Title	Autumn potato seedling failure due to potato dry rot in Nagasaki Prefecture, Japan, caused by Fusarium acuminatum and Fusarium commune
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Footnotes;

26	The nucleotide sequence data reported are available in the DDBJ/EMBL/GenBank
27	databases under the accession numbers LC469781-LC469790.
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29	Author contributions;
30	All authors contributed to the study conception and design. Material preparation, data
31	collection and analysis were performed by Hisashi Osawa, Yu Sakamoto, Seishi Akino
32	and Norio Kondo. The first draft of the manuscript was written by Hisashi Osawa and
33	all authors commented on previous versions of the manuscript. All authors read and
34	approved the final manuscript.
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19	Abstract
50	Failure to sprout due to seed-tuber rot is a serious problem for autumn potato

51	seedling cultivation in Nagasaki Prefecture, Japan. In this study, five strains were
52	isolated from rotten seed tubers sampled in 2015; when tubers were inoculated with
53	these strains, the tubers developed rot and failed to sprout. We identified these strains as
54	Fusarium acuminatum, Fusarium commune, and the known agent of potato dry rot,
55	Fusarium oxysporum based on morphological and DNA sequencing analyses. F.
56	commune and F. acuminatum were identified as causal agents of potato dry for the first
57	time in the world and in Japan, respectively.
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59	Keywords
60	Dry rot, Potato, Fusarium commune, Fusarium oxysporum, Fusarium acuminatum
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74	Double cropping of potatoes is common in parts of southwestern Japan, such
75	as Nagasaki Prefecture in the Kyushu region, but seed-tuber rot has caused a shortage of

autumn seedlings (Sakamoto et al. 2016a), especially of cultivars Aiyutaka (Nakao et al. 2004) and Sanjumaru (Mukojima et al. 2012) and yield losses in the main cropping areas of Nagasaki. Sections of rotten seed tubers were covered with mycelia and occasionally have wrinkles. Sprouting disorders have also been observed (Sakamoto et al. 2016b). Internal necrotic areas of rotten tubers were shades of brown, ranging from fawn to dark chocolate. Some severely rotten tubers may exhibit a soft rot caused by secondary saprophytic microorganisms. When viewed with a microscope, tuber sections contained hyphae with crescent-shaped conidia characteristic of *Fusarium* spp. Here we identified the causal agents of this tuber rot and sprouting disorder.

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Rotten seed tubers were unearthed from areas where seedlings failed to appear in potato cropping fields of the Nagasaki Agricultural and Forestry Technical Development Center (Unzen, Nagasaki, Japan) in October 2015. The rotten tubers were placed in plastic containers with wet paper and incubated at room temperature for 2–7 days. Newly produced hyphal masses were sampled and cultured on Komada medium (Komada 1975). Eighteen single-spore isolates were obtained from colonies grown on Komada medium, and five strains (F1501, F1503, F1510, F1516, and F1514) were selected from each host cultivar. Each isolate was grown on potato-sucrose agar medium for 2 weeks to obtain conidia. They were tested for pathogenicity by dipping six cut tubers (cv. Irish Cobbler) in a suspension of the respective conidia  $(2.0 \times 10^5)$ conidia/mL) for approximately 10 s. The tubers were then dried, placed in plastic bags filled with noninfested potting soil mixture (Katakura & Co-op Agri, Tokyo, Japan), and incubated at 25°C for 8 days. Tubers were then checked for rot. Tubers for a negative control were treated the same way but dipped in distilled water instead of a conidial suspension. All inoculated tubers ad abundant mycelia on the surface (Fig. 1a), except for two tubers inoculated with F1516; and all tubers with mycelia were soft rot. Some

rotten tubers had symptoms similar to those of dry rot including wrinkles and gaps, in addition to internal lesions filled with mycelia. The rotten tissue caused by strain F1514 was dark brown, whereas rotten tissue caused by other strains was cream-colored to light brown. Sprouting and root growth were observed in tubers in the negative control plot. However, inoculated tubers roots rarely grew; sprouting was not observed in any of the five tested strains (Fig. 1a, b). The five pathogenic strains (F1501, F1503, F1510, F1516, and F1514) were provided to the NARO Genebank project, Japan, as MAFF246882–246886, respectively.

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DNA was extracted from approximately 100 mg of fresh mycelia, that had been grown in potato-sucrose broth for 2 weeks, using a FavorPrep Plant Genomic DNA Extraction Mini Kit (Favorgen Biotech, Ping-Tung, Taiwan). Translation elongation factor 1-α (TEF) (O'Donnell et al. 1998) and mitochondrial small subunit (mtSSU) rDNA regions (White et al. 1990) were amplified by PCR in a 25-µL reaction volume containing template DNA, 15 μL distilled water, 2.5 μL 10× Ex Taq buffer, 2.0 μL 2.5 mM dNTPs, 1.6 μL 25 mM MgCl<sub>2</sub>, 2.5 μL 0.1% (w/v) bovine serum albumin, 0.4 U TaKaRa Ex Taq polymerase (TaKaRa Bio, Kusatsu, Japan), and 10 μM of each primer as previously described (O'Donnell et al. 1998; White et al. 1990). The thermal cycling conditions for TEF were initial denaturation at 95°C for 5 min; 36 cycles of denaturation at 95°C for 30 s, annealing at 55°C for 30 s, and extension at 72°C for 1 min; and final extension at 72°C for 10 min. For mtSSU, the conditions were the same as for TEF except for annealing at 50°C. PCR products were purified using a NucleoSpin Gel and PCR Clean-up (Macherey-Nagel, Düren, Germany) and sequenced by a DNA sequencing service (Hokkaido System Science, Sapporo, Japan). Sequence data were aligned using ClustalW software (Thompson et al. 1994), and DNA sequences of closely related strains were obtained from GenBank and NARO Genebank.

Aligned sequences from TEF and mtSSU were connected, and neighbor-joining phylogenic trees were produced using the Kimura-2 parameter model (Kimura 1980) in MEGA7 (Kumar et al. 2016). Bootstrap values were determined from 1,000 replications of the calculations. All gaps and missing data were eliminated from the data sets. According to a phylogenic tree from the combined TEF and mtSSU data, the strains cluster into three clades: F1501 belongs to the clade of Fusarium commune; F1503, F1510, and F1516 to the clade of Fusarium oxysporum; and F1514 to the clade of Fusarium acuminatum (Fig. 2). The DNA sequences of the five tested strains were submitted to the DNA Data Bank of Japan (DDBJ), European Molecular Biology Laboratory (EMBL), and GenBank as accession numbers LC469781–LC469790.

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For morphological observations, the five strains were cultured on potato dextrose agar (PDA) at 25°C in darkness for 2 weeks. Colony morphologies were then Structural characteristics of the strains (e.g., conidia, chlamydospores, and examined. phialides) were observed in cultures grown on synthetic low-nutrient agar (SNA) (Nirenberg and O'Donnell 1998) at 25°C under continuous black light for 2 weeks.  $\frac{\text{Con}}{\text{idia}}$  and chlamydospores (n = 50 each) were measured to determine means and ranges. The microconidia of all strains were aseptate or 1-septate, and oval or reniform in shape. The conidia formed false heads on monophialides from short conidiophores. Considerable differences in morphological characteristics were observed between F1514 and the other strains. Colony growth of F1514 on PDA was slower and the underside of the colonies was carmine-colored, whereas the others lacked color or were purple or violet. The macroconidia produced by F1514 on SNA were generally 4— 5-septate, equilaterally curved, and slender, with distinct foot-shaped basal cells. The other strains formed mainly micro- and macroconidia. The macroconidia were generally 3-4-septate and moderately curved, with foot-shaped basal cells (Table S1, Fig. 3). The

teleomorph stage was not observed for any tested strains grown on SNA or PDA. The morphological characteristics of all fungal isolates were compared with published descriptions (Gerlach and Nirenberg 1982). F1514 and others (1503, F1510, and F1516) had characteristics similar to those of F. acuminatum and F. oxysporum, respectively. The characteristics of F1501, which was in the F. commune clade, were also similar to those reported for *F. commune* (Skovgaard et al. 2003).

The results of this study suggest that the lack of sprouting of autumn potato seedlings in Nagasaki was attributable to potato dry rot caused by Fusarium spp., such as F. acuminatum, F. commune, and F. oxysporum. A new taxonomic classification and name for F. oxysporum have been proposed (Lombard et al. 2019); however, we adopted the previous classification in this study because the new name remains controversial. Initially, inoculated tubers were covered with mycelia, had dry rot, wrinkles, and voids with internal lesions. In Japan, Fusarium avenaceum, F. oxysporum, Fusarium solani f. sp. eumartii, F. solani f. sp. radicicola, Fusarium caeruleum, Fusarium graminearum, Fusarium culmorum, Fusarium ventricosum, and Fusarium sambucinum have been identified as causal agents of potato dry rot (Kodama 2004). However, the present study is the first to report potato dry rot caused by F. commune and F. acuminatum globally and in Japan, respectively. F. commune has been identified as a causal agent of root rot and damping-off in forest nurseries (Kim et al. 2012; Stewart et al. 2006); damping-off, seed rot, and seedling root rot in soybeans (Ellis et al. 2013); and crown and root rot in tomatoes (Hamini-Kadar et al. 2010). Both F. commune and F. acuminatum, reported here, may be indigenous to Japan because imported seed tubers have not been grown in the fields surveyed.

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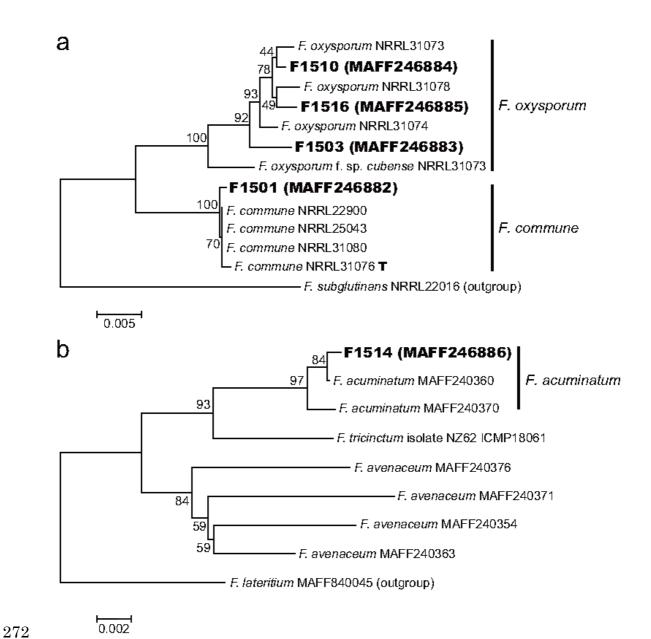
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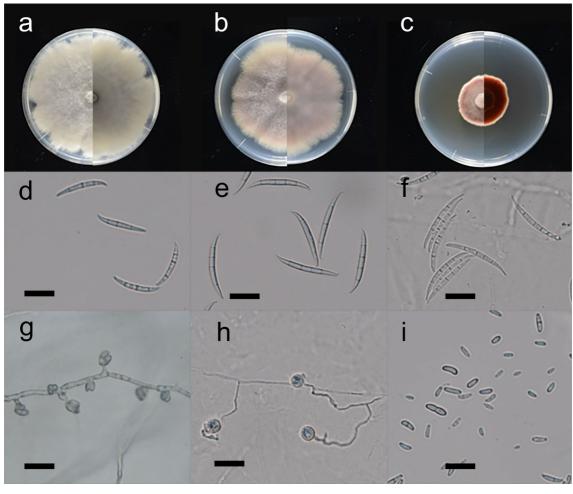
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181	Compliance with ethical standards
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183	Ethical approval: This article does not contain any studies with human participants or
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246	Figure legends
247	Fig. 1 Cut tubers (a) inoculated with pathogenic Fusarium strain F1510 or (b) Cut
248	tubers inoculated with distilled water as negative control.
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250	Fig. 2 Neighbor-joining phylogenic tree based on translation elongation factor $1-\alpha$ and

251	mitochondrial small subunit rDNA sequences. Numbers on branches indicate bootstrap
252	values obtained for 1,000 replicates. Bars indicate substitutions per site. (a) F1501,
253	F1503, F1510, F1516, and related species. T, ex-type strain. (b) F1514 and related
254	species.
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256	Fig. 3 Morphological characteristics of isolated strains. (a-c) Colonies on
257	potato-dextrose agar at 25°C after 2 weeks in darkness. (a-c) Left half of plate images,
258	top surface of plate and right half, reverse side of isolate (a) F1501 (Fusarium
259	commune), (b) F1510 (Fusarium oxysporum), and (c) F1514 (Fusarium acuminatum).
260	(d-f) Macroconidia of (d) F1501 (F. commune), (e) F1516 (F. oxysporum,), and (f)
261	F1514 (F. acuminatum). (g) Short conidiophores and microconidia produced false heads
262	with monophialidic phialides (F1516: F. oxysporum). (h) Smooth-walled
263	chlamydospores (F1501: F. commune). (i) Oval or reniform-shaped microconidia
264	(F1503: F. oxysporum). Scale bars, 20 μm.
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