



Title	Effects of biochar and litter on water relations of Japanese black pine (<i>Pinus thunbergii</i>) seedlings
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1 Effects of biochar and litter on water relations of Japanese black pine (*Pinus thunbergii*)
2 seedlings

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6
7 **Abstract**

8 Biochar is gaining increasing attention in the fields of forest rehabilitation, agriculture, etc. For evaluating biochar
9 application to improve the rhizosphere environment of Japanese black pine (*Pinus thunbergii*) at harsh coastal
10 environments, a pot experiment was carried out with a focus on ectomycorrhizal (ECM) symbiosis and water relations
11 of seedlings. Another characteristic recently observed at coastal forests is the invasion of the locust (*Robinia*
12 *pseudoacacia*), which can potentially increase soil nitrogen. In this study, four treatments were examined (biochar
13 addition, litter addition, biochar and litter addition, and no addition [control]) to determine the effects of biochar and/or
14 nitrogen-rich locust (*Robinia pseudoacacia*) litter application. Although effects of biochar and litter addition was not
15 observed on ECM symbiosis rate and species composition, treatments with biochar addition, maintained xylem water
16 potential (XWP) of needles for up to two weeks without irrigation, independent of litter addition. As biochar increased
17 relative fine root biomass (fine root biomass/total root biomass), it can be considered that biochar was able to maintain
18 needle XWP through increasing the relative amount of fine roots that can obtain water. Overall, these findings suggest
19 that biochar application can help to maintain water relations of Japanese black pine by enhancing fine root growth.

20
21 Keywords: *Robinia pseudoacacia*, Xylem water potential, ectomycorrhizal symbiosis, fine roots

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22 1 Introduction

23 Coastal sandy areas represent harsh growing conditions, such as drought conditions, lack of nutrients and
24 high salinity, all of which challenge newly planted seedlings. Field surveys have shown that Japanese black
25 pine (*Pinus thunbergii*, hereafter referred to as black pine) are associated with ectomycorrhizal (ECM)
26 species (Kataoka et al. 2008; Matsuda et al. 2009), which are suggested to play a vital role in improving
27 water and nutrient uptake from the soil (e.g. Smith and Read 1997), thereby enabling survival under harsh
28 coastal environments. Although a number of studies have examined the relationship between ECM
29 symbiosis and soil nutrient conditions (e.g. Marschner and Dell 1994; Choi et al. 2005; Zhang et al. 2017),
30 the role in water uptake and effects on water relations is much less clear (Lehto and Zwiazek 2011) . Several
31 studies, by comparing mycorrhizal and non-mycorrhizal seedlings, have showed that ECM symbiosis
32 improves seedling water relations (Dixon et al. 1980; Yin et al. 2018). Morte et al. (2001) reported that leaf
33 water potential was decreased by decreased soil moisture, however, the decrease was higher for non-
34 mycorrhizal than for mycorrhizal *Pinus halepensis* (Aleppo Pine) suggesting mycorrhizal seedlings may be
35 more tolerant to drought stress through enhanced water uptake. Furthermore, not only ECM infection but
36 ECM species composition may also affect seedling performance under water stress conditions (Kipfer et al.
37 2012).

38 Biochar, an impure form of elemental carbon with a highly porous structure and generated via
39 incomplete biomass combustion (Preston and Schmidt 2006) , is gaining increasing attention in the fields
40 of forestry, agriculture (Peng et al. 2019), and especially forest rehabilitation (e.g. Lehmann and Joseph
41 2015). Studies suggest that biochar has beneficial effects on plant growth and mitigates climate change
42 through enhancing carbon sequestration in managed ecosystems (Makoto et al. 2011b; Gale et al. 2017) as
43 well as natural regeneration after forest fires (Thomas and Gale 2015). Moreover, positive effects of biochar
44 on ECM symbiosis and fine root growth have also been reported (Ogawa 2007; Makoto et al. 2011a) via
45 increases in the availability of soluble/insoluble nutrients (e.g., Lehmann et al. 2003; DeLuca and Sala
46 2006; Makoto et al. 2011b) and improvements to the hydrological properties of sandy soils (Głąb et al.
47 2016). Biochar could thus work to enhance the rhizosphere environment and ECM symbiosis, potentially
48 improving water relations and nutrient conditions of black pine seedlings at coastal environments.

49 Another factor affecting ECM symbiosis at coastal forests is the presence of the woody legume: black
50 locust (*Robinia pseudoacacia*), which is in some cases planted as a “fertilizer tree” in coastal forests of
51 northern Japan (e.g. Masaka 2013). However, as it can spread rapidly, invasion by the locust is, in some
52 coastal forests, a recent common concern. It was reported by Taniguchi et al. (2007b) that the survival of
53 black pine seedlings growing in Locust dominant sites was significantly reduced by factors such light
54 intensity. Furthermore, *Locust* forms a symbiotic relationship with N-fixing rhizobia (Rice et al. 2004;
55 Marozas et al. 2015), and therefore can increase soil nitrogen (N) through the symbiosis and by shedding
56 high N content litter (Landgraf et al. 2005). Edaphic changes by the presence of black locust trees are
57 subsequently thought to affect ECM infection and ECM species composition (Aučina et al. 2007; Taniguchi
58 et al. 2007a) . Hence, environmental changes caused by Black locust may affect ECM species composition
59 and formation of black pine seedlings, especially at coastal forests.

60 The objective of this study was to evaluate the effects of biochar on black pine seedlings with and
61 without black locust leaf litter, focusing on plant water relations in order to evaluate biochar use at coastal
62 forests to mitigate drought stress. As fine roots play the vital role in absorbing water and nutrients from the
63 soil, and especially, new fine roots are where ECM colonize (Smith and Read 1997), investigations were
64 also made on ECM symbiosis. We hypothesised that addition of biochar would enhance plant water
65 relations by promoting ECM symbiosis and or changing species dominance and enhancing fine root growth.
66 Moreover, it was also hypothesized that the locust litter could work to decrease ECM symbiosis as it can
67 be expected that soil N will increase.

68

69 **2 Materials and methods**

70 **2.1 Plant material**

71 In September 2013, 40 two-year-old black pine seedlings produced in Niigata Prefecture, central Japan,
72 were planted in 17-L pots (27 cm diameter, 30 cm deep) containing river sand to resemble soil conditions
73 of Niigata coastal forests. Sand conditions were infertile, with only trace levels of N, phosphorus, and
74 potassium. The sand was commercially obtained from DALTEC Co., Ltd. (Sapporo, Japan) to uniform soil
75 conditions so that direct effects of biochar and litter on ECM symbiosis could be observed. The sand had a
76 particle diameter of 0.3–2.0 mm. Refer to Table 1 of Makoto et al. (2011a) for sand properties. The

77 seedlings were grown under natural sunlight (90% transmittance of natural sunlight; daytime photon flux
78 density on a fine day: 1300–1600 $\mu\text{mol m}^{-2} \text{s}^{-1}$; cut wavelength: <330 nm; almost no UV-B or C) inside a
79 greenhouse of the Graduate School of Agriculture, Hokkaido University from September 2013 to October
80 2014. Seedlings were grown in a greenhouse to avoid further natural ECM contamination. The temperature
81 and humidity inside the greenhouse were maintained at 23°C (± 8) and 45% (± 7), respectively. Seedlings
82 were watered with tap water once a week until mid-October 2014, when irrigation was stopped for XWP
83 measurements.

84

85 **2.2 Soil treatment**

86 To determine the individual and combined effects of biochar and litter on water relations and ECM infection
87 of black pine seedlings, four different soil treatments were examined: 1) biochar treatment, 2) litter
88 treatment, 3) biochar + litter treatment (both biochar and litter addition), and 4) no litter or biochar treatment
89 (control). The soil treatments began at the start of the experiment in September 2013.

90 The biochar which was used in this experiment was produced in the laboratory by incinerating Japanese
91 larch (*Larix kaempferi*) at 400°C from Forest Chemistry Lab. of Hokkaido University via residues of
92 harvested timbers of Sorachi Forest Office of Hokkaido Forest Management Bureau, following the method
93 of Makoto et al. (2011a). The produced biochar was crushed to a uniform size using a 5 mm mesh according
94 to methods of Makoto et al. (2010). In total, 25 g of biochar was added at a depth of approximately 5 cm.
95 The methodology of biochar addition followed results of Makoto et al. (2010), where addition of a layer of
96 charcoal in the middle of the soil profile promoted growth of the root system of the seedlings significantly
97 more than did pots without charcoal or with charcoal scattered throughout the soil. The amount of biochar
98 was set at a volume ratio of 1:4 (biochar: sand), which resulted in approximately 5 cm biochar layer. The
99 pH (H_2O) and density of the biochar was 6.0 ± 0.1 and $0.21 \pm 0.02 \text{ g mL}^{-1}$, respectively. Refer to Table 1
100 of Makoto et al. (2011a) for further chemical properties.

101 In the litter treatments, 7 g of litter, which is equivalent to approximately 20 mg of N when decomposed,
102 was added. This was based on the information that the average N content of black locust leaves is
103 approximately 20–30 mg g^{-1} (Choi et al. 2017). This value was also assumed, considering effects of
104 leaching. The litter used in this experiment was collected from a black locust-dominated area in Sapporo

105 Experimental Forest of Hokkaido University (Kanetoshi et al. 2009) in autumn, 2013 right before the
106 experiment started. The collected litter was fallen fresh yellow-brown leaves, however including about 20%
107 in volume of pale green leaves as leaves of black locust hardly changes color to yellow due to symbiotic
108 legume activities. The leaves were not yet decomposed. The N content of the locust litter was determined
109 using a NC analyser (NC-900; Sumica-Shimadzu, Osaka, Japan) as $18.5 \pm 0.6 \text{ mg g}^{-1}$. The litter was mixed
110 with the sandy soil and then applied as a top layer of approximately 5 cm thick, which resembled *in-situ*
111 conditions obtained from field survey at the Niigata costal forest. Locust litter was used instead of mix
112 planting black pine and black locust together to avoid complications of competition such as light, direct
113 effects on ECM symbiosis, and so-called “Allelopathy effects”, as suggested by Nasir et al. (2005) and
114 Taniguchi et al. (2007).

115 The initial mean stem base diameter (SBD) \pm standard error under biochar treatment, litter treatment,
116 litter + biochar treatment, and no treatment (control) were 4.1 ± 0.1 , 4.9 ± 0.3 , 4.1 ± 0.1 , and 4.4 ± 0.2 mm,
117 respectively.

118

119 **2.3 Measurements and analyses**

120 **2.3.1 XWP of the needles**

121 Xylem water potential of needles was measured to examine the water relations of black pine seedlings.
122 Four randomly selected seedlings from each treatment group were not given water from October 17, 2014,
123 to examine the XWP of the needles. Needles were sampled before dawn at approximately 2-3 day intervals
124 between October 20 and November 3, 2014 (total six measurements). The XWP of the needles were
125 immediately measured using a pressure chamber (Model-3000; Soil Moisture Equipment Corp., Santa
126 Barbara, CA).

127

128 **2.3.2 ECM identification**

129 ECM identification was done on five seedlings for each treatment (four of which were measured for XWP
130 and dry weight). After harvest, roots were immediately covered with wet paper towels, placed in a plastic
131 bag, and transferred to the laboratory, where they were stored in a refrigerator at 4°C until further analysis.
132 The roots were carefully washed to remove remaining soil clods and further washed with a paintbrush until

133 all soil was removed. A total of 200–300 root tips per seedlings (1200–1500 root tips were observed per
134 treatment) were randomly selected from the top, middle, and bottom of the root system. This method
135 followed the methods of Shinano et al. (2007) and Wang et al. (2015).

136 ECM taxa were first classified by microscopic observation (Olympus SZX-ILLK100, Japan) based on
137 their morphological characteristics, such as colour, texture, level of ramification, root tip shape and
138 morphology of the emanating hyphae. The observed root tips were then stored in a freezer at -20°C until
139 further analysis. Taxonomic classification based on the morphology was verified by DNA analysis using
140 three to five tips with identical morphotypes.

141 DNA was extracted using ISOPLANT (Nippon Gene Co., Ltd., Japan). Three cycles of centrifugal
142 separation were carried out at $12,000 \times g$ (11,441 rpm, 4°C) for 15, 4 and 1 minute, respectively. The
143 extracted DNA was amplified by polymerase chain reaction (PCR) using a thermal cycler (Applied
144 Biosystems 2720 Thermal Cycler). PCR was carried out using a primer set that reads the internal transcribed
145 spacer (ITS) region of ribosomal DNA with primer ITS1 (Fungal rDNA [ITS1] PCR Kit Fast; Takara Bio
146 Inc., Japan) under following conditions; one cycle at 94°C for 3 minutes, 25 cycles at 94°C for 20 seconds
147 $\rightarrow 50^{\circ}\text{C}$ for 20 seconds $\rightarrow 72^{\circ}\text{C}$ for 5 seconds and one cycle at 7°C for 7 minutes. The samples were then
148 stored at 8°C . DNA samples were confirmed by visual classification, using gel electrophoresis with a gel
149 made from 0.3 - 0.6 g of Agarose S (Nippon Gene Co., Ltd., Japan) and 15 - 30 mL of $0.5 \times$ Tris-borate-
150 EDTA buffer and were refined using the Fast Gene® Gel/PCR Extraction Kit (Nippon Gene Co., Ltd.,
151 Japan). The base sequence of each refined sample was decoded by a professional service using an ABI
152 3730xl DNA Analyser. Finally, obtained ECM sequences were compared with the GenBank database at the
153 DNA Nata Bank of Japan using the basic local alignment search tool (BLAST). If only one species had
154 high sequence homology, the sample was identified as that particular species. However, if several species
155 had high sequence homology, the sample was considered an unidentified species from a particular family.

156 From the obtained DNA identification, the ratio of dominance (P_s) of each species were calculated as
157 follows:

$$P_s = \frac{Q_{ECM}}{Q_{T-ECM}}, \quad \text{Eq. 1,}$$

158 where Q_{ECM} is the number of root tips infected by a particular ECM species in a given treatment and Q_{T-ECM}
159 is the total number of root tips infected by ECM fungi in a given treatment.
160 The total ECM symbiosis (R_E) rate was calculated by the following equation:

$$R_E = \frac{Q_T - Q_N}{Q_T} \times 100, \quad \text{Eq. 2,}$$

161 where Q_T is the total number of root tips observed per treatment and Q_N is the total number of root tips not
162 infected with ECM fungi.

163

164 **2.3.3 N content of the needles and soil**

165 In December 2014, needle and soil samples (taken approximately 5 cm below the surface) were collected
166 from four seedlings, which were measured for XWP and ECM analysis ($n=4$ for each treatment). The N
167 content (mg g^{-1}) of each sample ($n = 4$) was then measured using a CN analyser (Vario EL III; Elementar,
168 Ronkonkoma, NY).

169

170 **2.3.4 Biomass**

171 At the end of the experiment, the dry biomass and SBD were measured in the seedlings which were
172 measured for XWP and ECM fungi identification ($n=4$). Each plant was sorted into needles, buds, branches,
173 stems, roots, and fine roots (diameter < 2 mm) were separated from coarse roots (diameter > 2 mm). Relative
174 fine root biomass was also calculated by the following equation; fine root biomass/total root biomass $\times 100$.
175 All samples were dried at 70°C for at least 72 h prior to measurement.

176

177 **2.4 Statistical analysis**

178 The statistical difference between each treatment groups were analysed by one-way ANOVA, if significant,
179 Tukey's post-hoc test was carried out with a significance level of $p < 0.05$. It was also analysed that the
180 initial seedling size (initial stem base diameter) did not have a significant effect on the final results. The
181 data were assumed to follow a normal distribution. The relationship between needle XWP and the relative
182 fine root biomass was examined using linear regression analysis with a significance level of $p < 0.05$. All
183 analyses were performed using R (ver. 3.4.3).

184

185 **3 Results**

186 **3.1 XWP of needles**

187 XWP was measured six times at an interval of 2–3 days. Until 10 days after ceasing irrigation, no difference
188 was observed between treatment groups (Fig. 1). However, from 10 days, the biochar treatment and biochar
189 + litter treatment showed a higher XWP value compared to the control and litter treatment. This trend
190 continued till 17 days after stopping irrigation, and the biochar treatment and biochar + litter treatment
191 showed a more gradual decrease than the other two groups.

192

193 **3.2 ECM identification and species dominance**

194 *Wilcoxina mikolae*, *Amphinema* sp., *Russula* sp., *Suillus grandulatus*, and *Rhizopogon roseus* were
195 determined from the DNA identification (Table 1, Table 2). In all four treatments, *Wilcoxina mikolae*,
196 *Amphinema* sp. and *Russula* sp. were observed. At the biochar treatment, 6 species (including unknown
197 species) were observed, which was the most compared to other treatments. *Suillus grandulatus* was only
198 observed in this treatment and *Rhizopogon roseus* was only observed at the biochar treatment and the
199 biochar + litter treatment.

200 The ECM symbiosis rate (R_E) for each treatment was as follows; 99.93% in the biochar treatment,
201 98.50% in the litter treatment group, 99.12% in the biochar + litter treatment group, and 99.01% in the
202 control group (Table 2). Thus, neither litter nor biochar addition influenced R_E . In all treatments, P_S of
203 *Wilcoxina makolae* was the largest value, making it the most dominant species. At the biochar treatment,
204 although not statistically significant, P_S of *Amphinema* sp. was over 20%, meanwhile in the other treatments,
205 it was less than 5%. P_S of *Amphinema* sp. was second highest at the litter treatment. The P_S of other species
206 was all less than 5%.

207

208 **3.3 Needle and soil N contents**

209 The N content of needles and soil are shown in Table 3. For needle N content, difference between treatments
210 were not detected. On the other hand, for soil N content, it was increased at the biochar + litter treatment
211 compared to the other three treatments. Therefore, although soil N was not increased at the litter treatment,
212 it was increased at the biochar + litter treatment.

213 3.4 Growth and dry weight

214 Results of growth and, above- and below-ground dry weight are shown in Table 4 and 5. Concerning the
215 final stem base diameter, the smallest value was the biochar + litter treatment, and the other three treatments
216 showed similar results. A similar trend was also observed for final height.

217 Needle and branch dry weight were not affected by litter or biochar addition. For bud dry weight, the
218 litter treatment showed the highest value, and the biochar + litter treatment showed the smallest value.
219 Biochar addition did not have any effect on above-ground organ and dry weight. For stem dry weight, the
220 biochar + litter treatment showed the smallest value of all treatments. Concerning roots, biochar addition
221 had a significant increasing effect on fine root dry weight and also relative fine root (fine root dry
222 weight/total root dry weight). However, when both biochar and litter were added, neither fine root dry
223 weight and relative fine root were not increased. Furthermore, although from personal observation, the
224 increase in fine roots were especially observed at the middle part of the pot (5~8 cm from the surface),
225 relatively close to where the biochar was added.

226 As biochar addition increased fine root dry weight, consequently, the above-/below-ground (fine +
227 coarse root biomass) ratio showed the smallest value at the biochar treatment, however, the value was
228 highest at the biochar + litter treatment (Table 5).

229

230 4 Discussion

231 The addition of biochar significantly improved water relations of black pine seedlings, where XWP of the
232 needles were maintained for over 10 days regardless of litter addition (Fig. 1). However, in contrast with
233 our hypothesis, addition of biochar and litter had no effect on the rate of ECM symbiosis (R_E) and species
234 abundance (Table 2). That is, high rates of R_E were observed in all treatment groups, including those with
235 litter addition.

236 It was previously revealed that addition of N caused a decrease in both R_E and the number of ECM
237 species (Wallenda and Kottke 1998). In other studies, it has also been revealed that addition of litter had a
238 greater effect on the relative proportion of ECM symbionts than on species diversity (Aučina et al. 2007).
239 Furthermore, a decrease in R_E was previously observed in a pine stand rich in soil N due to the addition of
240 locust litter (Taniguchi et al. 2007a). However, in the latter study, 0.76 mg N g⁻¹ was detected at a depth of

241 0–5 cm, approximately three- to four-fold more than that observed under our litter treatment (Table 3,
242 approximately 0.20 mg N g⁻¹). We initially calculated that 20 mg N g⁻¹ would be added to each pot through
243 litter addition, however, this value assumed that all litter were decomposed, which may not have been the
244 case since this study ran for only one growing season and the given litter was fresh yellow-brown leaves
245 including pale green leaves collected right before the experiment. It is therefore possible that the amount of
246 litter added in our experiment was not sufficient to affect ECM fungal composition and diversity.
247 Furthermore, this may be due to the immobilization of nutrients by microorganism increased by the addition
248 of fresh leaf litter (undecomposed leaf litter). Litter addition did not decrease fine root dry weight, although
249 a decrease in fine roots (or increase in fine root turnover) is often reported with increasing soil N as the
250 necessity of roots decrease with the increase of relative nutrients, which also indicates that the added N
251 amount was relatively small. Nevertheless, it can be assumed that short-term litter accumulation at a soil
252 depth of 5 cm does not negatively affect R_E . However, effects of biochar and litter combined are still not
253 clear as in bud and stem dry weight, and final stem diameter and height showed the smallest value at the
254 biochar + litter treatment. No clear explanation could be made on this result, and further analyses must be
255 made on the combination effect of biochar and litter.

256 *W. mikolae* (Ascomycetes) was the dominant ECM species in all four treatment groups (Table 2). This
257 taxon is a generalist that is able to grow under a wide range of environmental conditions and often forms
258 symbiotic relationships with coniferous species (e.g. Baar et al. 1999; Ashkannejhad and Horton 2006). At
259 the initial stage of our experiment, morphological identification also identified *W. mikolae* as the dominant
260 species, suggesting that neither biochar nor litter addition affected species composition.

261 Although no effects of biochar and litter were observed on ECM symbiosis, significant effects were
262 observed on needle XWP (Fig. 1). The XWP of the seedlings under biochar treatment was consistently high
263 even without watering for 15 days, and independent of litter addition. Similar effects of biochar on leaf
264 water potential were observed under drought conditions in pear (*Pyrus ussuriensis* Maxim.) (Lyu et al.
265 2016), with a high XWP observed up until 11 days of drought. Meanwhile, the results of our study suggest
266 that this positive effect may last for up to 17 days. Concerning results of fine roots, both fine root dry weight
267 and relative fine root was increased by biochar addition. This trend is supported by previous findings in
268 which biochar enhanced root growth, particularly that of fine roots (e.g. Makoto et al. 2011a). Biochar may
269 have enhanced the soil's aeration property and induced fine root growth as roots of black pine is reported

270 to have a high demand in oxygen and prefer non-excessive moisture conditions (Karizumi 2012).
271 Additionally, since fine roots function to absorb water from the soil, it can be considered that biochar
272 addition was able to maintain needle XWP after ceasing irrigation as seedlings with biochar had relatively
273 more fine roots to obtain water. Accordingly, a significant correlation was observed between relative fine
274 root biomass and XWP at 17 days after stopping irrigation; seedlings with a high relative fine root biomass
275 also showing a high XWP (Fig. 2). These findings suggest that the addition of biochar improves plant water
276 relations by enhancing fine root growth (Prendergast-Miller et al. 2014).

277 Overall, findings of this study suggest that biochar has positive effects on water relations in black pine
278 seedlings as determined by observations of needle XWP and enhanced fine root growth. As the added
279 amount lead to increase in fine root biomass, it may also be the suggested amount for field application.
280 However, effects of biochar in combination with locust litter amount may need further investigations.
281 Furthermore, long-term and field studies may also be further required to further understand the combined
282 effects of locust litter (and the tree itself) and biochar.

283

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293

294 **Declaration on interest**

295 No potential conflict of interest was declared by the authors.

296

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