



Title	Unique Peripheral Antennas in the Photosystems of the Streptophyte Alga <i>Mesostigma viride</i>
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6   **Title:** Unique peripheral antennas in the photosystems of the streptophyte alga

7   *Mesostigma viride.*

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9   **Running Title:** Unique Photosystems in *Mesostigma viride*

10

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22    **Title:** Unique peripheral antennas in the photosystems of the streptophyte alga

23    *Mesostigma viride.*

24

25    **Running Title:** The Unique Photosystems in *Mesostigma viride*

26

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32    **Abbreviations:** CN, clear-native;  $\alpha$ -DDM,  $\alpha$ -dodecyl maltoside; Isoseq, isoform

33    sequencing; LHC, light-harvesting complex; PAGE, polyacrylamide gel electrophoresis

34

35

36

37 [Abstract]

38 Land plants evolved from a single group of streptophyte algae. One of the key factors  
39 needed for adaptation to a land environment is the modification of the peripheral  
40 antenna systems of photosystems. Here, the photosystems of *Mesostigma viride*, an  
41 earliest-branched streptophyte alga, were analyzed to gain insight into their evolution.  
42 Iso-seq and phylogenetic analyses of Light-Harvesting Complexes (LHCs) revealed that  
43 *M. viride* possesses three algae-specific LHCs, including algae-type LHCA2, LHCA9,  
44 and LHCP; while the streptophyte-specific LHCB6 was not identified. These data  
45 suggest that the acquisition of LHCB6 and the loss of algae-type LHCs occurred after  
46 the *M. viride* lineage branched off from other streptophytes. Clear-native (CN)-PAGE  
47 resolved the photosynthetic complexes, including the PSI-PSII megacomplex, PSII-  
48 LHCII, two PSI-LHCI-LHCIIIs, PSI-LHCI, and the LHCII trimer. Results indicated that  
49 the higher-molecular weight PSI-LHCI-LHCII likely had more LHCII than the lower-  
50 molecular weight one, a unique feature of *M. viride* photosystems. CN-PAGE coupled  
51 with mass spectrometry strongly suggested that the LHCP was bound to PSII-LHCII,  
52 while the algae-type LHCA2 and LHCA9 were bound to PSI-LHCI, both of which are  
53 different from those in land plants. Results of the present study strongly suggest that *M.*

54 *viride* photosystems possess unique features that were inherited from a common  
55 ancestor of streptophyte and chlorophyte algae.

56

57 **[Introduction]**

58 Photosystems (PSs) in oxygenic photosynthetic organisms are composed of core and  
59 peripheral antenna complexes. While the peripheral antenna complex in these organisms  
60 is highly divergent, the core complex is highly conserved (Green and Durnford, 1996;  
61 Neilson and Durnford, 2010). The diversity of the peripheral antenna system contributes  
62 to the ability of photosynthetic organisms to adapt to different light environments due to  
63 the multiple roles it plays in photosynthesis, including the harvesting of light energy and  
64 transferring it to the core complex. The variety of light spectra available in different  
65 environments is one of the driving forces behind the evolution of different peripheral  
66 antenna systems in photosynthetic organisms. (Croce and Amerongen, 2014; Stomp et  
67 al., 2007). The thermal dissipation of excess light energy, which occurs mainly in  
68 peripheral antennas (Ruban, 2018), is also necessary to avoid photooxidative damage.  
69 Therefore, photosynthetic organisms have also evolved a variety of thermal dissipation  
70 mechanisms in response to different environments (Giovagnetti and Ruban, 2018; Goss  
71 and Lepetit, 2015; Niyogi and Truong, 2013; Wobbe et al., 2016).

72 Notable differences in peripheral antenna systems have been reported, even  
73 within the green plants of photosynthetic organisms, and especially for PSI (Pan et al.,  
74 2020; Suga and Shen, 2020). In vascular plants, PSI retains four LHCI (Lhca1-Lhca4)  
75 proteins as a peripheral antenna and transiently gains a few extra LHCII proteins  
76 (Mazor et al., 2017; Pan et al., 2018; Qin et al., 2015). PSI in chlorophyte algae  
77 typically have a greater number of LHCI proteins forming their peripheral antenna than  
78 vascular plants. For example, PSI in *Chlamydomonas reinhardtii* has ten LHCI (two  
79 Lhca1 and one Lhca2-Lhca9) proteins and one mobile LHCII trimer that can transiently  
80 bind to the antenna (Su et al., 2019; Suga et al., 2019). A similar antenna structure has  
81 also been reported in *Bryopsis corticulans* (Qin et al., 2019). Chlorophyte algae may  
82 typically have a larger size PSI antenna than vascular plants as an adaptation to aquatic  
83 environments where light intensity is weaker than in terrestrial environments (Suga and  
84 Shen, 2020). In contrast, *Dunaliella salina* has a “mini-PSI” that does not possess a  
85 second LHCI belt unlike other reported chlorophyte algae (Perez-Boerema et al. 2020).  
86 Since *D. salina* has the ability to survive high salinity and high light environments  
87 (Perez-Boerema et al. 2020), the unique PSI structure may represent an adaptation to an  
88 extreme habitat. In the model moss plant *P. patens*, PSI can possess five (Pinnola et al.,  
89 2018) or eight (Iwai et al., 2018) more LHCI proteins than occurs in PSI-LHCI in

90 vascular plants, plus one additional mobile LHCII trimer. The number of LHC proteins  
91 in *P. patens* PSI can change in response its photosynthetic status (Iwai et al., 2015;  
92 Pinnola et al., 2018). The changes in the PSI antenna size reflect the variable light  
93 environment that is typically experienced by mosses, which is characterized by low  
94 irradiance complemented by sunflecks (Pinnola et al., 2018). These reports reveal that  
95 the peripheral antenna system of PSI in green plants exhibit large species-specific  
96 differences that apparently reflect their adaptation to different light environments.  
97 However, to the best of our knowledge, detailed investigations on the peripheral antenna  
98 system of PSI in streptophyte algae have not been reported so far.

99 Significant diversity is also present in thermal dissipation mechanisms in green  
100 plants. Thus far, PsbS- and LHCSR-dependent thermal dissipation mechanisms have  
101 been reported (Niyogi and Truong, 2013). The LHCSR-dependent mechanism is the  
102 primary process used for thermal dissipation in chlorophyte algae, although they  
103 possess both PsbS and LHCSR proteins (Correa-Galvis et al., 2016; Tibiletti et al.,  
104 2016). In contrast, vascular plants only possess PsbS-dependent thermal dissipation,  
105 having lost the LHCSR protein during their evolution (Giovagnetti and Ruban, 2018;  
106 Niyogi and Truong, 2013; Pinnola, 2019). Notably, both PsbS and LHCSR play an  
107 essential role in thermal dissipation in the model moss plant, *P. patens* (Pinnola, 2019).

108 Phylogenetically, *P. patens* represents an early-branched land plant and possesses  
109 thermal dissipation characteristics that are intermediate between chlorophyte algae and  
110 vascular plants. In addition, the contribution of PsbS to the thermal dissipating capacity  
111 in land plants is much higher than it is in chlorophyte algae. How thermal dissipation  
112 mechanisms have changed over the course of evolution from ancestral green algae to  
113 land plants, however, has not been fully elucidated. This can be mainly attributed to the  
114 absence of a suitable streptophyte alga species that could serve as a model for  
115 photosynthesis research.

116 *M. viride* is one of the earliest-branched freshwater streptophyte algae,  
117 although its exact phylogenetic position in green plants is still under debate (Lemieux et  
118 al., 2007, 2000; Li et al., 2020; Wang et al., 2020). Nevertheless, it is worth testing  
119 whether this basally branching alga retains a feature present in a common ancestor of  
120 streptophytes and chlorophytes. Therefore, in the present study, the photosystems of the  
121 earliest-branched *M. viride* were characterized to gain insight into the structure of  
122 photosystems of a common ancestor of streptophytes and chlorophytes. Results revealed  
123 that considerable changes occurred in photosystems after the *M. viride* lineage branched  
124 off from other streptophyte algae.

125

126 [Results]

127 **Separation of photosystems by ClearNative (CN)-PAGE**

128 Clear-Native (CN)-PAGE is a powerful technique that enables one to separate  
129 protein complexes while retaining their structure. Here, the *M. viride* photosystems  
130 were separated using amphipol-based CN-PAGE (Furukawa et al., 2019) after  
131 solubilization with a mild detergent, dodecyl maltoside ( $\alpha$ -DDM). As a result, the PSI-  
132 PSII megacomplex, the PSII-LHCII supercomplexes, the PSI-LHCl-LHCII bands, the  
133 PSI-LHCl, and the LHCII trimer were resolved (Fig. 1A). The identification of the  
134 separated bands was accomplished using 2D-CN/SDS-PAGE followed by immunoblot  
135 analysis (Fig. 1B) and silver-staining (Fig. 1C), as described in previous studies (Järvi et  
136 al., 2011; Takabayashi et al., 2011). The identification of two PSI-LHCl-LHCII bands  
137 were confirmed by further analysis described in a later section of this report. The overall  
138 band profile (Fig. 1A) was similar to the profile for *P. patens* presented by Furukawa et  
139 al. (2019), however, a substantial difference was evident for PSI-LHCl-LHCII. Two  
140 PSI-LHCl-LHCII bands were found in *M. viride* (Fig. 1A), whereas only one PSI-  
141 LHCl-LHCII band was found in the profile of *P. patens* presented by Furukawa et al.  
142 (2019). The presence of two PSI-LHCl-LHCII bands appears to be a unique  
143 characteristic of the PSs of *M. viride* since land plants also exhibit one PSI-LHCl-

144 LHCII band in BN-PAGE and CN-PAGE gels (Järvi et al., 2011; Pesaresi et al., 2009;  
145 Pinnola et al., 2018).

146

147 **Iso-seq analysis to provide a protein database for identification of *M. viride* LHC**

148 **proteins by MS**

149 No information is available on whether commercially-available LHC antibodies react

150 with *M. viride* LHCs. Therefore, mass spectrometry (MS) analysis of the protein

151 complexes resolved in the CN-PAGE was conducted, as described in our previous

152 papers (Takabayashi et al., 2017, 2013), to elucidate the composition of the LHC

153 proteins in the resolved PSI and PSII supercomplexes. This approach can be used to

154 estimate the positions of the bands of protein complexes separated by CN-PAGE,

155 especially for the relatively high expressed proteins. The number of *M. viride*

156 photosynthetic protein sequences available in public databases was limited at the time

157 of our MS analysis. Therefore, Isoform sequencing (Iso-seq) analysis was used to obtain

158 a transcriptome using PacBio sequencing, which provides longer and more complete

159 sequence information relative to short-read sequencing platforms such as Illumina

160 (Zhao et al., 2019). Full-length cDNA sequences enabled us to estimate full-length

161 amino acid sequences with high reliability, a feature that is advantageous for protein  
162 identification by MS.

163 The iso-seq analysis (see Materials and Methods) allowed us to identify nine  
164 PSI core proteins, 11 PSII core proteins, and 14 LHC proteins (Table S1). The small

165 subunits of PSI and PSII were not identified because the TransDecoder software  
166 identified ORFs that are at least 100 amino acids long using the default settings.

167 Notably, the rate of false positives drastically increased when a shorter minimum length  
168 was used. Therefore, default settings were used to predict ORFs in our analysis to obtain  
169 reliable results.

170

## 171 **Phylogenetic analysis of LHC proteins**

172 A BLAST query identified 14 putative *M. viride* LHC proteins (Table S1). The  
173 annotation of the LHC proteins derived from the BLAST query, however, may not be  
174 reliable as considerable sequence similarity has been reported among different LHC  
175 proteins. Therefore, a phylogenetic tree of LHC sequences was constructed based on the  
176 alignment shown in Fig. S1 and each LHC sequence of *M. viride* was annotated based  
177 on the nomenclature of *A. thaliana* LHC proteins, except for LHCBMs (corresponding  
178 to major LHCII (LHCB1, LHCB2, and LHCB3)) and three algae-specific LHCs (algae-

179 type LHCA2, LHCA9, and LHCP). Algae-type LHCA2 is conserved among  
180 chlorophytes, however, it is not closely related to plant-type LHCA2 (Fig. 2). Therefore,  
181 we annotated it as algae-type LHCA2 in this study to avoid any misrepresentation.

182 Based on the phylogenetic analysis, *M. viride* possesses LHCBs, LHCb4, LHCb5,  
183 LHCA1, two LHCA2, LHCA3, and three algae-specific LHCs as peripheral antenna  
184 proteins. The *M. viride* LHC sequences identified by the Iso-seq analysis exhibited good  
185 correspondence with previously reported *M. viride* LHCs (Koziol et al., 2007) (Fig. 2).

186 Some differences, however, were observed relative to the previous classification (Koziol  
187 et al., 2007). These differences are likely due to the considerable sequence similarities  
188 among LHCs, as the LHCs in green lineages rapidly diversified during the early  
189 evolution of green algae. LHCs that were classified differently in the previous report

190 (Koziol et al., 2007) and our present study have been marked with an asterisk (Fig. 2).

191 The Iso-seq analysis did not detect an LHCb6 gene, even though LHCb6 is  
192 highly conserved among streptophyte algae and land plants (Kouřil et al., 2016). This  
193 result is consistent, however, with a previous study by Koziol et al. (2007) in which  
194 LHCb6 sequences were also not identified in *M. viride*, and with the *M. viride* RNA-  
195 seq data generated in the 1000 plant transcriptomes (1kP) project (Carpenter et al.,  
196 2019; Leebens-Mack et al., 2019). In contrast, LHCP proteins were found in *M. viride*

197 (Fig.2), which is also consistent with the previous report by Koziol et al. (2007). LHCP  
198 is a unique LHC protein found in prasinophyte algae, including *Ostreococcus tauri* and  
199 *Micromonas pusilla*. Prasinophyte algae is a group of early-diverging chlorophyte algae,  
200 which gave rise to core chlorophyte algae including *C. reinhardtii*. It is very notable that  
201 *M. viride*, an early-divergent streptophyte alga, also encodes LHCP. Previous studies of  
202 a model prasinophyte alga, *O. tauri*, reported that it contains LHCP as the peripheral  
203 antenna of PSI-LHCI (Six et al., 2005; Swingley et al., 2010), although no detailed  
204 studies have been conducted on LHCP-containing photosystems. A possible loss of  
205 LHC B6 and the presence of LHCP in *M. viride* suggest that the LHC composition in *M.*  
206 *viride* is likely inherited from a common ancestor of streptophyte and chlorophyte algae.

207

### 208 **The distribution of LHC proteins in PSI and PSII**

209 No studies have been conducted on the distribution of *M. viride* LHCs in PSI and PSII,  
210 although they have been identified and classified (Koziol et al., 2007). Therefore, we  
211 determined the protein composition of the separated photosystems using MS. As  
212 expected, PSI, PSII, and LHC proteins were detected in the PSI-PSII megacomplex  
213 band (Table S2). Also as expected, PSI and LHC proteins were detected in the PSI-  
214 LHCI band (Table S3). Unexpected proteins were rarely detected in these two

215 photosystems, suggesting that contamination from the other protein complexes was  
216 limited. The PSII-LHCII band also contained PSII and LHC proteins, although some  
217 contamination from PSI subunits and the NDH-like complex subunits were observed to  
218 some extent (Table S4).

219 Importantly, the bands representing protein complexes on electrophoresis gels  
220 are often distorted by tailing at both ends. The protein complexes in the tailings are also  
221 detected by high-resolution of MS, which is the main source of contamination in a band  
222 of interest. To predict unknown protein complexes and to estimate the protein  
223 compositions of known protein complexes, a protein migration profile was used after  
224 the native-PAGE coupled with MS (Takabayashi et al., 2017, 2013). A protein migration  
225 profile is a plot where the y-axis represents the amount of protein estimated from MS  
226 data, while the x-axis represents the migration distance on a native-PAGE gel. A peak in  
227 the protein migration profile has been demonstrated to correspond to the position of the  
228 band (Helbig et al., 2009; Remmerie et al., 2011; Wessels et al., 2009). This approach  
229 allows one to determine if the proteins in a band detected by MS are contaminants from  
230 other protein complexes by verifying their peak positions in protein migration profiles  
231 (Helbig et al., 2009; Müller et al., 2016; Takabayashi et al., 2017; Wessels et al., 2009).

232 In the present study, migration profiles were constructed for PSI, PSII, and  
233 LHC proteins based on MS data. A normalized spectral abundance factor (NSAF)  
234 method, which is a label-free quantification method (Zybailov et al., 2006), was used to  
235 estimate the amount of protein from MS data. First, the migration profiles of PSI and  
236 PSII were compared. The distance between PSII-LHCII and PSI-LHCl-LHCII on the  
237 CN-PAGE gel was relatively close, and PSI-LHCl-LHCII bands contained considerable  
238 amounts PSI-LHCl-LHCII proteins. Nevertheless, it was possible to isolate those bands  
239 using their migration profiles (Fig. 3A). The PSII-LHCII peak was observed in the  
240 position of gel slices 8 and 9 (Fig. 3A), where the band was distinctly visible in the CN-  
241 PAGE gel (Fig. 1A). The PSI-LHCl-LHCII and the PSI-LHCII peaks were observed in  
242 the positions of gel slices 12 to 15 (Fig. 1A), where those bands were also visible in the  
243 CN-PAGE gel (Fig. 1A). These data indicate that it is possible to distinguish PSI-bound  
244 LHCs and PSII-bound LHCs by comparing their migration profiles with the migration  
245 profiles of PSI and PSII.

246 Comparing the migration profiles of LHCs with those of PSI and PSII resulted  
247 in the classification of LHCs into two groups. One group included LHCA1, LHCA2,  
248 LHCA3, algae LHCA2, and LHCA9 proteins, whose peaks overlapped with the peaks  
249 of PSI-LHCl and PSI-LHCl-LHCII (Fig. 3B). The other group included LHCB4,

250 LHC5, and LHCP proteins, whose peaks overlapped with the peak of PSII-LHCII  
251 (Fig. 3C). These data strongly suggest that PSI-LHCl includes the former LHCs,  
252 whereas PSII-LHCII includes the latter LHCs. Notably, the migration profile of  
253 LHCBM proteins appeared to be different from the profiles of other LHCs. Its highest  
254 peak corresponded with LHCII trimers, suggesting that LHCBM is the major  
255 component of the LHCII trimer.

256

257 **Separation of PSI-LHCl-LHCII complexes by sucrose density gradient**

258 Two PSI-LHCl-LHCII bands were observed on CN-PAGE gels. A “two-step”  
259 separation of those bands was performed to improve the purity of the bands observed on  
260 CN-PAGE using sucrose density gradient centrifugation as the first “rough” separation  
261 step prior to further separation by CN-PAGE. As a result, three bands (B1-B3)  
262 containing photosynthetic pigments were identified after separation by sucrose density  
263 gradient centrifugation (Fig. 4). After subsequent separation by CN-PAGE, PSI-LHCl-  
264 LHCII, in addition to PSI-LHCl, were found to be present in the middle band (B2) (Fig.  
265 4), while the bottom band (B3) primarily contained PSII-LHCII, and the upper band  
266 (B3) contained LHCII trimer (Fig. 4).

267       The three PSI-containing bands present in the CN-PAGE gel were further  
268       separated by SDS-PAGE after they were cut out of the CN-gel to compare their  
269       composition. Two replicates for each PSI-LHCl-LHCII band were subjected to 2D-  
270       SDS-PAGE (Fig. 5). Silver-staining of the resulting gels revealed the PsaA/PsaB  
271       heterodimer, the PsaA and the PsaB monomers, LHC monomers (c.a. 20kDa), and the  
272       small PSI subunits. A portion of PsaA and PsaB remained as a heterodimer and migrated  
273       slowly in the gel despite the inclusion of SDS (Fig. 5). A comparison of the three CN-  
274       PAGE bands containing PSI indicated that the stoichiometry of the PSI and LHCl  
275       subunits were similar, while the amounts of LHCII were greatest in the top band and  
276       lowest in the bottom band. These results indicate that variations in the size of the three  
277       CN-PAGE bands containing PSI were attributed to differences in the number of LHCII  
278       (mainly LHCBM) subunits bound to each PSI-LHCl complex. Importantly, the  
279       identification of the LHCII band was based on the separation pattern of the B1 and B3  
280       bands containing LHCII trimers by the subsequent 2D-SDS-PAGE (Fig. S2). These data  
281       suggest that the larger PSI-LHCl-LHCII possesses more LHCII than the smaller one.

282

### 283       [Discussion]

#### 284       Unique peripheral antenna systems of *M. viride*

285 We classified the LHCs in the peripheral antenna of PSI and PSII. LHC B4 and LHC B5  
286 were identified as the peripheral antenna of PSII-LHC II, similar to land plants. In  
287 contrast, LHC B6 was not identified in the Iso-seq analysis in this study. The absence of  
288 LHC B6 is consistent with the reported absence of LHC B6 sequences in *M. viride* LHC  
289 sequences (Koziol et al., 2007) and is also consistent with the *M. viride* RNA-seq data  
290 generated in the 1KP project (Carpenter et al., 2019; Leebens-Mack et al., 2019).  
291 Therefore, it is likely that *M. viride* does not possess LHC B6, although we do not  
292 exclude the possibility that the expression level of LHC B6 is relatively low compared to  
293 other LHCs, and thus was undetected. LHC B6 is widely distributed among streptophyte  
294 algae and land plants but has not been found in chlorophyte algae, including *C.*  
295 *reinhardtii* (Grebe et al., 2019; Kouřil et al., 2016). The putative loss of LHC B6 in *M.*  
296 *viride* suggests that LHC B6 was acquired during evolution after the divergence of *M.*  
297 *viride*.

298 *M. viride* PSII possesses LHCP (Koziol et al., 2007), the main peripheral  
299 antenna protein in the model prasinophyte alga, *O. tauri* (Six et al., 2005; Swingley et  
300 al., 2010), whereas LHCP has not been found in land plants. The peaks in the migration  
301 profile of LHCP corresponded with the PSI-PSII megacomplex and the PSII-LHC II  
302 (Fig. 3C). These data suggest that LHCP binds to PSII-LHC II in *M. viride*, although

303 further biochemical studies will be required to confirm this possibility. It should be  
304 noted that detection of the LHCP peak at the position of the PSI-PSII megacomplex  
305 could be explained by assuming co-migration of the PSI-PSII megacomplex and PSII-  
306 LHCII at the top part of CN-PAGE (Fig. 3C). Such co-migration of these complexes  
307 was actually observed in *A. thaliana* (Yokono et al. 2019).

308 A previous report revealed that LHCP, which is conserved among algae in the  
309 Mamiellophyceae, including *O. tauri* and *M. pusilla*, is a major LHC antenna protein in  
310 PSI-LHCI in the model prasinophyte algae, *O. tauri* (Swingley et al., 2010). It has also  
311 been suggested that LHCP functions as the peripheral antenna of PSII-LHCII, although  
312 no direct evidence has been provided (Six et al., 2005; Swingley et al., 2010). Our data  
313 support the idea that LHCP can function as the peripheral antenna of PSII-LHCII (Fig.  
314 3). Collectively, the data suggest that ancestral green algae possessed LHCP proteins as  
315 their peripheral antenna because the earlier-branched streptophyte *M. viride* and the  
316 prasinophyte *O. tauri* possess LHCP proteins in their photosystems. The LHCP-  
317 containing photosystem in *M. viride* was likely inherited from a common ancestor of  
318 chlorophyte and streptophyte algae that has been lost during the evolution of  
319 streptophyte algae to land plants. Considering that the loss of LHCP and the acquisition  
320 of the LHC6 seemed to occur concomitantly according to our phylogenetic analysis, it

321 is possible to hypothesize that LHCP in streptophyte PSII-LHCII have been replaced by  
322 LHC6 in land plants. Further investigation will be required, however, to confirm this  
323 hypothesis.

324 In regards to the peripheral antenna of PSI, two PSI-LHCl-LHCII  
325 supercomplexes were stably observed after their separation by CN-PAGE. LHCII was  
326 more abundant in the higher molecular weight band than it was in the lower molecular  
327 weight band, suggesting that the higher molecular PSI-LHCl-LHCII possesses at least  
328 two LHCII trimers. It is not presently known if at least two PSI-bound LHCII trimers  
329 change their binding to PSI in response to different light environments, i.e., if they are  
330 involved in state transitions. Since the binding of LHCII trimer to PSI was involved in  
331 the state transition according to previous studies in green algae and land plants (Drop et  
332 al., 2014; Pesaresi et al., 2009; Pinnola et al., 2018, 2018; Pribil et al., 2010; Takahashi  
333 et al., 2006), we hypothesize that the additional *M. viride* LHCII trimers are also  
334 involved in the state transition. We do not exclude the possibility, however, that the  
335 smaller form of PSI-LHCl-LHCII represents an artifact caused by the dissociation of  
336 LHCII from the larger form. Further studies are required to confirm this supposition.

337 In addition, PSI-LHCl in *M. viride* possesses algae-type LHCA2 and LHCA9  
338 proteins, which are conserved among chlorophytes including *C. reinhardtii* (Su et al.,

339 2019; Suga et al., 2019) and *Bryopsis corticulans* (Qin et al., 2019), but not conserved  
340 among streptophytes (Fig. 2). The loss of these algae-specific LHCs must have occurred  
341 after the *M. viride* lineage branched off from the other streptophyte algae, although  
342 further studies will also be required to confirm this possibility.

343 We constructed a hypothetical model of the antenna structure of PSI-LHCI-  
344 LHCII in *M. viride* using the reported structure of PSI-LHCI in chlorophyte algae and  
345 land plants as references (Fig. 6). Based on their sequence similarities, the binding  
346 manner of LHCA1, two LHCA2s, and LHCA3 proteins to the PSI core in *M. viride* may  
347 be similar to the binding of LHCA1, LHCA2, LHCA3, and LHCA4 proteins to the PSI  
348 core in vascular plants. Likewise, the binding manner of algae-type LHCA2 and  
349 LHCA2 to PSI-LHCI may be similar to the binding mechanism that occurs in PSI-LHCI  
350 in *C. reinhardtii* (Su et al., 2019; Suga et al., 2019) and *B. corticulans* (Qin et al., 2019).  
351 The binding site of the second LHCII trimer to PSI-LHCI, however, is unknown, as a  
352 PSI-LHCI with two LHCII trimers has not, to our knowledge, been reported. Similarly,  
353 the binding site of LHCP to PSII-LHCII is also unknown as no structural studies on  
354 LHCP-bound PSI-LHCI have been reported. Given the phylogenetic position of *M.*  
355 *viride* and its unique antenna system in PSI and PSII, further structural analyses of *M.*  
356 *viride* PSI-LHCI-LHCII are warranted and would significantly contribute to our

357 understanding of the changes that occurred in photosystems during the evolution of a  
358 common ancestor of chlorophytes and streptophytes to land plants.

359

360 **[Materials and Methods]**

361 **Algal strain and culture conditions**

362 *M. viride* strain, NIES-296, was obtained from the National Institute for Environmental  
363 Studies (NIES) (Ibaraki, Japan). The strain was cultured in 500 mL of C medium in a  
364 1L Erlenmeyer flasks at 22°C under a 14-h photoperiod of 20 µmol photons m<sup>-2</sup> s<sup>-1</sup>, as  
365 previously described by (Kunugi et al., 2016).

366

367 **RNA extraction**

368 One liter of a *M. viride* culture was centrifuged at 10,000 x g for 10 min at 4 °C. The  
369 pellet was resuspended in 0.5 mL RLT buffer (RNeasy Plant Mini Kit, Qiagen)  
370 supplemented with 1% 2-mercaptoethanol. The suspension was then transferred to a 2-  
371 ml vial containing 500 mg of glass beads (0.5 mm diameter). The vial was subjected to  
372 10s disruption treatments using a Mini-Bead Beater (Merck, Germany) and the  
373 suspension was then centrifuged at 21,600×g at 4 °C for 5 min. Supernatants from the

374 vials were used for total RNA isolation with a RNeasy Plant Mini Kit (Qiagen),  
375 according to the manufacturer's instructions.

376

377 **Full-length isoform sequencing (Iso-seq) using PacBio data**

378 The integrity of extracted total RNA was assessed using an Agilent 2100 Bioanalyzer  
379 (Agilent Technologies Inc). The Iso-seq library was prepared according to the protocol  
380 described by Pacific Biosciences (PN 101-070-200) using a SMARTer PCR cDNA  
381 Synthesis Kit (Clontech) and SMRTbell Template Prep Kit 1.0 SPv3 (Pacific  
382 Biosciences). Sequencing was performed on a PacBio Sequel platform.

383 Raw sequence data were processed using SMRT Link v6.0 (Pacific  
384 Biosciences) software and then further processed using IsoSeq3 (version 3.1) software  
385 tools (<https://github.com/PacificBiosciences/IsoSeq>) with default parameters. Circular  
386 consistency sequence (CCS) reads were generated from subread sequences. Full-length  
387 cDNA reads were then selected by finding the 5' and 3' primers or polyA tail using the  
388 lima tool in IsoSeq3. After trimming of the polyA tail and removal of the concatemer  
389 sequences, 84,545 full-length cDNA reads were obtained. Finally, 7,209 high-quality  
390 consensus full-length cDNA sequences were obtained after isoform clustering and  
391 polishing.

392

393 **Estimation of full-length amino acid sequences using the Iso-seq data**

394 TransDecoder (<https://github.com/TransDecoder/TransDecoder/>) software was used to

395 identify full-length protein sequences in the 7,209 full-length cDNAs. Candidate open

396 reading frames (ORFs) with a minimum length of 100 amino acids were identified using

397 the TransDecoder.LongOrfs module. Candidate ORFs were validated by BLASTP

398 queries using an e-value cutoff of  $10^{-5}$  against data protein database comprising proteins

399 of *Arabidopsis thaliana*, *Chara braunii*, *Chlamydomonas reinhardtii*, *Coccomyxa*

400 *subellipsoidea*, *Cyanidioschyzon merolae*, *Cyanophora paradoxa*, *Klebsormidium*

401 *flaccidum*, *Oryza sativa*, *Ostreococcus lucimarinus*, *Physcomitrella patens*, and

402 *Selaginella moellendorffii* within the Phytozome database (v12.1)(Goodstein et al.,

403 2012). After the blast queries, 5,829 protein sequences with significant blast hits were

404 obtained using the TransDecoder.Predict module. CD-HIT (Fu et al., 2012) software

405 was employed to remove redundant protein sequences using a cutoff value of 0.9. A

406 total of 3,198 protein sequences were obtained using CD-HIT software. These protein

407 sequences were used as a *M. viride* protein database for MS queries.

408

409 **Phylogenetic analysis of *M. viride* LHC proteins**

410 *M. viride* LHC proteins were identified by NCBI-BLASTP homology queries against  
411 the *M. viride* protein sequence database described above. *A. thaliana* and *C. reinhardtii*  
412 LHC proteins were used as query sequences in the BLASTP searches. The LHC  
413 sequences of *Arabidopsis thaliana*, *Marchantia polymorpha*, *Klebsormidium flaccidum*,  
414 *Ostreococcus lucimarinus*, and *Chlamydomonas reinhardtii* were obtained from the  
415 Phytozome database (<https://phytozome.jgi.doe.gov/>). The LHCs of *Cyanidioschyzon*  
416 *merolae* and *Pyropia yezoensis* were used as the outgroups in the phylogenetic tree  
417 analysis. Amino acid sequences of LHCs were aligned using the MAFFT algorithm  
418 (Katoh and Standley, 2013). Trimming of the alignment was done by a ClipKIT  
419 program (Steenwyk et al. 2020) with default parameters. A maximum likelihood (ML)  
420 phylogenetic tree including *M. viride* LHCs was constructed using W-IQ-TREE  
421 (Trifinopoulos et al., 2016) software under the best-fitting model (LG+F+I+G4).  
422 Ultrafast bootstrap values (1,000 replicates) are shown below the branches.  
423

#### 424 **Thylakoid membrane preparation**

425 A pellet obtained from the centrifugation (10,000 x g for 10 min at 4 °C) of 1 L of a *M.*  
426 *viride* culture was suspended in 2 mL BN- solubilization buffer (50 mM imidazole/HCl  
427 (pH 7.0), 20% glycerol) with 1% Protease Inhibitor Cocktail for plant cell lysate

428 (Merck, Germany). The suspension was transferred to a 2-ml vial containing 500 mg of  
429 glass beads (0.5 mm diameter). The vial was first immersed in liquid nitrogen and the  
430 cells were subsequently subjected to three 10s disruption treatments using a Mini-Bead  
431 Beater (Merck, Germany). The resulting sample was centrifuged at 21,600×g at 4 °C for  
432 5 min, and the supernatant was suspended in BN- solubilization buffer. The suspension  
433 was centrifuged at 200×g at 4 °C for 1 min, and the supernatant was centrifuged at  
434 21,600×g at 4 °C for 5 min to obtain the thylakoid membranes.

435

#### 436 **Clear-Native (CN)-PAGE**

437 CN-PAGE was performed as previously described (Furukawa et al. 2019). A linear 4–  
438 13% gradient polyacrylamide gel was used as the separation gel, and a 3.5%  
439 polyacrylamide gel was used as the sample gel. Thylakoid membranes were solubilized  
440 in 1%  $\alpha$ -DDM ( $\alpha$ -dodecyl maltoside) at 4 °C for 1 min. After centrifugation at 21,600×g  
441 at 4 °C for 5 min, the supernatant (roughly 5  $\mu$ g chlorophyll equivalent) was  
442 supplemented with 1% Amphipol A8-35 and loaded onto CN-PAGE gels. An anode  
443 buffer (25mM imidazole/HCl (pH 7.0)) and cathode buffer (50mM Tricine, 7.5mM  
444 imidazole) were used for the electrophoresis.

445

446    **Two-dimensional (2D)-CN/SDS-PAGE**

447    CN-gel strips were soaked in denaturation buffer (1% SDS, 50mM DTT) for 30 min,

448    and the 2D-SDS-PAGE was performed using a 14% polyacrylamide gel with 4M urea.

449    The proteins on the 2D-CN/SDS-gel were visualized by silver-staining using a Pierce

450    Silver Stain kit (ThermoFisher Scientific, USA), according to the manufacturer's

451    instructions. Alternatively, the obtained 2D-gels were used in the immunoblot analyses.

452

453    **Immunoblot analysis**

454    Proteins from the 2D-CN/SDS-gel were transferred to a polyvinylidene fluoride

455    membrane (PolyScreen PVDF transfer membrane, PerkinElmer Life Sciences, MA,

456    USA). Protein detection using specific antibodies was performed using Western

457    Lightning Plus-ECL (PerkinElmer Life science, MA, USA) reagent. All antibodies,

458    except anti-PsbB (AS04 038)), were purchased from Agrisera (Vännäs, Sweden).

459

460    **Sucrose density gradient**

461    *M. viride* thylakoid membranes were solubilized in 1%  $\alpha$ -DDM and then loaded onto a

462    continuous sucrose density gradient (0.3M sucrose to 1.3M sucrose in a buffer

463    containing 25mM MES-KOH (pH 6.5) and 0.1% GDN (GDN101, Anatrace)). The

464 samples were then subjected to ultracentrifugation at 72,000  $\times$  g using a S-65T rotor  
465 (Hitachi-koki, Japan) for 15 h at 4 °C.

466

467 **LC-MS/MS analysis**

468 CN-PAGE gel strips were cut horizontally as shown in Fig.1A. All gel pieces were  
469 subjected to in-gel digestion with trypsin and analyzed to identify their peptides by LC-  
470 MS/MS using Orbitrap Elite mass spectrometry (Thermo Fisher Scientific, Waltham,  
471 MA, USA) coupled with Thermo Easy-nLC (Thermo Fisher Scientific, Waltham, MA,  
472 USA). Each sample was loaded onto a C18-reversed phase EASY-Column (0.1 mm  $\times$   
473 20 mm, 5  $\mu$ m particle size, 120 Å pore size), before separation on a C18 Tip column  
474 (75  $\mu$ m  $\times$  120 mm; Nikkyo Technos, Tokyo, Japan). The samples were separated by a  
475 gradient formed by solvent A (0.1% formic acid) and solvent B (acetonitrile in 0.1%  
476 formic acid) at a flow rate of 300 nL/min. The gradient separation setting was as  
477 follows: 0-1min, 0–5% B; 1-12 min, 5–35% B; 12–25 min, 35%–90% B; 90% B; 25–45  
478 min.

479 Proteins in each sample were identified using SearchGUI (version 3.3.15)  
480 software and an andromeda search engine (Barsnes and Vaudel, 2018). The 3,198 M.  
481 *viride* protein sequences identified in the Iso-seq analysis were used for protein

482 identification. PeptideShaker (version 1.16.40) software with default settings was used  
483 to validate the protein identifications (Vaudel et al., 2015).

484 The normalized spectral abundance factor (NASF) (Zybailov et al., 2006)  
485 calculated by the PeptideShaker program was used as a label-free quantification method  
486 based on the LC-MS/MS data to estimate the abundances of the identified proteins in  
487 each gel slice. As previously described, protein migration profiles were then generated  
488 by plotting the NASF values on the y-axis, and plotting the gel slice number on the x-  
489 axis (Takabayashi et al., 2017, 2013).

490

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495

496 **Data availability**

497 Raw data of the Iso-seq analysis have been deposited to the DDBJ Sequence Read  
498 Archive (DRA) (<https://www.ddbj.nig.ac.jp/dra/index.html>) under the accession number  
499 PRJDB10366. In addition, the assembled full-length cDNA sequences of PSI, PSII, and

500 LHC sequences have been deposited to DDBJ/EMBL/GenBank as Transcriptome  
501 Shotgun Assembly (TSA) data under the accession numbers from ICQU01000001 to  
502 ICQU01000034.

503

504 **Disclosures**

505 Conflicts of interest: The authors declare no conflicts of interest.

506

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671

672 [Figure legends]

673 **Figure 1. Separation of photosynthetic complexes by CN-PAGE**

674 Separation of *M. viride* thylakoid membrane protein solubilized in 1%  $\alpha$ -DDM by  
675 amphipol A8-35-based CN-PAGE (A). The number in parentheses represents the  
676 number of the corresponding gel slice which was further subjected to mass spectrometry  
677 analysis (see Fig. 1A). Immunoblot analysis of PsaB (PSI), and PsbB (PSII) proteins  
678 after their separation by 2D-CN/SDS-PAGE (B). Anti-PsaB and anti-PsbB antibodies  
679 were purchased from Agrisera. 2D- CN/SDS-PAGE of *M. viride* thylakoid membrane  
680 protein complexes visualized by silver-staining (C).

681

682 **Figure 2. A phylogenetic tree of *M. viride* LHC proteins**

683 A maximum likelihood phylogenetic tree of *M. viride* LHC proteins. Sequences  
684 highlighted in yellow are clades containing the LHC from *M. viride*. The *M. viride* LHC

685 genes that are surrounded by a box represent the LHC genes identified in the Iso-seq  
686 analysis within the present study. LHCs that were classified differently between the  
687 previous report (Koziol et al., 2007) and our present study are marked with an asterisk.

688

689 **Figure 3. Comparison of LHC migration profiles in PSI and PSII**

690 *M. viride* thylakoid membranes were solubilized in  $\alpha$ -DDM and separated by CN-  
691 PAGE. After separation, gel regions containing the resolved photosystems were  
692 horizontally cut into approximately 1 mm slices. The number shown in parentheses in  
693 Fig. 1A represents the number of the corresponding gel slice. (A) Migration profiles of  
694 PSI and PSII. The X-axis indicates the number of the gel slice as indicated on the CN-  
695 PAGE gel (Fig. 1A). The Y-axis indicates the relative protein abundance estimated using  
696 a label-free quantification method on the data obtained in the MS analysis. The PSII  
697 peak shown in gel slices 8 and 9 likely correspond to the PSII-LHCII band on the CN-  
698 PAGE gel, while the PSI peaks shown in gel slices 11 to 15 likely correspond to the  
699 PSI-LHCl-LHCII band and the PSI-LHCl band. (B) Migration profiles of LHCA1,  
700 LHCA2, LHCA3, algae-type LHCA2, and LHCA9 proteins. Their highest peaks  
701 correspond to the PSI-LHCl-LHCII and PSI-LHCl bands. The protein data of the two  
702 algae-type LHCA2 protein were combined to construct the migration profile of the

703 algae-type LHCA2, as the two LHCA2 protein sequences were similar. (C) Migration  
704 profiles of LHC4, LHC5, and LHCBM proteins. The highest peaks in the LHC4  
705 and the LHC5 migration profiles correspond with the PSII-LHCII band, while the  
706 highest peak in the LHCBM migration profile corresponds to the LHCII trimer. All of  
707 the MS data obtained for the LHCBM proteins were combined due their sequence  
708 similarities.

709

710 **Figure 4. Separation of PSI-LHCl-LHCII supercomplexes using sucrose density**  
711 **gradient centrifugation followed by CN-PAGE.**

712 (A) Separation of *M. viride* thylakoid membrane protein complexes solubilized in 1%  $\alpha$ -  
713 DDM by sucrose density gradient centrifugation (0.3M-1.3M). The middle band (B2)  
714 contained the PSI-LHCl and the PSI-LHCl-LHCII, while the bottom band (B3)  
715 primarily contained PSII-LHCII and the upper band (B3) contained LHCII trimer. (B)  
716 Further separation of the PSI-enriched fraction (B2) in the sucrose gradient by CN-  
717 PAGE.

718

719 **Figure 5. Separation of two PSI-LHCl-LHCII bands and a PSI-LHCl band by 2D-**  
720 **SDS-PAGE**

721 Two PSI-LHCI-LHCII bands and a PSI-LHCI band were separated by CN-PAGE,  
722 followed by sucrose density gradient centrifugation (Fig. 4), and were then subjected to  
723 the 2D-SDS-PAGE. Protein bands were visualized by silver-staining.

724

725 **Figure 6. A hypothetical model of evolutionary changes in the PSI peripheral**  
726 **antenna system of green plants.**

727 During evolution, green plants have diverged into two groups: streptophytes and  
728 chlorophytes. Land plants are thought to have diverged from one group of freshwater  
729 streptophytes, while core chlorophytes, including *C. reinhardtii*, diverged from seawater  
730 chlorophytes (prasinophytes). The structure of PSI and its peripheral antennas have been  
731 previously reported in vascular plants (Qin et al. 2015; Mazor et al. 2017; Pan et al.,  
732 2018), moss plants (Iwai et al., 2018; Pinnola et al., 2018), and core chlorophytes,  
733 including *C. reinhardtii* (Su et al., 2019; Suga et al., 2019) and *D. salina* (Perez-  
734 Boerema et al. 2020). Notably, the “minimal” PSI-LHCI in *D. salina* lacks several core  
735 subunits in addition to the second LHCI belt. The PSI structure of streptophyte algae  
736 and prasinophyte algae, however, has not been reported. Based on the data obtained in  
737 the present study, *M. viride* PSI possesses LHCA1, two LHCA2s, LHCA3, algae-type  
738 LHCA2, and LHCA9. Since *M. viride* LHCA1, two LHCA2s, and LHCA3 exhibit

739 considerable sequence similarity with LHCA proteins in vascular plants, their binding  
740 site may be similar to the binding site of the LHCIs in vascular plants. Based on  
741 sequence similarities, the binding site of algae-type LHCA2 and LHA9 may be similar  
742 to the binding site of PSI-LHCI in chlorophytes. The binding site of LHCP to PSII-  
743 LHCII is unclear. *M. viride* PSI can bind at least two LHCII trimers, however, the  
744 binding site of the additional LHCII trimer is unknown.

745

746 **Fig. S1 A multiple sequence alignment of LHC sequences.** Below is the multiple  
747 sequence alignment used to construct a phylogenetic tree in Fig. 2. A MAFFT program  
748 (<https://mafft.cbrc.jp/alignment/software/>) was used to align LHC sequences and a  
749 ClipKIT program was used to do trimming of the alignment.

750

751 **Figure S2. 2D-SDS-PAGE of the high-molecular-weight PSI-LHCI-LHCII, PSI-**  
752 **LHCI, and LHCII trimer.**

753 The high-molecular-weight PSI-LHCI-LHCII, PSI-LHCI, and LHCII trimer bands  
754 separated by CN-PAGE after sucrose density gradient centrifugation were subjected to  
755 2D-SDS-PAGE. Protein bands were visualized by silver-staining.

756

757    **Supplementary Table Legends:**

758    Table S1. List of *M. viride* LHC proteins predicted in the Iso-seq analysis.

759    Table S2. Proteins identified in the *M. viride* PSI-PSII band by MS.

760    Table S3. Proteins identified in the *M. viride* PSII-LHCII band by MS.

761    Table S4. Proteins identified in the *M. viride* PSI-LHCI band by MS.

762

Fig. 1

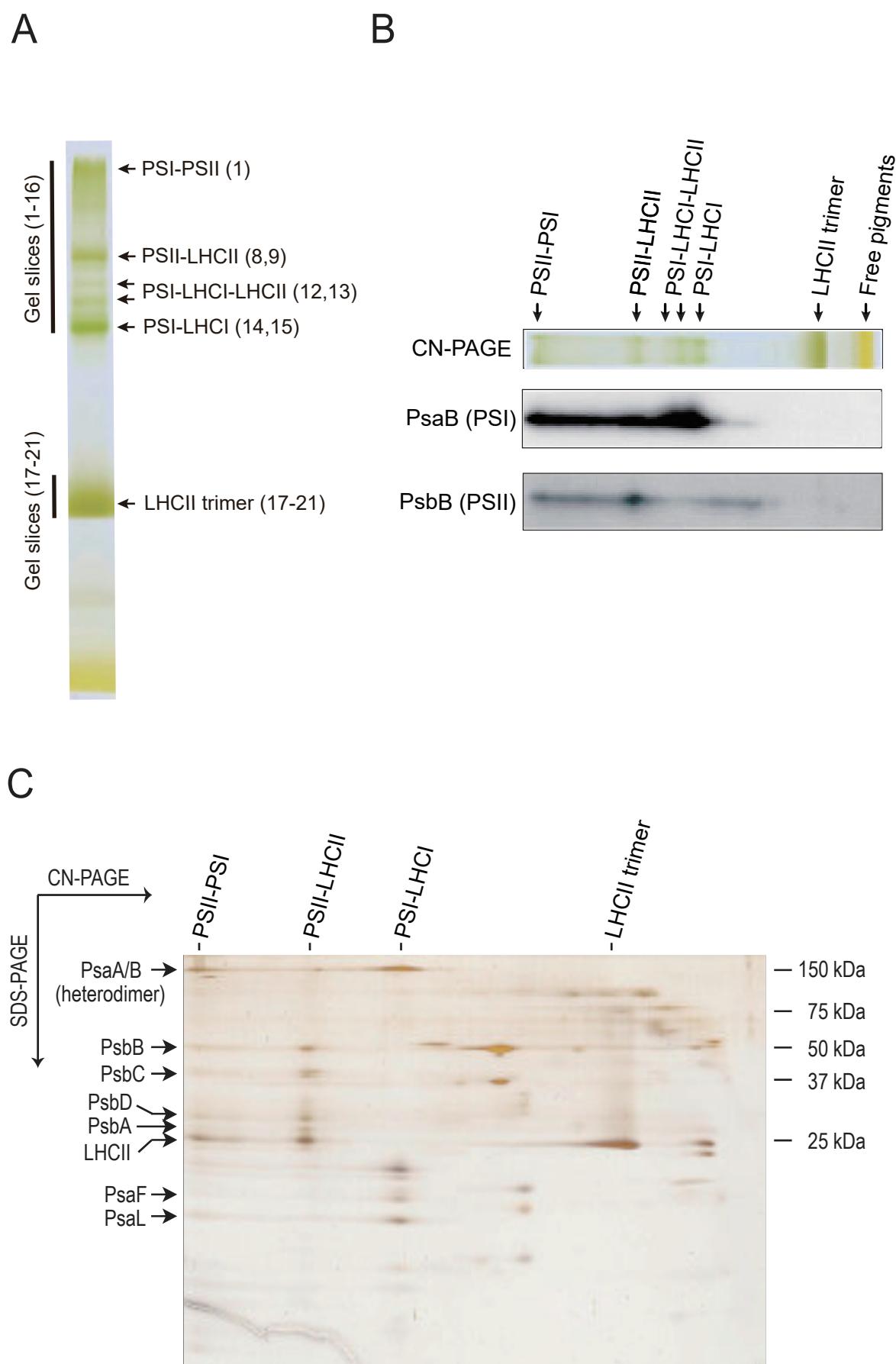


Fig. 2

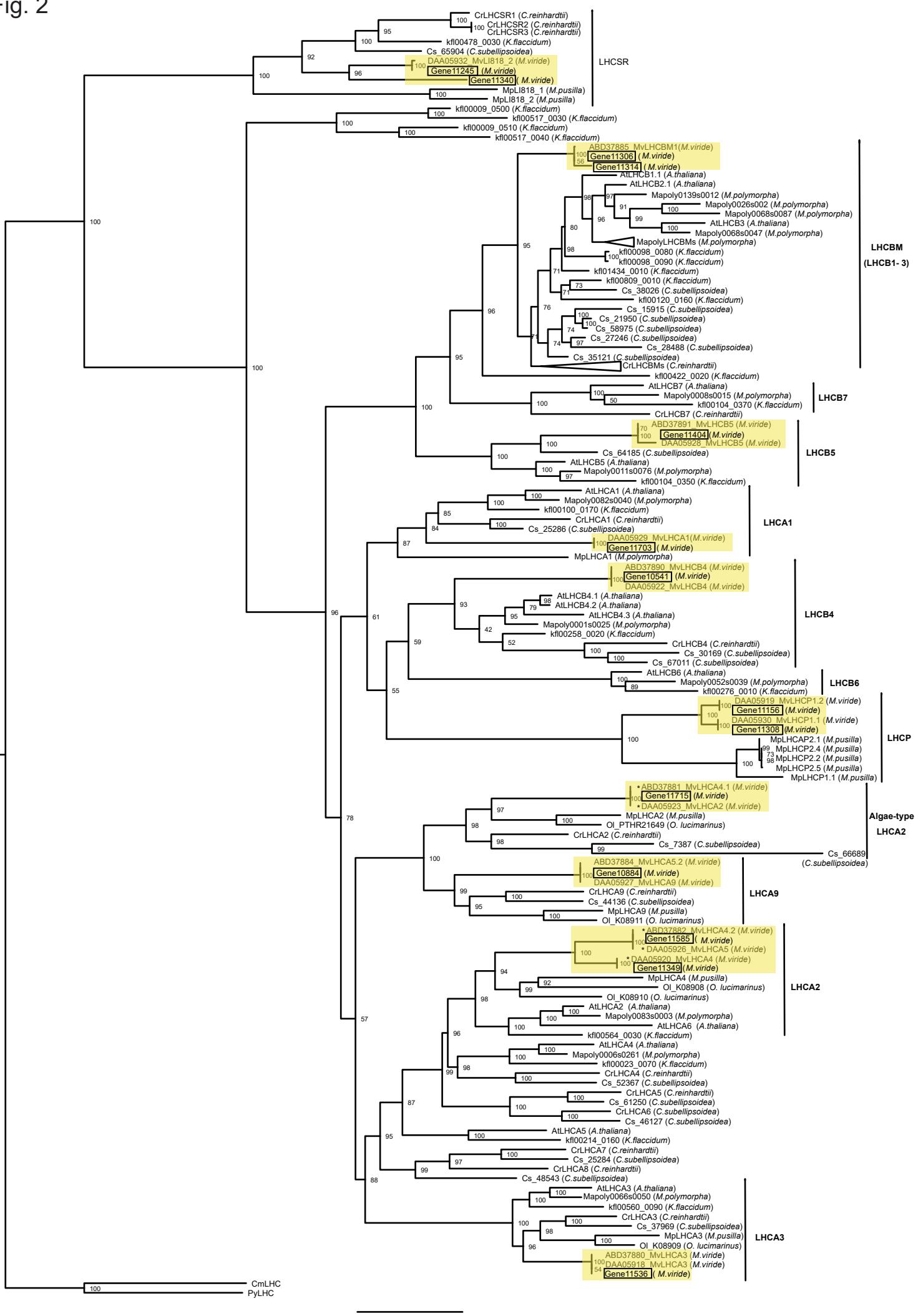
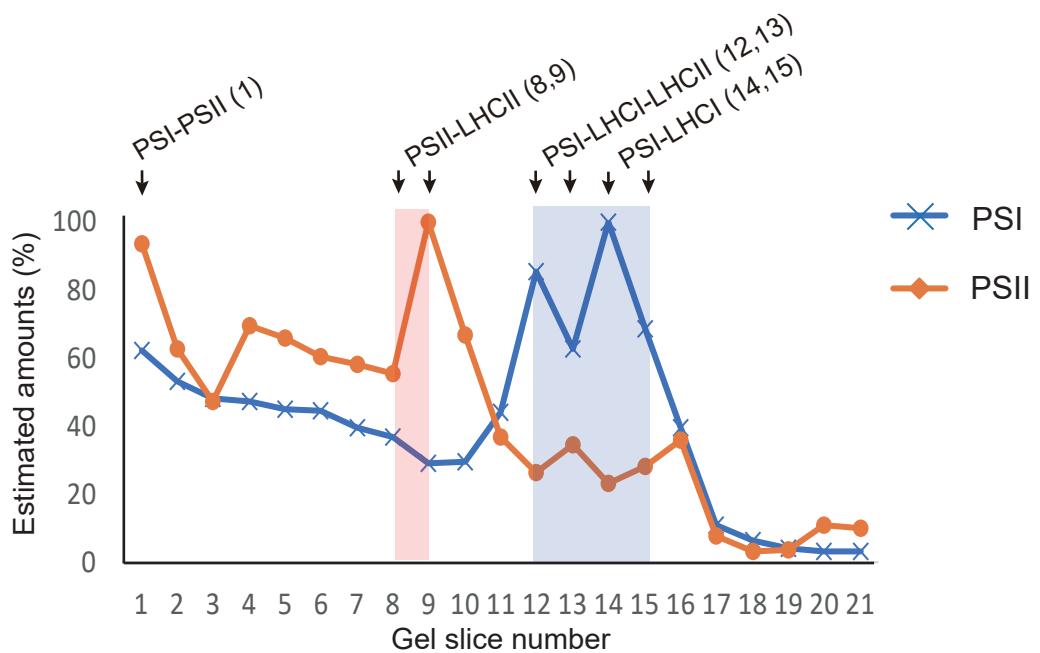
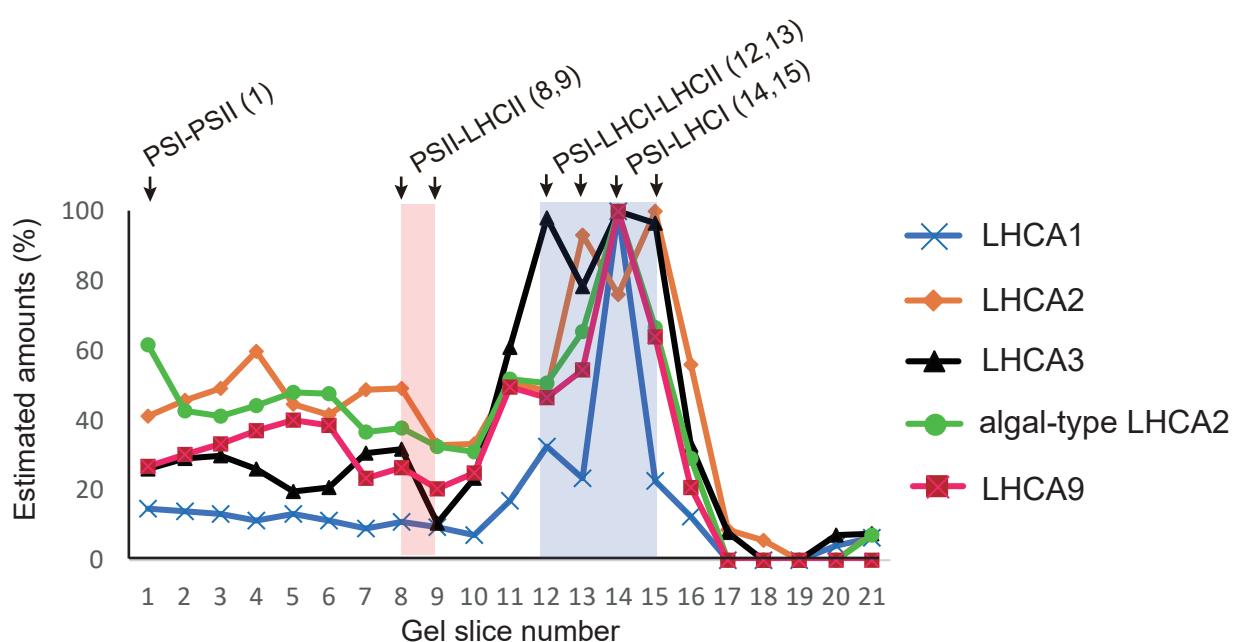


Fig. 3

A



B



C

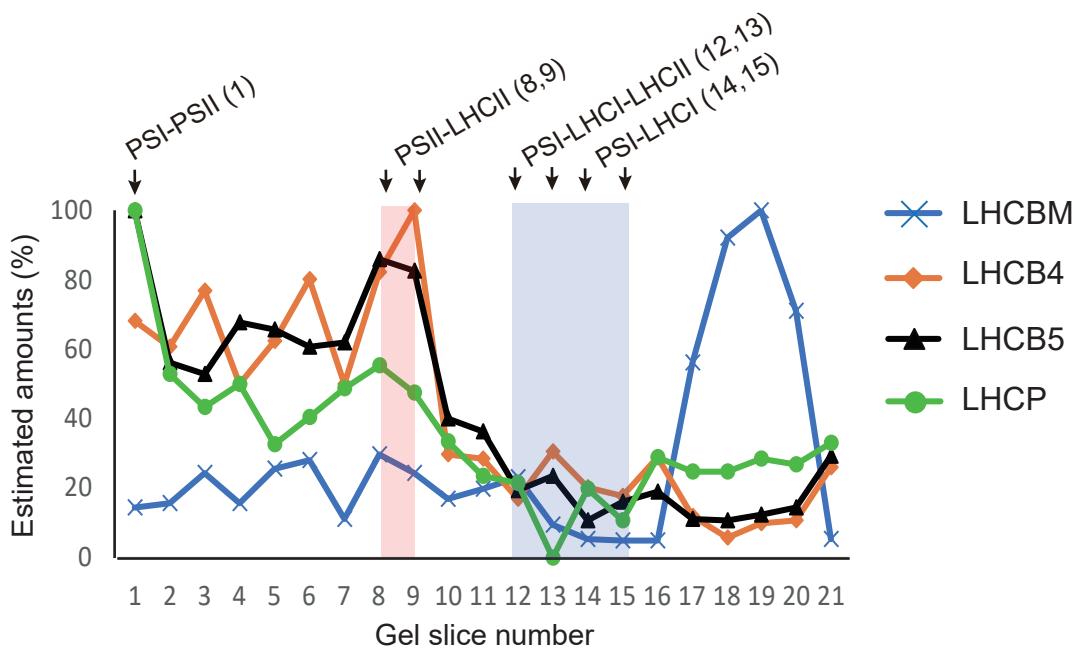


Fig. 4

A



- ← B3 (LHCII trimer and LHC monomer)
- ← B2 (PSI-LHCI(-LHCII))
- ← B1 (PSII-LHCII)

B

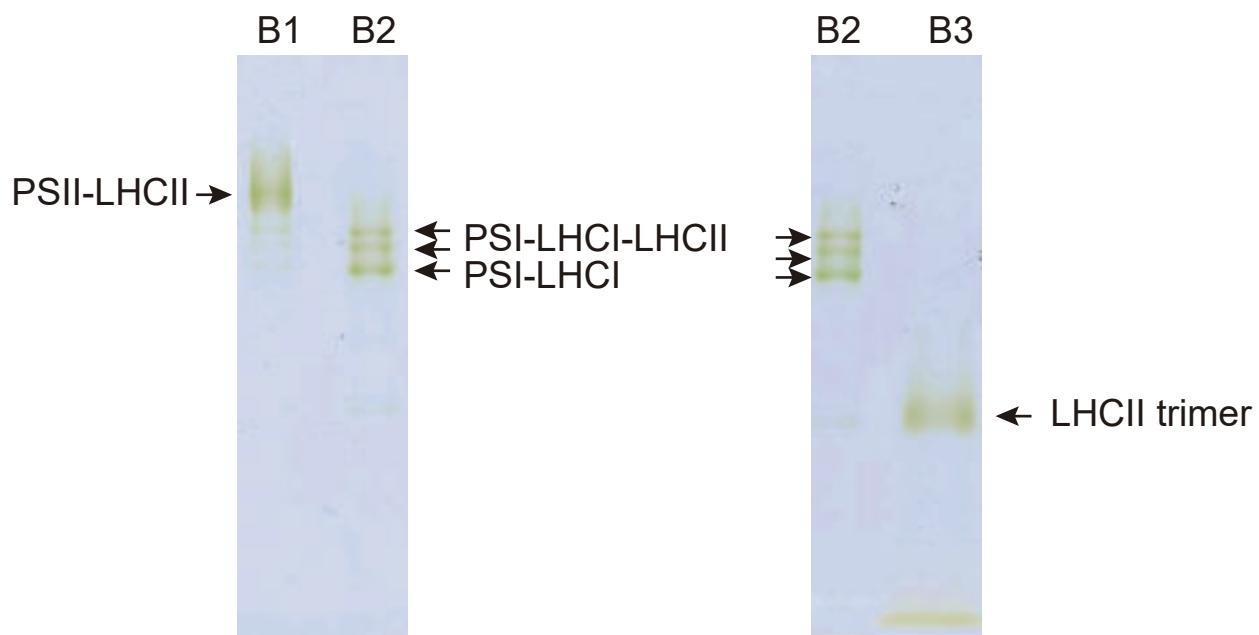


Fig. 5

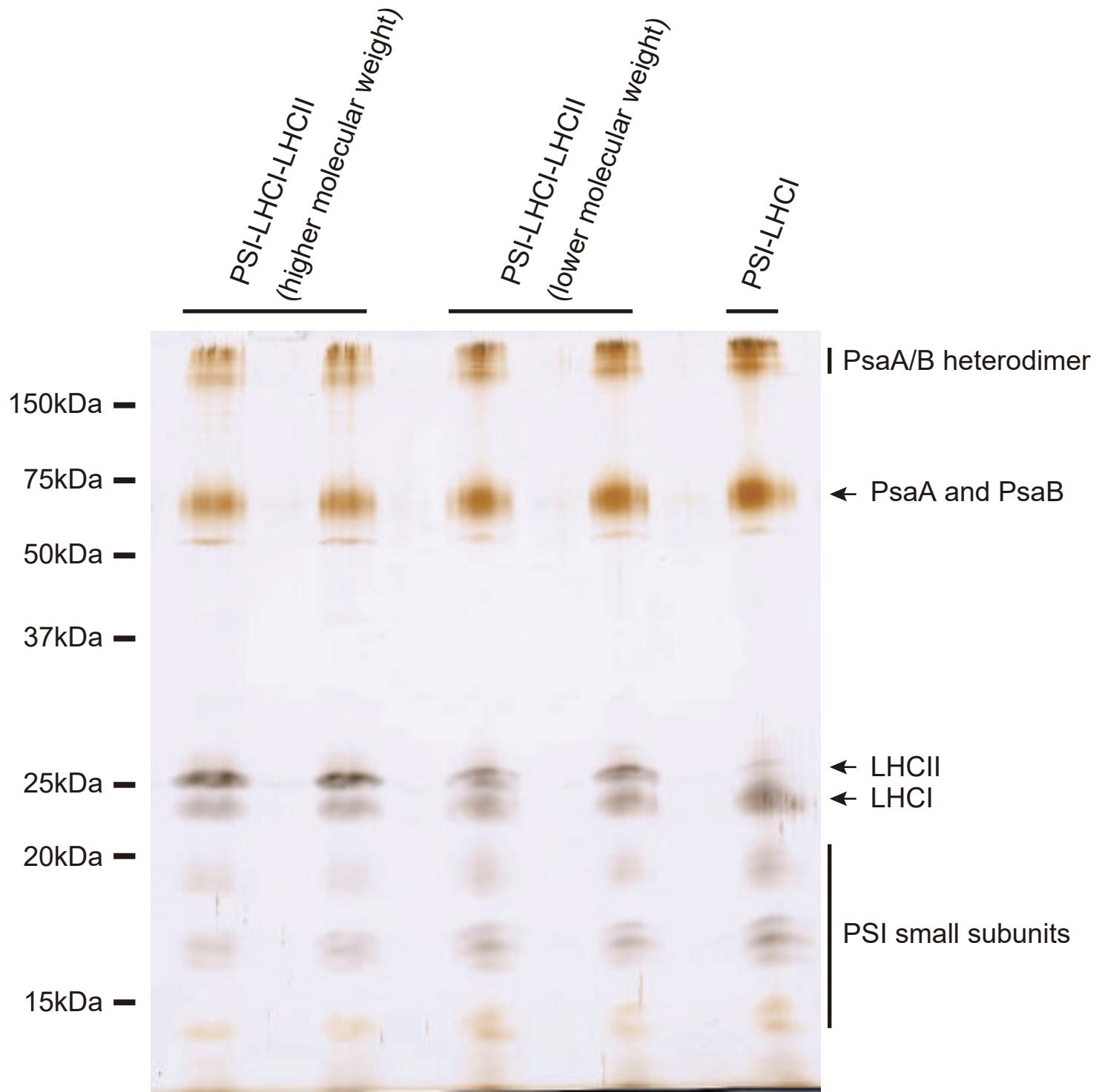
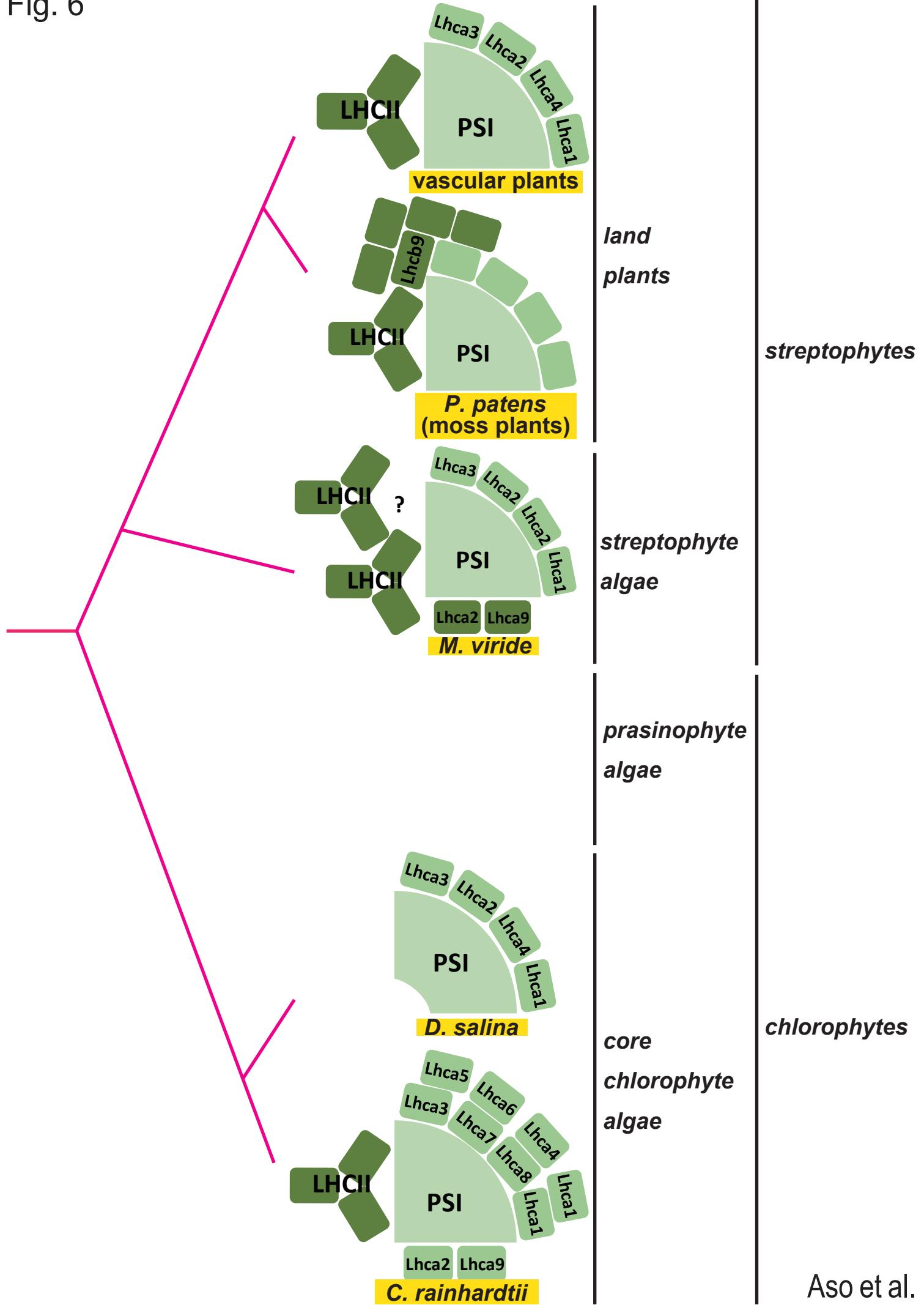
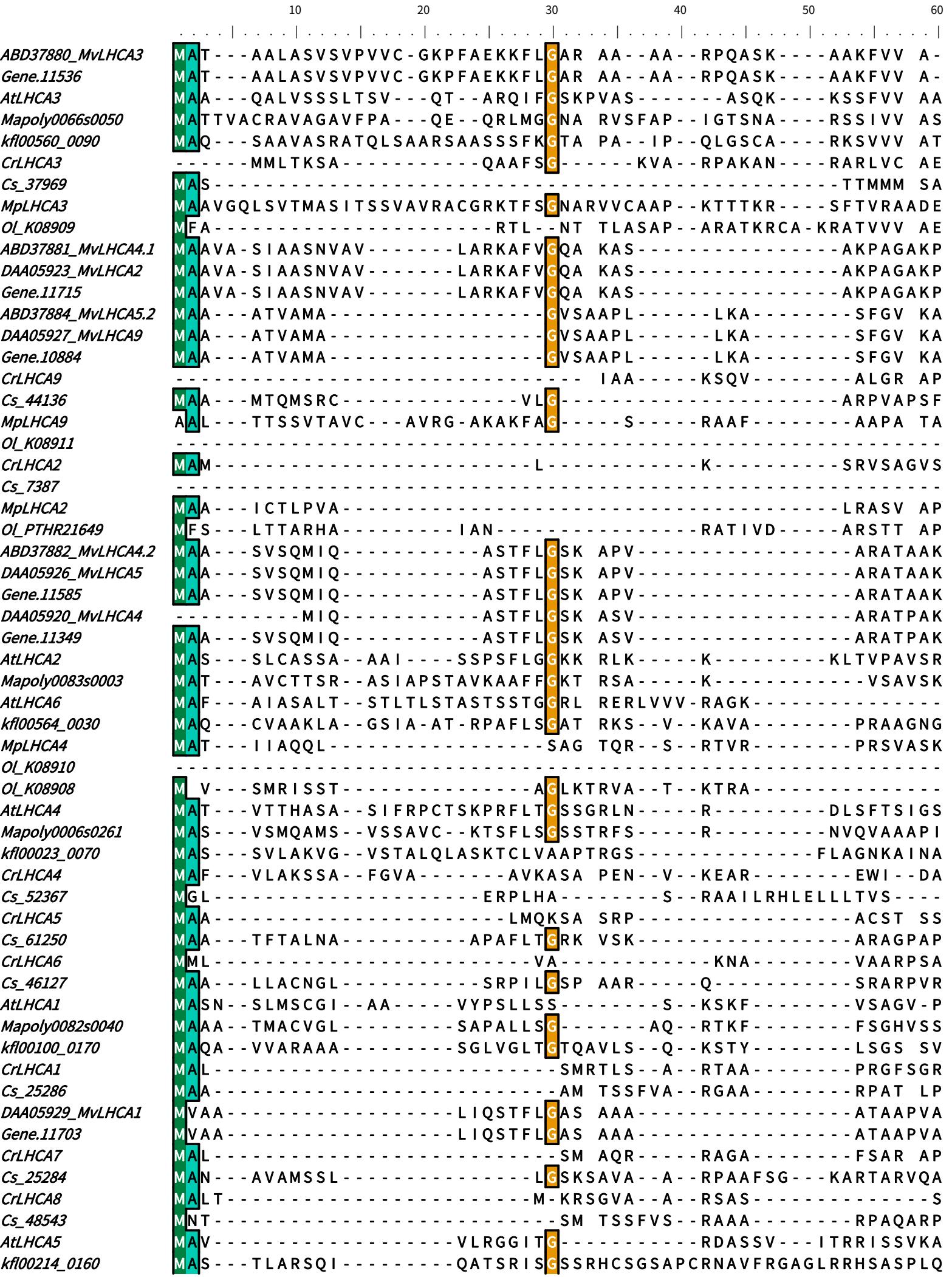


Fig. 6



**Fig. S1 A multiple sequence alignment of LHC sequences.** Below is the multiple sequence alignment used to construct a phylogenetic tree in Fig. 2. A MAFFT program (<https://mafft.cbrc.jp/alignment/software/>) was used to align LHC sequences and a ClipKIT program was used to do trimming of the alignment.

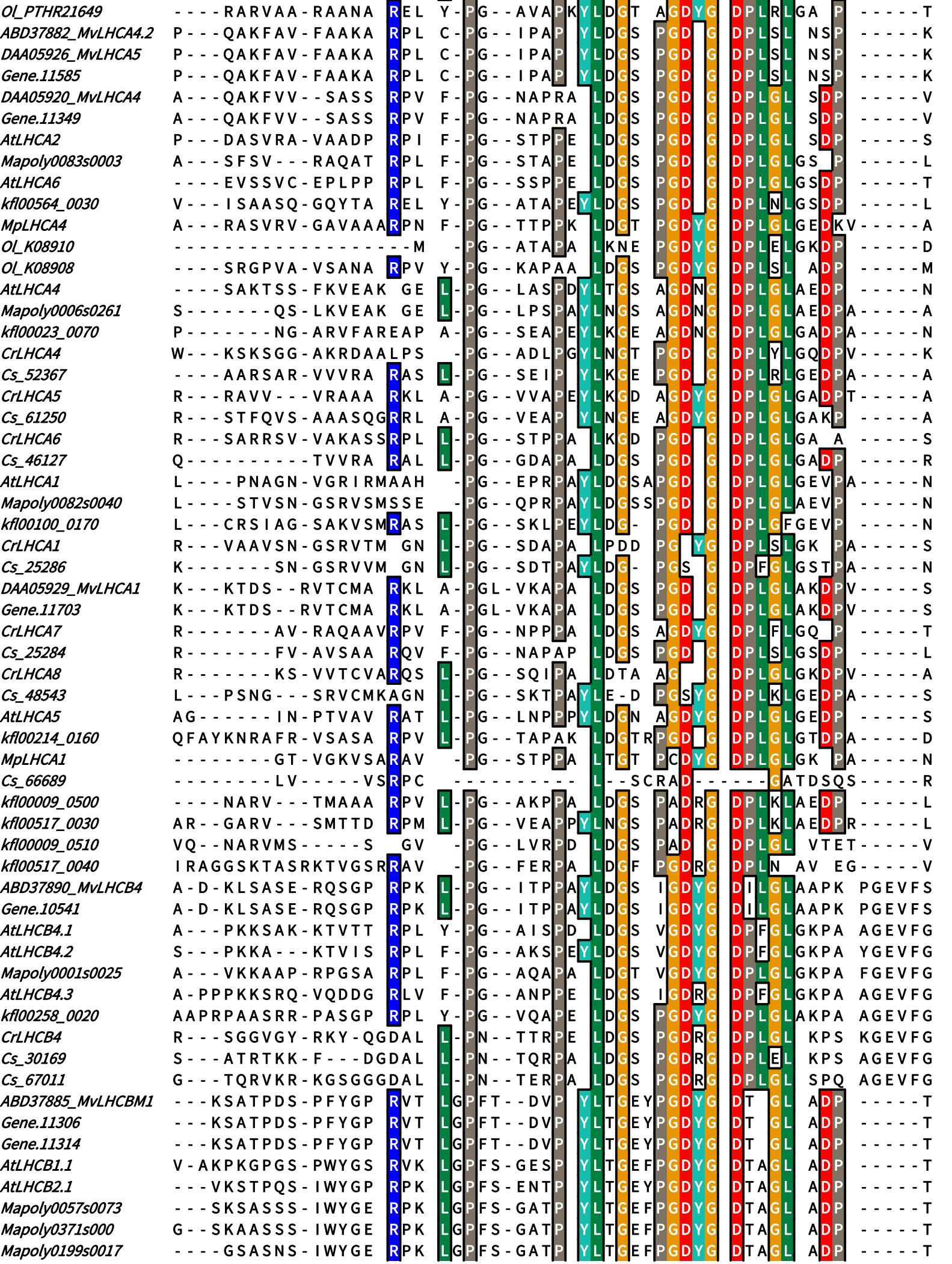


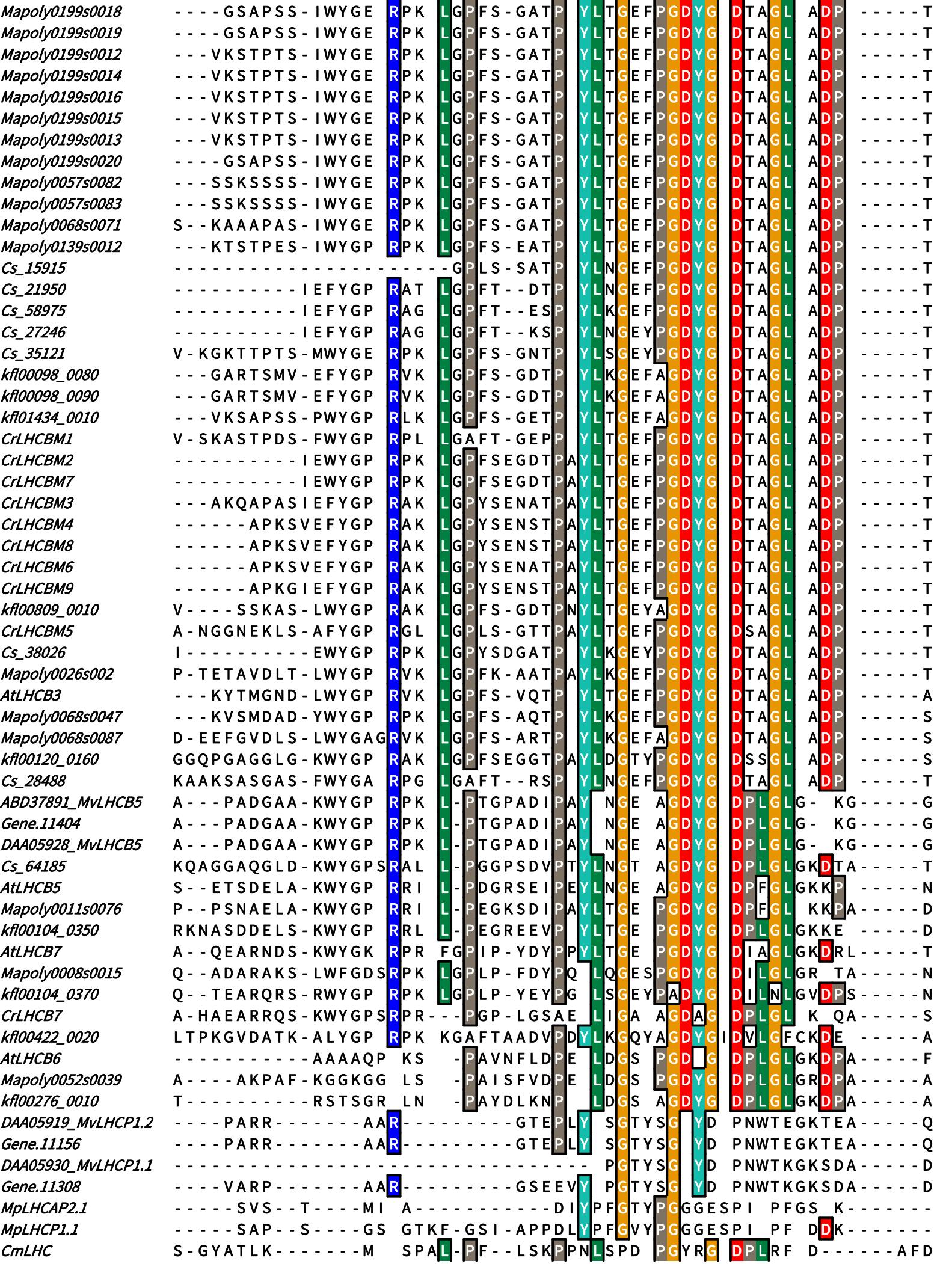
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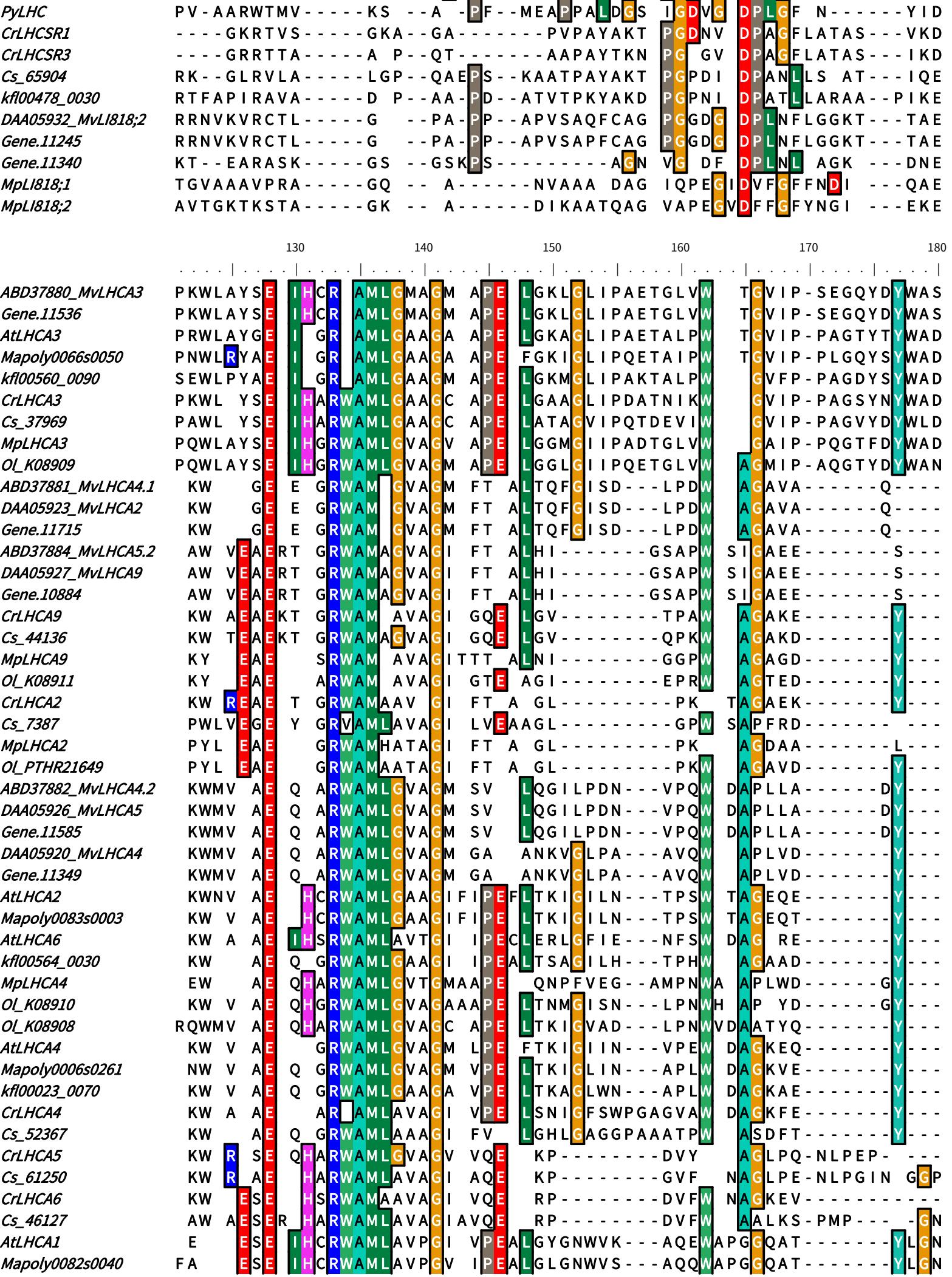
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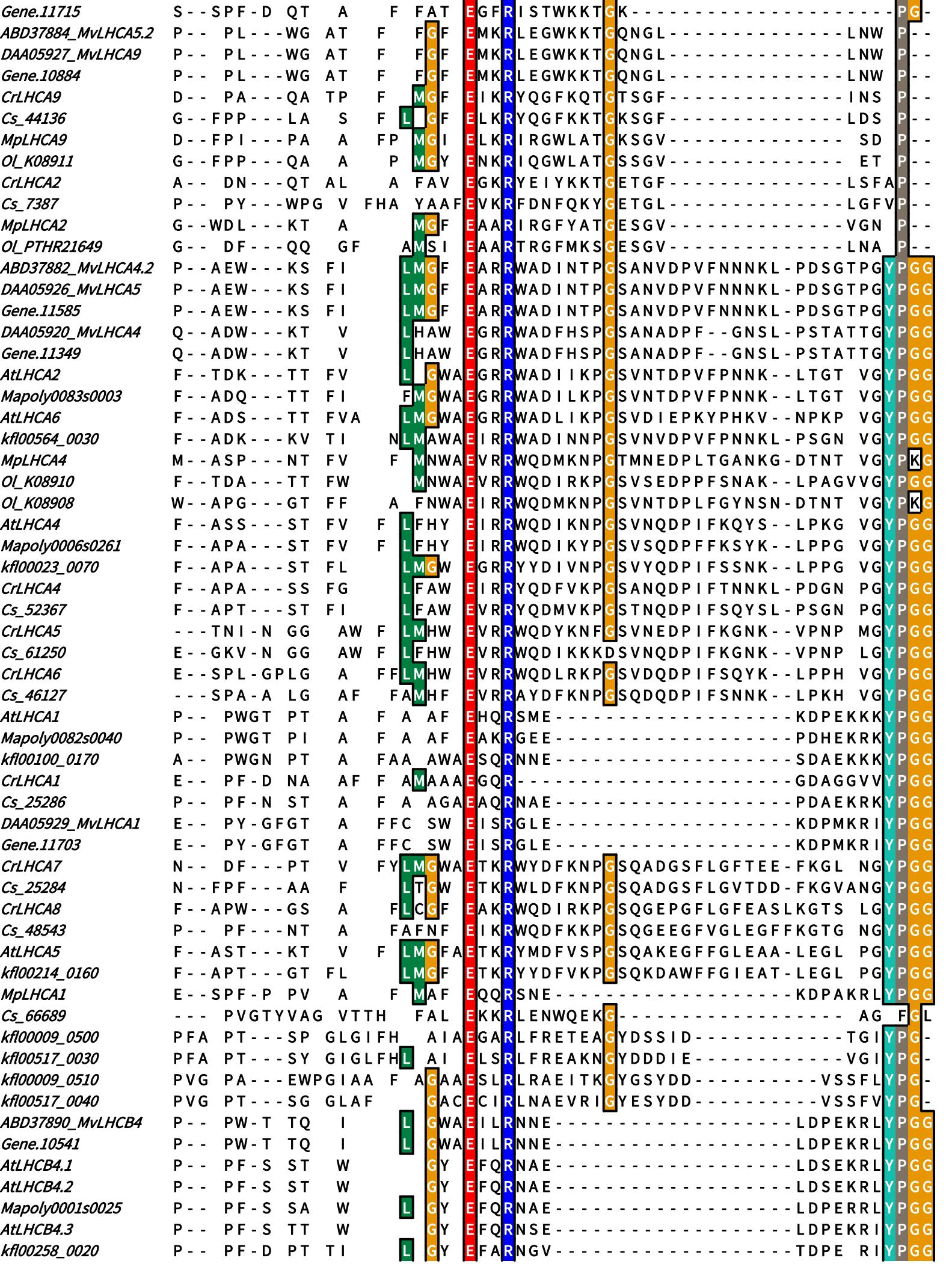




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 Mapoly0199s0019 FA NRELE I HARWAMLGA GC TPE LAKNG - VKFG - EAVW AG QIFS EGGLDYLG  
 Mapoly0199s0012 FA NRELE I HSRWAMLGA GC FPE LSKNG - VTFG - EAVW AG QIFAEGGLDYLG  
 Mapoly0199s0014 FA NRELE I HARWAMLGA GC FPE LSKNG - VSFG - EAVW AG QIFAEGGLDYLG  
 Mapoly0199s0016 FA NRELE I HARWAMLGA GC FPE LSKNG - VSFG - EAVW AG QIFAEGGLDYLG  
 Mapoly0199s0015 FA NRELE I HARWAMLGA GC FPE LSKNG - VSFG - EAVW AG QIFAEGGLDYLG  
 Mapoly0199s0013 FA NRELE I HARWAMLGA GC FPE LSKNG - VSFG - EAVW AG QIFAEGGLDYLG  
 Mapoly0199s0020 FA NRELE I HSRWAMLGA GC TPE LAKNG - VKFG - EAVW AG QIFAEGGLDYLG  
 Mapoly0057s0082 FA NRELE I HARWAMLGA GC TPE LAKNG - TKFG - EAVW AG QIFS EGGLDYLG  
 Mapoly0057s0083 FA NRELE I HARWAMLGA GC TPE LAKNG - TKFG - EAVW AG QIFS EGGLDYLG  
 Mapoly0068s0071 FA NRELE I HARWAMLGA GC FPE LAKNG - VKFG - EAVW AG QIFAEGGLDYLG  
 Mapoly0139s0012 FS NRELE I HSRWAMLGT GM FPE LAKNG - ITFG - EPIW AG QIFADGGLNLYLG  
 Cs\_15915 FA REIE I HARWAMLGA GC TPE LSQNG - FTFQ - EPVW AG QAQI LSSEGLDYLG  
 Cs\_21950 FA REIE I HARWAMLGA GC TPE LAKNG - VSFG - EAVW AG QAQIFGSDGLNLYLG  
 Cs\_58975 FA REIE I HARWAMLGA GC TPE LAKNG - VSFG - EAVW AG QAQIFGSDGLNLYLG  
 Cs\_27246 FA REIE I HARWAMLGA GC TPE LAKNG - VSFG - EAVW AG QIFAPGGLDYLG  
 Cs\_35121 FA REIE I HARWAMLGA GC TPE LAKNG - VSFG - EAVW AG QAQIFAPGGLDYLG  
 kfl00098\_0080 FA NRELE I HARWAMLGA GC FPE LAQNG - TKFG - EAVW AG QIFASGGLDYLG  
 kfl00098\_0090 FA NRELE I HARWAMLGA GC FPE LAQNG - TKFG - EAVW AG QIFASGGLDYLG  
 kfl01434\_0010 FA NRELE I HARWAMLGA GC TPE LAGNG - VKFG - ESVW AG QIFAPGGLDYLG  
 CrLHCBM1 FK RELE I HARWAMLGA GC FPE LGSYG - VPFG - EAVW AG QAQIFQEGGLDYLG  
 CrLHCBM2 FK RELE I HARWAMLGA GC TPE LAKNG - IPFG - EAVW AG QAQIFFAEGGLNLYLG  
 CrLHCBM7 FK RELE I HARWAMLGA GC TPE LAKSG - TQFG - EAVW AG QAQIFSEGGLDYLG  
 CrLHCBM3 FK RELE I HARWAMLGA GC TPE LAKSG - TQFG - EAVW AG QAQIFSEGGLDYLG

<i>CrLHCBM4</i>	FK	RELE	I	HARWAMLGA	GC	TPE	LAKNG	-TKFG-	EAVW	A	GAQIFSEGGLDYLGN			
<i>CrLHCBM8</i>	FK	RELE	I	HARWAMLGA	GC	TPE	LAKSG	-TKFG-	EAVW	A	GAQIFSEGGLDYLGN			
<i>CrLHCBM6</i>	FK	RELE	I	HARWAMLGA	GC	TPE	LAKSG	-TKFG-	EAVW	A	GAQIFSEGGLDYLGN			
<i>CrLHCBM9</i>	FK	RELE	I	HARWAMLGA	GI	TPE	LQKNG	-VQFG-	EAVW	A	GAQIFQEGGLNYLGN			
<i>kfl00809_0010</i>	FR	NRELE	I	HARWA <span style="background-color: yellow;">L</span> GA	GI	TPE	ALEKAG	-VKFG-	EAVW	A	GAQIFSADGLNYLGN			
<i>CrLHCBM5</i>	FK	RELE	I	HARWAMLGA	GC	TPE	LAKNG	-TPIV-	EPVW	A	GAQIFAEGGLDYLGN			
<i>Cs_38026</i>	FS	REIE	I	HSRW <span style="background-color: yellow;">A</span> LG	GI	TPE	LEKGG	-VKFQ-	EAVW	A	GAQIFSADGLNYLGN			
<i>Mapoly0026s002</i>	FA	NRELE	I	HSRWAMLGA	AGC	FPE	LAKNA	-VSFR-	EPVW	A	QIFSDGGLDYLGN			
<i>AtLHCB3</i>	FA	NRALE	I	HGRWAMLGA	AFGC	TPE	LQKVR	-VDFK-	EPVW	A	QIFSEGGLDYLGN			
<i>Mapoly0068s0047</i>	FA	NRALE	I	HGRWAMLGA	GC	LPEAL	VKS	-VTLK-	EAVW	A	QIFTDGGLDYLGN			
<i>Mapoly0068s0087</i>	FA	NRELE	I	HGRWAMLGA	WGACFFPE	LVKKS	CGLK	-EGVW		A	QIFTDGGLDYLGN			
<i>kfl00120_0160</i>	FR	NRELE	I	HSRWAMLGA	GM	LPE	LADS <span style="background-color: yellow;">G</span>	-IPIK-	EPVW	A	GAQIFDSDGLNYLGN			
<i>Cs_28488</i>	FS	REIE	I	HARWAMLGA	GI	VPE	LDQTN	-	-	DAGATIFGP <span style="background-color: yellow;">Q</span> YLG				
<i>ABD37891_MvLHCB5</i>	E	R PY E	I	HGRWAMLGV	GM	VPEGLYANGNTNIK	GAVW	D	GAVLLDPSTLT AGI					
<i>Gene.11404</i>	E	R PY E	I	HGRWAMLGV	GM	VPEGLYANGNTNIK	GAVW	D	GAVLLDPSTLT AGI					
<i>DAA05928_MvLHCB5</i>	E	R PY E	I	HGRWAMLGV	GM	VPEGLYANGNTNIK	GAVW	D	GAVLLDPSTLT AGI					
<i>Cs_64185</i>	E	RAYE	I	HARWAMLAAAGI	IPEGLQANG	-AAIK-	GGTW	TGAEMLN <span style="background-color: yellow;">G</span> GTLN	YF	AV				
<i>AtLHCB5</i>	FA	A FE	I	HARWAMLGAAGF	IPEALNKY	GANC	GP-EAVW	TGALLLDGNTLN	YFG	K				
<i>Mapoly0011s0076</i>	FD	AYE	I	HARWAMLGAAGF	IPEAFN	Y	GAVCGP-EAVW	TGALLLEGNTL	QYFG	GA				
<i>kfl00104_0350</i>	FD	R AAE	I	HARWAMLGAAGI	IPEAFNRSG	Y	GLPCGP-EAVW	TGAQLLLEGESLQ	C	CGI				
<i>AtLHCB7</i>	FD	FNFE	I	HARWAMLAA	GA	IPE	FDLTGTFHFA	-EPVW	RVGYSKLQGETL	Y	LG			
<i>Mapoly0008s0015</i>	FD	FNFE	I	HARWAMLGA	GA	IPE	LVRYGGLSFS	-EPVW	RVGYAKLQGETLD	Y	FGI			
<i>kfl00104_0370</i>	FQ	FNFE	I	HARWAMLGA	GA	IPE	LQYFSALDFT	-EPVW	VGYAKLQGEDLD	Y	FGI			
<i>CrLHCB7</i>	FA	EAE	I	HARWAMLGV	GC	VPE	LALRG	-VDLG-	EPVW	VGASKLNSDTLN	GGI			
<i>kfl00422_0020</i>	FA	LRAQE	F	ARWAMLGV	GM	YPEFFPAE <span style="background-color: yellow;">G</span>	-	-	FEVW	TGAQIFDPAGID	YLG	A		
<i>AtLHCB6</i>	KW	REAE	I	HGRWAMAAV	GIVFG	AWSG	-	-	VAW	A	GAQ-			
<i>Mapoly0052s0039</i>	KW	REAE	I	HGRWAMAAV	GIVFG	AWSG	-	-	IPW	A	GAD-			
<i>kfl00276_0010</i>	KW	REAE	I	HGRWA	AT	GC	IG	AFSG	-	A	GAA-			
<i>DAA05919_MvLHCP1.2</i>	AYNV	EV	I	HGRWAMLGCAGAWAA	E	QGTG	-	-	INW	A	GAICTPADGIH PGE			
<i>Gene.11156</i>	AYNV	EV	I	HGRWAMLGCAGAWAA	E	QGTG	-	-	INW	A	GAICTPADGIH PGE			
<i>DAA05930_MvLHCP1.1</i>	TYNV	SVE	I	HGRWAMLGCAGAWAA	E	QGTG	-	-	INW	A	GAICTPADGIH PGE			
<i>Gene.11308</i>	TYNV	SVE	I	HGRWAMLGCAGAWAA	E	QGTG	-	-	INW	A	GAICTPADGIH PGE			
<i>MpLHCAP2.1</i>	--	NAERE	I	HGRWAMLGV	TGAWAAA	ENG	TG	-	-	IPW	TAGTLCTPDDAVK	PGA		
<i>MpLHCAP1.1</i>	--	NAERE	I	HGRWAMLGV	TGAWAAA	ENG	TG	-	-	IPW	TAGTLCTPDDAVK	PGA		
<i>CmLHC</i>	NWL	E GE	K	GRVAMLAC	HFFVTE	E	FYQF	-	-	PF	AGAPKLA	-	AHDYFVK	
<i>PyLHC</i>	KFL	REAE	K	HCRVTMLAV	GLFVQE	E	FYTL	-	-	PF	SGGPALA	-	SHNYFVT	
<i>CrLHCSR1</i>	R	RESE	T	HGRVAMLAA	GF	VGEQLQD	-	-	FPL	FDGRVS	-	AIY FQQ		
<i>CrLHCSR3</i>	R	RESE	T	HGRVAMLAA	GF	VGEQLQD	-	-	FPL	WDGRVS	-	AIY FQQ		
<i>Cs_65904</i>	K	RESE	T	HGRVAMLAA	GW	VGEF	AD	-	-	KKLLSD <span style="background-color: yellow;">G</span> RT	-	AID FQQ		
<i>kfl00478_0030</i>	K	RESE	K	HGRVAMLAT	GF	VGEQ	ED	-	-	FPARFFPHVT	-	AIY FQQ		
<i>DAA05932_MvLI818;2</i>	KML	RESE	K	HGRVAMLAT	GF	VGETFNP	-	-	L	--FGG	IT	-AINQFQQ		
<i>Gene.11245</i>	KML	RESE	K	HGRVAMLAT	GF	VGE	TFNP	-	-	L	--FGG	IT	-AINQFQQ	
<i>Gene.11340</i>	LL	REAE	T	HGRVMLATAGF	AE	FNPL	-	-	-	FNG	IK	-AIN FQQ		
<i>MpLI818;1</i>	AQ	A VE	T	HGRVLAMLAS	GF	VGE	S	EG	-	SS	LFDSQVT	-	AIN FQQ	
<i>MpLI818;2</i>	AQ	A V	E	T HGRV <span style="background-color: yellow;">I</span> AMLAS	GF	VGE	Q	EG	-	-	SA	LFDANIT	-	AID FQQ

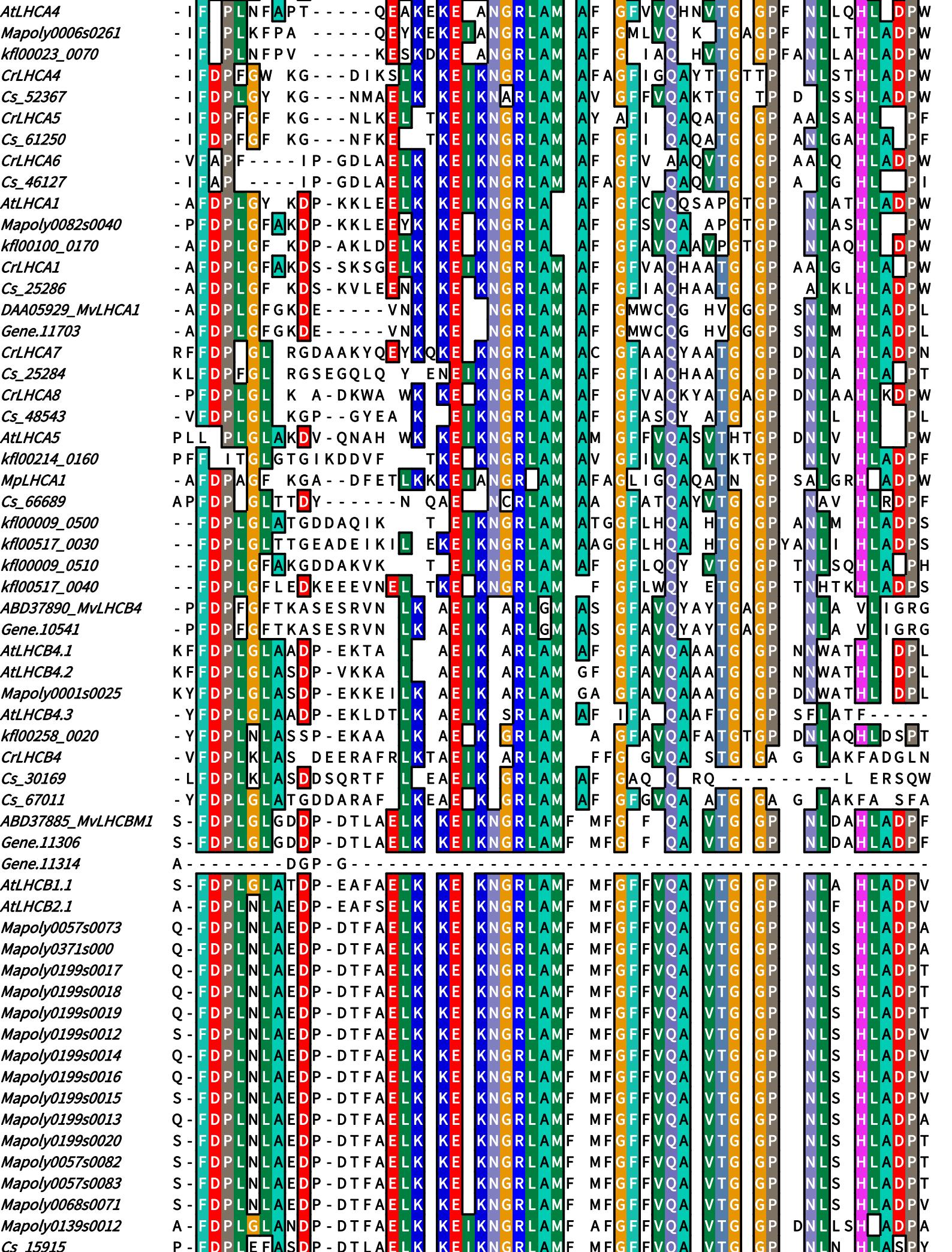
		190	200	210	220	230	240
<i>ABD37880_MvLHCA3</i>	P	-	LA FW	LMQFAE	LRRWQDYRHPG	SQS <span style="background-color: yellow;">K</span> QYFLGLEQFFGGSG	PS Y PGG
<i>Gene.11536</i>	P	-	LA FW	LMQFAE	LRRWQDYRHPG	SQS <span style="background-color: yellow;">K</span> QYFLGLEQFFGGSG	PS Y PGG
<i>AtLHCA3</i>	N	-	YT FV	ALMGFAE	HRR <span style="background-color: yellow;">L</span> QDWNP	SMG <span style="background-color: yellow;">K</span> QYFLGLEKGLAGSG	PA Y PGG
<i>Mapoly0066s0050</i>	P	-	YT FV	ALMGFAE	HRR <span style="background-color: yellow;">R</span> AQDYYKP	SMG <span style="background-color: yellow;">K</span> QYFLGFEKVLGGSG	PA Y PGG
<i>kfl00560_0090</i>	S	-	YT FG	LMAFAE	HKR <span style="background-color: yellow;">L</span> ADYRKPG	SQGKVFFLGMEKFLGGSG	PA Y PGG
<i>CrLHCA3</i>	P	-	YT FF	AMQFAE	LRR <span style="background-color: yellow;">L</span> QDFRP	SMG <span style="background-color: yellow;">Q</span> QYFLGLEAIFKGSG	AA Y PGG
<i>Cs_37969</i>	P	-	YS FF	L QFAE	LR <span style="background-color: yellow;">R</span> WQDFRNPG	SQG <span style="background-color: yellow;">K</span> QYFLGLEEVLKGS	PS Y PGG
<i>MpLHCA3</i>	P	-	TT FW	N LMNFAE	VKR <span style="background-color: yellow;">R</span> QDYWNPG	SQGETPLMGWEKGFA	GSAPA Y PGG
<i>OI_K08909</i>	P	-	FT FW	NAALMNFAE	L RRAQDYWNPG	SMG <span style="background-color: yellow;">K</span> QELIGWEKMLGGSG	PA Y PGG
<i>ABD37881_MvLHCA4.1</i>	S	--SPF-D	QT A	F FAT	EGFRISTWKKT <span style="background-color: yellow;">GK</span>	-	PG
<i>DAA05923_MvLHCA2</i>	S	--SPF-D	QT A	F FAT	EGFRISTWKKT <span style="background-color: yellow;">GK</span>	-	PG

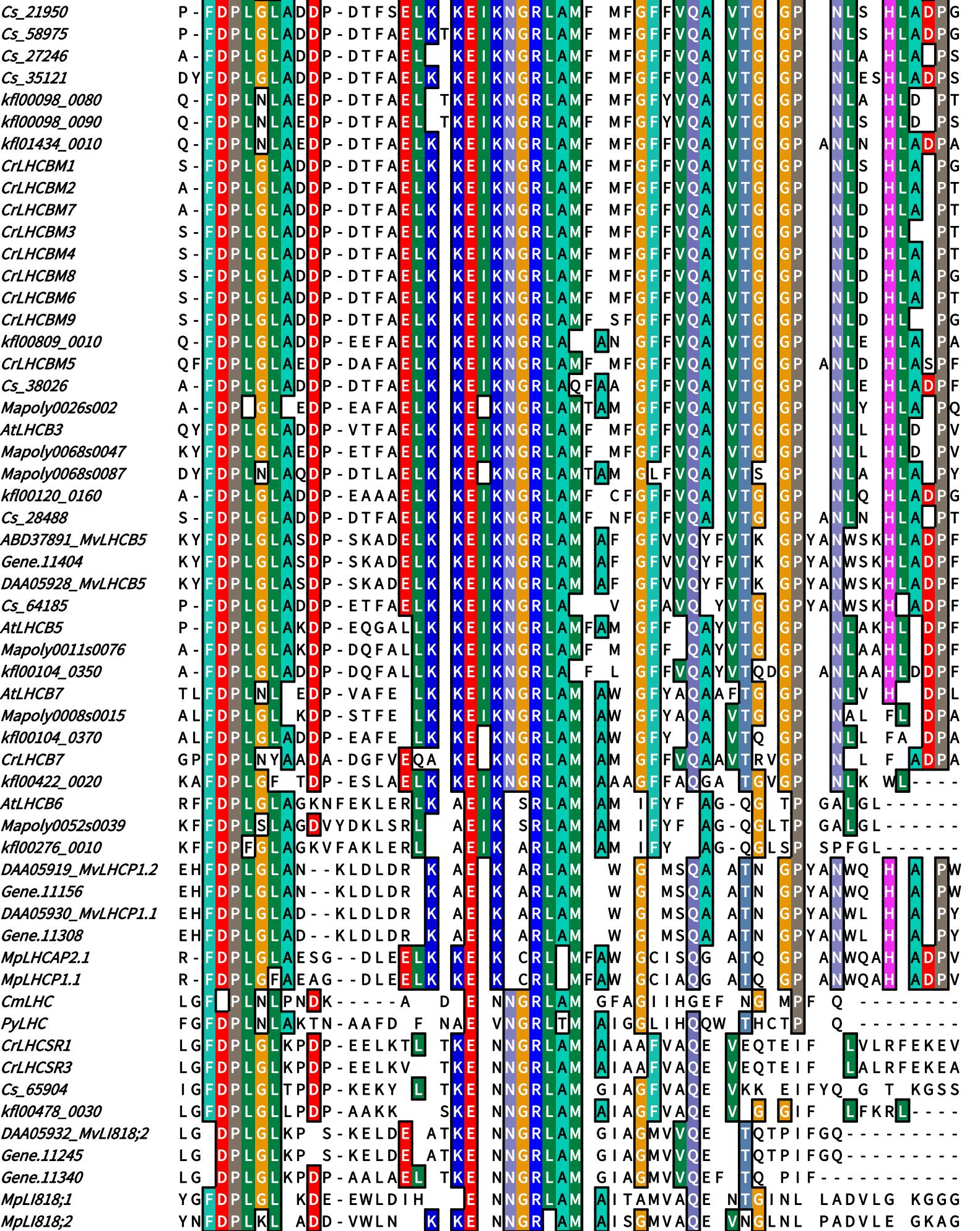


<i>CrLHCB4</i>	S -	P F - S	T Q	W	L	G G A E	F Y R N S E	-	T N P E K R C Y P G G
<i>Cs_30169</i>	S -	P F - S	S Q	W	L	G G A E	I Y R N R S	-	T D L Q R I Y P G G
<i>Cs_67011</i>	S -	P F - S	S Q	W	A	L G G A E	I Y R N R E	-	L E P Q A R I Y P G G
<i>ABD37885_MvLHCBM1</i>	PSL	H A Q S	I A T	F	A	L M G L	E G Y R V G G	G P L G	-
<i>Gene.11306</i>	PSL	H A Q S	I A T	F	A	L M G L	E G Y R V G G	G P L G	-
<i>Gene.11314</i>	P -	-	-	-	-	P G A R	-	P V H	-
<i>AtLHCB1.1</i>	PSL	H A Q S	L A	W A T	L M G A	E G Y R V A G N	G P L G	-	E A E L L Y P G G
<i>AtLHCB2.1</i>	PNL	H A Q S	L A	W A	L M G F	E G Y R I G G	G P L G	-	E G L P L Y P G G
<i>Mapoly0057s0073</i>	S G L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	E V S P I Y P G G
<i>Mapoly0371s000</i>	S G L	H A Q S	L A	W A C	L M G A	E G Y R V S G	G P L G	-	E V S P I Y P G G
<i>Mapoly0199s0017</i>	S G L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	E V S P I Y P G G
<i>Mapoly0199s0018</i>	S G L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	E V S P I Y P G G
<i>Mapoly0199s0019</i>	S G L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	E V S P I Y P G G
<i>Mapoly0199s0012</i>	S S L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	D V V P I Y P G G
<i>Mapoly0199s0014</i>	S S L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	E V S P I Y P G G
<i>Mapoly0199s0016</i>	S S L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	E V S P I Y P G G
<i>Mapoly0199s0015</i>	S S L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	D V V P I Y P G G
<i>Mapoly0199s0013</i>	S S L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	E V S P I Y P G G
<i>Mapoly0199s0020</i>	S S L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	D V V P I Y P G G
<i>Mapoly0057s0082</i>	S S L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	D V V P I Y P G G
<i>Mapoly0057s0083</i>	S S L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	D V V P I Y P G G
<i>Mapoly0068s0071</i>	S S L	H A Q S	L A	W A C	L M G A	E G Y R V A G	G P L G	-	D V V P I Y P G G
<i>Mapoly0139s0012</i>	E N L	H A Q S	L A	G C	L M G L	E G Y R V G G	G P L G	-	A D L P I Y P G G
<i>Cs_15915</i>	P N L	H A Q S	V A T	A	L M G S A E	A A Y R A A G S	- A P G V	-	D G L K L Y P G G
<i>Cs_21950</i>	S S L	H A Q S	I A T	A C	L M G G A E	A A Y R A A G E	- G P G L	-	E G L S L Y P G G
<i>Cs_58975</i>	P S L	H A Q S	I A T	A C	L M G G A E	A A Y R A N G E	- G P G V	-	E G L S L Y P G G
<i>Cs_27246</i>	P S L	H A Q S	I A	F S	L M G L	E G Y R V N G	- G P A G	-	E G L A L Y P G E
<i>Cs_35121</i>	P S L	H A Q S	I A	A S	L M G A	E G Y R V Y G	- G P G G	-	E G L K V Y P G G
<i>kfl00098_0080</i>	P S L	H A Q S	L A	A S	L M G A	E A Y R V N G	- G P L G	-	E V E P L Y P G G
<i>kfl00098_0090</i>	P S L	H A Q S	L A	A S	L M G A	E A Y R V N G	- G P L G	-	E V E P L Y P G G
<i>kfl01434_0010</i>	P S L	H A Q S	L A	G S	L M G A	E G Y R V N G	- G P L G	-	E I E P L Y P G G
<i>CrLHCBM1</i>	P N L	H A Q S	L A	G T	L M G A	E G Y R V N G	- G P L G	-	E G L K L Y P G G
<i>CrLHCBM2</i>	E N L	H A Q S	I A T	A F	M G L A E	A A Y R A N G	- G P L G	-	E G L P L Y P G G
<i>CrLHCBM7</i>	E N L	H A Q S	I A T	A F	M G L A E	A A Y R A N G	- G P L G	-	E G L P L Y P G G
<i>CrLHCBM3</i>	P S L	H A Q N	V A T	A	L M G L	E G Y R V N G	- G P A G	-	E G L P L Y P G E
<i>CrLHCBM4</i>	P S L	H A Q N	V A T	A	L M G L	E G Y R V N G	- G P A G	-	E G L P L Y P G E
<i>CrLHCBM8</i>	P S L	H A Q N	V A T	A	L M G L	E G Y R V N G	- G P A G	-	E G L P L Y P G E
<i>CrLHCBM6</i>	P S L	H A Q N	V A T	A	L M G L	E G Y R V N G	- G P A G	-	E G L P L Y P G E
<i>CrLHCBM9</i>	P S L	H A Q N	V A T	A	L G L	E G Y R V N G	- G P A G	-	E G L P L Y P G E
<i>kfl00809_0010</i>	P G L	H A Q S	L A T	A T	L G A	E G Y R V N G	- G P L G	-	E V E P L Y P G G
<i>CrLHCBM5</i>	P G L	H A Q S	L A T	A	L M G A	E G Y R V N G	- G P A G	-	E G L K L Y P G G
<i>Cs_38026</i>	P S L	H A Q S	I A T	A T	L G L A E	E G Y R V N G	- G P A G	-	E I T P L Y P G G
<i>Mapoly0026s002</i>	P N L	H A Q S	L A	W A S	L M G L	E G Y R I G G	- G P L G	-	D A G G L Y P G G
<i>AtLHCB3</i>	P N L	H A Q S	L A	G F	L M G L	E G F R I N G L	- D G V G	-	E G D L Y P G G
<i>Mapoly0068s0047</i>	P N L	H A Q S	L A	W A	L M G A	E G Y R Q N G L	- P G I G	-	E G G E L Y P G G
<i>Mapoly0068s0087</i>	P N L	H A Q S	L A	W A C	L M G L	E G Y R S G G	- G P L G	-	K V T P L Y P G G
<i>kfl00120_0160</i>	S S L	H A Q S	L A T	A C	L M G A	E G Y R V N G L	- E G F Q	-	E R D P L Y P G G
<i>Cs_28488</i>	P G L	N A K N	I A T	A	L M G A	E G Y R V N G	- G P A G	-	E G L K V Y P G E
<i>ABD37891_MvLHCB5</i>	P -	N P V - P	A L A	I F	A F F L	E N Y R Y Q Q D	- G P W G	-	T G L P L Y P G G
<i>Gene.11404</i>	P -	N P V - P	A L A	I F	A F F L	E N Y R Y Q Q D	- G P W G	-	T G L P L Y P G G
<i>DAA05928_MvLHCB5</i>	P -	N P V - P	A L A	I F	A F F L	E N Y R Y Q Q D	- G P W G	-	T G L P L Y P G G
<i>Cs_64185</i>	P W G N N P L P	F L	V	A L	G A	E R Y R Q S G E	- G P P G Y S P V G K F D S S I F S G L	N L Y P G G	-
<i>AtLHCB5</i>	N -	P I - N	V L A	V A	L G G A E	E Y Y R I T N G L	-	D F E K L Y P G G	
<i>Mapoly0011s0076</i>	S -	P V - N	A A A	I A	L G G A E	E Y Y R S T N K	- S P L G	-	S D L R L Y P G G
<i>kfl00104_0350</i>	T -	P L - N	A A A T I A	I A	L G G A E	E Y Y R S A N K	- S P L G	-	S D L P L Y P G G
<i>AtLHCB7</i>	P G L A G S Q G	I V	A I C	A	L M V G P E	E Y A R Y C G I E A L E P L G I Y L	-	P G I N Y P G G	
<i>Mapoly0008s0015</i>	P G L A G A Q G	L V	A F C	A	L M V G P E	E Y A R Y C G I E A L E P L G V F L	-	P G I N Y P G G	
<i>kfl00104_0370</i>	P G L A G G Q G	L I	A F C	A	L M L G P E	E Y A R A T G I A A L E P V G L Y L	-	P G E N P G G	
<i>CrLHCB7</i>	E G F A G K Q G	G L	A A C	A	L M G G P E	E Y A R Y V G I R S L E P V G V F L	-	P G Q N Y P G G	

<i>kf00422_0020</i>	PNV	NAHS	IA	AAF	A	LMGGAE	FARL	KAPK	- - - - -	EM	GLY	PGG	
<i>AtLHCB6</i>	PDA	APFSFGS	GT			LMGW	E	SKRWVDFFNPDSQSVEWATPWSKT	FANTG	QGY	PGG		
<i>Mapoly0052s0039</i>	PSA	APFSFGT	GT			LMGW	E	GKRWADYVNPNSQLVDWATPWSRT	FGNTGLQGY	PGG			
<i>kf00276_0010</i>	PGA	APFTFGT	GT			LMSW	E	GKRWVDFYNPSSQSVEWATPWSKT	FANTGQQGY	PGG			
<i>DAA05919_MvLHCP1.2</i>	PAGSGFPNFYIQ	A	ST	AMGLAE	GYRGG	GLIDSC	- - - - -	- - - - -	FPETVGDL	PGG			
<i>Gene.11156</i>	PAGSGFPNFYIQ	A	ST	AMGLAE	GYRGG	GLIDSC	- - - - -	- - - - -	FPETVGDL	PGG			
<i>DAA05930_MvLHCP1.1</i>	PEGSGFPNFYIQ	G	S	MGLAE	GYRGG	GLIDNV	- - - - -	- - - - -	FPEEVGDL	PGG			
<i>Gene.11308</i>	PEGSGFPNFYIQ	G	S	MGLAE	GYRGG	GLIDNV	- - - - -	- - - - -	FPEEVGDL	PGG			
<i>MpLHCAP2.1</i>	PEGSGYPSFWA	A		L	GLAE	AYRTGLTDPA	- - - - -	- - - - -	FDETVGDVS	PGG			
<i>MpLHCP1.1</i>	PEGSGYPSFWA	A	FL	GSAE	CYRTGLFENP	- - - - -	- - - - -	- - - - -	FPEEL-SVT	PGG			
<i>CmLHC</i>	--	SGAMQ	LAF	GFLE	FL	HRG	VLYSDMEW	- - - - -	- - - - -	KGRKP	PGE		
<i>PyLHC</i>	--	QGAMQ	LLW	CGLE		GVPAVL	M-MQG	- - - - -	- - - - -	SGRRP	PGE		
<i>CrLHCSR1</i>	--	GQG	--	FWEPL	A	GVAE	SYRVAVGWTPTGTGFN	- - - - -	- - - - -	SLK	DYE	PGD	
<i>CrLHCSR3</i>	--	GQG	--	FWEPL	A	GVAE	SYRVAVGWTPTGTGFN	- - - - -	- - - - -	SLK	DYE	PGD	
<i>Cs_65904</i>	--	EYK	GA	I	FWEPL	FS	GLAE	AWRIGVGWNPSSDKFN	- - - - -	QLR	DYSPGE		
<i>kf00478_0030</i>	--	ENE	GA	I	FWEPL	FA	ALA	ESYRVGLGWTPDSTTNFN	- - - - -	TLR	DYE	PGN	
<i>DAA05932_MvLI1818;2</i>	--	PQP	--	FWE	V	A	GLAE	GFRINRGWSPAEEAFFSI	- - - - -	GVLK	SYTPGT		
<i>Gene.11245</i>	--	PQP	--	FWE	V	A	GLAE	GFRINRGWSPAEEAYFSI	- - - - -	GVLK	SYTPGT		
<i>Gene.11340</i>	--	PQP	--	FWE	VC	G	AMA	EAYRLQEGWNPRDGYY	- - - - -	KLRPGYQ	PGD		
<i>MpLI1818;1</i>	--	PLP	--	FWLGLGA	L	FI	EASRVQIAWSPDASRLG	- - - - -	- - - - -	LMK	DHT	PGD	
<i>MpLI1818;2</i>	--	PGT	--	FWG	LGA	FT	EANR	VQTAWNPEADKL	F	- - - - -	LLK	DYT	PGD

	250	260	270	280	290	300											
<i>ABD37880_MvLHCA3</i>	A	I	F	FLGYGK	D	E	- - - - -	- - - - -	NLT	H	LA	D	PF				
<i>Gene.11536</i>	A	I	F	FLGYGK	D	E	- - - - -	- - - - -	NLT	H	LA	D	PF				
<i>AtLHCA3</i>	P	F	F	PLGFGK	D	E	- - - - -	- - - - -	NLL	H	LA	D	PF				
<i>Mapoly0066s0050</i>	P	F	F	FLGFGR	D	E	- - - - -	- - - - -	NLL	H	LA	D	PF				
<i>kf00560_0090</i>	P	F	F	FAGFGK	D	E	- - - - -	- - - - -	NLL	H	LA	D	PF				
<i>CrLHCA3</i>	P	F	F	LFNLGKTE	- - -	AAMKE	ELK	KEIKNGRLAM	AV	G	F	Q	Q	TA	GP		
<i>Cs_37969</i>	Q	F	F	LFNLGKSD	- - -	LKE	ELK	KEIKNGRLAM	AV	F	V	Q	G	TG	GP		
<i>MpLHCA3</i>	KYF	F	ANL	GKTD	- - -	MA	KKEIKNGRLAM	AF	GIAVQA	ATG	GP	NLT	H	LA	D	PF	
<i>OL_K08909</i>	-	F	F	GQGKSD	- - -	MA	KKEIKNGRLAM	ACFACGQA	ATG	GP	NLI	H	LA	D	PF		
<i>ABD37881_MvLHCA4.1</i>	-	FLD	I	G	DS	- - -	P	KEIKNGRLAM	FAF	GFVAQY	F	VNGMGP	GLL	H	LE	PQ	
<i>DAA05923_MvLHCA2</i>	-	FLD	I	G	DS	- - -	P	KEIKNGRLAM	FAF	GFVAQY	F	VNGMGP	GLL	H	LE	PQ	
<i>Gene.11715</i>	-	FLD	I	G	DS	- - -	P	KEIKNGRLAM	FAF	GFVAQY	F	VNGMGP	GLL	H	LE	PQ	
<i>ABD37884_MvLHCA5.2</i>	-	FDPLG	- - -	NSKE	E	M	KEIKNGRLAM	FAWC	GFMVQA	VTR	GP	TN	LEKH	A	D	PF	
<i>DAA05927_MvLHCA9</i>	-	FDPLG	- - -	NSKE	M	KEIKNGRLAM	FAWC	GFMVQA	VTR	GP	TN	LEKH	A	D	PF		
<i>Gene.10884</i>	-	FDPLG	- - -	NSKE	M	KEIKNGRLAM	FAWC	GFMVQA	VTR	GP	TN	LEKH	A	D	PF		
<i>CrLHCA9</i>	-	FDPLG	- - -	NSPS	ATKE	KEIKNGRLAM	AF	GFCVQA	ATRTQP	GP	TN	LEKH	A	D	PF		
<i>Cs_44136</i>	-	FDPANL	D	DS	- - -	EANA	EKEIKNGRLAM	AF	GFVGAA	VCR	GP	ALQSHL	H	LA	D	PF	
<i>MpLHCA9</i>	-	DP	G	- - -	NNDA	A	KEIKNGRLAM	AF	GVVVQQA	VYRTGP	GP	AALK	H	V	D	PF	
<i>OL_K08911</i>	-	FDPLG	G	GS	- - -	KDE	K	KEIKNGRAAM	AF	GIVVQG	VYR	GP	AALK	H	A	P	PF
<i>CrLHCA2</i>	-	FDPLG	G	KS	- - -	E	E	KEIKNGRLAM	AF	GFCSQAA	VY	GP	TLQLH	A	LA	D	PG
<i>Cs_7387</i>	-	FDPLN	RDDY	- - -	K	QSE	E	NGRLAM	AF	GFASQA	ANTG	GP	NLK	H	AD	P	PT
<i>MpLHCA2</i>	-	FDPSGQ	- - -	DSPA	K	KEIKNGRLAM	F	GMVSQYAV	TGTS	SP	GLEAH	A	P	PT			
<i>OL_PTHR21649</i>	-	FDPAGL	- - -	DAP	K	KEIKNGRLAM	AF	GMVSQYAV	TGTLSP	SP	GLEAH	A	P				
<i>ABD37882_MvLHCA4.2</i>	-	MF DPLGF	KG	--	NLEES	SKW	KE	NGRLAM	AS	GMF	QYDATG	SP	TNLA	H	LA	D	PW
<i>DAA05926_MvLHCA5</i>	-	MF DPLGF	KG	--	NLEES	SKW	KE	NGRLAM	AS	GMF	QYDATG	SP	TNLA	H	LA	D	PW
<i>Gene.11585</i>	-	MF DPLGF	KG	--	NLEES	SKW	KE	NGRLAM	AS	GMF	QYDATG	SP	TNLA	H	LA	D	PW
<i>DAA05920_MvLHCA4</i>	-	PF DPLGF	KG	--	NLP	ELK	KEIKNGRLAM	AS	GLFVQYSATG	ASP	DNLAAH	H	LA	D	PG		
<i>Gene.11349</i>	-	PF DPLGF	KG	--	NLP	ELK	KEIKNGRLAM	AS	GLFVQYSATG	ASP	DNLAAH	H	LA	D	PG		
<i>AtLHCA2</i>	-	LF DPLG	WGSGSPA	KL	KEL	TKEIKNGRLAM	AV	GAWFQH	YTGTGP	DNLFAH	H	LA	D	PG			
<i>Mapoly0083s0003</i>	-	FDPLG	WGAGGA	AKV	KEL	TKEIKNGRLAM	AV	GAWFQA	YTGTGP	DNLFAH	H	LA	D	PG			
<i>AtLHCA6</i>	-	FDPLG	GYAKD	S	SKAG	ELK	KEIKNGRLAM	AF	GFCFQATYT	TS	DPL	NLMAH	H	LA	D	PG	
<i>kf00564_0030</i>	-	FDPLG	GYAKD	S	SKAG	ELK	KEIKNGRLAM	AF	GFFFQA	YTGVGP	DNLTT	H	LA	D	PG		
<i>MpLHCA4</i>	-	FDPLG	GAKD	P	EKV	V	KL	KEIKANGRLAM	AV	GCI	NEAT	GVGP	ANLKAH	H	LA	D	PA
<i>OL_K08910</i>	-	FDPLG	GYAKD	G	DLK	T	KA	KEIKANGRLAM	AF	GIMVQYDHT	GVGP	ANLVSH	H	AD	PA		
<i>OL_K08908</i>	-	FDKFGWAKD	E	KT	TA	ELK	KEIKNGRLAM	AF	GICAQY	QTGVGP	NLF	FSH	G	PG			





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320

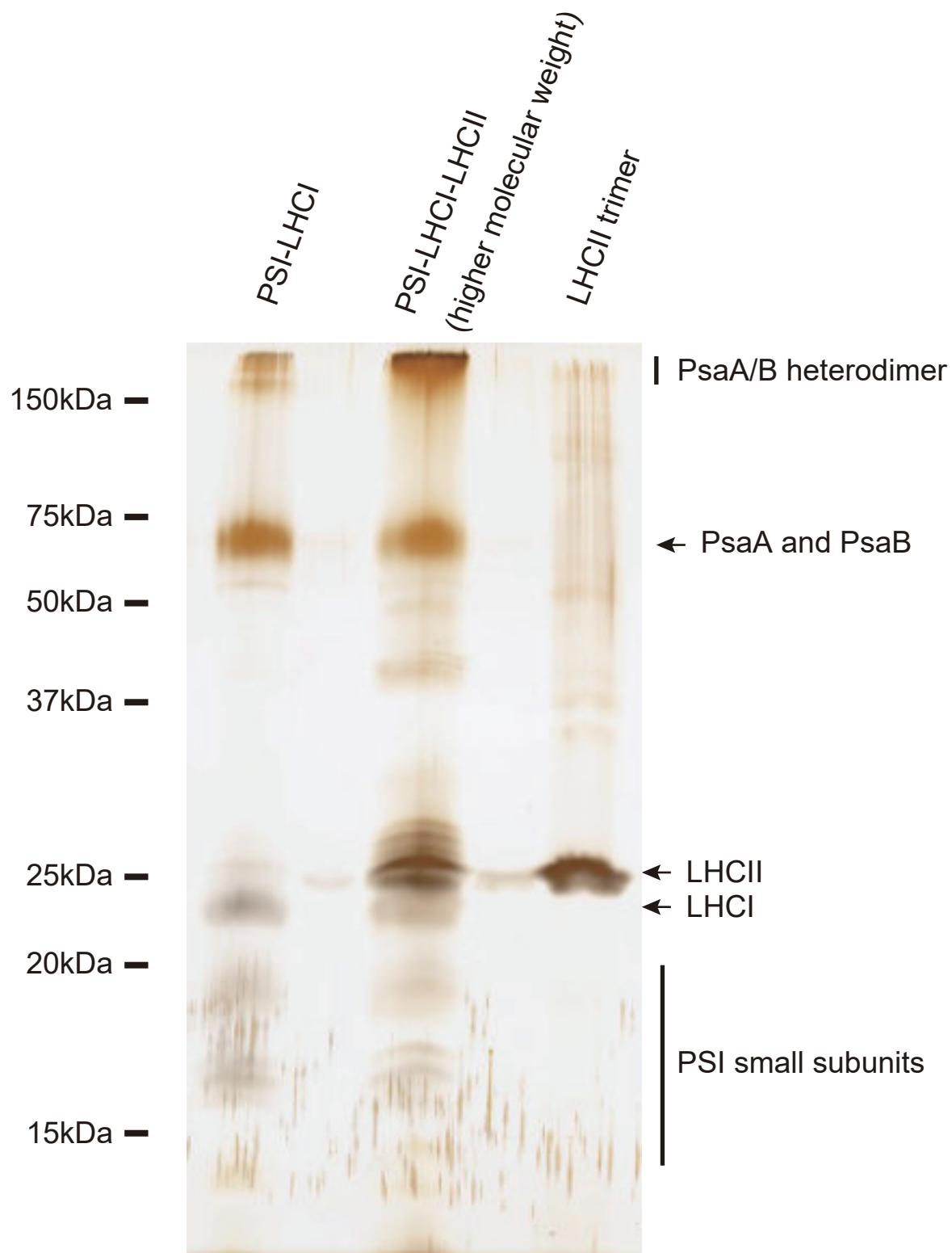
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*ABD37880\_MvLHCA3* NNNILT-TFGKIGGSF-----  
*Gene.11536* NNNILT-TFGKIGGSF-----  
*AtLHCA3* NNNVLT-SLKFH-----  
*Mapoly0066s0050* NNNVLT-NLKIH-----  
*kfl00560\_0090* ANNILT-TWGTPPGL-----  
*CrLHCA3* NNNILT-NFGKLVA-----  
*Cs\_37969* GILT-NFGHPAL-----  
*MpLHCA3* ANNMLA-GFGAIGGVSP-----  
*OL\_K08909* GLLV-NFQNIGGVSP-----  
*ABD37881\_MvLHCA4.1* QNV S-TEYAPLFFVATVWGLIFRPI  
*DAA05923\_MvLHCA2* QNV S-TEYAPLFFVATVWGLIFRPI  
*Gene.11715* QNV S-TEYAPLFFVATVWGLIFRPI  
*ABD37884\_MvLHCA5.2* NNNFIT-SIANLPNVVGK-----  
*DAA05927\_MvLHCA9* NNNFIT-SIANLPNVVGK-----  
*Gene.10884* NNNFIT-SIANLPNVVGK-----  
*CrLHCA9* GKNITY-YLTHLPETLGSA-----  
*Cs\_44136* SNNIIG-SIARLPETIGATAPPA-----  
*MpLHCA9* GCN MAT-NIMHIGSTF-----  
*OL\_K08911* GCN MAT-NIMNIPVNLA-----  
*CrLHCA2* HNNI T-SSVGVPETAVTVALPMIYFPW  
*Cs\_7387* HNNI A-SKVGPEVTLAVITPIV-----  
*MpLHCA2* QVNI T-SSVGNEFVAIIAPCYFRPI  
*OL\_PTHR21649* AVNL T-SSVGGEAVAFIAAPTFPRPI  
*ABD37882\_MvLHCA4.2* HNNVCALLPF-----  
*DAA05926\_MvLHCA5* HNNVCALLPF-----  
*Gene.11585* HNNVCALLPF-----  
*DAA05920\_MvLHCA4* HANI ALDKSGL-----  
*Gene.11349* HANI ALDKSGL-----  
*AtLHCA2* HATI A-AFTP-----  
*Mapoly0083s0003* HATV A-ALDKLQ-----  
*AtLHCA6* HCNV S-AFTSH-----  
*kfl00564\_0030* HNTI A-QL-----  
*MpLHCA4* HNTI T-NFVPIQF-----  
*OL\_K08910* HNNV A-AFIGF-----  
*OL\_K08908* QVGV -----  
*AtLHCA4* HNTIVQ-TFN-----  
*Mapoly0006s0261* HTTIVQ-TLAN-----  
*kfl00023\_0070* HVTVVS-SIEKFVGSS-----  
*CrLHCA4* STTV QNDLARL-----  
*Cs\_52367* SNNV GIEHARL-----  
*CrLHCA5* GNNILK-NIGTCT-----VPHSTIPL  
*Cs\_61250* GNNITK-NIGTCA-----IPSSTIPL  
*CrLHCA6* GTTI S-KAAVVPGQAVAPIPASEIPT  
*Cs\_46127* GTTI S-KAVVIPGQAIVPIPSSSTIPT  
*AtLHCA1* HNNIGD---IVIPFN-----  
*Mapoly0082s0040* ANNIAN---IIIPRSVL-----  
*kfl00100\_0170* HNTVAD---VFIPRSIL-----  
*CrLHCA1* GANFAT-NGISVPFF-----  
*Cs\_25286* GANFAT-NGVSIPYLT-----  
*DAA05929\_MvLHCA1* HNNAAAR-----LPL  
*Gene.11703* HNNAAAR-----LPL  
*CrLHCA7* HVNFAT-NGVSIPIA-----  
*Cs\_25284* AVTFAT-NGVSLPFVH-----  
*CrLHCA8* HVNYAT-NGVSLPFL-----  
*Cs\_48543* ANNFTT-NGTSLPAQAHPLKP-----  
*AtLHCA5* HKTIIQ-TLFTSTS-----  
*kfl00214\_0160* HVTVAE-TLGRA-----  
*MpLHCA1* HVNAAV-N-----  
*Cs\_66689* GQNI T-QGEKGTAVVIAFAEGA-----

*kfl00009\_0500* HNNWY G - N - - -  
*kfl00517\_0030* HNNWY G - S - - -  
*kfl00009\_0510* ANWF SQS - - - - QL  
*kfl00517\_0040* ANWL NGS - - - - PI  
*ABD37890\_MvLHCB4* QTIVQ - - - - -  
*Gene.10541* QTIVQ - - - - -  
*AtLHCB4.1* HTTIID-TFSSS - - - - -  
*AtLHCB4.2* HTTIID-TFSSS - - - - -  
*Mapoly0001s0025* HTTIID-TFSK - - - - -  
*AtLHCB4.3* - - - N - N - - -  
*kfl00258\_0020* TN T I D - TLSK - - -  
*CrLHCB4* NGKGL - - -  
*Cs\_30169* P - - -  
*Cs\_67011* PELVED - - IEKAAGVI - - -  
*ABD37885\_MvLHCBM1* VNNNG A - Y - - ATSSGLVG - - -  
*Gene.11306* VNNNG A - Y - - ATSSGLVG - - -  
*Gene.11314* - - -  
*AtLHCB1.1* NNNA A - F - - ATNFVPGK - - -  
*AtLHCB2.1* ANNA S - Y - - ATNFVPGN - - -  
*Mapoly0057s0073* VNNNA A - Y - - ATNFTPGN - - -  
*Mapoly0371s000* VNNNA A - Y - - ATNFTPGN - - -  
*Mapoly0199s0017* VNNNA A - Y - - ATNFTPGN - - -  
*Mapoly0199s0018* VNNNA A - Y - - ATNFTPGN - - -  
*Mapoly0199s0019* VNNNA A - Y - - ATNFTPGN - - -  
*Mapoly0199s0012* ANNA A - Y - - ATNFTPGS - - -  
*Mapoly0199s0014* ANNA A - Y - - ATNFTPGN - - -  
*Mapoly0199s0016* ANNA A - Y - - ATNFTPGN - - -  
*Mapoly0199s0015* ANNA A - Y - - ATNFTPGN - - -  
*Mapoly0199s0013* VNNNA A - Y - - ATNFTPGN - - -  
*Mapoly0199s0020* VNNNA A - Y - - ATNFTPGS - - -  
*Mapoly0057s0082* VNNNA A - Y - - ATNFTPGN - - -  
*Mapoly0057s0083* VNNNA A - Y - - ATNFTPGN - - -  
*Mapoly0068s0071* ANNA A - Y - - ATTFTPGN - - -  
*Mapoly0139s0012* TNNA A - Y - - ATAFTPQ - - -  
*Cs\_15915* ANNG A - A - - ATKFVP - - -  
*Cs\_21950* VNNNG A - A - - ATKFAP - - -  
*Cs\_58975* VNNNG A - A - - ATKFAP - - -  
*Cs\_27246* VNNNG A - A - - ATKFVPS - - -  
*Cs\_35121* VNNNG A - A - - ATKFVP - - -  
*kfl00098\_0080* VNNNA A - Y - - ATAFTPQ - - -  
*kfl00098\_0090* VNNNA A - Y - - ATAFTPQ - - -  
*kfl01434\_0010* VNNNA A - Y - - ATQFAPGQ - - -  
*CrLHCBM1* TNNA A - Y - - ATKFTPQ - - -  
*CrLHCBM2* AVNA A - Y - - ATKFTPSA - - -  
*CrLHCBM7* AVNA A - Y - - ATKFTPSA - - -  
*CrLHCBM3* VNNNA A - F - - ATKFTPSA - - -  
*CrLHCBM4* VNNNA A - F - - ATKFTPSA - - -  
*CrLHCBM8* VNNNA A - F - - ATKFTPSA - - -  
*CrLHCBM6* VNNNA A - F - - ATKFTPSA - - -  
*CrLHCBM9* VNNNA A - F - - ATKYTPSA - - -  
*kfl00809\_0010* VNNNA A - Y - - ATAFTPQ - - -  
*CrLHCBM5* TNNA T - Y - - AQKFTPQ - - -  
*Cs\_38026* AVNG T - I - GAQKFVPGN - - -  
*Mapoly0026s002* INNA A - Y - - ATNFVPRS - - -  
*AtLHCB3* ANNA A - F - - ATKFAPGA - - -  
*Mapoly0068s0047* ANNA A - Y - - ATNFVPGA - - -  
*Mapoly0068s0087* VNNNA A - Y - - ATNFVPVR - - -  
*kfl00120\_0160* ANNG A - Y - - ATAFTSPGQ - - -  
*Cs\_28488* VNNNG A - A - - ATKFTP - - -

*ABD37891\_MvLHC5* GYNFLT - ILGSGSERVPTL -----  
*Gene.11404* GYNFLT - ILGSGSERVPTL -----  
*DAA05928\_MvLHC5* GYNFLT - ILGSGSERVPTL -----  
*Cs\_64185* GYNLLT - IIGAEDRVPTL -----  
*AtLHC5* GNNLLT - VIAGTAERAPTL -----  
*Mapoly0011s0076* GNNLLT - VLQGSAERVPSL -----  
*kfl00104\_0350* ANNIIIS - VIGGNIERSPVL -----  
*AtLHC7* HNNNLLIA - MLQT -----  
*Mapoly0008s0015* HQNL A - YATSS -----  
*kfl00104\_0370* HNNNI T - AFNSS -----  
*CrLHC7* HNNN I - NLAHLQ -----  
*kfl00422\_0020* ----- H -----  
*AtLHC6* -----  
*Mapoly0052s0039* -----  
*kfl00276\_0010* -----  
*DAA05919\_MvLHC1.2* VENV KYA ----- FNQ -  
*Gene.11156* VENV KYA ----- FNQ -  
*DAA05930\_MvLHC1.1* VENV KYT ----- FQ -  
*Gene.11308* VENV KYT ----- FQ -  
*MpLHCAP2.1* HANVLT - NAASGFGFY -----  
*MpLHC1.1* HANVLT - NAASGFGFY -----  
*CmLHC* ----- T - N ----- FQPL  
*PyLHC* ----- LSGKLFP -----  
*CrLHCSR1* ILELEGPLTPLPDNLKAI -----  
*CrLHCSR3* ILELDGLPVTPLPDNLKSL -----  
*Cs\_65904* --- SP -----  
*kfl00478\_0030* GL -----  
*DAA05932\_MvLI18;2* -----  
*Gene.11245* -----  
*Gene.11340* -----  
*MpLI18;1* ALKAMSLDEAACSKAFEATAVL -----  
*MpLI18;2* ALEAMATNEAACAKAFEAAVFAA -----

Fig. S2



**Figure S2. 2D-SDS-PAGE of the high-molecular-weight PSI-LHCl-LHCII, PSI-LHCl, and LHCII trimer.**

The high-molecular-weight PSI-LHCl-LHCII, PSI-LHCl, and LHCII trimer bands separated by CN-PAGE after sucrose density gradient centrifugation (Fig. 7) were subjected to 2D-SDS-PAGE. Protein bands were visualized by silver-staining.

**Table S1-1. The list of the PSI proteins predicted by the Iso-seq analysis in this study**

	Gene annotation in this study	Accession No.	The best-hit gene in <i>A.thaliana</i>			The best-hit gene in <i>C.reinhardtii</i>		
Gene3244	PSAA	ICQU01000001	ATCG00350.1	PSAA	Photosystem I, PsaA/PsAB protein	NP_958375.1	PSAA	Photosystem I P700 chlorophyll a apoprotein A1
Gene3504	PSAB	ICQU01000002	ATCG00340.1	PSAB	Photosystem I, PsaA/PsAB protein	NP_958404.1	PSAB	Photosystem I P700 chlorophyll a apoprotein A2
Gene9022	PSAD	ICQU01000006	AT1G03130.1	PSAD-2	photosystem I subunit D-2	Cre05.g238332.t1.1	PSAD	Photosystem I reaction center subunit II, 20 kDa
Gene11867	PSAE	ICQU01000029	AT2G20260.1	PSAE-2	photosystem I subunit E-2	Cre10.g420350.t1.2	PSAE	Photosystem I 8.1 kDa reaction center subunit IV
Gene11501	PSAF	ICQU01000021	AT1G31330.1	PSAF	photosystem I subunit F	Cre09.g412100.t1.2	PSAF	Photosystem I reaction center subunit III
Gene11916	PSAG	ICQU01000031	AT1G55670.1	PSAG	photosystem I subunit G	Cre12.g560950.t1.2	PSAG	Photosystem I reaction center subunit V
Gene11865	PSAH	ICQU01000028	AT1G52230.1	PSAH-2	photosystem I subunit H2	Cre07.g330250.t1.2	PSAH	Subunit H of photosystem I
Gene11650	PSAL	ICQU01000024	AT4G12800.1	PSAL	photosystem I subunit I	Cre12.g486300.t1.2	PSAL	Photosystem I reaction center subunit XI
Gene11942	PSAO	ICQU01000034	AT1G08380.1	PSAO	photosystem I subunit O	Cre07.g334550.t1.2	PSAO1	Photosystem I subunit O

**Table S1-2. The list of the PSII proteins predicted by the Iso-seq analysis in this study**

	Gene annotation in this study	Accession No.	The best-hit gene in <i>A.thaliana</i>					The best-hit gene in <i>C.reinhardtii</i>					
Gene10292	PSBA	ICQU01000008	ATCG00020.1	PSBA	photosystem II reaction center protein A	NP_958377.1	PSBA	photosystem II protein D1	Cre09.g396213.t1.1	PSBO	Oxygen-evolving enhancer protein 1 of photosystem II		
Gene4825	PSBB	ICQU01000003	ATCG00680.1	PSBB	photosystem II reaction center protein B	NP_958388.1	PSBB	photosystem II 47 kDa protein	Cre12.g550850.t1.2	PSBP1	Oxygen-evolving enhancer protein 2 of photosystem II		
Gene6781	PSBC	ICQU01000005	ATCG00280.1	PSBC	photosystem II reaction center protein C	NP_958422.1	PSBC	photosystem II 44 kDa protein	Cre08.g372450.t1.2	PSBQ	Oxygen evolving enhancer protein 3		
Gene4841	PSBD	ICQU01000004	ATCG00270.1	PSBD	photosystem II reaction center protein D	NP_958420.1	PSBD	photosystem II protein D2	Cre06.g261000.t1.2	PSBR	10 kDa photosystem II polypeptide		
Gene10723	PSBO	ICQU01000010	AT3G50820.1	PSBO-2	photosystem II subunit O-2								
Gene11499	PSBP	ICQU01000020	AT1G06680.1	PSBP-1	photosystem II subunit P-1								
Gene11808	PSBQ	ICQU01000027	AT4G05180.1	PSBQ-2	photosystem II subunit Q-2								
Gene11938	PSBR	ICQU01000033	AT1G79040.1	PSBR	photosystem II subunit R								
Gene11934	PSBW	ICQU01000032	AT2G30570.1	PSBW	photosystem II reaction center W								
Gene11880	PSBX	ICQU01000030	AT2G06520.1	PSBX	photosystem II subunit X								
Gene9164	PSBY	ICQU01000007	AT1G67740.1	PSBY	photosystem II BY								
						Cre10.g452100.t1.1	PSBY	Ycf32-related polyprotein of photosystem II					

**Table S1-3. The list of the LHC proteins predicted by the Iso-seq analysis in this study**

	Gene annotation in this study	Accession No.	The best-hit gene in <i>A.thaliana</i>				The best-hit gene in <i>C.reinhardtii</i>			
Gene11245	LHCSR	ICQU01000013	AT1G15820.1	LHC86	light harvesting complex photosystem II subunit 6		Cre08.g367500.t1.1	LHCSR3.1	Stress-related chlorophyll a/b binding protein 2	
Gene11340	LHCSR	ICQU01000017	AT3G61470.1	LHCA2	photosystem I light harvesting complex gene 2		Cre08.g367500.t1.1	LHCSR3.1	Stress-related chlorophyll a/b binding protein 2	
Gene11306	LHCBM	ICQU01000014	AT2G05100.1	LHC82.1	photosystem II light harvesting complex gene 2.1		Cre01.g066917.t1.1	LHCBM1	Chlorophyll a/b binding protein of LHCII	
Gene11314	LHCBM	ICQU01000016	AT2G05070.1	LHC82.2	photosystem II light harvesting complex gene 2.2		Cre01.g066917.t1.1	LHCBM1	Chlorophyll a/b binding protein of LHCII	
Gene10541	LHCB4	ICQU01000009	AT2G40100.1	LHC84.3	light harvesting complex photosystem II		Cre17.g720250.t1.2	LHC84	Chlorophyll a/b binding protein of photosystem II	
Gene11404	LHC85	ICQU01000019	AT4G10340.1	LHC85	light harvesting complex of photosystem II 5		Cre16.g673650.t1.1	LHC85	Minor chlorophyll a/b binding protein of photosystem II	
Gene11156	LHCP	ICQU01000012	AT5G54270.1	LHC83	light-harvesting chlorophyll B-binding protein 3		Cre03.g156900.t1.2	LHCBM5	Chlorophyll a/b binding protein of LHCII	
Gene11308	LHCP	ICQU01000015	AT5G54270.1	LHC83	light-harvesting chlorophyll B-binding protein 3		Cre04.g232104.t1.1	LHCBM3	Light-harvesting complex II chlorophyll a/b binding protein M3	
Gene11703	LHCA1	ICQU01000025	AT3G54890.1	LHCA1	photosystem I light harvesting complex gene 1		Cre06.g283050.t1.2	LHCA1	Light-harvesting protein of photosystem I	
Gene11349	LHCA2	ICQU01000018	AT3G61470.1	LHCA2	photosystem I light harvesting complex gene 2		Cre16.g687900.t1.2	LHCA7	Light-harvesting protein of photosystem I	
Gene11585	LHCA2	ICQU01000023	AT3G47470.1	LHCA4	light-harvesting chlorophyll-protein complex I subunit A4		Cre10.g452050.t1.2	LHCA4	Light-harvesting protein of photosystem I	
Gene11536	LHCA3	ICQU01000022	AT1G61520.1	LHCA3	photosystem I light harvesting complex gene 3		Cre11.g467573.t1.1	LHCA3	Chlorophyll a/b binding protein of photosystem I, type III	
Gene11715	algae-type LHCA2	ICQU01000026	AT1G45474.1	LHCA5	photosystem I light harvesting complex gene 5		Cre12.g508750.t1.2	LHCA2	Light-harvesting protein of photosystem I	
Gene10884	LHCA9	ICQU01000011	AT1G45474.1	LHCA5	photosystem I light harvesting complex gene 5		Cre07.g344950.t1.2	LHCA9	Light-harvesting protein of photosystem I	

**Table S2. The identified proteins in the PSI-PSII band by MS**

Gene ID	NSAF (fmol)	Category	Gene annotation in this study	Accession No.	Best-Hit Gene in Arabidopsis		Best-Hit Gene in Chlamydomonas	
					Gene ID	Gene Name	Gene ID	Gene Name
Gene11306	4526	LHC	LHCBM	ICQU01000014	AT2G05100.1	LHCB2.1	Cre01.g066917.t1.1	LHCBM1
Gene10292	4034	PSII	PSBA	ICQU01000008	ATCG00020.1	PSBA	NP_958377.1	PSBA
Gene11501	3411	PSI	PSAF	ICQU01000021	AT1G31330.1	PSAF	Cre09.g412100.t1.2	PSAF
Gene11156	3322	LHC	LHCP	ICQU01000012	AT5G54270.1	LHCB3	Cre03.g156900.t1.2	LHCBM5
Gene11585	2942	LHC	LHCA2	ICQU01000023	AT3G47470.1	LHCA4	Cre10.g452050.t1.2	LHCA4
Gene4825	2883	PSII	PSBB	ICQU01000003	ATCG00680.1	PSBB	NP_958388.1	PSBB
Gene11867	2773	PSI	PSAE	ICQU01000029	AT2G20260.1	PSAE-2	Cre10.g420350.t1.2	PSAE
Gene11865	2654	PSI	PSAH	ICQU01000028	AT1G52230.1	PSAH-2	Cre07.g330250.t1.2	PSAH
Gene4841	2605	PSII	PSBD	ICQU01000004	ATCG00270.1	PSBD	NP_958420.1	PSBD
Gene3547	2468	Others			AT5G59970.1		Cre12.g506350.t1.2	HFO18
Gene11916	2444	PSI	PSAG	ICQU01000031	AT1G55670.1	PSAG	Cre12.g560950.t1.2	PSAG
Gene9022	2440	PSI	PSAD	ICQU01000006	AT1G03130.1	PSAD-2	Cre05.g238332.t1.1	PSAD
Gene11404	2412	LHC	LHCB5	ICQU01000019	AT4G10340.1	LHCB5	Cre16.g673650.t1.1	LHCB5
Gene10541	2301	LHC	LHCB4	ICQU01000009	AT2G40100.1	LHCB4.3	Cre17.g720250.t1.2	LHCB4
Gene11715	2235	LHC	algae-type LHCA2	ICQU01000026	AT1G45474.1	LHCA5	Cre12.g508750.t1.2	LHCA2
Gene6781	1782	PSII		ICQU01000005	ATCG00280.1	PSBC	NP_958422.1	PSBC
Gene11340	1707	LHC	LHCSR	ICQU01000017	AT3G61470.1	LHCA2	Cre08.g367500.t1.1	LHCSR3.1
Gene6617	1577	Others			AT3G61320.1		Cre06.g261750.t1.2	
Gene3504	1569	PSII	PSAB	ICQU01000002	ATCG00340.1	PSAB	NP_958404.1	PSAB
Gene7944	1382	Others			AT3G61320.1		Cre06.g261750.t1.2	
Gene11703	1355	LHC	LHCA1	ICQU01000025	AT3G54890.1	LHCA1	Cre06.g283050.t1.2	LHCA1
Gene10723	1232	PSII	PSBO	ICQU01000010	AT3G50820.1	PSBO-2	Cre09.g396213.t1.1	PSBO
Gene11536	1228	LHC	LHCA3	ICQU01000022	AT1G61520.1	LHCA3	Cre11.g467573.t1.1	LHCA3
Gene10884	1219	LHC	LHCA9	ICQU01000011	AT1G45474.1	LHCA5	Cre07.g344950.t1.2	LHCA9
Gene7515	1169	Others			AT5G04180.1	ATACA3	Cre09.g415700.t1.2	CAH3
Gene11308	1066	LHC	LHCP	ICQU01000015	AT5G54270.1	LHCB3	Cre04.g232104.t1.1	LHCBM3
Gene7423	858	Others			AT5G17170.1	ENH1	Cre12.g510400.t1.1	CPLD30
Gene8340	839	Others			AT4G38970.1	FBA2	Cre05.g234550.t1.2	FBA3
Gene9362	780	Others			AT5G23860.1	TUB8	Cre12.g549550.t1.2	TUB2
Gene8224	529	Others			ATCG00490.1	RBCL	NP_958405.1	RBCL
Gene8089	512	Others						

**Table S3. The identified proteins in the PSII-LHCII band by MS**

Gene ID	NSAF(fmol)	Category	Gene annotation in this study	Accession No.	Best-Hit Gene in Arabidopsis		Best-Hit Gene in Chlamydomonas	
					Gene ID	Gene Name	Gene ID	Gene Name
Gene11306	7753	LHC	LHCBM	ICQU01000014	AT2G05100.1	LHCB2.1	Cre01.g066917.t1.1	LHCBM1
Gene10292	6458	PSII	PSBA	ICQU01000008	ATCG00020.1	PSBA	NP_958377.1	PSBA
Gene10541	3371	LHC	LHCB4	ICQU01000009	AT2G40100.1	LHCB4.3	Cre17.g720250.t1.2	LHCB4
Gene4825	3021	PSII	PSBB	ICQU01000003	ATCG00680.1	PSBB	NP_958388.1	PSBB
Gene11404	1991	LHC	LHCB5	ICQU01000019	AT4G10340.1	LHCB5	Cre16.g673650.t1.1	LHCB5
Gene11865	1757	PSI	PSAH	ICQU01000028	AT1G52230.1	PSAH-2	Cre07.g330250.t1.2	PSAH
Gene6781	1495	PSII	PSBC	ICQU01000005	ATCG00280.1	PSBC	NP_958422.1	PSBC
Gene11938	1341	PSII	PSBR	ICQU01000033	AT1G79040.1	PSBR	Cre06.g261000.t1.2	PSBR
Gene3547	1317	Others			AT5G59970.1		Cre12.g506350.t1.2	HFO18
Gene11585	1159	LHCI	LHCA2	ICQU01000023	AT3G47470.1	LHCA4	Cre10.g452050.t1.2	LHCA4
Gene11867	1142	PSI	PSAE	ICQU01000029	AT2G20260.1	PSAE-2	Cre10.g420350.t1.2	PSAE
Gene11916	1105	PSI	PSAG	ICQU01000031	AT1G55670.1	PSAG	Cre12.g560950.t1.2	PSAG
Gene4841	1095	PSII	PSBD	ICQU01000004	ATCG00270.1	PSBD	NP_958420.1	PSBD
Gene7779	1091	Others			ATCG00430.1	PSBG	Cre12.g492300.t1.2	NUO10
Gene11308	1084	LHC	LHCP	ICQU01000015	AT5G54270.1	LHCB3	Cre04.g232104.t1.1	LHCBM3
Gene9022	1080	PSI	PSAD	ICQU01000006	AT1G03130.1	PSAD-2	Cre05.g238332.t1.1	PSAD
Gene11715	976	LHC	algae-type LHCA2	ICQU01000026	AT1G45474.1	LHCA5	Cre12.g508750.t1.2	LHCA2
Gene10884	937	LHC		ICQU01000011	AT1G45474.1	LHCA5	Cre07.g344950.t1.2	LHCA9
Gene11501	926	PSI	PSAF	ICQU01000021	AT1G31330.1	PSAF	Cre09.g412100.t1.2	PSAF
Gene11349	902	LHC	LHCA2	ICQU01000018	AT3G61470.1	LHCA2	Cre16.g687900.t1.2	LHCA7
Gene11703	858	LHC	LHCA1	ICQU01000025	AT3G54890.1	LHCA1	Cre06.g283050.t1.2	LHCA1
Gene7515	822	Others			AT5G04180.1	ATACA3	Cre09.g415700.t1.2	CAH3
Gene11156	789	LHC	LHCP	ICQU01000012	AT5G54270.1	LHCB3	Cre03.g156900.t1.2	LHCBM5
Gene2597	781	Others			ATCG01100.1	NDHA		
Gene11934	741	PSII	PSBW	ICQU01000032	AT2G30570.1	PSBW		
Gene4851	714	Others			ATCG01110.1	NDHH	Cre09.g405850.t1.1	NUO7
Gene11718	690	Others			ATCG00420.1	NDHJ	Cre07.g327400.t1.1	NUO9
Gene9021	683	Others			ATCG01070.1	NDHE	Cre09.g402552.t1.1	NUO11
Gene11761	676	Others			AT3G61320.1		Cre06.g261750.t1.2	
Gene6617	618	Others			AT4G12800.1	PSAL	Cre12.g486300.t1.2	PSAL
Gene11650	605	PSI	PSAL	ICQU01000024	AT1G20020.1	ATLFNR2	Cre11.g476750.t1.2	FNR1
Gene8813	572	Others			AT2G28720.1		Cre13.g590750.t1.2	HTB21
Gene11139	565	Others			AT3G50820.1	PSBO-2	Cre09.g396213.t1.1	PSBO
Gene10723	565	PSII	PSBO	ICQU01000010	AT4G05180.1	PSBQ-2	Cre08.g372450.t1.2	PSBQ
Gene11808	539	PSII		ICQU01000027	AT5G17170.1	ENH1	Cre12.g510400.t1.1	CPLD30
Gene8340	531	Others			AT5G58260.1			
Gene10856	527	Others			AT3G61470.1	LHCA2	Cre08.g367500.t1.1	LHCSR3.1
Gene11340	526	LHC	LHCSR	ICQU01000017	AT1G15980.1	NDF1		
Gene5104	522	Others						

**Table S4. The identified proteins in the PSI-LHCI band by MS**

Gene ID	NSAF(fmol)	Category	Gene annotation in this study	Accession No.	Best-Hit Gene in Arabidopsis		Best-Hit Gene in Chlamydomonas	
					Gene ID	Gene Name	Gene ID	Gene Name
Gene11703	9063	LHC	LHCA1	ICQU01000025	AT3G54890.1	LHCA1	Cre06.g283050.t1.2	LHCA1
Gene11916	6111	PSI	PSAG	ICQU01000031	AT1G55670.1	PSAG	Cre12.g560950.t1.2	PSAG
Gene11867	5182	PSI	PSAE	ICQU01000029	AT2G20260.1	PSAE-2	Cre10.g420350.t1.2	PSAE
Gene11501	5101	PSI	PSAF	ICQU01000021	AT1G31330.1	PSAF	Cre09.g412100.t1.2	PSAF
Gene11536	4709	LHC	LHCA3	ICQU01000022	AT1G61520.1	LHCA3	Cre11.g467573.t1.1	LHCA3
Gene10884	4541	LHC	LHCA9	ICQU01000011	AT1G45474.1	LHCA5	Cre07.g344950.t1.2	LHCA9
Gene11865	2669	PSI	PSAH	ICQU01000028	AT1G52230.1	PSAH-2	Cre07.g330250.t1.2	PSAH
Gene11349	2593	LHC	LHCA2	ICQU01000018	AT3G61470.1	LHCA2	Cre16.g687900.t1.2	LHCA7
Gene9022	2205	PSI	PSAD	ICQU01000006	AT1G03130.1	PSAD-2	Cre05.g238332.t1.1	PSAD
Gene11585	1980	LHC	LHCA2	ICQU01000023	AT3G47470.1	LHCA4	Cre10.g452050.t1.2	LHCA4
Gene11715	1861	LHC	algae-type LHCA2	ICQU01000026	AT1G45474.1	LHCA5	Cre12.g508750.t1.2	LHCA2
Gene11306	1695	LHC		ICQU01000014	AT2G05100.1	LHCB2.1	Cre01.g066917.t1.1	LHCBM1
Gene3547	1333	Others	PSAB	ICQU01000002	AT5G59970.1		Cre12.g506350.t1.2	HFO18
Gene3504	1286	PSI			ATCG00340.1	PSAB	NP_958404.1	PSAB
Gene11650	1115	PSI	PSAL	ICQU01000024	AT4G12800.1	PSAL	Cre12.g486300.t1.2	PSAL
Gene11761	914	Others	PSBD	ICQU01000004	AT2G28720.1		Cre13.g590750.t1.2	HTB21
Gene11139	868	Others			ATCG00270.1	PSBD	NP_958420.1	PSBD
Gene4841	822	PSII	PSAA	ICQU01000001	ATCG00350.1	PSAA	NP_958375.1	PSAA
Gene3244	820	PSI			ATCG00020.1	PSBA	NP_958377.1	PSBA
Gene10292	783	PSII	PSBA	ICQU01000008	AT3G61320.1		Cre06.g261750.t1.2	
Gene9567	782	Others			AT5G04180.1	ATACA3	Cre09.g415700.t1.2	CAH3
Gene6617	765	Others	PSBB	ICQU01000003	ATCG00680.1	PSBB	NP_958388.1	PSBB
Gene7515	730	Others			AT2G40100.1	LHCB4.3	Cre17.g720250.t1.2	LHCB4
Gene4825	682	PSII	LHC	ICQU01000009	AT4G38970.1	FBA2	Cre05.g234550.t1.2	FBA3
Gene10541	675	LHC			ATCG00280.1	PSBC		
Gene5134	575	Others	PSBC	ICQU01000005				
Gene9362	538	Others						
Gene6781	535	PSII						