FREEZING, MELTING AND LIGHT STRESS ON THE PHOTOPHYSIOLOGY OF ICE ALGAE: *EX SITU* INCUBATION OF THE ICE ALGAL DIATOM *FRAGILARIOPSIS CYLINDRUS* (BACILLARIOPHYCEAE) USING AN ICE TANK[1]

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RUNNING HEAD: Ice algal physiology in an ice tank

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ABBREVIATIONS:

cDNA, complementary DNA; Chl *a*, chlorophyll *a*; ChlF, active chlorophyll *a* fluorescence; DD, diadinoxanthin; DT, diatoxanthin; DES, de-epoxidation state index of DD-DD xanthophyll cycle; ETR, electron transport rate; Fe', inorganic Fe species; FRRf, fast repetition rate fluorometry; HL, high light; LL, low light; NPQ, non-photochemical quenching; PQ, plastoquinone; *PP*, primary productivity; PPC, photoprotective carotenoid; PSC, photosynthetic carotenoid; Q_A, the first quinone electron acceptor; Q_B, secondary quinone electron acceptor; Tchl *a*, total chl *a*; UHPLC, ultra-high performance liquid chromatography

ABSTRACT

47 Sea-ice algae contribute up to 25% of the primary productivity of polar seas and seed
48 large-scale ice-edge blooms. Fluctuations in temperature, salinity, and light associated
49 with the freeze/thaw cycle can significantly impact the photophysiology of ice-
50 associated taxa. The effects of multiple co-stressors (i.e., freezing temperature and high
51 brine salinity or sudden high light exposure) on the photophysiology of ice algae were
52 investigated in a series of ice tank experiments with the polar diatom *Fragilariopsis*
53 *cylindrus* under different light intensities. When algal cells were frozen into the ice, the
54 maximum quantum yield of photosystem II photochemistry (PSII) (F_v/F_m) decreased
55 possibly due to the damage of PSII reaction centres and/or high brine salinity stress
56 suppressing the reduction capacity downstream of PSII. Expression of the *rbcL* gene
57 was highly upregulated, suggesting that cells initiated strategies to enhance survival
58 upon freezing in. Algae contained within the ice-matrix displayed similar levels of
59 F_v/F_m regardless of the light treatments. Upon melting out, cells were exposed to high
60 light (800 μmol photons m^{-2} s^{-1}), resulting in a rapid decline in F_v/F_m and significant
61 upregulation of non-photochemical quenching (NPQ). These results suggest that ice
62 algae employed safety valves (i.e., NPQ) to maintain their photosynthetic capability
63 during the sudden environmental changes. Our results infer that sea ice algae are highly
64 adaptable when exposed to multiple co-stressors and that their success can, in part, be

explained by the ability to rapidly modify their photosynthetic competence; a key factor contributing to algal bloom formation in the polar seas.

67 INTRODUCTION

68 Sea ice algae are significant primary producers in sea ice zones. They contribute
69 5−25% of the total primary production (*PP*) in seasonally ice-covered regions
70 (Legendre et al. 1992, Lizotte 2001, Michel et al. 2006, Loose et al. 2011, Arrigo 2017)
71 and ~50% of the annual *PP* in perennially ice-covered regions (Satoh et al. 1989,
72 McMinn et al. 2010, Fernandez-Mendez et al. 2015). Ice-associated algae are of high
73 ecological importance as not only the primary energy source for sea ice biota such as
74 zooplankton, krill, larval fish and benthos (Lizotte 2001, Boetius et al. 2013, 2015,
75 Kohlbach et al. 2017, Moteki et al. 2017, Bernard et al. 2018) but also as seed biomass
76 for ice-edge blooms in marginal sea ice zones (e.g., Smith and Nelson 1986, Syvertsen
77 1991, Arrigo and van Dijken 2003, Arrigo 2014). Ice-edge blooms, which ice algae may
78 seed, contribute ~50% of the total *PP* in marginal sea ice zone of the Southern Ocean
79 (Smith and Nelson 1986, Sakshaug and Slagstad 1991, Deppeler and Davidson 2017),
80 although it is still controversial which ice algae or phytoplankton actually contribute to
81 the blooms (see review of van Leeuwe et al. 2018). Upon freezing in, algae can form
82 aggregates within microstructures of the ice, which channel excluded liquid brine from
83 sea ice to under-ice seawater when ice grows (i.e., brine channels) (Melnikov and
84 Bondarchuk 1987, Assmy et al. 2013, Boetius et al. 2013, Katlein et al. 2015,

85 Fernandez-Mendez et al. 2014, Boetius et al. 2015). During the sea ice melt, these algal
86 aggregates can rapidly sink and transport carbon down to the sea floor. This process
87 contributes to the biological carbon pump (Riebesell et al. 1991, Taguchi et al. 1997,
88 Boetius et al. 2013, Katlein et al. 2015) and underlines the important contribution of ice
89 algae to both the ecology and biogeochemistry of sea ice zones. It is thus important to
90 understand how ice algae survive in the dynamic and heterogeneous sea ice
91 environment. Sea ice environments are subject to large fluctuations in temperature
92 (~−20 ºC to −1.8 ºC; Petrich and Eicken 2017), salinity (~25 to 170; Arrigo et al. 2010)
93 and light availability (~0 to 1000 μmol photons m^{-2} s^{-1}; Galindo et al. 2017). Variation
94 in these abiotic parameters can influence the photosynthetic efficiency of ice algae,
95 however, they are not independent (i.e. multiple co-stressors; McMinn et al. 2017). Few
96 studies have addressed the influence of multiple co-stressors on the photophysiology of
97 sea ice algae (Ralph et al. 2005, 2007, Ryan et al. 2011, Martin et al. 2011, McMinn et
98 al. 2014; see review of McMinn 2017), and little is known about how co-stressors
99 influence ice algal photosynthesis and production during freeze/thaw cycles (McMinn
100 2017). Despite their significant ecological and biogeochemical importance, it remains
101 difficult to directly obtain the photophysiological properties of sea ice taxa *in situ.*
102 Kennedy et al. (2012) were the first to use sea ice tanks to undertake short-term

incubation of the haptophyte *Phaeocystis antarctica* within the laboratory. This method has the advantage that ice algae can be incubated in an environment mimicking pack ice conditions. Although many studies have addressed ice algal photosynthesis and production using isolates resuspended in water (e.g., Yan et al. 2019), it remains difficult to incubate them in the laboratory (e.g. in a culturing flask), while simulating realistic environmental conditions. Using a previous ice tank model, Kennedy et al. (2012) found that a freezing event suppressed the photochemical efficiency of PSII. Although these experiments were able to reveal the effects of freezing on the photophysiology of ice algae, their short-term nature did not address the longer-term mechanism of freezing stress. Understanding how ice algae maintain photosynthetic capacity and survive during freeze/thaw cycles is of considerable importance to better assess primary productivity in sea ice environments. In this study, a novel 70 L ice tank was constructed for the long-term incubation of ice algae under ecologically realistic conditions. It was used to investigate the effects of multiple co-stressors on ice algal photophysiology:

(1) freezing stress with low temperature and high brine salinity;

(2) chronic low light availability under low temperature and high salinity;

(3) melting stress with exposure to less saline meltwaters and sudden increase in light.

The sea ice diatom *Fragilariopsis cylindrus* was maintained in the ice tank for 20 days followed by ice melt and light exposure experiments to reproduce the environment that released ice algae experience. This study aims to investigate the effects of multiple co-stressors on ice algal photophysiology in an artificial sea ice environment during a freeze/thaw cycle. It provides new insights into how ice algae are able to tolerate environmental fluctuations that contribute to their dominance of ice-edge blooms.

MATERIALS AND METHODS

Ice tank incubation

The polar pennate diatom *Fragilariopsis cylindrus* (isolated from Antarctic sea ice in 2015, Davis station, East Antarctica; Kennedy et al. 2019), was incubated in a purpose designed ice tank (Island Research, Tasmania). The algal culture was less than two years old at the start of the experiments. The ice tank was contained within a freezer (–20 ºC), and the ice thickness and temperature gradient of the ice were controlled by interactions between a basal heater and the adjustable ambient freezer temperature (Fig. 1). This enabled an ice thickness of approximately 5.5 cm to be maintained during the experiment. *F. cylindrus* was incubated in Aquil media (Price et al. 1989), buffered with

138 ethylenediaminetetraacetic acid (EDTA) (final concentration 5.00 μM) at a salinity of
139 35, and a Fe concentration of 400 nM. The concentration of total inorganic forms of Fe
140 (Fe') was 1.54 nM, this being continuously supplied from the Fe-EDTA complexes to
141 the medium calculated using the software Visual MINTEQ, ver. 3.1
142 (https://vminteq.lwr.kth.se). Light intensities beneath the ice, measured with a spherical
143 photosynthetically active radiation (PAR) sensor (QSL-100, Biospherical Instrument
144 Inc.), were set at 150 and 30 μmol photons m^{-2} s^{-1} (White LED, PANEL-300-18W,
145 LED Lighting Products, Australia; Fig. S1) as high and low light (HL and LL)
146 treatments, respectively. Two discrete ice tank runs were conducted for each light
147 treatment. The diatom *F. cylindrus* was inoculated with low cell densities (HL: 4011 ±
148 151 cells mL^{-1}; LL: 3933 ± 132 cells mL^{-1}) to avoid the possibilities of self-shading
149 and nutrient starvation during the incubation experiments. They were maintained in the
150 ice tank for 3 days for acclimation where seawater temperature was maintained at 2.5
151 ºC. Prior to freezing in, a sample was taken to assess the original physiological state of
152 the algae (day −05, hereafter). The temperature lowered to −1.8 ºC to initiate ice
153 formation. Once ice had formed after 3 days, the under-ice water was partially replaced
154 with ultrapure water to reduce the salinity down to 35; as the salinity of the underlying
155 water had increased (to ~38) as a result of brine rejection. Two-day acclimation period

156 was used prior to sampling. Ice sampling was conducted every 5 days for 20 days (days
157 00, 05, 10, 15, and 20). Brine salinity and brine fraction were estimated following Cox
158 and Weeks (1983) and Eicken (2009). To minimize heterogeneity among ice cores, ice
159 samples were randomly collected from the tank chamber (pseudo-replicate) with a trace
160 metal-free hand drill (2 cm in diameter) from randomly annotated grids on the ice
161 surface, following normal random sampling numbers generated by the software R
162 (https://www.r-project.org/). To assess both the effects of melting and high light
163 exposure, ice samples were melted at 2.5 ºC and salinity 25 for 2 days (melt samples,
164 hereafter). The melt samples were then exposed to intense light (light samples,
165 hereafter) (800 μmol photons m^{-2} s^{-1}) to mimick the typical summer surface
166 (MODIS/Aqua).

167

168 *Fast repetition rate (FRR) fluorometry*

169 To monitor the photophysiology of *F. cylindrus* during the freezing and melting
170 processes, variable chlorophyll *a* fluorescence (ChlF) measurements were conducted
171 using a bench-top Fast Repetition Rate fluorometer (FRRf) (FastOcean Act2Run
172 Systems, Chelsea Technologies) with Act2Run software (Chelsea Technologies).
173 Thawed ice samples were buffered with filtered seawater (ice:seawater ratio = 1: 1) at

174 2.5 ºC in the dark for 30 min, and the slushy melted ice samples were placed in a quartz
175 cuvette to measure fluorescence. A single turnover protocol was applied for ChlF
176 measurement; 100 flashlets with 1 μs duration at a wavelength 450 nm and 2 μs
177 intervals for excitation of reaction centres of PSII and 20 flashlets with 1 μs duration
178 and 20 flashlets with 1 μs duration and 100 μs intervals for relaxation. Eighteen light
179 steps were applied to generate a rapid light curve (RLC) from 0 to 800 μmol photons
180 m^{-2} s^{-1}, taking <5 min to complete one RLC. At each light step (~15 s), at least five
181 induction and relaxation curves were averaged to obtain ChlF yields, described in Table
182 1, after calibrating the ChlF yields with filtered seawater. According to the models
183 proposed by Kolber et al. (1998), photosynthetic parameters of chlorophyll *a* (chl *a*)
184 induction curves were calculated based on the ChlF yields as shown in Table 1. Electron
185 transport rate through the reaction centres of PSII (RCII) (ETR_{RCII}) was calculated as
186 follows:

188 $$\mathrm{ETR_{RCII}} = E \times \sigma_{PSII} \times \frac{F'_\mathrm{q}/F'_\mathrm{m}}{F_\mathrm{v}/F_\mathrm{m}} \times \Phi_\mathrm{RCII} \times 6.022 \times 10^{-1}$$

187

189 following Suggett et al. (2011) and Schuback et al. (2016), an alternative approach to
190 calculating ETR_{RCII} of Kolber et al. (1998), wherein Φ_{RCII} is the quantum yield of RCII

191 assumed as 1, and the following number is a conversion factor to mol quanta mol
192 $RCII^{-1}$ s^{-1}. The calculated ETR_{RCII} at each light step were fitted to the model of Platt et
193 al. (1980) to obtain photosynthesis−irradiance (ETR_{RCII}−*E*) parameters (Table 1). To
194 assess the heat dissipation of algal cells, non-photochemical quenching based on the
195 normalized Stern-Volmer (S-V) coefficient (NPQ_{NSV}') under the actinic light equivalent
196 to the incubation light intensities was also calculated following McKew et al. (2013):

197 $$\mathrm{NPQ_{NSV}}' = F_o'/F_v'$$

198

199 where F_o' is a minimum ChlF yield after a relaxation sequence calculated following
200 Oxborough and Baker (1997) (Table 1).

201

202 *Validation experiments of melting procedures of ice samples for ChlF measurement*
203 While ChlF has been widely used on seawater samples (e.g., Suggett et al. 2011), it
204 has not been broadly applied to ice algae research (Miller et al. 2015) because of the
205 need for the ice samples to be melted. Melting the ice samples changes the
206 photophysiology of the algal cells (e.g., Garrison and Buck 1986, Mikkelsen and
207 Witkowski 2010, Rintala et al. 2014, Campbell et al. 2019). Here, multiple comparisons

were conducted on melt procedures of an ice sample from the ice tank. Measurements of ChlF were performed on three treatments of ice samples; (1) intact ice, (2) slushy melted ice suspended in filtered seawater (ice: seawater ratio=1: 1), (3) slowly (24 h) and directly melted samples. The blank calibration for F_o was conducted with filtered seawater as described, however, an empty quartz tube was used for the calibration because it was impossible to prepare a blank ice sample for the treatment 1. It was thus assumed that the ice samples for the treatment had a high enough biomass without F_o calibration. All samples were stored or melted in the dark for 30 min for the treatments 1 and 2, whereas the other samples (i.e. the treatment 3) were kept at 2.5 ºC during the overnight melting. The FRRf was used for the ice samples from the ice tank as described above.

Cell abundance of ice algae

Triplicated microscopic samples were fixed with neutralized formaldehyde solution (final concentration 2%). Cell abundance of *F. cylindrus* was determined with an inverted microscope (Axiovert 25, Zeiss) and a Sedwick-Rafter counting chamber (Pyser-SGI Lmd.). At least 500 cells were enumerated to determine cell abundance (LeGresley and McDermott 2010). Growth rate was calculated following Wood et al.

226 (2005):

228 $$\mu = \frac{\ln(N_t/N_0)}{t - t_o}$$

227

229 *Pigment composition*

230 Pigment concentrations were determined to quantify chlorophylls, photosynthetic and
231 photoprotective carotenoids, and chlorophyll derivatives with a Ultra-High Performance
232 Liquid Chromatography (UHPLC) (Suzuki et al. 2015). On each sampling day,
233 triplicated ice or seawater samples were filtered onto 25 mm GF/F filters (Whatman)
234 using gentle vacuum (<0.013MPa) via a 25 mm polypropylene in-line filter holder
235 (Swinnex, Merck). Rintala et al. (2014) confirmed that fast and direct melting was a
236 reliable method for pigment analysis in ice algae. The filter was flash frozen in liquid
237 nitrogen and stored in a deep freezer (−80 ºC). After thawing, the filter was blotted dry
238 with a filter paper, and the pigments were extracted using the *N*, *N*-dimethylformamide
239 (DMF) bead-beating method (Suzuki et al. 2015). The extracted pigments were
240 suspended in DMF and then injected into an UHPLC for pigment determination. The
241 ratio of diadinoxanthin (DD) and diatoxanthin (DT) was calculated to assess the
242 xanthophyll epoxidation-de-epoxidation cycle and photoprotective strategies of algae
243 (Katayama et al. 2017), assessed as a de-epoxidation state (DES; DT/(DD+DT)). Total

chl *a* (Tchl *a*: sum of chl *a*, chlorophyllide *a*, chl *a*-allomer, and chl *a*-epimer) concentrations were calculated as an indicator of algal biomass. Contributions of chlorophyllide (chllide *a*) *a* to Tchl *a* were also calculated as an index of the breakdown of chl *a*. Following Schuback et al. (2016) and Yan et al. (2019), ratios of photoprotective carotenoids (PPC: DD, DT, and β,β-carotene) to photosynthetic carotenoids (PSC: only fucoxanthin here) in diatoms and the xanthophyll pool size ((DD+DT)/Tchl *a*) were calculated to quantify photoprotective potentials of the diatom *F. cylindrus*.

Gene expression of photosynthesis-related genes; psbA *and* rbcL

To investigate the mechanisms of freezing and melting stress on cells of *F. cylindrus* in water and in the ice, gene expression of the photosynthesis-related genes, *psbA* and *rbcL*, was measured. To stabilize the RNA, 250 μL of RNAlater (Sigma) was immediately added to the ice core after collection. Triplicated melted ice and seawater samples were filtered onto two 25 mm, 2 μm polycarbonate Isopore membrane filters (Millipore) with gentle vacuum (<0.013MPa) passing through a 25 mm polypropylene in-line filter holder (Swinnex, Merck) for DNA and RNA samples. The DNA samples were placed in a cryotube and flash frozen in liquid nitrogen and stored at −80 ºC. RNA

262 samples were suspended in 600 μL RLT buffer (Qiagen) in a cryotube, to which 10 μL
263 of β-mercaptoethanol (Sigma-Aldrich) was added. The RNA sample then was flash
264 frozen in liquid nitrogen prior to storage at −80 ºC until further analysis. DNA and RNA
265 were extracted following Endo et al. (2013) and Endo et al. (2015), respectively. The
266 extracted RNA was reverse-transcribed to complementary DNA (cDNA) with the
267 PrimeScriptTM RT Master Mix (RR036, Takara) reagent according to the
268 manufacturer's specification. DNA and cDNA copy numbers of the *psbA* and *rbcL*
269 genes were quantified by quantitative PCR (qPCR). The primer sets and PCR conditions
270 used in this study are shown in Table S1 in the Supporting Information. Gene
271 expression was defined as the ratio of copy numbers of cDNA to DNA.

272

273 *Statistical analysis*

274 Statistical analyses were conducted using the SigmaPlot software program ver. 11.0
275 (SystStat Software, Inc.). One-way ANOVA with Tukey's test was performed on the
276 individual sampling days in each light treatment to identify variations in ChlF
277 parameters, pigment concentrations and gene expression. The differences in each
278 parameter were considered significant when $p<0.05$. Two-way ANOVA with Tukey's

tests was performed on the individual sampling days and light treatment to identify the differences in ChlF and pigment data to assess the effects of differences in photosynthetic ability and pigment composition between light treatments. Data was transformed for normalization prior to the ANOVA tests with the R statistical software using the R package MASS. Normality and variance of the data were checked by performing Shapiro-Wilk's test and Levene's test using the SigmaPlot software program after the data transformation, respectively.

RESULTS

Ice physics and ice algal growth

During the experiments, ice thickness remained stable at 5.5 cm with little basal melting or sublimation. Temperature within the ice increased with depth (ice surface: −22.5 ºC, ice-water interface: −2.2 ºC) (Fig. 2) and the underlying seawater was stable at ca. −1.8 ºC throughout the incubations. Brine salinity decreased with depth from 293.7 ± 0.0 to 39.4 ± 0.5, whereas the brine fraction increased (3.7 ± 0.0% to 34 ± 0%, assuming no gas bubbles were present). Nutrient concentrations did not significantly change during the incubations (data not shown). The ice formation concentrated *F. cylindrus* cells in the ice (HL: 8883 ± 224 cells mL^{-1}; LL; 8600 ± 573 cells mL^{-1})

297 compared with under-ice seawater (3551 ± 95 cells mL^{-1}; LL: 3639 ± 135 cells mL^{-1}).
298 Cells of *F. cylindrus* were thus successfully incorporated into the ice matrix where slow
299 but positive growth in both treatments was observed (0.020 ± 0.001 and 0.020 ± 0.002
300 under HL and LL, respectively); there was no significant difference in growth rate
301 between the treatments (t=0.00, p>0.05, Welch's t-test).
302
303 *Variable chl a fluorescence*
304 *Dark values (*F_v/F_m *and* σ_{PSII}*)*
305 At the beginning of the incubations (day −05), initial F_v/F_m of *F. cylindrus* showed
306 comparable and relatively high values in both high light (HL: 0.47 ± 0.05) and low light
307 (LL: 0.45 ± 0.06) conditions (t=0.63, p>0.05, Welch's t-test) (Fig. 3A, B). Upon
308 freezing in (day 00), F_v/F_m decreased significantly to 0.2−0.3 in both HL and LL (HL:
309 $F_{(7,40)}$=143.27, q=31.26, p<0.001; LL; $F_{(7,40)}$= 501.93 q=35.77, p<0.001, one-way
310 ANOVA, Tukey's test) (Fig. 3A, B), while σ_{PSII} did not change (HL: $F_{(7,40)}$=10.59,
311 q=2.01, p>0.05; LL: $F_{(7,40)}$= 12.97, q=3.65, p>0.05, one-way ANOVA) (Fig. 3C, D).
312 Once incorporated into the ice matrix the F_v/F_m of *F. cylindrus* cells remained stable
313 from day 00 to day 20 regardless of the light availability (HL: 0.20 ± 0.03; LL:0.16 ±
314 0.05, t=1.68, p>0.05, Welch's t-test), until initiation of the melt cycle. The lowest F_v/F_m

315 was observed on day 15 in both HL and LL (HL: 0.18 ± 0.02; LL: 0.09 ± 0.02) (Fig.
316 3A, B). The σ_{PSII} values at both light levels were relatively stable throughout the
317 incubations with slight decreases with time (Fig. 3C, D). Once the ice had melted,
318 F_v/F_m values seemed to be recovered from the values at day 20 in LL (HL: 0.18 ± 0.02;
319 LL: 0.19 ± 0.02) to almost equivalent levels on day −05 (see above), respectively. There
320 were, however, significant differences in F_v/F_m values between day −05 and melt
321 samples both in HL and LL (HL: q=35.57, p<0.001; q=35.77, p<0.001, one-way
322 ANOVA, Tukey's test) (39 ± 1% lower and 15 ± 1% higher, respectively) (Fig. 3A, B).
323 The responses of σ_{PSII} differed to F_v/F_m, which was significantly different on day −05,
324 but similar to σ_{PSII} values on day 20 regardless of the light availability (HL: q=1.07,
325 p>0.05; LL: q=1.08, p>0.05, one-way ANOVA) (Fig. 3C, D). Immediately after light
326 exposing the melted samples at 800 μmol photons m^{-2} s^{-1}, F_v/F_m significantly
327 decreased in both light treatments (HL: q=13.09, p<0.001; LL: q=53.11, p<0.001, one-
328 way ANOVA, Tukey's test) (39 ± 3% and 29 ± 2% for HL and LL treatments,
329 respectively). Despite the large decreases in F_v/F_m, σ_{PSII} showed minimal variation.
330
331 *Non-photochemical quenching (NPQ_{NSV}')*
332 Values of non-photochemical quenching (NPQ_{NSV}') in both light treatments were low

on day −05 prior to freezing in. Upon ice incorporation, NPQ_{NSV}' significantly increased in both HL and LL treatments (HL: $F_{(7,40)}$= 52.55, q=44.54, p<0.001; LL: $F_{(7,40)}$= 149.81, q=10.10, p<0.001, one-way ANOVA, Tukey's test) (Fig. 4). NPQ_{NSV}' in HL plateaued immediately after ice formation (i.e., day 00), while NPQ_{NSV}' in LL was observed to gradually increase and reach a maximum on day 05 (Fig. 4). Post ice melt, NPQ_{NSV}' in the LL tank decreased to the identical levels as those observed on day −05 (q=1.36, p>0.05, one-way ANOVA), whereas the HL treatment showed a significant increase in NPQ_{NSV}' (q=6.676, p<0.001, one-way ANOVA, Tukey's test). The light exposure after the melting event significantly enhanced NPQ_{NSV}' (almost doubled) in both HL and LL treatments (HL: q=11.39, p<0.001; LL: q=5.22, p=0.012, one-way ANOVA) (Fig. 4).

Photosynthesis-irradiance (ETR_{RCII}-E) curve

The initial slopes of ETR_{RCII}-E curve, α (regarded as light utilization efficiency), dropped significantly when the algae were frozen into the ice (HL: $F_{(7,40)}$= 8.20, q=6.84, p<0.001; LL: $F_{(7,40)}$= 11.27, q=8.24, p<0.001, one-way ANOVA, Tukey's test) but did not significantly differ thereafter in HL treatment (p>0.05, one-way ANOVA) (Fig. 5A, B). A similar trend was observed in the LL treatment, although α increased significantly

351 towards the end of the incubation (p<0.05, one-way ANOVA, Tukey's test). After the
352 ice melted, α values did not significantly change in HL (q=0.97, p>0.05, one-way
353 ANOVA), while, in LL, α returned the initial levels at day −05. Maximum electron
354 transport rates (ETR_{max}) in both treatments significantly decreased upon freezing in
355 (HL: $F_{(7,40)}$= 7.63, q=11.85, p<0.001; LL: $F_{(7,40)}$= 86.30, q=18.52, p<0.001, one-way
356 ANOVA, Tukey's test) (from 281 ± 37 and 300 ± 67 to 111 ± 53 and 38 ± 13 in HL and
357 LL treatments, respectively). Upon melting out, ETR_{max} in LL returned (335 ± 57) to
358 the initial levels at day −05 (q=2.40, p>0.05, one-way ANOVA), whereas, in HL,
359 ETR_{max} was not recovered (q=14.41, p<0.001, one-way ANOVA, Tukey's test) (Fig.
360 5C, D). Light saturation indices, E_k, gradually decreased during the course of the
361 incubation experiments without any conspicuous variation during the frozen period
362 (Fig. 5E, F). The HL treatment showed higher E_k throughout the frozen period from day
363 05 to day 20 compared to LL ($F_{(1,40)}$=9.02, p=0.004, two-way ANOVA, Tukey's test).
364 When melted out, E_k values returned to the initial levels on day −05 in both treatments
365 although the LL treatment show a higher E_k (HL: $F_{(7,40)}$= 7.63, q=0.63, p>0.05; LL:
366 $F_{(7,40)}$= 66.51, q=12.30, p<0.001, one-way ANOVA, Tukey's test). On the other hand,
367 E_k significantly decreased in both HL and LL treatments (HL: q=8.20, p<0.001; LL:
368 q=7.91, p<0.001, one-way ANOVA, Tukey's test) (Fig. 5E, F).

369

370 *Validation experiments for sample melt procedures*

371 Variable ChlF parameters varied among the different melt procedures of the ice sample
372 (Fig. S2 in the Supporting Information). When the intact crushed ice sample was packed
373 directly into a quartz tube, F_v/F_m values were lower, followed by that of the slushy
374 melted sample. These values were almost the same and did not show any significant
375 difference ($F_{(2,7)}$=10.76, q=2.90, p>0.05, one-way ANOVA), whereas the slow-direct
376 melt sample showed a significantly higher value compared to its counterparts (q=4.94,
377 p=0.024, one-way ANOVA, Tukey's test). Unlike F_v/F_m values, σ_{PSII} varied little
378 regardless of the melt methods and did not show any significant enhancement or
379 suppression ($F_{(2,7)}$=3.31, p>0.05, one-way ANOVA) (Fig. S2 in the Supporting
380 Information).

381

382 *Pigment composition*

383 Initial Tchl *a* concentrations were at the same level between incubations at day−05
384 ($F_{(1,31)}$=2.29, p>0.05, Two-way ANOVA) (Fig. 6A, B). After *F. cylindrus* cells were
385 incorporated into the ice, ice algae in the HL treatment maintained stable Tchl *a*
386 biomass ($F_{(7,16)}$=2.27, q=0.09, p>0.05, one-way ANOVA) although Tchl *a* in the LL

387 treatment showed a gradual increase. Contributions of chllide *a* to Tchl *a* were slightly
388 higher in the HL treatment than in the LL tank throughout the incubations
389 ($F_{(1,31)}$=59.96, p<0.001, Two-way ANOVA, Tukey's test; 48.0−62.5% and 34.0−54.9%,
390 respectively) (Fig. 6A, B). The cellular DD-DT pool size in both the HL and LL
391 treatments at the beginning of the incubations was comparable ($F_{(1,31)}$=73.27, q=1.76,
392 p>0.05, Two-way ANOVA), whereas, during the frozen period the cellular pool size
393 grew faster was significantly larger in the HL ice tank than those in the LL tank
394 ($F_{(7,16)}$=73.27, q=12.11, p<0.001, Two-way ANOVA, Tukey's test; 17.3−25.3% and
395 8.8−14.7% to Tchl *a*, respectively) (Fig. 6C, D). During the frozen period from day 05
396 to day 20, DES was also higher in the HL ice tank (0.293−0.552) compared to the LL
397 ice tank (0.107−0.231) ($F_{(1,31)}$=35.65, p=0.027, Two-way ANOVA, Tukey's test) (Fig.
398 6C, D). Light exposure enhanced the DES in both treatments (4.20- and 5.23-fold
399 increases in the HL and LL ice tank, respectively). Higher PPC/PSC ratios were present
400 in the HL ice tank than in LL throughout the incubations ($F_{(1,31)}$=69.22, p<0.001, Two-
401 way ANOVA, Tukey's test) (Fig. 6E, F). The PPC/PSC ratios showed little variation
402 before and after light exposure in both light treatments (HL: $F_{(7,16)}$=3.82, q=0.04,
403 p>0.05; LL: $F_{(7,16)}$=3.86, q=0.85, p>0.05, one-way ANOVA).
404

405 *Gene expression of photosynthesis-related genes*

406 rbcL

407 Prior to incubation there was no significant difference in the expression of the *rbcL*

408 gene in either HL or LL (t=0.33, p>0.05, Welch's t-test) (Fig. 7A, B). Transcriptional

409 activity of the *rbcL* gene was highly upregulated when algae were frozen into the ice

410 (HL: $F_{(7,16)}$=7.65, q=5.30, p=0.03; LL: $F_{(7,16)}$=7.39, q=6.89, p=0.003, one-way

411 ANOVA, Tukey's test) in both HL and LL (Fig. 7A, B). After day 05 and day 10, gene

412 expression of the *rbcL* gene decreased significantly in the HL and LL treatments (HL:

413 q=7.290, p=0.002; LL: q=6.79 p=0.040, one-way ANOVA, Tukey's test), respectively

414 (Fig. 7A, B). This low level was sustained during the frozen period. Comparing the

415 results of melt and light periods, the *rbcL* gene expression did not show any

416 conspicuous change in either treatment (HL: q=2.173, p>0.05; LL: q=0.31, p>0.05,

417 one-way ANOVA).

418

419 psbA

420 Unlike the *rbcL* gene, the expression of the *psbA* gene differed with respect to light

421 availability (Fig. 7C, D). The initial value in the HL treatment was ~2-fold higher than

422 that in LL. In HL, transcription of the *psbA* gene was highly upregulated during freezing

($F_{(7,16)}$=19.90, q=10.28, p<0.001, one-way ANOVA, Tukey's test); which was similar to the *rbcL* gene. In LL, upregulation of *psbA* was not evident ($F_{(7,16)}$=2.02, p>0.05, one-way ANOVA). Expression of *psbA* in HL showed a similar trend to the *rbcL* gene, whereas, under LL, the *psbA* gene expression did not change throughout the experiment (Fig. 7C, D).

DISCUSSION

Ice tank incubation

We here, for the first time, successfully demonstrated the use of a purpose designed ice tank to study the photophysiology of ice algae for a prolonged period (i.e. 20 days) under environmentally relevant conditions. When ice formed, cooling of the water body from the surface in the ice tank, in a −20 ºC freezer, realistically mimicked *in situ* ice formation events in wintertime, with cells of *F. cylindrus* successfully incorporated into the ice. The temperature profile of the artificial sea ice indicates that the ice properties simulated winter ice environments, inferred from a sharp decrease in temperature with depth (Fig. 2) (Petrich and Eicken 2017). It suggests that the ice tank was well suited to investigate survival strategies of ice algae under chronic multiple stress in wintertime.

The identical initial F_v/F_m values at day−05 (Fig. 3A, B) indicate that the two discrete ice tank runs (i.e. HL and LL treatments) started with the similar physiological states of *F. cylindrus.*

Freezing event

The freezing events in the ice tank experiments reported herein suppressed photochemical quantum efficiency of PSII (Fig. 3A, B). Drawdowns in F_v/F_m were also observed in the previous ice tank study using *Phaeocystis antarctica* (Kennedy et al. 2012) during ice formation. This suppression has also been noted in natural sea ice, inferred from lower F_v/F_m of ice algae than pelagic phytoplankton (McMinn et al. 2008, Yamamoto et al. 2014).

A significant amount of chl *a* breakdown products has been reported in ice algae of both landfast and pack-ice in the Arctic and Antarctic (e.g., Horner 1985, Alou-Font et al. 2013). One might interpret the here observed increase in chllide *a* (Fig. 6A, B) as inactivation of PSII reaction centres. Chllide *a*, lacking the phytol chain, has a less pigment aggregation potential (Fiedor et al. 2003), leading to reduced coupling of excitons (inefficient energy transfer between pigments) (Rosenbach-Belkin et al. 1991).

457 However, the stable amount of chllide *a* during the freezing events (Fig. 6A, B)
458 indicates that the existence of chllide *a* had only a minor effect on the drawdown in
459 F_v/F_m. Also, not all inactive pigments (i.e. chllide *a*) are involved in reaction centres
460 but function as photosynthetic antennae pigments.

461 Suggett et al. (2009) demonstrated that chronic stress decreases F_v/F_m with a
462 concomitant enhancement of σ_{PSII} because of the absorbed light energy being funnelled
463 to fewer active PSII reaction centres if some reaction centres are damaged/inactive
464 (Suggett et al. 2009, 2011, Falkowski and Raven 2013). In this study, however, little
465 variation in σ_{PSII} was observed during the freezing event (Fig. 3C, D). This suggests that
466 photoinactivation could not be significant during ice formation.

467 Freezing stress, combined with low temperatures and high brine salinity, could be the
468 main cause of the reduction in the photochemistry of PSII and the capacity of electron
469 sinks (Ensminger et al. 2006, Ralph et al. 2007). Low temperatures reduce membrane
470 viscosity, which suppresses photochemical reactions in the electron transport chain
471 (Morgan-Kiss et al. 2006). At the same time, high brine salinity might suppress the
472 reduction capability of electron donors around PSII (i.e., the first quinone electron
473 acceptor Q_A and the plastoquinone (PQ) pool; Ralph et al. 2007). Indeed, the here

474 observed high brine salinity (55 on average and >80 in the upper part of the ice) was
475 high enough to suppress the downstream components of PSII (Ralph et al. 2007). This
476 combined stress of low temperature and high salinity has been found to stagnate
477 electron transfer along the electron transport chain down to ferredoxin and cause
478 “electron clogging” (e.g., Lazár et al. 2005). This “electron clogging” consequently can
479 lead to (a) the low F_v/F_m (e.g., Schwarz et al. 2017), (b) overreduction of the PQ pool
480 (Maxwell et al. 1995, Allen and Nilson 1997, Pfannschmidt 2003) and (c) an imbalance
481 of the photochemical electron transport (“photostasis”; Öquist and Huner 2003). We
482 interpret the here observed increase in NPQ_{NSV}’ (Fig. 4) as a measure for alleviating
483 excess excitation pressure (e.g. Caron et al. 1987, Ting and Owens 1993, Goss and
484 Jakob 2010).

485 The HL treatment exhibited a larger DD-DT pool and higher PPC/PSC compared with
486 the LL counterpart (Fig. 6C, D, E, F). These light responses can be interpreted as up-
487 front protection to mitigate the production of reactive oxygen species under high light
488 (McMinn and Hegseth 2004, Katayama and Taguchi 2013, Kuczynska et al. 2015). This
489 notion is supported by field observations of (Kropuenske et al. 2009, Schuback et al.
490 2016, Galindo et al. 2017) and emphasized light-photoprotection relationships (Lavaud
491 et al. 2002, Domingues et al. 2012).

492 Photosynthetic performance, assessed with ETR_{RCII}–E curves, showed rapid
493 photoacclimation responses during freezing; a greater decrease in ETR_{max} than that of α
494 led to apparent dark acclimation with a lower E_k after freezing (Fig. 5). However, the
495 concomitant decrease of α does not coincide with classical shade photoacclimatisation
496 strategies (i.e. higher α and lower E_k under lower light would be expected; MacIntyre et
497 al. 2002). The decrease in α suggests a decrease in n_{PSII}, the active fraction of PSII to
498 chl *a* (e.g. Falkowski and Raven 2013). Values of α are represented as a product of σ_{PSII}
499 and n_{PSII}; however, σ_{PSII} values were stable during the freezing event, suggesting
500 photoinactivation of RCII (i.e., lowered n_{PSII}). It is thus suggested that both the
501 combination of direct photoinactivation and electron clogging deactivated the
502 photochemical reactions at RCII. The freezing event, therefore, suppressed RCII
503 photochemistry both directly and indirectly.

504 The strong upregulation of expression of the *psbA* gene in the HL treatment
505 accelerated repair of damaged PSII under overexcitation pressure in the HL condition
506 and vice versa (Fig. 7C, D) (Petrou et al. 2010, Galindo et al. 2017). The significant
507 upregulation of the *rbcL* gene (Fig. 7A, B) could be evidence of a cold acclimation
508 strategy to increase cellular RuBisCO and compensate for the low catalytic activity of
509 the temperature-sensitive enzyme by increasing its concentration (Devos et al. 1998,

510 Lyon and Mock 2014, Young et al. 2015). However, Mock and Hoch (2005) reported
511 downregulation of *rbcL* transcription in *F. cylindrus* under low temperatures.
512
513 *During the frozen period*
514 ChlF parameters remained relatively stable during the frozen period and little variation
515 in F_v/F_m, σ_{PSII}, and NPQ_{NSV}' values were observed within the ice (Figs. 3 and 4),
516 indicating that sea ice is a stable platform for algal photosynthesis (Maccario et al.
517 2015, Arrigo 2017). The comparable levels of F_v/F_m and σ_{PSII} in both ice tanks suggest
518 that the *F. cylindrus* cells sustained or optimized their photosynthesis within the ice
519 regardless of light availability (Fig. 3). This physiological acclimation to ice
520 environments may play a key role in the successful survival and proliferation of ice
521 algae within and at the bottom of sea ice (Kropuenske et al. 2010, Lacour et al. 2018).
522 McMinn and Hegseth (2004) suggested that ice diatoms can grow out in sea ice if
523 F_v/F_m is >0.125. In this study, the observed F_v/F_m values during the frozen period were
524 high enough to actively grow in the ice. Indeed, this study, for the first time, reported
525 positive growth in the ice tank. Although the growth rates in both treatments (~0.02 d^{-1})
526 were lower than other reported values from culture experiments (e.g., Mock and Hoch
527 2008, Kropuenske et al. 2010). Interestingly, the growth rates in both the HL and LL

528 treatments were comparable regardless of the under-ice light intensities. This indicates
529 that light availability was not a limiting factor for algal growth in the artificial sea ice.
530 Evidently, photosynthetic and photoprotective abilities between the HL and LL ice
531 tanks were comparable during the frozen periods, as inferred from the identical levels of
532 F_v/F_m and NPQ_{NSV} (Fig. 3A, B; Fig. 4). As discussed above, the electron clogging
533 caused by multiple co-stressors could be the major determinant of algal growth. The
534 low temperature lowered the enzymatic activity of RuBisCO (Young et al. 2015), which
535 could be another possible cause of the identical growth rates observed in both HL and
536 LL treatments. The gradual increase in light utilization index α in the LL ice tank (Fig.
537 5A, B) was consistent with the classical concept of photoacclimation (MacIntyre et al.
538 2002). As noted above, σ_{PSII} was stable in the LL treatment (Fig. 3C, D). Cells under
539 LL may increase the proportion of active RCII relative to pigmentation (chl *a*) in order
540 to enhance light capture during freezing in (Fig. 5E, F). Only algae in the LL ice tank
541 showed a gradual increase in Tchl *a* concentration, in spite of the same algal growth rate
542 in both HL and LL treatments; this could be further evidence of dark acclimation (e.g.,
543 Falkowski and Owens 1980). The smaller PPC/PSC in the LL ice tank also supports this
544 suggestion by indicating preferential synthesis of photosynthetic chlorophylls and
545 carotenoids rather than PPC (Fig. 6E, F). Cells in the HL treatment strengthened their

546 photoprotective capability, reflected in higher values of PPC/PSC and DES compared to
547 cells in LL (Fig. 6C, D, E, F) (Kropuenske et al. 2009, 2010). However, both HL and
548 LL treatments showed increased their DD-DT pool and DES, suggesting again a highly
549 reduced PQ pool due to the electron clogging (i.e., the development of ΔpH) (Leptit et
550 al. 2010, 2013). Interestingly, NPQ_{NSV}' levels were comparable between the light
551 treatments regardless of the difference in DES (Figs. 4 and 5C, D). Nymark et al. (2013)
552 suggested that low-light acclimated cells have more *LHCX* proteins, which enlarges
553 their NPQ capacity (Bailleul et al. 2010). Interestingly, the transcriptional activity of the
554 *rbcL* gene greatly decreased after the sharp increase on day 00 (onset of freezing),
555 suggesting that cells had used the high abundance of RuBisCO (or its mRNA) that was
556 synthesized during the freezing event (Fig. 7A, B). The constant but lower expression of
557 *psbA* could keep the repair of PSII to a 'business as usual' rate (Fig. 7C, D), whereas the
558 downregulation of the gene after initial significant upregulation in the HL treatment
559 might be caused to alleviate the photochemical imbalance in the electron transport chain
560 due to the upregulation when shocked by freezing stress (e.g. changes in PSII to PSI
561 ratio).
562
563 *Light exposure after ice melt*

564 Light exposure to released algal cells in spring time suppressed photosynthetic
565 activities. Exposure to irradiance following melting decreased F_v/F_m due to the high
566 excitation pressure (Fig. 3A, B), despite ice melt before light exposure increasing
567 photochemical activity. The increase in F_v/F_m after ice melt was consistent with results
568 of the validation experiments (Fig. S2 in the Supporting Information). The degree of
569 drawdown in F_v/F_m was comparable regardless of the incubation light intensity after
570 light exposure (Fig. 3A, B). NPQ_{NSV}' was downregulated when the ice melted but was
571 enhanced after light exposure (Fig. 4). This reverse response to light exposure between
572 F_v/F_m and NPQ_{NSV}' might be evidence for fast activation of photoprotection (Figs. 3A,
573 B and 4) especially in the HL treatment with the higher DES (Fig. 6C, D). Damage of
574 PSII, however, was likely because of the considerable increase in the ratio of chllide *a*
575 to Tchl *a* after light exposure in both ice tanks (Fig. 6A, B). One might suspect that the
576 melting process prior to sample filtration caused the significant breakdown of chl *a* with
577 activation of chlorophyllase (Jeffery and Hallegraeff 1987); however, a stable level of
578 chllide *a* to Tchl *a* observed throughout the incubation period even from day −05 before
579 ice formation (Fig. 5A, B) suggests the melting process (osmotic stress) could not be
580 responsible for the high concentrations of chllide *a* and other degraded pigments.
581 However, the significant difference in chllide *a* contributions between HL and LL

582 throughout the incubations might relate to the breakdown of chl *a* because high light to
583 *F. cylindrus* leads a high qI (non-photochemical quenching by photoinactivation)
584 (Kropuenske et al. 2009). After ice melt, PPC/PSC decreased while F_v/F_m increased
585 (Figs. 3A, B and 6E, F). This suggests a re-optimization of photosynthetic performance
586 to a water environment after release from freezing stress. Light exposure, however,
587 activated the DD-DT xanthophyll cycle (i.e., de-epoxidation) (Fig. 6C, D) (Petrou et al.
588 2010), although the size of the DD-DT pool was little changed (Fig. 6C, D). Activation
589 of the DD-DT cycle is the fastest photoprotective strategy and occurs at a scale of
590 seconds to minutes (Goss and Jakob 2010), whereas the size of the pool actually
591 changes much slower. Interestingly, NPQ_{NSV}' in the HL treatment was much higher
592 than that of the LL treatment with differences in their DD-DT pool size (Figs. 4 and 6C,
593 D), suggesting that pigmentation and light history were largely responsible for their
594 potential photoprotective ability (Szechyńska- Hebda et al. 2010, Galindo et al. 2017). It
595 seems to be unusual that E_k values significantly decreased (i.e. dark acclimation) upon
596 light exposure, contrary to the expected response from high-light acclimation (Fig. 5E,
597 F). Transcriptional activity of the *psbA* gene was stable in both treatments after light
598 exposure (Fig. 7C, D). It is suggested that algal cells did not immediately start to
599 synthesize mRNA for repair of the damaged RCII, possibly because post-transcriptional

600 regulation is significant for D1 protein synthesis (Malnoë et al. 1988, Danon and
601 Mayfield 1991) with a large mRNA pool (Mulo et al. 2012). The dark reaction might
602 also be independent or to have been slow to respond to the light exposure event because
603 the gene expression of the *rbcL* gene was invariant before and after light exposure (Fig.
604 6A, B).
605
606 CONCLUSIONS
607 This study demonstrates how the combined effects of ice formation and melting
608 together with light exposure affect the photophysiology of *F. cylindrus*. This study is
609 the first attempt to conduct the *ex situ* incubation of the ice algal *F. cylindrus* in
610 artificial sea ice to quantify the effects of multiple co-stressors in sea ice and to provide
611 new insights into the fate of ice algae. The ice tank successfully mimicked ice formation
612 events as experienced in the field (Figs. 1 and 2). This enabled investigation into the
613 effects of multiple co-stressors on the capacity of *F. cylindrus* to survive this ephemeral
614 habitat. We successfully observed, for the first time, positive growth of ice algal cells in
615 artificial sea ice. Ice formation events suppressed the photochemical process of RCII,
616 despite optimal irradiance prior to freezing in (Fig. 3A, B). This was likely a result of
617 high brine salinity and low temperature, which led to electron clogging in the electron

618 transport chain. Ice algal cells rapidly optimized their photophysiology to the dark
619 environment within the ice, inferred from decreases in E_k (Fig. 3 and 5E, F). Light
620 availability could affect the photoacclimative ability of ice algae, possibly by increasing
621 active RCII. Further investigation on the connectivity of photosynthetic antennae to
622 RCII would be needed to address the efficiency of energy transfer to RCII (e.g.,
623 D'Haene et al. 2015, Yoshida et al. 2018). Although ice melt stimulated algal
624 photosynthesis, the subsequent light exposure resulted in photoinhibition (Fig. 3A, B)
625 with significant chllide *a* accumulation and NPQ_{NSV}' enhancement. However,
626 photoprotection was rapidly activated. We conclude that the diatom *F. cylindrus*
627 possesses a high photosynthetic plasticity that contributes to the ecological success of
628 this species in both pelagic and sea-ice habitats.

629

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635 Scientific Research (JP18H03352, JP17H00775).

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LIST OF FIGURES

651 Left (A and C) and right (B and D) panels indicate data from the HL and LL treatment,
652 respectively. Open, closed and shaded bars indicate values of seawater samples before
653 freezing, ice and seawater samples after melting, respectively. Different letters above
654 bars in a panel indicate significant differences in values between sampling days with
655 one-way ANOVA with Tukey's test; there is no significant difference between values if
656 a given letter is shown in the combination of letters of the counterparts. The D stands
657 for 'day', while Melt and Light indicate values after melting and light exposure
658 experiments, respectively. Error bars show 1 standard deviation ($n \geq 5$).

659

660 Figure 4. Non-photochemical quenching based on the normalized Stern-Volmer
661 quenching coefficient under the actinic light (150 and 30 μmol photons m^{-2} s^{-1} for HL
662 and LL, respectively) (NPQ_{NSV}') during the ice tank incubation experiments. Panels A
663 and B indicate data from the HL and LL treatments, respectively. Different letters above
664 bars in a panel indicate significant differences in values between sampling days with
665 one-way ANOVA with Tukey's test; there is no significant difference between values if
666 a given letter is shown in the combination of letters of the counterparts. The D stands
667 for 'day', while Melt and Light indicate values after melting and light exposure
668 experiments, respectively. Error bars show 1 standard deviation ($n \geq 5$).

669

670 Figure 5. Photosynthesis−irradiance (ETR_{RCII}−E) parameters during the ice tank
671 incubation experiments. Top panels (A and B) show light utilization efficiency under
672 dim light (α) as initial slopes of the ETR_{RCII}-E curves, middle panels (C and D) indicate
673 maximum electron transport rate (ETR_{max}), and bottom panels (E and F) show light
674 saturation index (E_k). Left (A, C and E) and right (B, D, and F) panels show data from
675 the HL and LL treatments. Different letters above bars in a panel indicate significant
676 differences in values between sampling days with one-way ANOVA with Tukey's test;
677 there is no significant difference between values if a given letter is shown in the
678 combination of letters of the counterparts. The D stands for 'day', while Melt and Light
679 indicate values after melting and light exposure experiments, respectively. Error bars
680 show 1 standard deviation ($n \geq 5$).

681

682 Figure 6. Variations in pigment concentrations during the ice tank incubation
683 experiments. Top panels (A and B) show total chl *a* concentration (Tchl *a*) [mg m^{-3}]
684 and contribution of chlorophyllide *a* to Tchl *a*, middle panels (C and D) indicate the size
685 of DD-DT pool ((DD+DT)/Tchl *a*) and de-epoxidation state of xanthophyll pigments

686 (DES; DT/(DD+DT)), and bottom panels (E and F) show ratio of photoprotective
687 carotenoids (PPC) to the photosynthetic carotenoid (PSC) quantified with UHPLC. Left
688 (A, C, and E) and right (B, D, and F) show data from the HL and LL treatments,
689 respectively. The D stands for 'day', while Melt and Light indicate values after melting
690 and light exposure experiments, respectively. Results of one-way and two-way ANOVA
691 tests performed on the pigment data were described in the text. Error bars show 1
692 standard deviation (*n*=3).

693

694 Figure 7. Gene expression [cDNA/DNA] of photosynthesis-related genes. Top (A and
695 B) and bottom (C and D) panels show gene expression of the *rbcL* and *psbA* genes
696 during the ice tank incubation experiments. Left (A and C) and right (B and D) show
697 data from the HL and LL treatments, respectively. Different letters above bars in a panel
698 indicate significant differences in values between sampling days with one-way ANOVA
699 with Tukey's test; there is no significant difference between values if a given letter is
700 shown in the combination of letters of the counterparts. The D stands for 'day', while
701 Melt and Light indicate values after melting and light exposure experiments,
702 respectively. Error bars show 1 standard deviation (*n*=3).

LIST OF SUPPLEMENTAL MATERIALS

Table S1. Primer sets and PCR conditions of qPCR and qRT-PCR for the *psbA* and *rbcL* genes

Figure S1. A photo of the ice tank with a light panel.

The light panel has a white LED with light diffuser to homogenize the light field over the ice.

Figure S2. Chlorophyll *a* fluorescence parameters with different melting procedures.

(a): Maximum quantum yield of PSII photochemistry (F_v/F_m); (b): functional absorption cross-section (σ_{PSII}), determined with FRR fluorometry.

Intact ice: intact ice placed into the cuvette for ChlF measurement; slush; ice sample partially melted in filtered seawater; Direct slow: melted ice sample overnight at 2.5 ºC.

Different letters indicate significant differences between the given values. Error bars indicate 1 standard deviation. *n*=6.

Table 1.

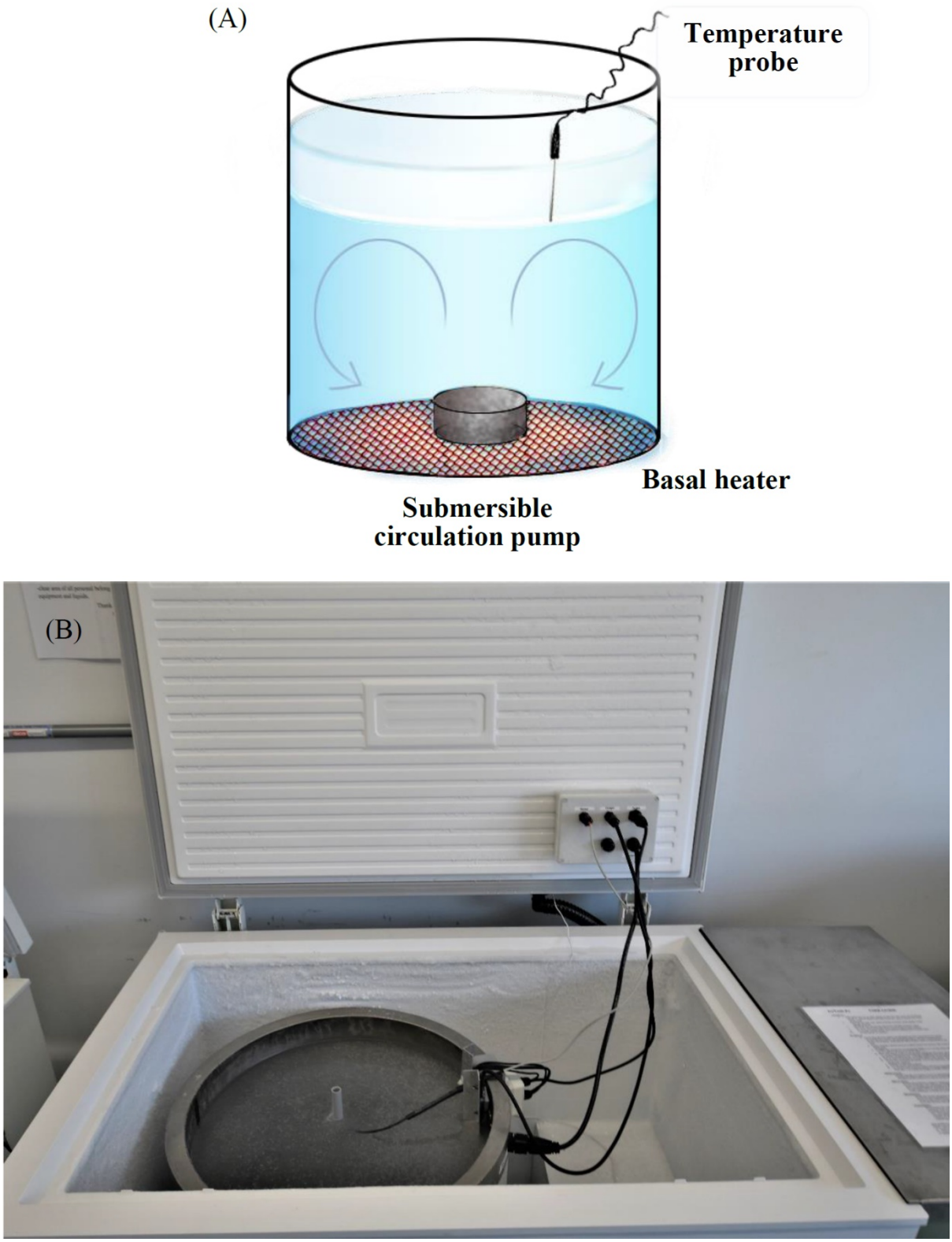
Yield and parameter		Unit	Derivation or reference
Fluorescence yield			
F	Fluorescence yield	Unitless	
F_o	Minimum fluorescence yield of dark regulated cells	Unitless	
F_m	Maximum fluorescence yield of dark regulated cells	Unitless	
F_v	Maximum variable fluorescence yield	Unitless	$F_m - F_o$
F'	Fluorescence yield under actinic light	Unitless	
F_o'	Minimum fluorescence yield of actinic light-acclimated cells	Unitless	Oxborough and Baker (1997)
F_m'	Maximum fluorescence yield of actinic light-acclimated cells	Unitless	
F_v'	Variable fluorescence yield under actinic light	Unitless	$F_m' - F_o'$
F_q'	Difference in fluorescence yields between F' and F_m'	Unitless	$F_m' - F'$
Fluorescence parameters in the dark			
σ_{PSII}	Functional absorption cross-section of PSII in the dark	nm^2 $RCII^{-1}$	Kolber et al. (1998)
F_v/F_m	Maximum quantum yield of PSII photochemistry	Unitless	$(F_m - F_o)/F_m$
τ	Time constant for Q_A reoxidation	ms	Kolber et al. (1998)
Fluorescence parameters under actinic light			
$F_{q'}/F_{m'}$	Effective photochemical efficiency of PSII under actinic light	Unitless	$(F_m' - F')/F_m'$
NPQ_{NSV}’	Non-photochemical quenching based on the S-V approach	Unitless	McKew et al. (2013)
ETR_{RCII}-*E* curve parameters			
ETR_{RCII}	Absolute electron transport rate through RCII	mol e^{-1} s^{-1}	Schuback et al. (2016)
E	Actinic light irradiance	mol photons m^{-2} s^{-1}	
α	Light utilization index under dim light	(mol e^{-1} s^{-1}) (μmol photons m^{-2} s^{-1})$^{-1}$	Platt et al. (1980)
β	Light inhibition index under high light caused PSII photoinhibition	(mol e^{-1} s^{-1}) (μmol photons m^{-2} s^{-1})$^{-1}$	Platt et al. (1980)
ETR_{max}	Maximum electron transport rate through RCII	mol e^{-1} s^{-1}	Platt et al. (1980)
E_k	Light saturation index	μmol photons m^{-2} s^{-1}	Platt et al. (1980)

719

Table S1

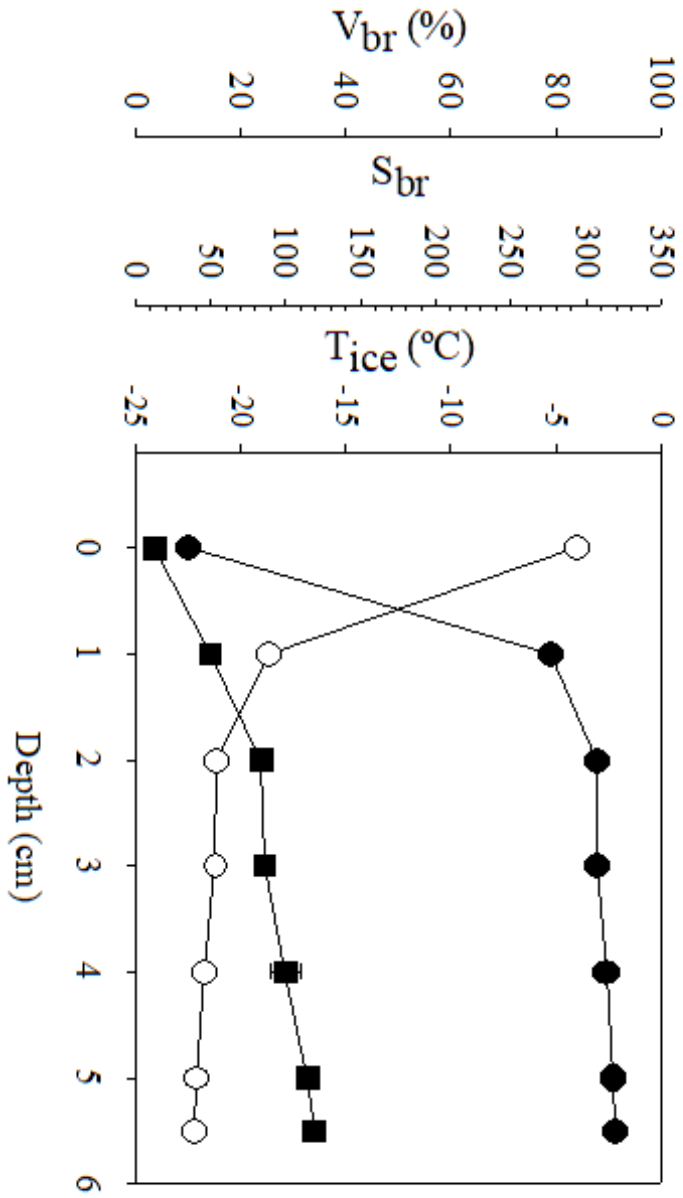
psbA	
Forward primer	5'-AGAACCACCAAATACACCAGCAA-3'
Reverse primer	5'-TCCAAGCTGAGCACAACATCTT-3'
Amplicon size	71
PCR condition	94 °C for 60 s; 40 cycles under 98 °C for 10 s, 62 °C for 60 s, and 72 °C for 60s
Baseline gene	Gene fragment of *Fragilariopsis cylindrus* from Antarctic sea ice[*1]
Reference	Krell et al. (2007)
rbcL	
Forward primer	5′-GATGATGARAAYATTAACTCW-3′
Reverse primer	5′-TAWGAACCTTTWACTTCWCC-3′
Amplicon size	113
PCR condition	94 °C for 60 s; 40 cycles under 98 °C for 10 s, 55 °C for 60 s, and 72 °C for 60s
Baseline gene	Gene fragment of *Thalassiosira weissflogii* (CCMP1336)
Reference	John et al. (2007); Endo et al. (2015)

*1: The isolate of *F.cylindrus* was obtained from Antarctic sea ice in the Weddel Sea in 1991 (Krell et al., 2007).


(A)
Temperature probe
Basal heater
Submersible circulation pump
(B)

722

723 Fig. 1


V_{br} (%)
0
20
40
60
80
100
S_{br}
0
50
100
150
200
250
300
350
T_{ice} (°C)
-25
-20
-15
-10
-5
0
Depth (cm)
0
1
2
3
4
5
6

724

725 Fig.2

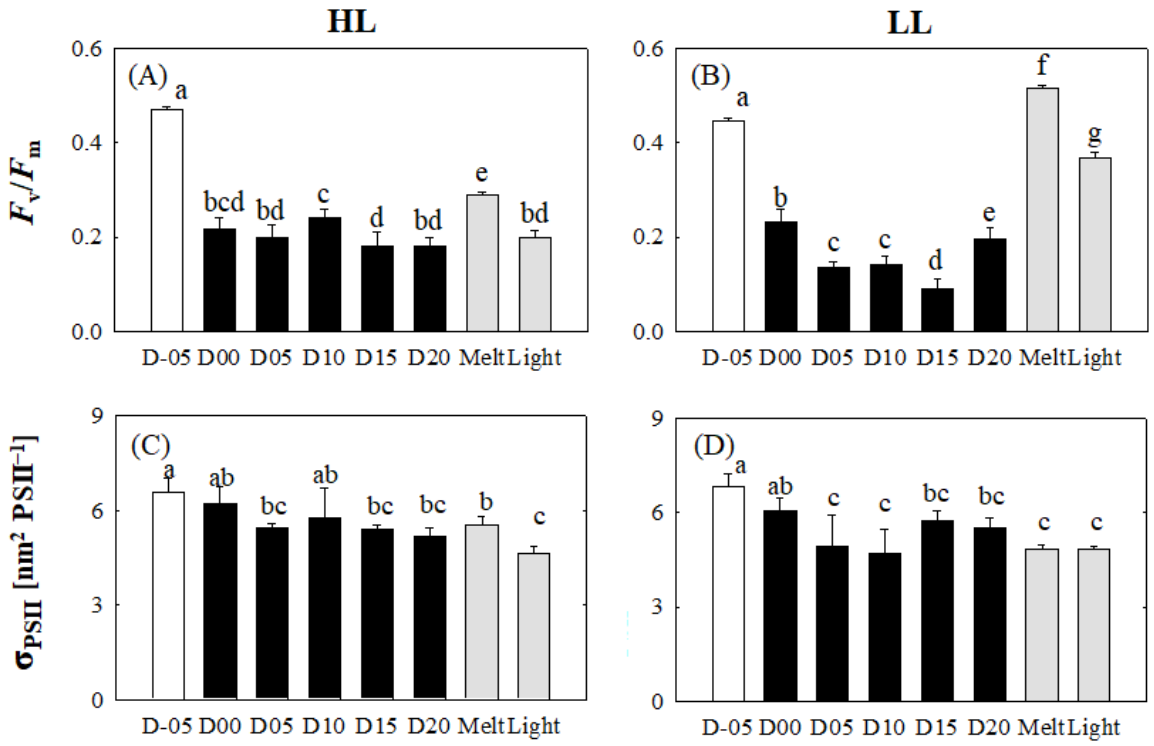
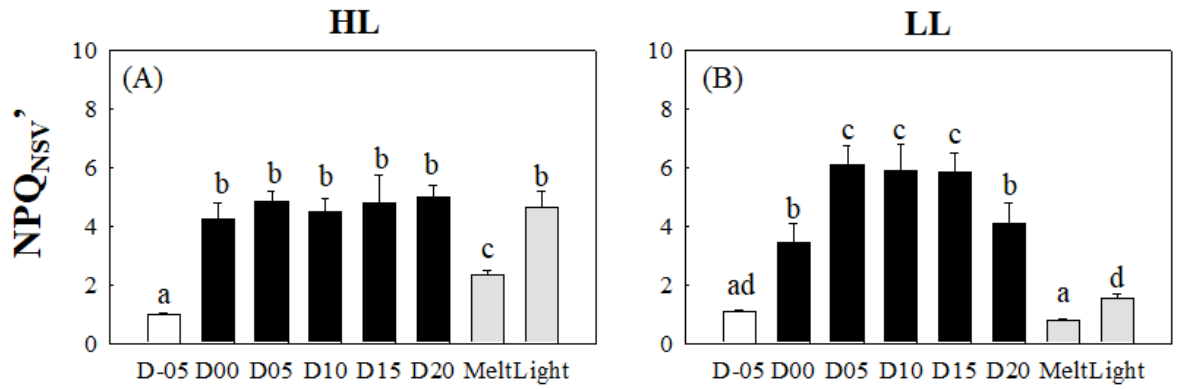

HL
LL
(A)
(B)
(C)
(D)
F_v/F_m
σ_{PSII} [nm² PSII⁻¹]
D-05 D00 D05 D10 D15 D20 Melt Light

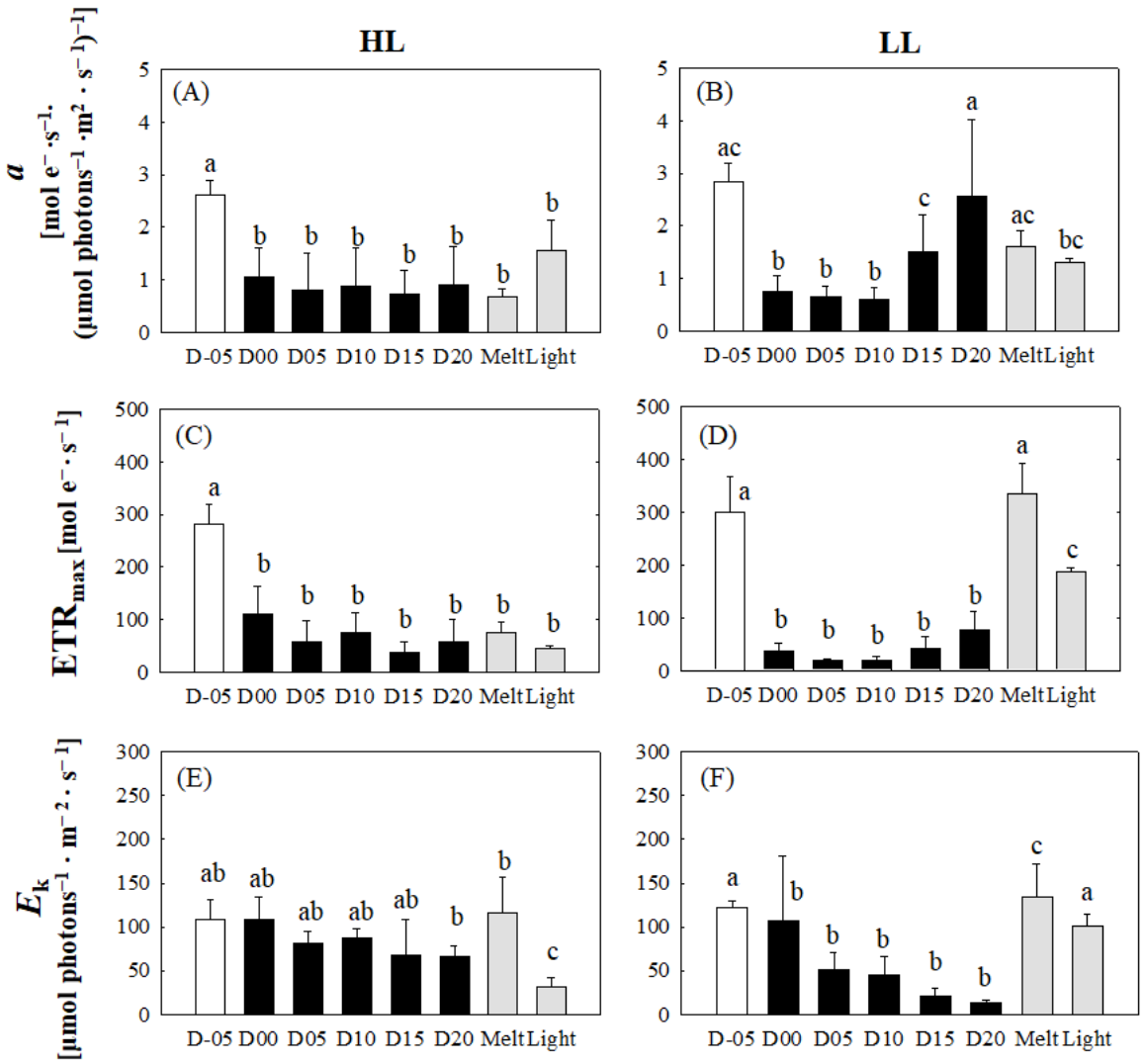
Fig. 3


HL
LL
(A)
(B)
NPQNSV'
10
8
6
4
2
0
a
b
b
b
b
b
c
b
ad
b
c
c
c
b
a
d
D-05 D00 D05 D10 D15 D20 MeltLight
D-05 D00 D05 D10 D15 D20 MeltLight

728

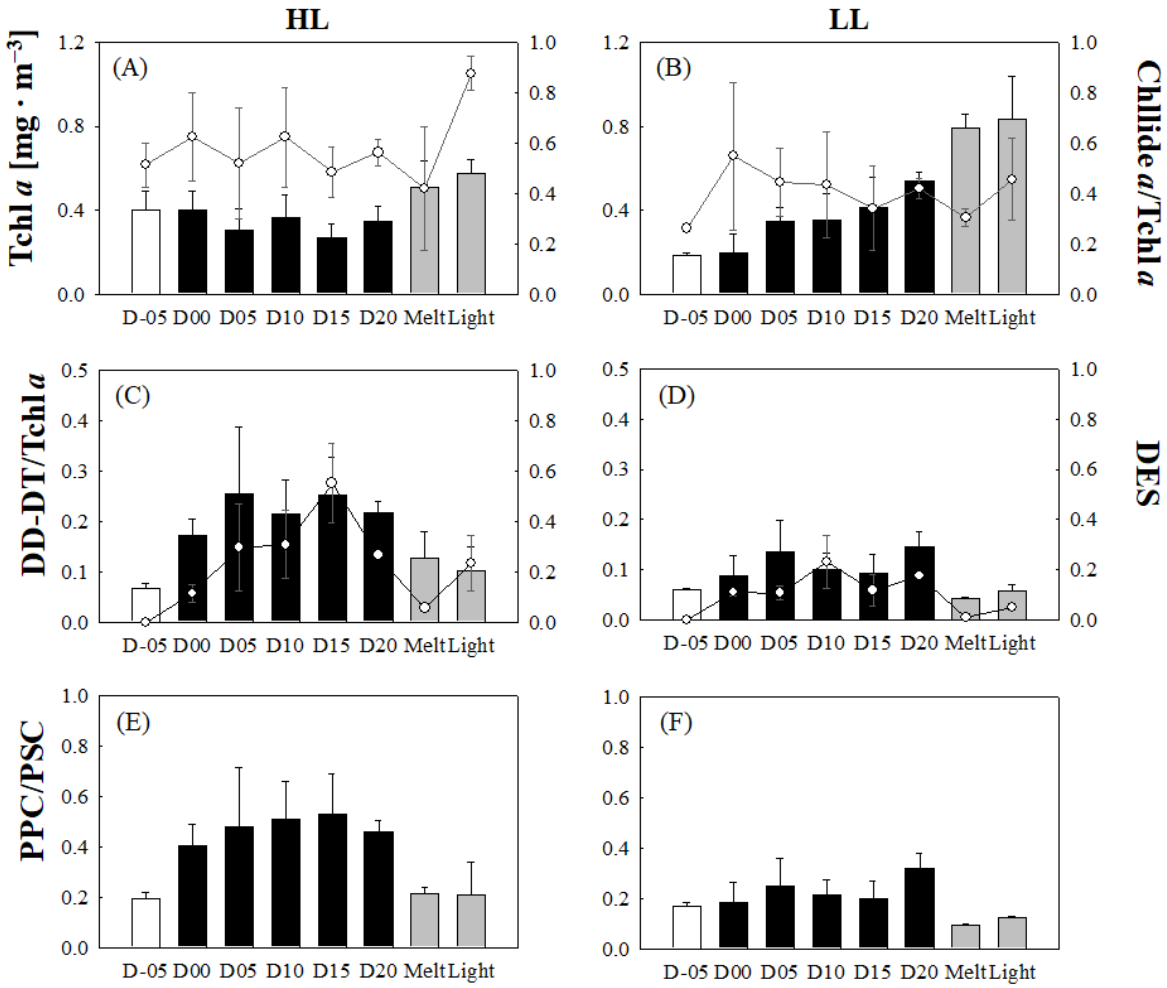
729 Fig. 4

730



731

732 Fig. 5


HL
LL
(A)
(B)
(C)
(D)
(E)
(F)
Tchl a [mg · m⁻³]
Chllide a/Tchl a
DD-DT/Tchl a
DES
PPC/PSC
D-05 D00 D05 D10 D15 D20 Melt Light

733

734 Fig. 6

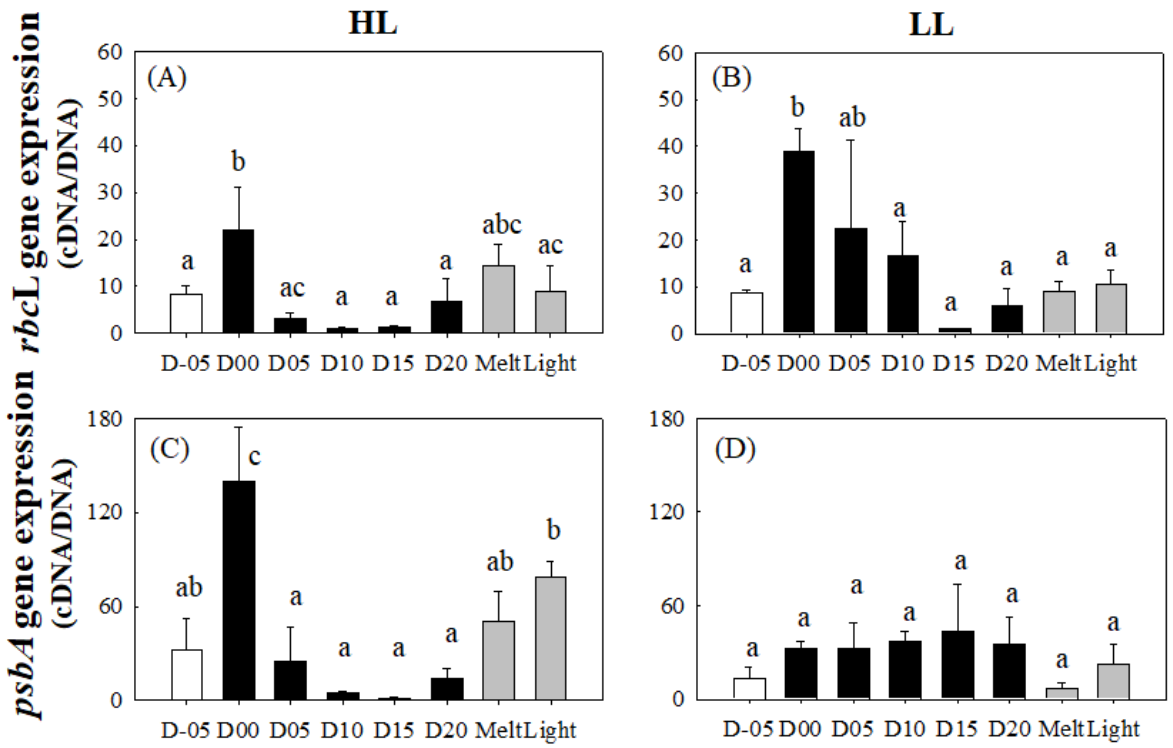

HL
LL
rbcL gene expression (cDNA/DNA)
psbA gene expression (cDNA/DNA)
(A)
(B)
(C)
(D)
D-05 D00 D05 D10 D15 D20 Melt Light

Fig. 7

738 Figure S2

739

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