exposure and occurrence of respiratory diseases; assess the relationship

87 between metal exposure and incidence of oxidative stress; find the association between  
88 urinary concentrations of metals, MDA, 8-OHdG with age and sex.

89

90 **2. Materials and methods**

91 *2.1. Sampling*

92 Urine is considered the main excretory pathway for metals and a better medium for  
93 biomonitoring metal exposure (Smolders et al. 2014). Heavy metals and metalloid  
94 concentrations in urine could be an indication of both long and short term exposures  
95 (Crinnion 2010). In view of this, human urine ( $n = 190$ ; 57 males and 133 females) was  
96 collected in the morning from the general population of three urban sites (Atonsu, Manhyia  
97 and Tafo) in Kumasi (Fig. 1). Samples were collected into corning tubes (Corning  
98 Incorporated, New York, USA) in January to February of 2015. Manhyia is in close  
99 proximity to Kejetia (1.1 km apart), Adum (1.5 km apart) and Romanhill (1.2 km apart),  
100 where soils were polluted with metals (Akoto et al. 2017). In previous studies,  
101 concentrations of metals were highest in the livers of wild rats trapped in Adum compared  
102 to other sites in Kumasi (Bortey-Sam et al. 2015). Tafo is also 2.3 and 2.6 km from Suame  
103 and Mbrom, respectively, whose soils were polluted with metals (Akoto et al. 2017).

104 Moreover, 12 human urine samples (7 males and 5 females) were collected from  
105 Kwame Nkrumah University of Science and Technology campus (KNUST) and used as  
106 reference/control samples, even though metal exposure via consumption or inhalation was  
107 possible. KNUST, a university in Kumasi, has minimal vehicular motion and no industrial

108 activities. In previous studies, heavy metals and metalloid levels in KNUST soils were low  
109 compared to recommended levels (Akoto et al. 2016; Akoto et al. 2017). In addition,  
110 particulate matter and soil samples from KNUST have been used as controls in previous  
111 studies of environmental contaminants (Bortey-Sam et al. 2013; Bortey-Sam et al. 2014; N  
112 Bortey-Sam et al. 2015).

113 For quality control purposes, urine was collected from 4 children in residential areas of  
114 KNUST to form a composite. Composite samples were used to give a more representative  
115 measure and also to account for any variabilities in heavy metals and metalloid  
116 concentrations. Since humans could be exposed to metals through various sources, the  
117 sample was measured several times to confirm the concentration.

118 During the sampling process, participant's information, including age, gender, body  
119 weight, height, place of residence, occupation, and personal lifestyle including smoker/non-  
120 smoker, were obtained through face-to-face interviews. Further, information on respiratory  
121 symptoms related to metals such as asthma, wheeze, tachycardia, bronchitis and rhinitis  
122 (Gehring et al. 2015; Huang et al. 2016) were collected. The Ethical/Institutional Review  
123 Board of Ghana Health Service (GHS) and Council for Scientific and Industrial Research  
124 (CSIR), Accra, Ghana, approved this study. Written and informed consent was obtained  
125 from each participant and parents gave consent and completed questionnaires on behalf of  
126 their children. The samples collected were kept frozen at the Department of Chemistry,  
127 KNUST, Ghana. Later the samples were transported to the Toxicology laboratory of the  
128 Graduate School of Veterinary Medicine, Hokkaido University, Japan, and stored at -30 °C  
129 until analysis.

130    2.2. *Sample extraction and analysis*

131    2.2.1. *Heavy metals and a metalloid*

132    Method described by Yabe et al. (Yabe et al. 2015) was used for the extraction of heavy  
133    metals and a metalloid from the urine samples collected. Briefly, 1 mL of each urine was  
134    transferred into a digestion vessel and 5 mL of 60 % nitric acid (Kanto Chemical) and 1 mL  
135    of 30 % hydrogen peroxide (Kanto Chemical) were added. Sample digestion (Speedwave  
136    MWS-2; Berghof) was for 52 min and up to 190 °C. The digested samples were transferred  
137    into corning tubes and diluted to 10 mL with de-ionized water (Milli-Q). Concentrations of  
138    arsenic (As), cadmium (Cd), cobalt (Co), chromium (Cr), copper (Cu), lead (Pb),  
139    manganese (Mn), nickel (Ni) and zinc (Zn) in each urine were measured by Inductively  
140    Coupled Plasma-Mass Spectrometer (ICP-MS; 7700 series, Agilent technologies, Tokyo,  
141    Japan).

142    2.2.2. *Malondialdehyde, MDA (Elisa kit)*

143    Concentrations of urinary MDA were measured (based on instructions from  
144    manufacturer) using a UV-VIS Spectrophotometer (UV-2600 Shimadzu Corporation,  
145    Kyoto, Japan). Briefly, 10 uL of butylated hydroxytoluene (BHT) reagent was transferred  
146    into a vial and 250 uL of calibrator (0, 1, 2, 3 and 4 µM) or urine was added. After the  
147    addition of 250 uL each of 1 M phosphoric acid and 2-thiobarbituric acid (TBA) reagent,  
148    the solution was vortexed vigorously and incubated at 60 °C for 1 hr. The mixture was  
149    transferred into a cuvette and spectra was recorded from 400-700 nm after it was  
150    centrifuged at 10000 x g for 2-3 min. 3<sup>rd</sup> derivative analysis was performed at 514 nm.

151    2.2.3. *8-hydroxy-2-deoxy-guanosine (8-OHdG)*

152    Extraction and analysis of urine sample for 8-OHdG followed the method described by  
153    Bortey-Sam et al. (Bortey-Sam et al. 2017). Briefly, urine (1 mL) was diluted with HPLC  
154    grade water (2 mL) after spiking with 25 ng/mL of (15N5) 8-OHdG (internal standard).  
155    Prior to sample loading, the Oasis HLB cartridge (3cc, 60 mg; Waters Corporation, Milford,  
156    MA, USA) was primed with 1 mL each of methanol and water. The solid-phase extraction  
157    cartridge was then washed with 3 mL of water and the target analyte (8-OHdG) eluted with  
158    3 mL of water: acetonitrile (1:1, v/v). The eluate was evaporated to near dryness under  
159    nitrogen gas. The residue was re-dissolved in water (100 µL), and 10 µL injected into the  
160    LC–MS/MS. A Phenomenex Gemini 3u C18 110A column (150 mm × 2 mm i.d., 4 micron,  
161    Phenomenex, California, USA) with a guard column was used for the separation of 8-  
162    OHdG and (15N5) 8-OHdG in urine. Gradient elution was as follows: 0.0–1.0 min, 5% B;  
163    1.01–3.00 min, 50% B; 3.01–6.00 min, 5% B. Multiple reaction monitoring (MRM) in  
164    negative ionization mode was used to identify the target analytes at a column temperature  
165    of 40 °C. Mobile phases A (0.1% formic acid) and B (100% methanol) were pumped at a  
166    flow rate of 250 µL/min (Bortey-Sam et al. 2017).

167    2.3. *Creatinine concentrations in human urine*

168    To compensate for variations in urine dilution, urinary creatinine was used to adjust  
169    concentrations of metals, MDA and 8-OHdG. Concentrations of creatinine in urine were  
170    determined based on the manufacturer's instructions (Arbor Assays, Michigan, USA).  
171    Briefly, 100 µL of DetectX® Creatinine Reagent (Arbor Assays, Michigan, USA) was  
172    added to 50 µL of sample, blank (water), or standards into clear plate wells. Prior to 30 min

173 incubation (at room temperature), the sides of the plate was tapped for adequate mixing and  
174 the plate was covered with a plate sealer and pressed to seal adequately. The optical density  
175 produced from the plate reader well (Muiltiskan GO, Thermo Scientific, Vantaa, Finland)  
176 was read at 490 nm. To calculate the creatinine concentrations, a 4PLC built-in software  
177 was used and results expressed in g/L. Obtained creatinine concentrations (g/L) (mean;  
178 [range]) in human urine in Atonsu ( $1.57 \pm 0.969$ ; [0.0398-4.35]), Manhyia ( $1.76 \pm 1.33$ ;  
179 [0.202-5.96]), Tafo ( $2.30 \pm 1.59$ ; [0.0185-7.43]) and KNUST ( $2.71 \pm 1.45$ ; [1.46-5.58])  
180 were used to adjust the respective urinary metal concentrations.

181 *2.4. Quality control and quality assurance*

182 *2.4.1. Heavy metals and a metalloid*

183 After every 10 sample analyses, blanks and duplicates were analysed and the Relative  
184 Standard Deviation (RSD) obtained for duplicate runs was  $\leq 4\%$ . Calibration curves using  
185 standard solutions were run and the linearity obtained for each metal was greater than 0.999.  
186 Analytical-reagent grade chemicals and standard stock solutions were used (Wako Pure  
187 Chemicals). The detection limits ( $\mu\text{g/L}$ ) were 0.009 (As), 0.001 (Cd), 0.003 (Co), 0.003  
188 (Cr), 0.008 (Cu), 0.002 (Pb), 0.034 (Mn), 0.003 (Ni) and 0.019 (Zn). Heavy metals and  
189 metalloid concentrations in human urine were expressed in  $\mu\text{g/g}$  creatinine.

190 The urine samples of children collected from residential areas of KNUST was used for  
191 quality control and recovery tests. The samples were spiked with standard solutions of  
192 metals and digested using the method described. The recovery rates of the spiked urine  
193 ranged from 95 (Pb) - 98% (Cd). Concentrations of metals in urine sample used for this

194 purpose were below the respective limits of detection (LODs) and differences between  
195 concentrations before and after spiking was used to calculate the recoveries.

196 *2.4.2. MDA and 8-OHdG*

197 For 8-OHdG, (15N5) 8-OHdG was used as internal standard and spiked into urine prior  
198 to sample preparation and extraction. Internal standard method with a five-point calibration  
199 (1, 5, 10, 50 and 100 ng/mL) was used for quantification. The average linearity of the  
200 calibration standards for both MDA and 8-OHdG were greater than 0.99. The LOD and  
201 limit of quantification (LOQ) for 8-OHdG were 0.0196 and 0.6 ng/mL, respectively, and  
202 average recovery ([15N5] 8-OHdG) was  $86 \pm 9.8\%$ . After every 10 samples, spiked solvent  
203 blanks (with 8-OHdG only) and duplicate samples were analysed and average internal  
204 standard recovery for spiked solvents blanks was  $104 \pm 8.7\%$ . The %RSD for duplicate  
205 samples were less than or equal to 10% (MDA and 8-OHdG). LOD and LOQ for MDA  
206 were 0.205 and 0.63 uM, respectively.

207 *2.4.2. Creatinine*

208 Quantitation was performed based on a seven–point calibration (0.3125, 0.625, 1.25, 2.5,  
209 5, 10, and 20 mg/dL) and the average linearity of the calibration standard was greater or  
210 equal to 0.9996. LODs and LOQs were calculated based on  $3SD/S$  and  $10SD/S$ ,  
211 respectively (SD is the standard deviation of five replicate measurements of the target  
212 analyte and S is the slope of the calibration curve). LOD and LOQ of creatinine were  
213 0.00151 and 0.00505 g/L, respectively. Duplicate samples were run after every batch of 10

214 samples and the %RSD was  $2.99 \pm 2.30$ . Blank samples were also run after every 11  
215 samples.

216 *1.5. Data analysis*

217 IBM SPSS v 20 (SPSS Inc., Illinois, USA) was used for statistical analyses and the  
218 normality of the data was tested using Kolmogorov–Smirnov (K–S) and Shapiro-Wilks  
219 tests. A value of LOD/2 was assigned to metals/metalloid concentrations below their  
220 respective LODs. The central tendency of the analyte concentrations was illustrated with  
221 the geometric mean concentrations (Ott 1990). To compare urinary concentrations of  
222 metals from the study areas, ANOVA and Tukey tests were performed, after data was  
223 normalized by log transformation, and a  $p$  value less than 0.05 was considered significant.  
224 Pearson's correlation of logged data was used to determine the association between metals,  
225 MDA, 8-OHdG and age. The distribution of metals/metalloid, MDA and 8-OHdG between  
226 male and female participants was done using Student's T-Test and statistical significance  
227 was at  $p$  less than 0.05. Odds ratios (ORs) at 95% confidence interval (CI) was used to  
228 determine the association between exposure to As and occurrences of respiratory symptoms.  
229 This was derived using logistic regression model. As was treated as a continuous variable  
230 in the logistic regression. Regression models were adjusted for covariates such as age and  
231 sex. Statistical significance was set at  $p < 0.05$  and performed with JMP 10 statistical  
232 software (SAS Institute).

233 **3. Results and discussion**

234 *3.1. Urinary levels of heavy metals and a metalloid*

235       The order of the geometric mean concentrations (adjusted by urinary creatinine; GM<sub>creat</sub>)  
236       of heavy metal and metalloid from all study sites in Kumasi was Zn ( $335 \pm 340$ ) > As (49.8  
237        $\pm 52.2$ ) > Cu ( $14.7 \pm 28.9$ ) > Ni ( $2.36 \pm 6.33$ ) > Cr ( $0.825 \pm 7.32$ ) > Pb ( $0.716 \pm 7.59$ )  $\geq$  Co  
238       ( $0.712 \pm 3.57$ ) > Mn ( $0.276 \pm 2.35$ )  $\geq$  and Cd ( $0.240 \pm 2.22$ )  $\mu\text{g/g}$  creatinine. The urinary  
239       concentrations of all metals measured varied significantly ( $p < 0.01$ ) (K-S and S-W's tests).  
240       The results of GM<sub>creat</sub> indicated that Co, Cu, Zn and As were detected in all samples  
241       (100%) while the detection rate for urinary Cr was 78%, Mn (89%), Ni (79%), Cd (99%),  
242       Pb (76%), MDA (95%) and 8-OHdG (59%).

243       Urinary concentrations of Ni and Cr (except Manhyia) were significantly higher ( $p =$   
244       0.0002-0.0145) in participants who lived in Atonsu compared to other sites including  
245       KNUST (Table 1). Additionally, urinary concentrations of As and Cd in participants who  
246       lived in urban sites were significantly higher ( $p = 0.0001$ -0.01) compared to KNUST  
247       participants (Table 1). Although not significant ( $p = 0.0798$ -0.838), concentrations of Mn,  
248       Co and Cd (significantly lower in KNUST) in urine were higher in Atonsu participants  
249       while highest levels of Pb and Cu (significantly lower in KNUST) were detected in Tafo  
250       participants. The high urinary metals could be due to high metal exposure to residents.  
251       Previous studies reported that although metal concentrations in Atonsu soils were below  
252       recommended levels, the soils were enriched with Zn, Cd, Cr and Pb (Akoto et al. 2017).  
253       Tafo on the other hand is close to communities (Suame and Mbrom) polluted with metals  
254       and filled with light scale industries (Akoto et al. 2017).

255       Metals and metalloid concentrations detected in urine of participant's from the present  
256       study were compared to studies conducted in the US (Caldwell et al. 2009; CDC 2005),

257 Czech Republic (Benes et al. 2002), Spain (Aguilera et al. 2010), France (de Burbure et al.  
258 2006), Italy (Alimonti et al. 2000) China (Huang et al. 2016; Lu et al. 2016) and  
259 Democratic Republic of Congo (Banza et al. 2009) (Table S1). The results (Table S1)  
260 showed that all metal measured in this study were comparable or lower than previous  
261 studies with the exception of urinary As concentrations which were higher in this study.  
262 From Table S2, the unadjusted urinary concentrations (ng/mL) of metals (except Co) from  
263 all sites were on the average (11.3 (Zn) - 83.3% (As)) higher compared to the unadjusted  
264 reference values suggested by the Canadian Health Measure (Saravanabhan et al. 2016);  
265 Fourth National Report on Human Environmental Chemicals, USA (Crinnion 2010) and  
266 Human Biomonitoring Commission, Germany (Schulz et al. 2009).

267 In addition to water and food consumption, soil and dust are other possible ways of  
268 exposure to metals/metalloids especially children via hand-to-mouth practices (Berglund et  
269 al. 2011). Residents of various countries including Ghana, consume large amounts of  
270 geophagic white clay for religious, cultural, nutritional, and medicinal reasons as well as in  
271 response to famine and pregnancy-related cravings (Mathee et al. 2014; Nkansah et al.  
272 2016b). Also, exposure to metals could be through the intake of medicinal herbs (Nkansah  
273 et al. 2016a) frequently used in the treatment of various ailments. Lower levels of urinary  
274 metals in KNUST participants could be due to the low vehicular movement and industrial  
275 activities, and point source of metal pollution was low.

276 The high levels of urinary As in participants could be attributed to the gold mining  
277 activities in some parts of Kumasi. Residents could be exposed because of the composition  
278 of the ore containing the gold. After blasting the gold bearing rock, miners roast the ore and

279 this leads to the production and distribution of arsenic trioxide gas (Amonoo-Neizer et al.  
280 1996). In addition, exposure to organic As was associated with consumption of sea foods  
281 such as shellfish (Aguilera et al. 2010), although the recommended total urinary As  
282 concentration was estimated to be 27 ng/mL by the Canadaian Health Measures  
283 (Saravanabhan et al. 2016). Highest urinary concentrations of As (336, 297 and 234 µg/g  
284 creatinine) were detected in urine of participants who complained of asthma, diabetes,  
285 rhinitis and tachycardia, symptoms which have been associated with As exposure (Maull et  
286 al. 2012; Parvez et al. 2008; Parvez et al. 2010; Parvez et al. 2011; Saha et al. 1999).

287 *3.2. Association between urinary metals, MDA, 8-OHdG with age*

288 As shown in Table 2, there was no significant association ( $p = 0.424-0.928$ ) between  
289 urinary metal concentrations and age from KNUST participants and participants who lived  
290 in urban areas. Previous studies reported a positive correlation between urinary Cd levels  
291 and age while a negative association was observed between other metals and age (Banza et  
292 al. 2009). Results of this study further showed that urinary MDA was highest in age group  
293 3-20 years. The effects of metal exposure is more pronounced in children than adults  
294 because their immune and nervous systems are not fully developed (Olsen 2000). Moreover,  
295 the breathing rate of children is higher and their consumption rate per body weight is also  
296 higher than adults (Schwartz 2004).

297 Although significant correlation ( $p = 0.443$ ) between age and urinary Cd concentrations  
298 was not observed (Table 2), levels were highest in ages 61-85 years compared to the other  
299 age groups (Table 3). This is possibly because concentrations of Cd in urine indicates long-  
300 term accumulation and consequently higher in the elderly (Paschal et al. 2000).

301    *3.3. Association between urinary metals, MDA, 8-OHdG with sex*

302    As shown in Table 4, significantly higher levels ( $p = 0.0089\text{-}0.017$ ) of Co, As and Cd  
303    were detected in urine of female participants compared to males. However, urinary MDA  
304    and 8-OHdG were comparable (Table 4) and this is similar to outcomes obtained by Lu et  
305    al. (Lu et al. 2016). Although not significant ( $p = 0.054\text{-}0.383$ ), urinary concentrations of  
306    Cr, Mn, Cu and Zn were also higher in females than males which is similar to results  
307    obtained by Banza et al. (Banza et al. 2009). The significantly higher urinary Cd in females  
308    was similar to results obtained in other studies (Berglund et al. 2011; Castano et al. 2012;  
309    Paschal et al. 2000; Vahter et al. 2007). Iron deficiency has mainly been related to gender  
310    differences in urinary Cd excretion. This influences high levels of duodenal divalent  
311    transporter which results in the rise in transport and absorption of Cd (Berglund et al. 2011;  
312    Paschal et al. 2000; Vahter et al. 2007).

313    Sex differences in Cu levels have been reported and females recorded higher  
314    concentrations than males (Benes et al. 2002). The higher urinary Cu levels in women  
315    could be attributed to hormonal changes that occur in puberty (Wapnir 1998). Additionally,  
316    significantly higher urinary Mn was detected in women than men and this trend could be  
317    attributed to biological differences in the way females and males handle Mn (Berglund et al.  
318    2011). Lindberg et al. reported that women had significantly higher dimethylarsinic acid  
319    (DMA) concentrations than men, because they (women) can more efficiently methylate  
320    arsenic than men (Lindberg et al. 2008). Although not significant ( $p = 0.371$ ), urinary  
321    concentrations of Pb were higher in males than females. In a study by Berglund et al.,  
322    higher levels of Pb were detected in urine of men than women (Berglund et al. 2011). This

323 results was opposite from previous studies where women excreted higher urinary Pb than  
324 men (Castano et al. 2012). These in addition to other factors (such as differences in  
325 exposure levels, lifestyle etc.) could explain the sex difference in urinary excretion of  
326 metals although the mechanisms underlying these sex differences remain unknown (Howe  
327 et al. 2016).

328 *3.4. Association between metal exposure and occurrence of respiratory symptoms*

329 OR was performed for all metals, however, the study focused on As since it was the  
330 most toxic substance (ATSDR 2015) and the second most abundant metal detected in  
331 human urine in Kumasi. In addition, concentrations of urinary As in this study was higher  
332 than previously conducted studies in China, Congo, Spain and USA (Table S1). Moreover,  
333 urinary As was on average 83% higher (from all study sites) compared to recommended  
334 values.

335 The study revealed that exposure to As was significantly ( $p = 0.041-0.043$ ) associated  
336 with increased odds of asthma (OR = 2.76, CI: 1.11-6.83) and tachycardia (OR = 3.93, CI:  
337 1.01-15.4) in Kumasi residents (Table 5). In a study by Huang et al. urinary concentrations  
338 of As were significantly and positively associated with the occurrence of asthma when the  
339 metalloid was considered as a continuous variable and when divided into quantiles (Huang  
340 et al. 2016). Similarly, As exposure has been associated with (i) occurrence of lung  
341 dysfunction (ii) increased mortality due to respiratory diseases (Parvez et al. 2011).  
342 Additionally, inhalation of As dust or fumes during milling of ores or mining (which is a  
343 common practice in Kumasi) often resulted in chronic cough, laryngitis, bronchitis and  
344 rhinitis (Saha et al. 1999). In other cohort studies, chronic cough, chest sounds, shortness of

345 breath, blood in sputum and other respiratory symptoms were observed due to As exposure  
346 (Parvez et al. 2010). Additionally, positive association between urinary As and serum Clara  
347 cell protein (CC16; a novel biomarker of respiratory illness) was found, with urinary As  
348 levels inversely related with lung function (Parvez et al. 2008).

349 Environmental factors have been associated with the occurrence of asthma (Huang et al.  
350 2016) and as a results, living in metal contaminated areas could increase the risks of  
351 respiratory disease among residents.

### 352 *3.5. Human health risk implications*

353 As shown in Table 2, there was no association ( $p = 0.180\text{-}0.972$ ) between urinary  
354 concentrations of metals, MDA and 8-OHdG in KNUST participants. However, in urine of  
355 participants at urban sites, significant association ( $p = < 0.0001\text{-}0.0063$ ) was noted between  
356 metals and MDA. Additionally, concentrations of urinary As also correlated significantly ( $p$   
357 = 0.0003) with 8-OHdG. This trend indicates the possibility of lipid peroxidation or DNA  
358 damage although these products have also been associated with the presence of  
359 cardiovascular diseases, atherosclerosis, diabetes and cancers (Wu et al. 2004). Aflanie et al.  
360 concluded that human exposure to As and Cd could increase MDA levels and cause  
361 oxidative stress and inflammation (Aflanie et al. 2015). Lu et al. also reported a correlation  
362 between As and 8-OHdG (Lu et al. 2016). Arsenic could bind to thiols and this has been  
363 considered crucial in increasing 8-OHdG levels (Valko et al. 2007). In previous reports,  
364 urinary As was associated with diabetes (Maull et al. 2012). Several metals (e.g. As, Cd,  
365 Pb) are also known to impair kidney function and increase different cancer risks, while  
366 others could affect the nervous and cardiovascular systems (e.g., As, Cd, Mn, Pb) (EFSA

367 2011; Straif et al. 2009; WHO 2008). In previous studies, oxidative stress correlated with  
368 allergic inflammatory diseases (Bartsch and Nair 2004), obesity and atherosclerosis  
369 (Kobayashi et al. 2011; Wu et al. 2004) and these were some symptoms participants  
370 complained of during the face-to-face interview. 10% of participants in this study were  
371 diabetics and 2% had arthritis. Of the 33 participants that we calculated body mass index,  
372 11 were overweight and 7 were obese.

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375 *3.6. Limitations of the study*

376 The major limitation to this study was the small sample size which could have resulted  
377 in wide CI in the regression analysis. Additionally, participants were not medically  
378 examined and this could have resulted in false information.

379 **4. Conclusions**

380 Urinary metal/metalloid concentrations were studied in Kumasi residents, and although  
381 Zn was the most abundant, urinary As was higher in 83% of participants compared to  
382 recommended levels. The study revealed that urinary concentrations of metals, MDA and  
383 8-OHdG were higher in urban sites participants compared to the control site. Females  
384 excreted significantly higher levels of Co, As and Cd than males. As exposure was  
385 significantly associated with increased odds of asthma and tachycardia. Although no  
386 relationship was found between urinary metals and age, Cd and MDA levels were highest

387 in age groups 61-85 and 3-20 years, respectively. Exposure of Kumasi residents to heavy  
388 metals and a metalloid increased the occurrences of lipid peroxidation and/or DNA damage.

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621 **Figure captions:**

622 **Fig. 1** Map showing human urine sampling locations in Kumasi, Ghana (yellow pins  
623 indicate city centre and environs contaminated with metals; red pins indicates human urine  
624 sites; white pin indicate KNUST campus). Obtained from (Bortey-Sam et al. 2017).

625

626 Table 1: Creatinine adjusted metal concentrations ( $\mu\text{g/g}$  creatinine) in human urine, collected in 2015, from four sites in  
 627 Kumasi, Ghana  
 628

Sample site	n		Cr	Mn	Co	Ni	Cu	Zn	As	Cd	Pb	MDA	8-OHdG
Atonsu	82	GM <sub>creat</sub>	1.50 <sup>a</sup>	0.375 <sup>a</sup>	0.855 <sup>a</sup>	4.20 <sup>a</sup>	15.7 <sup>a</sup>	386 <sup>a</sup>	67.3 <sup>a</sup>	0.289 <sup>a</sup>	0.615 <sup>a</sup>	54.9 <sup>a</sup>	1.02 <sup>a</sup>
		SD	2.21	2.94	4.00	6.30	25.6	291	61.1	0.980	8.27	68.2	1.99
Manhyia	51	GM <sub>creat</sub>	0.961 <sup>ab</sup>	0.220 <sup>a</sup>	0.734 <sup>a</sup>	1.19 <sup>b</sup>	16.1 <sup>a</sup>	350 <sup>a</sup>	43.5 <sup>b</sup>	0.227 <sup>a</sup>	0.636 <sup>a</sup>	40.1 <sup>ab</sup>	0.876 <sup>a</sup>
		SD	1.17	2.01	1.17	5.66	13.6	409	31.6	0.254	0.897	55.9	1.82
Tafo	57	GM <sub>creat</sub>	0.442 <sup>c</sup>	0.259 <sup>a</sup>	0.569 <sup>a</sup>	1.76 <sup>b</sup>	16.7 <sup>a</sup>	290 <sup>a</sup>	42.8 <sup>b</sup>	0.231 <sup>a</sup>	0.962 <sup>a</sup>	32.7 <sup>b</sup>	0.911 <sup>a</sup>
		SD	3.84	1.45	4.36	6.58	23.2	357	45.5	3.861	9.601	48.7	3.78
KNUST	12	GM <sub>creat</sub>	0.180 <sup>bc</sup>	0.115 <sup>a</sup>	0.481 <sup>a</sup>	0.524 <sup>b</sup>	5.67 <sup>b</sup>	180 <sup>a</sup>	15.9 <sup>c</sup>	0.074 <sup>b</sup>	0.312 <sup>a</sup>	23.7 <sup>ab</sup>	0.779 <sup>a</sup>
		SD	0.132	0.310	0.359	0.970	2.88	80.9	18.1	0.046	0.302	11.9	0.270

629 n: number of samples; nd: below limits of detection/quantification; GM<sub>creat</sub>: geometric mean concentration adjusted by  
 630 creatinine; SD: standard deviation; different letters (a, b and c) within a column indicates significant differences ( $p < 0.05$ )

631      Table 2: Correlation analysis between urinary concentrations ( $\mu\text{g/g}$  creatinine) of metals,  
 632      MDA, 8-OHdG and age of participants in Kumasi, Ghana

**Urban sites**

Variables	Age/years	Cr	Mn	Co	Ni	Cu	Zn	As	Cd	Pb	MDA	8-OHdG
Age/years	1	-0.0643	-0.0585	-0.337*	-0.0174	0.0698	0.0493	0.0466	0.11	-0.0314		
MDA	-0.181	0.532**	0.381**	0.669**	0.592**	0.660**	0.585**	0.777**	0.593**	0.586**	1	
8-OHdG	0.0812	0.171	0.217	0.218	0.223	0.360*	0.224	0.488**	0.267	0.108	0.453**	1

**KNUST  
(Control site)**

Variables	Age/years	Cr	Mn	Co	Ni	Cu	Zn	As	Cd	Pb	MDA	8-OHdG
Age/years	1	0.0352	-0.142	-0.305	-0.077	-0.250	-0.205	-0.217	-0.254	-0.707		
MDA	0.0451	0.357	0.339	0.271	0.311	0.332	0.348	0.371	0.263	0.182	1	
8-OHdG	-0.567	0.490	-0.013	0.411	0.077	0.340	0.242	0.270	0.224	0.221	0.340	1

633      \*: Indicates significance at  $p < 0.05$ ; \*\*: Indicates significance at  $p < 0.01$

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639 Table 3: Age differences in urinary concentrations ( $\mu\text{g/g}$  creatinine) of metals, MDA, and 8-OHdG among participants in  
 640 Kumasi, Ghana

Age groups	n		Cr	Mn	Co	Ni	Cu	Zn	As	Cd	Pb	MDA	8-OHdG
3-20	36	GM <sub>creat</sub>	1.66 <sup>a</sup>	0.400 <sup>a</sup>	1.24 <sup>a</sup>	3.26 <sup>a</sup>	20.3 <sup>a</sup>	425 <sup>a</sup>	51.6 <sup>a</sup>	0.296 <sup>a</sup>	1.18 <sup>a</sup>	50.5 <sup>a</sup>	0.557 <sup>a</sup>
		SD	1.02	0.43	4.59	4.74	17.2	572	51.2	0.619	8.18	98.9	0.558
21-40	102	GM <sub>creat</sub>	0.801 <sup>a</sup>	0.249 <sup>a</sup>	0.700 <sup>ab</sup>	2.36 <sup>a</sup>	14.2 <sup>a</sup>	297 <sup>a</sup>	49.3 <sup>a</sup>	0.224 <sup>a</sup>	0.664 <sup>a</sup>	43.0 <sup>a</sup>	1.07 <sup>a</sup>
		SD	3.39	2.75	2.82	5.61	24.9	256	51.1	3.08	7.87	48.2	3.48
41-60	33	GM <sub>creat</sub>	0.554 <sup>a</sup>	0.279 <sup>a</sup>	0.472 <sup>b</sup>	2.99 <sup>a</sup>	17.4 <sup>a</sup>	336 <sup>a</sup>	63.9 <sup>a</sup>	0.299 <sup>a</sup>	0.758 <sup>a</sup>	42.2 <sup>a</sup>	1.12 <sup>a</sup>
		SD	2.57	0.653	0.414	7.12	27.3	238	57.8	0.269	1.39	70.3	2.21
61-85	31	GM <sub>creat</sub>	1.25 <sup>a</sup>	0.421 <sup>a</sup>	0.572 <sup>ab</sup>	2.59 <sup>a</sup>	17.6 <sup>a</sup>	528 <sup>a</sup>	82.7 <sup>a</sup>	0.400 <sup>a</sup>	0.728 <sup>a</sup>	47.9 <sup>a</sup>	0.604 <sup>a</sup>
		SD	1.76	0.330	8.69	5.93	6.93	291	36.6	2.02	16.7	36.9	0.539

641 n: number of samples; nd: below limit of detection/quantification; GM<sub>creat</sub>: geometric mean concentration adjusted by  
 642 creatinine; SD: standard deviation; different letters (a and b) within a column indicates significant differences ( $p < 0.05$ )  
 643 among age groups

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658 Table 4: Sex differences in urinary metal, MDA and 8-OHdG concentrations ( $\mu\text{g/g}$  creatinine) in Kumasi, Ghana

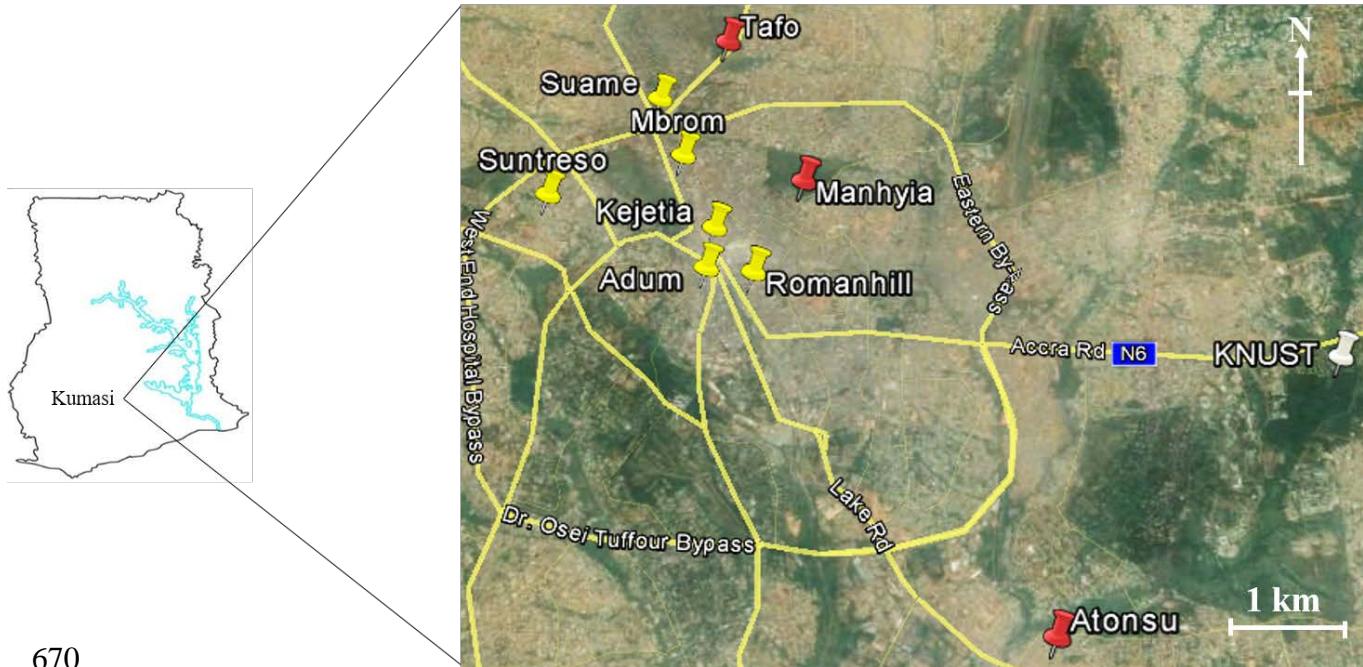
Sex	n		Cr	Mn	Co	Ni	Cu	Zn	As	Cd	Pb	MDA	8-OHdG
Female	138	GM <sub>creat</sub>	0.903 <sup>a</sup>	0.331 <sup>a</sup>	0.812 <sup>a</sup>	2.24 <sup>a</sup>	16.5 <sup>a</sup>	362 <sup>a</sup>	56.1 <sup>a</sup>	0.283 <sup>a</sup>	0.687 <sup>a</sup>	42.3 <sup>a</sup>	0.962 <sup>a</sup>
		SD	2.84	2.67	3.92	11.5	22.6	334	57.9	2.692	8.46	60.1	2.86
Male	64	GM <sub>creat</sub>	0.678 <sup>a</sup>	0.187 <sup>a</sup>	0.547 <sup>b</sup>	3.00 <sup>a</sup>	14.2 <sup>a</sup>	291 <sup>a</sup>	40.8 <sup>b</sup>	0.175 <sup>b</sup>	0.838 <sup>a</sup>	45.0 <sup>a</sup>	0.910 <sup>a</sup>
		SD	2.13	0.529	2.80	7.63	21.5	215	36.3	0.314	5.08	59.4	2.33

659 n: number of samples; nd: below limit of detection/quantification; GM<sub>creat</sub>: geometric mean concentration adjusted by  
 660 creatinine; SD: standard deviation; different letters (a and b) within a column indicate significant differences (Student's T-  
 661 Test);  $p <$  0.05)

662 Table 5: Adjusted odds ratios (OR; 95% CI) for the presence or absence of respiratory  
663 symptoms in Kumasi residents due to arsenic exposure

Metalloid	Clinical symptom	OR	CI		p value
As	Asthma	2.76	1.11	6.83	0.043
	RTI	2.42	0.075	7.86	0.618
	Tachycardia	3.93	1.01	15.4	0.041
	Rhinitis	0.848	0.145	4.97	0.855
	Dyspnea	1.09	0.344	3.47	0.880

664 OR: Odds ratio; CI: 95% Confidence Interval; RTI: respiratory tract infection; The models  
665 were adjusted for age and sex.  
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**Fig. 1**

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