

HOKKAIDO UNIVERSITY

Title	Unexpected low genetic differentiation between Japan and Bering Sea populations of a deep-sea benthic crustacean lacking a planktonic larval stage (Peracarida: Tanaidacea)			
Author(s)	Kakui, Keiichi; Nomaki, Hidetaka; Komatsu, Hironori; Fujiwara, Yoshihiro			
Citation Biological Journal of the Linnean Society, 131(3), 566-574 https://doi.org/10.1093/biolinnean/blaa106				
Issue Date	2020-11			
Doc URL	http://hdl.handle.net/2115/83124			
Rights	This is a pre-copyedited, author-produced version of an article accepted for publication in Biological Journal of the Linnean Society following peer review. The version of record Volume 131, Issue 3, 2020, Pages 566-574 is available online at: https://doi.org/10.1093/biolinnean/blaa106.			
Туре	article (author version)			
Additional Information	There are other files related to this item in HUSCAP. Check the above URL.			
File Information	Biological Journal of the Linnean Society_2020.pdf			



This is a pre-copyedited, author-produced version of an article accepted for publication in "Biological Journal of the Linnean Society" following peer review. The version of record of this article is available online at: https://doi.org/10.1093/biolinnean/blaa106.

- 1 Unexpected low genetic differentiation between Japan and Bering Sea
- 2 populations of a deep-sea benthic crustacean lacking a planktonic larval
 3 stage (Peracarida: Tanaidacea)
- KEIICHI KAKUI^{1*}, HIDETAKA NOMAKI², HIRONORI KOMATSU³, and YOSHIHIRO
 FUJIWARA⁴
- 8 ¹ Faculty of Science, Hokkaido University, Sapporo 060-0810, Japan
- 9 ² SUGAR, X-star, Japan Agency for Marine-Earth Science and Technology (JAMSTEC),
- 10 Yokosuka 237-0061, Japan
- ³ Department of Zoology, National Museum of Nature and Science, Tsukuba 305-0005, Japan
- ⁴*Research Institute for Global Change (RIGC), Japan Agency for Marine-Earth Science and*
- 13 Technology (JAMSTEC), Yokosuka 237-0061, Japan
- 14

 $\overline{7}$

- 15 *Corresponding author: Keiichi Kakui; kakui@eis.hokudai.ac.jp; Faculty of Science,
- 16 Hokkaido University, N10 W8 Kita-ku, Sapporo 060-0810, Japan
- 17
- 18 Short running title: low genetic differentiation of a deep-sea tanaidacean
- 19

20 Information on the extent, diversity, and connectivity of populations is lacking for most

- 21 deep-sea invertebrates. Species in the order Tanaidacea, one of the most diverse and abundant
- 22 macrofaunal groups in the deep sea, are benthic, lack a planktonic larval stage, and thus
- 23 would be expected to have narrow distributional ranges. But here we show that, with
- 24 molecular evidence from the COI gene, the deep-sea tanaidacean *Carpoapseudes spinigena*
- has a distributional range spanning at least 3700 km, from off northern Japan to the
- southeastern Bering Sea. Living individuals found in a sediment core indicated that the
- 27 species is a sedentary burrower. COI analyses revealed a low level of genetic diversity overall
- for *C. spinigena*, and low differentiation (*p*-distance, 0.2–0.8%) between the Japan and
- 29 Bering Sea populations. One hypothesis to explain the low genetic diversity over a broad
- 30 region is the Japan population was founded by individuals transported by ocean currents from
- 31 the Bering Sea; however, due limited data, other explanations cannot be ruled out. Our results
- 32 underscore that continued sampling is of fundamental importance to understanding how
- 33 genetic and taxonomic diversity originate and are maintained in the deep sea.
- 34

35 ADDITIONAL KEYWORDS: COX1 - Crustacea - direct development - Malacostraca -

- 36 Pacific phylogeography population genetics.
- 37

38	INTRODUCTION
39	
40	Sea bottoms deeper than 200 m occupy about 70% of the Earth's surface, yet the population
41	genetic data necessary to understand the extent, diversity, and connectivity of deep-sea
42	invertebrate populations have been very limited (Taylor & Roterman, 2017). Population
43	genetic studies are now appearing for various groups (e.g. Boavida et al., 2019; Janssen et al.,
44	2019; Cheng et al., 2020; Guggolz et al., in press). As with shallow-water species, dispersal
45	of planktonic larvae by ocean currents is probably the main factor maintaining connectivity
46	between populations of most deep-sea invertebrates (e.g. Yahagi et al., 2017; Kobayashi et al.,
47	2018). However, the migration of adults in species with high swimming capability (e.g.
48	Winkelmann et al., 2013) and the transport of epibenthic adults by bottom currents (e.g.
49	Braby et al., 2009; Hamel et al., 2019) could also enable gene flow between populations.
50	Dispersal barriers proposed for deep-sea animals include strong currents crossing between
51	populations, low-oxygen zones, topographical factors (e.g. shallow straits, mid-ocean ridges,
52	depth differences), and long distances between patches of suitable habitat (McClain & Hardy,
53	2010).
54	The superorder Peracarida is the most speciose crustacean group with ca. 17000
55	described species (Appeltans et al., 2012) and is well represented in the deep sea (e.g. Grassle
56	& Maciolek, 1992). Although peracarids show diverse modes of living, such as benthic,
57	planktonic (including symbionts on gelatinous plankton), and parasitic (Ohtsuka et al., 2009;
58	Kakui, 2016; Castellani et al., 2017; Smit et al., 2019), peracarids generally lack a primary
59	planktonic larval stage but instead undergo direct development, although parasitic species
60	tend to have highly mobile stages (Smit et al., 2019). The innate dispersal capability of
61	peracarids thus depends largely on their mode of living, and sedentary, benthic species are
62	expected to have low dispersal capability.
63	Some deep-sea benthic peracarids show restricted distributions (Riehl & De Smet,

Some deep-sea benthic peracards show restricted distributions (Riehl & De Smet,
2020), but others have much broader distributions (Brandt *et al.*, 2012) than expected from
their mode of living; in some cases, conspecificity has been confirmed by molecular
phylogeographic studies based on mitochondrial gene markers, i.e., cytochrome *c* oxidase
subunit I (COI), 12S rRNA, and/or 16S rRNA genes (e.g. Brix *et al.*, 2011; Riehl & Kaiser,
2012; Riehl *et al.*, 2018). Riehl & Kaiser (2012) discussed the importance of bottom currents
and other erosion-deposition events on continental shelves as factors allowing deep-sea
infaunal peracarids to achieve broad distributions.

The peracarid order Tanaidacea contains about 1500 described species worldwide (Anderson, 2020), most of which are small (up to a few millimeters long) and are free living in benthic habitats (Kakui, 2016). Tanaidaceans comprise one of the most diverse and abundant macrofaunal groups in the deep sea (Larsen *et al.*, 2015), but little is known of their biology there, and no previous study has examined their population structure in that

76	environment.

92

Carpoapseudes spinigena Bamber, 2007, a member of the relatively rare, deep-sea 77tanaidacean genus Carpoapseudes Lang, 1968 (Apseudoidea: Apseudidae), has not been 78reported since its original description which was based on fixed specimens collected from a 79single site in the Pacific Ocean off northern Japan (Fig. 1; 974–965 m depth; Bamber, 2007). 80 During research cruises in 2007 and 2017, we collected C. spingena from two localities 81 82 several thousand kilometers apart, one being the type locality around northern Japan (1028– 83 1075 m depth) and the other located in the southeastern Bering Sea (1536–1569 m depth). In 84 addition, we observed the behavior of living C. spinigena trapped in a core sampler, and 85 examined the morphology of juvenile individuals at stage of release from the maternal brood pouch. Here we show the genetic relationship between these two *C. spinigena* populations 86 based on partial COI gene sequences, describe the mode of living of this species, and discuss 87 the wide distribution of *C. spinigena*. 88 89 90

MATERIAL AND METHODS

93Tanaidaceans were collected with a beam trawl having a 3 m opening, a dredge installed on a 94Deep Tow Camera System, and a multiple corer during research cruises of the research vessel (R/V) Tansei-maru and R/V Mirai (Japan Agency for Marine-Earth Science and Technology, 95 96 JAMSTEC) (Table 1), near the type locality in northern Japan and in the eastern Aleutian 97Islands, USA, respectively (Fig. 1). Specimens from the two Bering Sea sites were regarded as samples from a single population, because the two sites were only ca. 1.2 km apart, shorter 98 than the distance between net-in and net-out (ca. 2.7 km) of the beam trawl at station 99 100 KT-07-29-K1 in northern Japan. Living individuals collected with the multiple corer, and some fresh specimens collected during the R/V Mirai cruise, were photographed before 101 fixation. Whole bodies or dissected parts of specimens were fixed in 80% ethanol, and later 102 transferred to 99% ethanol for preservation. See Supporting Information S1 for additional 103 104 details.

105Morphological terminology follows Larsen (2003). The identification of specimens was based on Bamber (2007) and Hansknecht & Santos (2008). Female tanaidaceans have a 106 marsupium (brood pouch) for brooding eggs (cf. Kakui et al., 2017) and release juveniles at 107 the "manca II" stage directly from the marsupium (Larsen, 2003); manca II individuals 108 109 resemble adults, but are unisex and lack percopod 6 (6th walking leg) and pleopods 110 (abdominal legs used for ventilating the burrow or for swimming). Male apseudoid tanaidaceans have a genital cone whereas females lack it. Specimens were identified to stage 111 (manca II, female, or male) under a Nikon SMZ1500 stereomicroscope, and deposited with 112JAMSTEC (JAMSTEC nos. 1170055975, 1170055978, 1170055979, 1170055985, 113

114 1170055986) and in the Invertebrate Collection of the Hokkaido University Museum
115 (ICHUM), Sapporo (ICHUM-6092–6112).

Total DNA was extracted from the cheliped or whole body of adults, and from an 116 embryo collected from a marsupium, by using a NucleoSpin Tissue XS Kit (TaKaRa Bio, 117118 Japan); after extraction, each exoskeleton was recovered and preserved in 80% ethanol. Part of the COI gene was amplified by PCR using primers LCO-1490 and HCO-2198 (Folmer et 119 al., 1994). PCR amplification conditions with TaKaRa Ex Taq DNA polymerase (TaKaRa 120Bio) were 94°C for 1 min; 35 cycles of 98°C for 10 sec, 42 or 50°C for 30 sec, and 72°C for 12112250 sec; and 72°C for 2 min. Nucleotide sequences were determined by direct sequencing with 123a BigDye Terminator Kit ver. 3.1 and a 3730 Genetic Analyzer (Life Technologies, USA). Sequences (658 nt, encoding 219 amino acids) were aligned by using MEGA7 (Kumar et al., 1242016) (the dataset contained no indels), and *p*-distances among sequences were calculated 125with MEGA7. All sequences we determined were deposited in the International Nucleotide 126127Sequence Database (INSD) through the DNA Data Bank of Japan (DDBJ), under accession numbers LC545527-545558. 128

129 An integer neighbor-joining (IntNJ) network (Leigh & Bryant, 2015) for the COI 130 sequences was constructed with PopART v. 1.7 (Leigh & Bryant, 2015) at 0.50 reticulation 131 tolerance. Haplotype diversity (*h*) and nucleotide diversity (π) were calculated with DnaSP v. 132 6.12.03 (Rozas *et al.*, 2017). Tajima's *D* (Tajima, 1989), Fu's *F*_S (Fu, 1997), and Φ_{ST} were 133 calculated with Arlequin v. 3.5.2.2 (Excoffier & Lischer, 2010).

134

135

136

137

RESULTS

138Male and female *C. spinigena* were similar in general body shape and had narrow, rod-shaped pleopods; manca II specimens were generally similar to males and females but lacked 139pleopods (Fig. 2A–F, f1). In one core from the multiple corer, we found three living 140 individuals on the sediment surface, each of which was positioned at the entrance to a burrow, 141 142with the posterior part of the body in the burrow (Fig. 2G; also see Kakui, 2020a). 143We identified 10 COI haplotypes (H1-10) among 32 C. spinigena individuals (21 from Japan and 11 from the Bering Sea). The aligned sequences (658 nt) contained 10 144polymorphic sites (Table 2) involving only synonymous substitutions. In the IntNJ network 145(Fig. 3), haplotypes H1–4 from the Japan population and H5–10 from the Bering Sea 146population formed separate groups; the two populations shared no haplotypes. Eighteen of 147148the 21 sequences from Japan were H4. The most common haplotypes in the Bering Sea population were H5 (three individuals) and H7 (four individuals). The most nucleotide 149substitutions observed in pairwise comparisons among haplotypes was five (H1, H2, H3 vs. 150H10), corresponding to a *p*-distance of 0.8%. The fewest nucleotide substitutions between 151

152 haplotypes from the two populations was one (H4 vs. H5).

153	Overall <i>h</i> for <i>C</i> . <i>spinigena</i> was moderately high (0.673) but π was low (0.0020) (Table
154	2). In the Japan population, both <i>h</i> and π values were low (0.271 and 0.0004, respectively); in
155	the Bering Sea population, h was high (0.836) but π was low (0.0022). Tajima's D value for
156	the Japan population was a significant, negative value (-1.727 ; $p = 0.024$), whereas other
157	values were not significant. Fu's $F_{\rm S}$ values for C. spinigena overall and for both populations
158	were significant ($p < 0.02$; Holsinger, 2017), negative values (Table 2). Φ_{ST} between Japan
159	and Bering Sea populations was significant and high (0.6797; $p < 0.001$).
160	
161	
162	DISCUSSION
163	
164	Low dispersal capability of Carpoapseudes spinigena
165	The life cycle, morphology, and mode of living of Carpoapseudes spinigena indicate that its
166	native dispersal capability is quite low. As a tanaidacean (cf. Kakui et al., 2017), the species
167	lacks planktonic larval stages, and females directly release manca II individuals from their
168	marsupium. In both males and females, as described by Bamber (2007), the pleopods are
169	narrow and rod-shaped, rather than wide and leaf-shaped (cf. Shiino, 1966: fig. 7G),
170	indicating that both sexes have poor swimming ability. Manca II individuals are similar in
171	body shape to males and females but lack pleopods. None of the developmental stages we
172	observed show apparent swimming adaptations such as the enormously long or
173	paddle-shaped pereopods known in deep-sea swimming isopods (Marshall & Diebel, 1995).
174	Our multiple corer sample showed that C. spinigena lives as a burrower (Fig. 2G). All of
175	these observations indicate that C. spinigena individuals are likely sedentary throughout their
176	life.
177	
178	Genetic diversity in C. spinigena overall and in two populations
179	Our phylogeographic analysis of the COI sequences revealed low genetic diversity in C.
180	spinigena. The maximum p-distance of 0.8% that we detected among 32 C. spinigena
181	sequences from two populations ca. 3700 km apart is much lower than that observed within a
182	single population in a shallow-water apseudoid species, Mesokalliapseudes macsweenyi
183	(Drumm, 2003) (ca. 3% <i>p</i> -distance; Drumm & Kreiser, 2012). The moderately high <i>h</i> (0.673)
184	and low π (0.0020) and significantly negative Fu's F_s values suggest that the overall
185	population we sampled underwent a population reduction followed by rapid population
186	growth and accumulation of mutations (Grant & Bowen, 1998). The Japan and Bering Sea
187	populations shared no haplotypes, and the Φ_{ST} value between the two populations was
188	significant and high, indicating low connectivity between the two populations.
189	The Japan and Bering Sea populations appear to have different histories. The two

190 clusters in our haplotype network are separated by one nucleotide substitution. The Japan 191cluster shows a star-like pattern, with haplotype H4 comprising 18 of 21 sequences (86%) 192and three haplotypes each connected to H4 by one mutational step (Fig. 3). Both the h and π values were low (0.271 and 0.0004, respectively), and Tajima's D and Fu's F_S values were

193significant and negative, indicating that the Japan population underwent a recent population 194

- bottleneck or was established by a founder event (Grant & Bowen, 1998). The Bering Sea 195
- cluster was more branched and more diverse, with high h (0.836), low π (0.0022), and a 196 significant, negative Fu's $F_{\rm S}$ value. These indices indicate that the Bering Sea population 197 underwent a past reduction in size followed by expansion, but may have been affected by
- 199 200

198

How did C. spinigena become widely distributed? 201

small sample size (n = 11).

The low genetic distance between the two widely separate populations we examined is 202203difficult to reconcile with the low innate dispersal capability expected for a small, deep-sea fossorial crustacean lacking planktonic larval stages. One possible explanation is that the 204205Japan population originated through a founder from the Bering Sea population via transport by deep ocean currents, a mechanism that Riehl & Kaiser (2012) suggested for dispersal by 206 207deep-sea macrostylid isopods. In our study area, bottom currents at the depths where we 208found C. spinigena (1000–1500 m) are complex and have been minimally investigated, but 209 do occur along the Aleutian Ridge (flowing eastward) and Bering Sea Slope (flowing northwestward) (Kinder et al., 1975; Roden, 1995); additionally, a southward flow dominated 210by the East Kamchatka Current occurs at depths of 1500 m or shallower on the western side 211of the Kamchatka Strait (Stabeno et al., 1999; the maximum depth of the strait is 4420 m), 212and a southwestward flow of the Oyashio Current has been observed at the depths of 1000 m 213214or shallower off the eastern coast of northern Japan (Uehara & Miyake, 1999) (Fig. 1).

A pattern similar to our COI haplotype network, where the Bering Sea cluster is more 215branched and more diverse than the Japan cluster, was observed in the crangonid shrimp 216Argis lar (Owen, 1839) (Fujita et al., 2017). In that species, however, more nucleotide 217218substitutions (six or 14 substitutions in 571 nt) were detected between a Bering cluster 219(Lineage C) and Japan clusters (Lineages A and B), indicating older divergences in A. lar than in *C. spinigena* and that the two species have different evolutionary histories. *Argis lar* 220differs from *C. spinigena* in having a relatively short (15–20 days at 5°C; Nakano, 1993) 221222pelagic larval phase in its life cycle and in inhabiting shallower depths (10–350 m; Komai & 223Komatsu, 2009), both of which would increase the feasibility of dispersal by surface currents. 224Other hypotheses than a single long-distance dispersal event are possible to account

for the low genetic diversity of C. spinigena between the southeastern Bering Sea and 225226northern Japan. The haplotype pattern and diversity values apparently indicating a founder event for the population in northern Japan could also have resulted from a local bottleneck. 227

228Furthermore, it is unknown whether C. spinigena is continuously distributed in the relevant depth interval between northern Japan and the southeastern Bering Sea. A continuous 229distribution along the Kurile Islands, eastern Kamchatka, Aleutian Basin, and Aleutian 230 archipelago could have been achieved by relatively rapid but stepwise dispersal mediated by 231deep-sea currents, or by slower, stepwise range expansion. Even poorly dispersing bottom 232dwellers can expand their range gradually by local currents (e.g. Martel & Chia, 1991; 233Norkko et al., 2001). Finally, even if a founder population of C. spinigena was established in 234northern Japan by long-distance dispersal, the source population could have been closer than 235236the Bering Sea, i.e., the Kuriles or Kamchatka. Data from across the putatively broad range of 237C. spinigena are necessary to determine the most likely among these various possibilities.

It is unknown whether other deep-sea tanaidaceans show similarly wide distributions. Species in *Protanais*, one of the main indicators of successional stages in deep-sea wood-fall communities (McClain & Barry, 2014), may show narrower distributions restricted to areas where sunken wood is abundant. Species in some groups producing males adapted for swimming (natatory males; Błażewicz-Paszkowycz *et al.*, 2014) possibly show wider distributions and more connectivity among populations.

The main results of our study are 1) that the deep-sea tanaidacean species *Carpoapseudes spinigena* is a burrower, which together with the lack of swimming appendages and a larval phase, indicate poor dispersal capability, and 2) that two populations thousands of kilometers apart show remarkably low genetic divergence. Explaining these incongruent results is of fundamental importance to understanding how genetic and taxonomic diversity originate and are maintained in the deep sea, and our results underscore the need for continued sampling in this environment.

- 251
- 252
- 253
- 254

ACKNOWLEDGMENTS

255We thank the captains and crews of R/V Tansei-maru and R/V Mirai, technicians from Marine Work Japan and Nippon Marine Enterprises, and researchers on board for support 256257during cruises KT-07-29 and MR17-04 Leg2; Yuki Oya for help with molecular work; Takumi Onishi and Aoi Tsuyuki for assistance with literature; Matthew H. Dick for reviewing 258the manuscript and editing our English; and the editor and three anonymous reviewers for 259260critical comments on the manuscript. This study is an outcome of the following project 261directed by YF: "Aleutian Magic" observations in the south-eastern Bering Sea. It was 262funded in part by KAKENHI grant JP16K18597 to KK from the Japan Society for the Promotion of Science (JSPS). 263

- 264
- 265

266	REFERENCES
267	
268	Amante C, Eakins BW. 2009. ETOPO1 1 arc-minute global relief model: procedures, data
269	sources and analysis. NOAA Technical Memorandum NESDIS NGDC-24. National
270	Geophysical Data Center, NOAA. Available at: http://dx.doi.org/10.7289/V5C8276M.
271	Anderson G. 2020. Tanaidacea—Forty Years of Scholarship, Version 3.0. Available at:
272	https://aquila.usm.edu/tanaids30/5/
273	Appeltans W, Ahyong ST, Anderson G, Angel MV, Artois T, Bailly N, Bamber R, Barber
274	A, Bartsch I, Berta A, Błażewicz-Paszkowycz M, Bock P, Boxshall G, Boyko CB,
275	Branão SN, Bray RA, Bruce NL, Cairns SD, Chan T-Y, Cheng L, Collins AG,
276	Cribb T, Curini-Galletti M, Dahdouh-Guebas F, Davie PJF, Dawson MN, de Clerck
277	O, Decock W, de Grave S, de Voogd NJ, Domning DP, Emig CC, Erséus C,
278	Eschmeyer W, Fauchald K, Fautin DG, Feist SW, Fransen CHJM, Furuya H,
279	Garcia-Alvarez O, Gerken S, Gibson D, Gittenberger A, Gofas S, Gómez-Daglio L,
280	Gordon DP, Guiry MD, Hernandez F, Hoeksema BW, Hopcroft RR, Jaume D, Kirk
281	P, Koedam N, Koenemann S, Kolb JB, Kristensen RM, Kroh A, Lambert G,
282	Lazarus DB, Lemaitre R, Longshaw M, Lowry J, Macpherson E, Madin LP, Mah
283	C, Mapstone G, McLaughlin PA, Mees J, Meland K, Messing CG, Mills CE,
284	Molodtsova TN, Mooi R, Neuhaus B, Ng PKL, Nielsen C, Norenburg J, Opresko
285	DM, Osawa M, Paulay G, Perrin W, Pilger JF, Poore GCB, Pugh P, Read GB,
286	Reimer JD, Rius M, Rocha RM, Saiz-Salinas JI, Scarabino V, Schierwater B,
287	Schmidt-Rhaesa A, Schnabel KE, Schotte M, Schuchert P, Schwabe E, Segers H,
288	Self-Sullivan C, Shenker N, Siegel V, Sterrer W, Stöhr S, Swalla B, Tasker ML,
289	Thuesen EV, Timm T, Todaro MA, Turon X, Tyler S, Uetz P, van der Land J,
290	Vanhoorne B, van Ofwegen LP, van Soest RWM, Vanaverbeke J, Walker-Smith G,
291	Walter TC, Warren A, Williams GC, Wilson SP, Costello MJ. 2012. The magnitude
292	of global marine species diversity. <i>Current Biology</i> 22: 2189–2202.
293	Bamber RN. 2007. Suborders Apseudomorpha Sieg, 1980 and Neotanaidomorpha Sieg,
294	1980. In: Larsen K, Shimomura M, eds. Tanaidacea (Crustacea: Peracarida) from
295	Japan III. The deep trenches; the Kurile-Kamchatka Trench and Japan Trench. Zootaxa
296	1599. Auckland: Magnolia Press, 13–40.
297	Błażewicz-Paszkowycz M, Jennings RM, Jeskulke K, Brix S. 2014. Discovery of
298	swimming males of Paratanaoidea (Tanaidacea). Polish Polar Research 35: 415–453.
299	Boavida J, Becheler R, Choquet M, Frank N, Taviani M, Bourillet J-F, Meistertzheim
300	A-L, Grehan A, Savini A, Arnaud-Haond S. 2019. Out of the Mediterranean?
301	Post-glacial colonization pathways varied among cold-water coral species. <i>Journal of</i>
302	Biogeography 46: 915–931.
303	Braby CE, Pearse VB, Bain BA, Vrijenhoek RC. 2009. Pycnogonid-cnidarian trophic

304 interactions in the deep Monterey Submarine Canyon. Invertebrate Biology 128: 359-363. 305Brandt A, Błażewicz-Paszkowycz M, Bamber RN, Mühlenhardt-Siegel U, Malyutina 306 MV, Kaiser S, de Broyer C, Havermans C. 2012. Are there widespread peracarid 307308 species in the deep sea (Crustacea: Malacostraca)? Polish Polar Research 33: 139–162. Brix S, Riehl T, Leese F. 2011. First genetic data for species of the genus *Haploniscus* 309 310Richardson, 1908 (Isopoda: Asellota: Haploniscidae) from neighbouring deep-sea basins in the South Atlantic. Zootaxa 2838: 79-84. 311312Castellani C, Lehtiniemi M, Meland K. 2017. Crustacea: Lophogastrida and Mysida. In: 313 Castellani C, Edwards M, eds. Marine plankton: a practical guide to ecology, methodology, and taxonomy. Oxford: Oxford University Press, 471–489. 314Cheng J, Hui M, Li Y, Sha Z. 2020. Genomic evidence of population genetic differentiation 315in deep-sea squat lobster Shinkaia crosnieri (Crustacea: Decapoda: Anomura) from 316 317Northwestern Pacific hydrothermal vent and cold seep. Deep-Sea Research Part I 156: 318 103188. Drumm DT, Kreiser B. 2012. Population genetic structure and phylogeography of 319 Mesokalliapseudes macsweenvi (Crustacea: Tanaidacea) in the northwestern Atlantic 320 321and Gulf of Mexico. Journal of Experimental Marine Biology and Ecology 412: 58–65. 322Excoffier L, Lischer HEL. 2010. Arlequin suite ver 3.5: a new series of programs to perform 323 population genetics analyses under Linux and Windows. *Molecular Ecology Resources* **10:** 564–567. 324Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R. 1994. DNA primers for amplification 325of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. 326 Molecular Marine Biology and Biotechnology 3: 294–299. 327 Fu Y-X. 1997. Statistical tests of neutrality of mutations against population growth, 328 hitchhiking and background selection. Genetics 147: 915–925. 329Fujita J, Drumm DT, Iguchi A, Ueda Y, Yamashita Y, Ito M, Tominaga O, Kai Y, Ueno 330 331M, Yamashita Y. 2017. Deep-sea phylogeographic structure shaped by 332paleoenvironmental changes and ongoing ocean currents around the Sea of Japan in a 333 crangonid shrimp, Argis lar. Zoological Science 34: 406–413. Grant WS, Bowen BW. 1998. Shallow population histories in deep evolutionary lineages of 334marine fishes: insights from sardines and anchovies and lessons for conservation. 335336 Journal of Heredity 89: 415–426. 337 Grassle TF, Maciolek NJ. 1992. Deep-sea species richness: regional and local diversity 338estimates from quantitative bottom samples. American Naturalist 139: 313–341. Guggolz T, Meißner K, Schwentner M, Dahlgren TG, Wiklund H, Bonifácio P, Brandt A. 339 340 in press. High diversity and pan-oceanic distribution of deep-sea polychaetes: Prionospio and Aurospio (Annelida: Spionidae) in the Atlantic and Pacific Ocean. 341

342	Organisms Diversity & Evolution. doi: 10.1007/s13127-020-00430-7.
343	Hamel J-F, Sun J, Gianasi BL, Montgomery EM, Kenchington EL, Burel B, Rowe S,
344	Winger PD, Mercier A. 2019. Active buoyancy adjustment increases dispersal potential
345	in benthic marine animals. Journal of Animal Ecology 88: 820–832.
346	Hansknecht T, Santos KC dos. 2008. Carpoapseudes heardi n. sp. (Tanaidacea:
347	Apseudomorpha) from Caribbean waters near Tobago. Gulf and Caribbean Research
348	20: 67–74.
349	Holsinger K. 2017. Lecture Notes in Population Genetics. figshare. Available at:
350	https://doi.org/10.6084/m9.figshare.100687.v3
351	Janssen A, Stuckas H, Vink A, Martinez Arbiz P. 2019. Biogeography and population
352	structure of predominant macrofaunal taxa (Annelida and Isopoda) in abyssal
353	polymetallic nodule fields: implications for conservation and management. Marine
354	<i>Biodiversity</i> 49: 2641–2658.
355	Kakui K. 2016. Review of the taxonomy, diversity, ecology, and other biological aspects of
356	order Tanaidacea from Japan and surrounding waters. In: Motokawa M, Kajihara H. eds.
357	Species diversity of animals in Japan. Berlin: Springer, 603–627.
358	Kakui K. 2020a. Living Carpoapseudes spinigena individual in a surface core sampler.
359	figshare. Available at: https://doi.org/10.6084/m9.figshare.12038886 (to editor and
360	reviewers: not yet public; private link: https://figshare.com/s/881d35c56be3629c9916)
361	Kakui K, Hayakawa Y, Katakura H. 2017. Difference in size at maturity in annual and
362	overwintering generations in the tanaidacean Zeuxo sp. in Oshoro Bay, Hokkaido, Japan.
363	Zoological Science 34: 129–136.
364	Kinder TH, Coachman K, Galt JA. 1975. The Bering Slope Current system. Journal of
365	Physical Oceanography 5: 231–244.
366	Kobayashi G, Mukai R, Alalykina I, Miura T, Kojima S. 2018. Phylogeography of benthic
367	invertebrates in deep waters: A case study of Sternaspis cf. williamsae (Annelida:
368	Sternaspidae) from the northwestern Pacific Ocean. Deep-Sea Research Part II 154:
369	159–166.
370	Komai T, Komatsu H. 2009. Deep-sea shrimps and lobsters (Crustacea: Decapoda) from
371	northern Japan, collected during the project "Research on Deep-sea Fauna and
372	Pollutants off Pacific Coast of Northern Japan". In: Fujita T, ed. Deep-sea fauna and
373	pollutants off Pacific coast of northern Japan. National Museum of Nature and Science
374	Monographs 39. Tokyo: National Museum of Nature and Science, 495–580.
375	Kumar S, Stecher G, Tamura K. 2016. MEGA7: Molecular evolutionary genetics analysis
376	version 7.0 for bigger datasets. Molecular Biology and Evolution 33: 1870-1874.
377	Larsen K. 2003. Proposed new standardized anatomical terminology for the Tanaidacea
378	(Peracarida). Journal of Crustacean Biology 23: 644-661.
379	Larsen K, Guțu M, Sieg J. 2015. Order Tanaidacea Dana, 1849. In: von Vaupel Klein JC,
	11

380 Charmantier-Daures M, Schram FR, eds. The Crustacea. Revised and updated, as well as extended from the Traité de Zoologie 5. Leiden: Brill, 249–329. 381Leigh JW, Bryant D. 2015. PopART: full-feature software for haplotype network 382construction. *Methods in Ecology and Evolution* **6:** 1110–1116. 383384Marshall NJ, Diebel C. 1995. 'Deep-sea spiders' that walk through the water. Journal of Experimental Biology 198: 1371–1379. 385 Martel A, Chia F-S. 1991. Drifting and dispersal of small bivalves and gastropods with 386 direct development. Journal of Experimental Marine Biology and Ecology 150: 131– 387 388 147. 389 McClain C, Barry J. 2014. Beta-diversity on deep-sea wood falls reflects gradients in energy availability. Biology Letters 10: 20140129. 390 McClain CR, Hardy SM. 2010. The dynamics of biogeographic ranges in the deep sea. 391392Proceedings of the Royal Society B 277: 3533–3546. 393 Nakano S. 1993. Kurozakoebi no horan-oyaebi no yosei to fusyutu, shiikukekka ni tsuite [Rearing results of ovigerous females and hatchlings of Argis lar]. Contributions to the 394*Fisheries Researches in the Japan Sea Block* **29:** 77–91. [in Japanese] 395Norkko A, Cummings VJ, Thrush SF, Hewitt JE, Hume T. 2001. Local dispersal of 396 397 juvenile bivalves: implications for sandflat ecology. Marine Ecology Progress Series 398 **212:** 131–144. Ohtsuka S, Koike K, Lindsay D, Nishikawa J, Miyake H, Kawahara M, Mulyadi, 399 Mujiono N, Hiromi J, Komatsu H. 2009. Symbionts of marine medusae and 400 ctenophores. Plankton & Benthos Research 4: 1–13. 401 Riehl T, Kaiser S. 2012. Conquered from the deep sea? A new deep-sea isopod species from 402 403 the Antarctic Shelf shows pattern of recent colonization. PLoS ONE 7: e49354. 404 Riehl T, De Smet B. 2020. Macrostylis metallicola spec. nov.—an isopod with geographically clustered genetic variability from a polymetallic-nodule area in the 405Clarion-Clipperton Fracture Zone. PeerJ 8: e8621. 406 Riehl T, Lins L, Brandt A. 2018. The effects of depth, distance, and the Mid-Atlantic Ridge 407 408 on genetic differentiation of abyssal and hadal isopods (Macrostylidae). Deep-Sea 409 Research Part II 148: 74–90. Roden GI. 1995. Aleutian Basin of the Bering Sea: thermohaline, oxygen, nutrient, and 410 current structure in July 1993. Journal of Geophysical Research 100: 13539–13554. 411 Rozas J, Ferrer-Mata A, Sánchez-DelBarrio JC, Guirao-Rico S, Librado P, 412413Ramos-Onsins EE, Sánchez-Gracia A. 2017. DnaSP 6: DNA sequence polymorphism 414 analysis of large data sets. *Molecular Biology and Evolution* **34:** 3299–3302. Shiino SM. 1966. On Kalliapseudes (Kalliapseudes) tomiokaensis sp. nov. (Crustacea: 415Tanaidacea) from Japanese waters. *Report of Faculty of Fisheries, Prefectural* 416 University of Mie **5:** 473–488. 417

418	Smit NJ, Bruce NL, Hadfield KA. 2019. Parasitic Crustacea, Zoological Monographs 3.
419	Berlin: Springer.
420	Stabeno PJ, Schumacher JD, Ohtani K. 1999. The physical oceanography of the Bering
421	Sea: a summary of physical, chemical, and biological characteristics, and a synopsis of
422	research on the Bering Sea. In: Loughlin TR, Ohtani K, eds. Dynamics of the Bering
423	Sea: a summary of physical, chemical, and biological characteristics, and a synopsis of
424	research on the Bering Sea. British Colombia: North Pacific Marine Science
425	Organization, 1–28.
426	Tajima F. 1989. Statistical method for testing the neutral mutation hypothesis by DNA
427	polymorphism. Genetics 123: 585–595.
428	Taylor ML, Roterman CN. 2017. Invertebrate population genetics across Earth's largest
429	habitat: the deep-sea floor. Molecular Ecology 26: 4872–4896.
430	Uehara K, Miyake H. 1999. Deep flows on the slope inshore of the Kuril-Kamchatka
431	Trench southeast off Cape Erimo, Hokkaido. Journal of Oceanography 55: 559-573.
432	Wessel P, Smith WHF, Scharroo R, Luis J, Wobbe F. 2013. Generic Mapping Tools:
433	improved version released. Eos, Transactions, American Geophysical Union 94: 409-
434	410.
435	Winkelmann I, Campos PF, Strugnell J, Cherel Y, Smith PJ, Kubodera T, Allcock L,
436	Kampmann M-L, Schroeder H, Guerra A, Norman M, Finn J, Ingrao D, Clarke M,
437	Gilbert MTP. 2013. Mitochondrial genome diversity and population structure of the
438	giant squid Architeuthis: genetics sheds new light on one of the most enigmatic marine
439	species. Proceedings of the Royal Society B 280: 20130273.
440	Yahagi T, Watanabe HK, Kojima S, Kano Y. 2017. Do larvae from deep-sea hydrothermal
441	vents disperse in surface waters? Ecology 98: 1524–1534.
442	

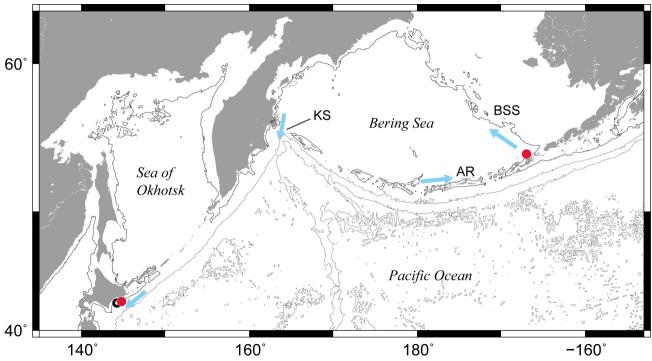
SUPPORTING INFORMATION

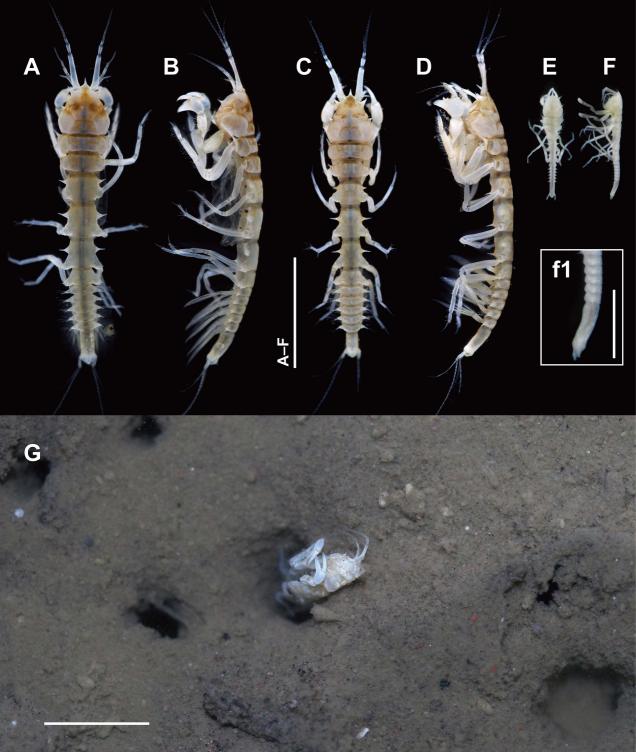
Supporting information S1. Details on sampling, sorting, and sample fixation.

- 446 **Figure legends**
- 447

448 **Figure 1.** Map showing our sampling sites for *Carpoapseudes spinigena* (red circles) and the

- type locality (black open circle; Bamber, 2007). Bathymetric contours are 200 m (thicker
- lines) and 5000 m (thinner lines). Map and plots were generated using GMT5 (Wessel *et al.*,
- 451 2013) based on data publicly available from ETOPO1 (Amante & Eakins, 2009). Bottom
- 452 currents around 1000–1500 m depth (thick blue arrows) were drawn with reference to Kinder
- 453 et al. (1975), Roden (1995), Stabeno et al. (1999), and Uehara & Miyake (1999).
- 454 Abbreviations: AR, Aleutian Ridge; BSS, Bering Sea Slope; KS, Kamchatka Strait.
- 455
- 456 Figure 2. *Carpoapseudes spinigena* collected from the Bering Sea. A, B, Fresh female,
- 457 dorsal (A) and left (B) views; C, D, fresh male, dorsal (C) and left (D) views; E, F, f1,
- 458 ethanol fixed manca II, dorsal (E) and left (F, f1) views (f1, pleon and pleotelson), most of
- 459 uropod lost; **G**, living individual without major disturbance in collection, with its posterior
- 460 half in a burrow, found in a sediment core sample. Scale: f1, 1 mm; others, 5 mm.
- 461
- 462 **Figure 3.** IntNJ network for COI haplotypes (658 nt) from 21 and 11 *Carpoapseudes*
- 463 *spinigena* individuals from the Japan and Bering Sea populations, respectively. Labeled
- 464 circles indicate haplotypes, with size of the circle proportional to frequency of the haplotype;
- 465 number of individuals > 1 also labeled inside circles. Smaller, black circle indicates an
- 466 intermediate haplotype not observed; each line between circles indicates a single mutational
- 467 substitution. H1–10, haplotypes 1–10.





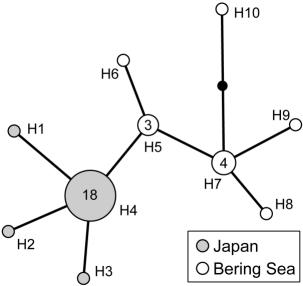


Table 1. List of stations by R/V *Tansei-maru* (KT) and R/V *Mirai* (MR). BT, beam trawl with 3 m opening; D, dredge installed on a Deep Tow Camera System; MC, multiple corer. ^a, net in; ^b, net out.

Station no.	Gear	Date	Start position	End position	Depth (m)
Japan population					
KT-07-29-K1	BT	7 Nov 2007	42°35.0′N	42°34.7 <i>´</i> N	1028–1075
			144°48.0'E ^a	144°49.9´E ^b	
Bering Sea population					
MR17-04_Leg2 St. G	D	15 Aug 2017	54°11.6378´N	54°11.6336′N	1566–1569
			166°59.3735′W	166°59.4198´W	
MR17-04_Leg2 St. G	MC	15 Aug 2017	54°11.7635 <i>°</i> N		1536
			166°58.3413′W		

Table 2. Genetic diversity indices for two *Carpoapseudes spinigena* populations. *N*, number of individuals; N_h , number of haplotypes; N_p , number of polymorphic sites; *h*, haplotype diversity; π , nucleotide diversity.

Population	N	N_h	Np	H	π	Tajima's <i>D</i>	Fu's F _S
Japan	21	4	3	0.271	0.0004	-1.727 (p = 0.024)	$-2.820 \ (p < 0.001)$
Bering Sea	11	6	6	0.836	0.0022	-1.218 (<i>p</i> = 0.115)	-2.508 (<i>p</i> = 0.013)
Overall	32	10	10	0.673	0.0020	-1.465 (p = 0.064)	-5.076 (p = 0.001)