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Elevated Myeloperoxidase-DNA Complex Levels in Sera of Patients with IgA Vasculitis

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Keywords

IgA vasculitis · Neutrophil extracellular traps · Antineutrophil cytoplasmic antibody · Renal complications

Abstract

Introduction: IgA vasculitis is a systemic disease that results from the entrapment of circulating IgA-containing immune complexes in small-vessel walls in the skin, kidneys, and gastrointestinal tract. An excessive formation of neutrophil extracellular traps (NETs) is involved in the pathogenesis of vasculitis, especially in antineutrophil cytoplasmic antibody (ANCA)-associated vasculitis. This study aimed to clarify whether NETs are implicated in IgA vasculitis. **Methods:** Twenty-two patients with IgA vasculitis and 4 healthy volunteers were enrolled in this study. Serum levels of myeloperoxidase (MPO)-DNA complex, a fragment derived from NETs, were determined by enzyme-linked immunosorbent assay (ELISA), and the association between MPO-DNA complex levels and clinical parameters was examined. The presence of the ANCA was also assessed by ELISA specific for MPO and proteinase 3 (PR3) and indirect immunofluorescence (IIF), followed by assessing the differences in clinical parameters with and without the ANCA. **Results:** Serum MPO-DNA com-

plex levels were significantly higher in patients with IgA vasculitis than those in healthy controls. A significant positive correlation between the serum MPO-DNA complex and IgA levels was noted. Interestingly, 63.6% of IgA vasculitis patients were ANCA-positive in IIF with an atypical pattern, whereas neither MPO-ANCA nor PR3-ANCA was detected by ELISA. These findings indicated that some IgA vasculitis patients possessed the so called minor ANCA. Serum IgA and MPO-DNA complex levels and the frequency of hematuria in the minor ANCA-positive group were significantly higher than in the minor ANCA-negative group. **Conclusion:** The collective findings suggested that NETs are certainly involved in the pathogenesis of IgA vasculitis.

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Introduction

Henoch-Schönlein purpura, IgA vasculitis as adopted by the 2012 Chapel Hill consensus conference (CHCC 2012), is a multisystem disease believed to be a consequence of the entrapment of circulating IgA-containing immune complexes in small-vessel walls in the skin, kidneys, and gastrointestinal tract [1].

Neutrophil extracellular traps (NETs) are web-like DNA decorated with antimicrobial proteins, including histones, myeloperoxidase (MPO), and proteinase 3 (PR3), which are released from microorganism-triggered neutrophils [2]. Accumulating evidence indicates that an excessive formation of NETs is involved in the pathogenesis of vasculitis, especially in antineutrophil cytoplasmic antibody (ANCA)-associated vasculitis [3, 4]. In ANCA-associated vasculitis, the ANCA binds with MPO and PR3, expressed on neutrophils activated by pro-inflammatory stimuli; consequently, excessive activation of neutrophils occurs. These neutrophils release reactive oxygen species, proteolytic enzymes, and NETs, injuring the endothelium of small vessels.

An excessive formation of NETs induces microinflammation in the small-vessel wall via immunothrombus formation [5, 6]. In contrast, Aleyd et al. [7] have demonstrated that IgA immune complexes in the plasma and synovial fluid of patients with rheumatoid arthritis induced NETs. Based on this evidence, it is hypothesized that NETs are implicated in the pathogenesis of IgA vasculitis.

To verify the hypothesis, 22 patients with IgA vasculitis and 4 healthy volunteers were enrolled in this study. Serum levels of the MPO-DNA complex, a fragment derived from NETs, were determined by enzyme-linked immunosorbent assay (ELISA), and the association between MPO-DNA complex levels and clinical parameters was examined. The presence of the ANCA was also assessed by ELISA specific for MPO and PR3 and indirect immunofluorescence (IIF), followed by assessing the differences in clinical parameters with and without the ANCA.

Patients and Methods

Patients

This study reviewed the sequential records of 22 patients with IgA vasculitis seen at the Department of Dermatology, St. Marianna University School of Medicine between 2010 and 2016. Patients presented with initial cutaneous manifestations characteristic of IgA vasculitis, with palpable purpura over the lower extremities. Based on the presence of leukocytoclastic vasculitis, vascular IgA depositions, and the absence of thrombocytopenia or known hematologic or connective tissue disorders, they were diagnosed with IgA vasculitis. The diagnosis was made according to the criteria defined by the American College of Rheumatology [8] and the KAWAKAMI algorithm [9]. None of the patients demonstrated any coexisting malignancy, other autoimmune diseases, or viral hepatitis and was positive for mixed cryoglobulinemia.

Table 1. Clinical and serological findings in IgA vasculitis

Characteristic	(n = 22)
Age, years	38.90±19.25
Gender (M/F)	8/14
Arthralgia, n (%)	11 (50.0)
Abdominal pain, n (%)	8 (36.4)
Proteinuria, n (%)	14 (63.6)
Hematuria, n (%)	15 (68.2)
CRP, mg/dL	1.62±2.33 (normal, <0.3)
IgA, mg/dL	340.7±175.6 (normal, 95–360)
ANCA in IIF, n (%)	14 (63.6)

ANCA, antineutrophil cytoplasmic antibody; CRP, C-reactive protein; IIF, indirect immunofluorescence.

Serum Samples

All blood samples were immediately centrifuged at 1,500 g for 30 min at 4°C. Serum samples were collected and stored at –80°C until analysis. Immunological assessments were performed on sera collected when each patient presented with palpable purpura over the lower extremities. For controls, 4 healthy volunteers were included.

Detection of MPO-ANCA, PR3-ANCA, C-Reactive Protein, and IgA

Serum MPO-ANCA and PR3-ANCA were determined by ELISA (Medical & Biological Laboratories, Tokyo, Japan) according to the manufacturer's protocol. Serum IgA and C-reactive protein (CRP) levels were assayed using laser nephelometry (Nihon Kohden, Tokyo, Japan).

IIF for ANCA

A commercially available IIF kit (Medical & Biological Laboratories) was used for ANCA detection according to the manufacturer's instructions. In brief, ANCA detection by IIF was performed on ethanol-fixed and formalin-fixed neutrophils. Serum samples were diluted in phosphate-buffered saline and screened at a dilution of 1:20. Interpretation of immunofluorescence results was based on the characteristics of the ethanol- and formalin-fixed specimens.

Quantification of MPO-DNA Complex

The amount of in vivo NET formation can be monitored as MPO-DNA complexes in the serum [3]. ELISA for MPO-DNA complexes was carried out, as described previously [10, 11]. In brief, microtiter plates (Thermo Fisher Scientific, Waltham, MA, USA) were first coated with the monoclonal antihuman MPO antibody (5 µg/mL; AbD Serotec, Kidlington, UK) and incubated overnight at 4°C. After blocking with 1% bovine serum albumin, the sera (final dilution at 1:3) and horseradish peroxidase-conjugated anti-DNA antibody in the Cell Death Detection kit (Roche Diagnostics, Tokyo, Japan) were applied to the wells, followed by color development according to the manufacturer's instructions. The serum from a patient with MPO-ANCA-associated vasculitis served as a positive control.

Statistical Analysis

Differences in patient parameters were assessed using the Mann-Whitney U-test and χ^2 test for independent samples. Cor-

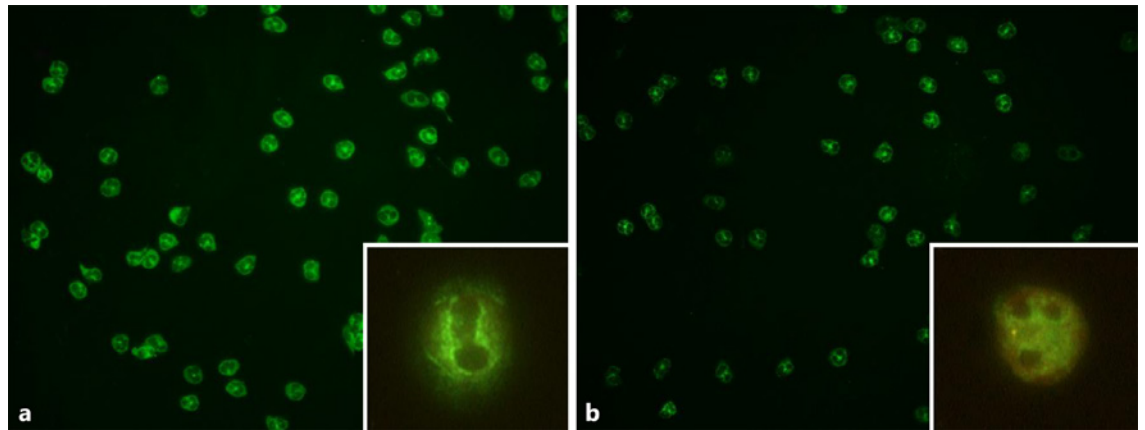


Fig. 1. IIF detection of ANCA in IgA vasculitis. IIF demonstrated an atypical pattern with fine rim-like staining of the nuclear periphery in ethanol-fixed neutrophils (**a**) and a cytoplasmic fluorescence in formalin-fixed neutrophils (**b**). **insets** High-power field of view. IIF, indirect immunofluorescence; ANCA, antineutrophil cytoplasmic antibody.

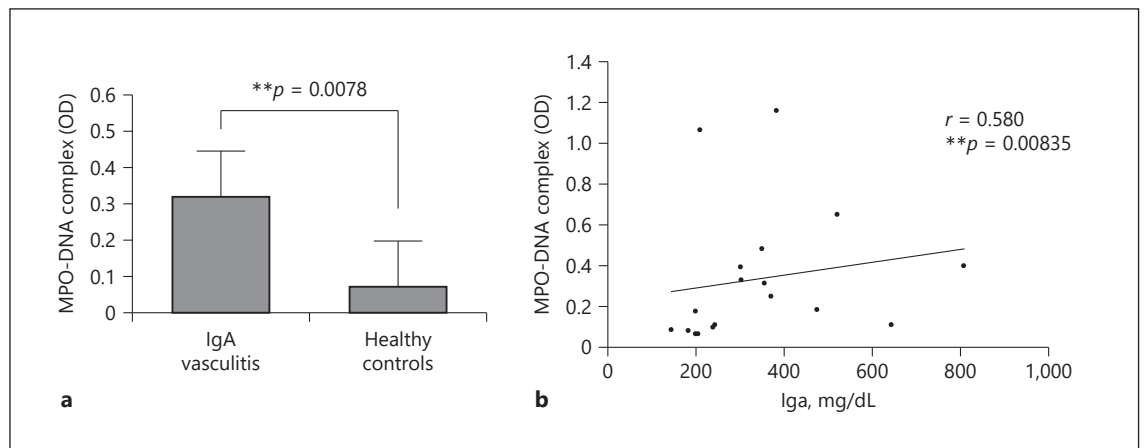


Fig. 2. Correlation between the serum MPO-DNA complex and IgA levels in IgA vasculitis. **a** Serum MPO-DNA complex levels were significantly higher in IgA vasculitis patients than in healthy controls. **b** A significant positive correlation was noted between the serum MPO-DNA complex and IgA levels in IgA vasculitis patients. MPO, myeloperoxidase.

relations between quantitative parameters were measured by Spearman's rank correlation coefficient. A p value < 0.05 was considered statistically significant. All data are expressed as mean \pm standard deviation.

Results

Clinical and Serological Findings in Patients with IgA Vasculitis

The clinical and serological features of 22 IgA vasculitis patients are listed in Table 1. Fourteen (63.6%) patients

were positive for serum CRP. Serum MPO-ANCA and PR3-ANCA were negative according to ELISA in all patients. In contrast, the ANCA was detected in 14 (63.6%) patients based on IIF. When ethanol-fixed neutrophils were subjected to IIF, positive fluorescence was detected as fine rim-like signals of the nuclear periphery distinct from the typical perinuclear-ANCA pattern (Fig. 1a), whereas the typical cytoplasmic-ANCA pattern was observed on formalin-fixed neutrophils (Fig. 1b). ANCAs that demonstrate such characteristics are atypical ANCAs [12]. All sera from ANCA-positive IgA vasculitis patients enrolled in this study demonstrated the atypical ANCA

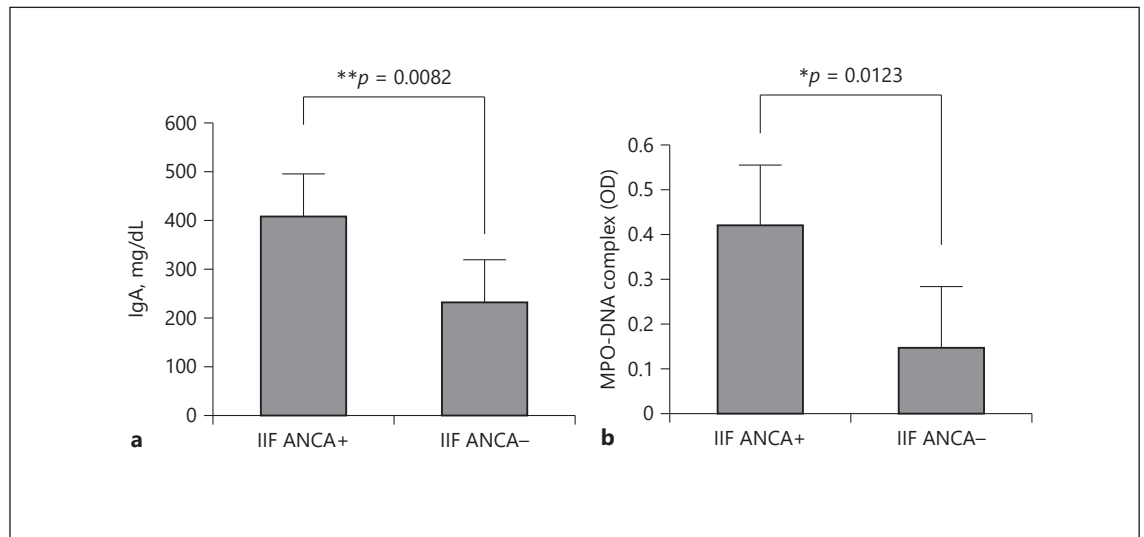


Fig. 3. Differences in serum IgA and MPO-DNA complex levels in IgA vasculitis based on ANCA production. **a** Comparison of serum IgA levels between the minor ANCA-positive and ANCA-negative groups. **b** Comparison of serum MPO-DNA complex levels between the minor ANCA-positive and ANCA-negative groups. MPO, myeloperoxidase; ANCA, antineutrophil cytoplasmic antibody; IIF, indirect immunofluorescence.

pattern. It has been shown that ANCAs recognize neutrophil cytoplasmic antigens other than MPO and PR3, namely, minor ANCAs, which usually exhibit the atypical pattern [12]. These findings indicated that some IgA vasculitis patients possessed the so called minor ANCA.

Correlation between Serum MPO-DNA Complex and IgA Levels in IgA Vasculitis

Circulating MPO-DNA complex levels in the serum of 22 IgA vasculitis patients and 4 healthy volunteers were determined by ELISA. Serum MPO-DNA complex levels were significantly higher in IgA vasculitis patients than healthy controls (Fig. 2a). A significant positive correlation between the serum MPO-DNA complex and IgA levels in 22 IgA vasculitis patients was noted (Fig. 2b; $r = 0.580$, $p = 0.00835$).

Differences in Serum IgA and MPO-DNA Complex Levels in IgA Vasculitis Patients Based on ANCA Production

Twenty-two IgA vasculitis patients were divided into 2 subgroups: minor ANCA-positive and ANCA-negative groups. Both serum IgA and MPO-DNA complex levels in the minor ANCA-positive group were significantly higher than those in the minor ANCA-negative group (Fig. 3a, b). In addition, the frequency of hematuria in the minor ANCA-positive group was significantly higher than that in the minor ANCA-negative group ($p = 0.0195$).

Similar trends were noted in the frequency of proteinuria, although the difference did not reach statistical significance ($p = 0.0540$). Any correlation among clinical and laboratory parameters, such as serum CRP levels, arthralgia, and abdominal pain, between the minor ANCA-positive and minor ANCA-negative groups was not observed.

Discussion

This study demonstrated that serum levels of the MPO-DNA complex, a fragment derived from NETs, were significantly higher in IgA vasculitis patients than in healthy controls. A significant positive correlation was noted between the serum MPO-DNA complex and IgA levels in IgA vasculitis patients. It is generally accepted that increased serum IgA concentration is a characteristic laboratory finding during the acute stage of IgA vasculitis [13–15]. Thus, it is suggested that NETs may be associated with the disease activity of IgA vasculitis.

The 2 major categories of small-vessel vasculitis are ANCA-associated vasculitis and immune complex vasculitis, both of which often affect small vessels in the skin [16]. Interestingly, minor ANCA that recognized neutrophil cytoplasmic antigens other than MPO and PR3 was detected in 63.6% of IgA vasculitis patients in this study. These findings suggested the presence of a common im-

munological basis underlying ANCA-associated vasculitis and immune complex vasculitis, including IgA vasculitis, which can explain similar manifestations in the skin, rather than the complication of the 2 diseases.

Serum IgA and MPO-DNA complex levels and the frequency of hematuria in the minor ANCA-positive group were significantly higher than those in the minor ANCA-negative group. Similar trends were noted in the frequency of proteinuria between the groups. The collective findings suggested the association between minor ANCA production and disease activity of IgA vasculitis, including renal manifestations.

It is debatable whether the elevated NET formation is a result or cause of IgA vasculitis. IgA immune complexes have been demonstrated to induce NETs [7]. MPO-ANCA and PR3-ANCA can perpetuate a vicious circle of NET production that maintains the delivery of antigen-chromatin complexes to the immune system [3]. Similarly, some minor ANCAs can enhance NET formation [17, 18]. Although further studies are needed to identify antigens recognized by minor ANCAs in IgA vasculitis patients and determine whether the minor ANCA induces NETs, this study suggested that NETs are certainly involved in the pathogenesis of IgA vasculitis.

Acknowledgment

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Statement of Ethics

This study was approved by the St. Marianna University Ethics Committee (No. 3755), and written informed consent was obtained from all patients and healthy volunteers.

Conflict of Interest Statement

The authors have no conflicts of interest to declare.

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Author Contributions

Sora Takeuchi, Tamihiko Kawakami, and Akihiro Ishizu designed the research. Sora Takeuchi, Tatsuro Okano, Haruki Shida, and Daigo Nakazawa performed the experiments. Sora Takeuchi, Tatsuro Okano, Haruki Shida, Daigo Nakazawa, Utano Tomaru, and Akihiro Ishizu analyzed the data. Sora Takeuchi, Tamihiko Kawakami, and Akihiro Ishizu wrote the manuscript. Takafumi Kadono supervised this study. All the authors discussed the results and contributed to the preparation of the manuscript.

Data Availability Statement

All data generated or analyzed during this study are included in this article. Further inquiries can be directed to the corresponding author.

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