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<tr>
<td>Citation</td>
<td>Japanese Journal of Veterinary Research, 39(2-4): 167-177</td>
</tr>
<tr>
<td>Issue Date</td>
<td>1991-12-26</td>
</tr>
<tr>
<td>DOI</td>
<td>10.14943/jjvr.39.2-4.167</td>
</tr>
<tr>
<td>Doc URL</td>
<td><a href="http://hdl.handle.net/2115/8940">http://hdl.handle.net/2115/8940</a></td>
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<td>Type</td>
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<td>File Information</td>
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CHARACTERIZATION OF DNA FRAGMENT FROM
CHLAMYDIA PSITTACI AVIAN STRAIN WHICH SHOWS
HIGH HOMOLOGY WITH HYPB GENE OF CHLAMYDIA

Chiaki SATO¹, Atushi KATUMATA², Ikuo TAKASHIMA¹
and Nobuo HASHIMOTO¹

SUMMARY
A study was performed to characterize DNA fragment No. 17 of C. psittaci
strain P-1041 which encoded 42 KD β-galactosidase fusion protein with type­
specific antigenecity. Sequence determination identified a partial open reading
frame that spanned about 1,200bp. nucleotides. Screening the literatures for
the nucleotide and deduced amino acid sequences revealed extensive similarity
between the DNA fragment of P-1041 and two chlamydial hypB genes. This
DNA showed 91.5% homology with C. psittaci GPIC hypB gene in nucleotide
sequence and 96.4% homology in deduced amino acid sequence. The hypB gene
of C. trachomatis serovar A and the P-1041 DNA fragment showed 81.2% and
91.3% homology in nucleotide and amino acid sequences, respectively. Dot
enzyme-linked immunosorbent assay, for the products of deleted DNA fragments
defined the coding region for type-specific antigenic polypeptide. In addition, the
P-1041 DNA fragment carried a sequence highly homologous (>49%) with other
bacterial and plant genes called chaperonin which responds to various stress in
cells. From these results, the P-1041 DNA fragment was found to be a part of
hypB gene and to encode the region critical for type-specific antigenicity.

Key words: Chlamydia psittaci, heat-shock gene

INTRODUCTION
The genus Chlamydia is a procaryotic pathogen which is an obligate intracellular
parasite of eucaryotic cells and causes a wide variety of syndromes in mammalian and
avian hosts (2, 12, 13). This gram negative bacteria has a unique life cycle in which
two distinct forms, an infectious, metabolically inactive form elementary body (EB) and
a non-infectious, metabolically active form reticulate body (RB) can be defined (1, 9).

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Despite this unique developmental cycle and considerable medical importance, relatively little is known of the molecular basis for biological properties of chlamydia. At present, only a limited number of chlamydial genes have been cloned.

Recently, Morrison et al. cloned and sequenced hyp genes of *Chlamydia psittaci* GPIC strain and *Chlamydia trachomatis* serovar A and suggested that hyp gene products were responsible for the hypersensitivity of ocular chlamydial infections (6, 7, 8). It was shown that the hyp gene contained two open reading frames (ORFs), hypA and hypB, coding 20KD and 57KD proteins, respectively. These two proteins have been identified as members of a stress response protein family which is widely conserved among bacteria and plants with common antigenic properties. Proteins such as *Escherichia coli* GroEL, *Coxella burnetii* 60KD Hsp, and plant Rubisco subunit binding protein are included in this family which is classified as chaperonin (3, 15).

Although the avian strain of *C. psittaci* is mostly responsible for human psittacosis, little has been published about molecular cloning of the gene of this species. Furthermore, hypB was reported to encode the antigenic epitope common to the chlamydial genus.

We recently cloned DNA fragments from *C. psittaci* pigeon strain (P-1041) expressing 42KD β-galactosidase fusion protein with type-specific antigenicity, as revealed by the reactivity only with homologous antiserum to the *C. psittaci* P-1041 strain (11). In this report, sequence analysis was performed to define the characteristics of the P-1041 DNA fragment expressing 42KD protein, showing the DNA fragment to have highly homologous sequence with the chlamydial hypB genes. Furthermore, deletion fragments were generated from the P-1041 DNA fragment and their products were examined for antigenicity, to reveal coding region critical for type specific antigen.

**MATERIALS AND METHODS**

**Bacteria, bacteriophages and plasmids:** *Chlamydia psittaci* avian strain P-1041 was isolated from feral pigeons in our laboratory. Cultivation and purification of chlamydia and extraction of DNA has been followed by ref. 4 and 16. Plasmid pUC19 was used initially for cloning of chlamydial DNA fragments. Subclones in the M13 phages, mp18 and mp19, were used for DNA sequencing as well as plasmids pUC18 and pUC19. The plasmids, phages, and *E. coli* host strain JM109 were kindly provided by the Department of Genetic Biochemistry, Faculty of Science, Hokkaido University, Sapporo.

**Antisera to chlamydia:** Antisera against chlamydia were prepared in mice. An emulsion of purified EB suspension was injected subcutaneously. The immunizations were carried out at weeks 0, 2 and 3. Antisera were collected one week after the final immunization.

**DNA preparation and cloning:** The methods of preparation and cloning of chlamy-
Hyp gene of *C. psittaci* avian strain have been followed by ref. 4. The purified EBs were treated with 1% (W/V) sodium dodecyl sulfate (SDS) and 20mM EDTA for 1hr at 37°C, followed by digestion with proteinase K (0.1mg/ml) for 2hr at 37°C. The DNA was isolated by phenol-chloroform extractions and ethanol precipitation. RNA was removed by treatment with 0.1mg/ml ribonuclease A for 1hr at 37°C. Phenol-chloroform extraction and ethanol precipitation were repeated, and the final DNA pellet was suspended 10mM Tris-HCl buffer containing 1mM EDTA, pH 8.0. Total DNA from *C. psittaci* P-1041 was partially digested with *Sau3AI* and ligated to *BamHI*-digested plasmid pUC19. These recombinant plasmids were introduced to *E. coli* JM109. Ampicillin-resistant white colonies were screened for expression of chlamydial proteins with dot-ELISA.

Dot-ELISA for immuno-screening: Standard methods were used for enzyme-linked immunoassay (17). Briefly, the transformed *E. coli* were transferred to the nitrocellulose filter. After lysis by chloroform, products of each colony were allowed to react with anti-chlamydia homologous antiserum that was absorbed with host *E. coli* proteins. The filter was then washed, and the bound antibodies were reacted with peroxidase-labeled antimouse immunoglobulin goat serum (CAPPEL, Co., Ltd., West Chester, PA.). The chromogen-3, 3'-diaminobenzidine tetrahydro-chloride was used for colour development.

Enzymes and radiolabeled compounds: Restriction endonucleases were purchased from TAKARA SHUZO Co., Ltd (Kyoto, Japan) or NIPPON GENE Co., Ltd (Toyama, Japan), and the manufacturer’s directions for their use were followed. The radiolabeled triphosphate, \([\alpha-32p]\) dCTP, was obtained from AMERSHAM JAPAN Co., Ltd (Tokyo, Japan). All enzymes were used according to the instructions provided by the manufacturers.

DNA sequencing and sequence analysis: Both strands of the DNA were sequenced by the primer extention dideoxy-chain termination procedure by Sangar et al. (10). The nested deletions were generated by the DNase I/Shotgun cloning method and each deleted fragment was inserted in pUC18 and pUC19 for double strand sequencing (5). In addition, some of fragments were cloned into M13 phages for sequencing. Sequence analysis was done using the DNASIS programs (HITACHI Software Engineering Co., Ltd. Yokohama, Japan), and the homologous sequence was searched among the sequences submitted in the GenBank and EMBL nucleic acid sequence data bank.

Determination of coding region for chlamydial antigenic polypeptide: Each of the recombinant plasmids containing sequence-deleted fragment generated by DNase I/Shotgun cloning methods was tested for the expression of antigenic proteins. Characterization of the products of these clones was carried out by dot ELISA as described above for immuno-screening using homologous and heterologous antisera (17).
RESULTS

Characterization of recombinant clones: Chlamydial DNA was inserted into pUC19, and recombinant plasmid was used to transform *E. coli*. Ampicillin-resistant white colonies expressing β-galactosidase fusion protein were screened by dot ELISA. Only two of over 8,000 colonies tested were positive giving strong signal, and one of them was further characterized. The product of the recombinant was previously shown to be 42KD β-galactosidase fusion protein by western blotting and to have type-specific antigenicity by dot ELISA (11).

The plasmid containing chlamydial DNA inserts at approximately 1.7Kbp was

![Restriction Map of P-1041 DNA Fragment No. 17](image)

Fig. 1 Restriction map of P-1041 DNA fragment No.17 and antigenic expression of the deleted fragments. Deleted fragments from pUC19 produced immuno-reactive polypeptide (+) or nonimmunoreactive polypeptide (−).
digested with several restriction enzymes and mapped as followed by ref. 14. The restriction map of this fragment is shown in Fig. 1.

Nucleotide sequence and deduced amino acid sequence: The nucleotide and deduced amino acid sequences are shown in Fig. 2. A partial ORF of ca. 1,250bp nucleotides was identified and an amino acid sequence of 417 residues was predicted approximately as a 44.8KD polypeptide. This is close to the estimated molecular weight (47.3KD).
mass of 42KD obtained by SDS-polyacrylamide gel electrophoresis of the product previously shown (11). There was no putative termination structure after the stop codon of the ORF. In addition, there seemed to be another ORF started, from a distance of 139 bases downstream from the stop codon of the first ORF. Furthermore, this suppositional second ORF had no putative promoter site nor ribosome binding site.

Antigen-coding region of the fragment and the cross-reactivity of the expressed antigen with the antisera against the other chlamydial strains: To investigate the antigen coding region, the nested deletions were generated by the DNase I/shotgun cloning methods and each of the products of the deletion DNA fragment was tested on the antigenicity (Fig. 1). The dot-ELISA analysis revealed that 79110 of the fragment, a region containing ca. 800 bases starting from 5'end, expressed an immunoreactive product. However, nearby 79124 of the P-I041 DNA fragment, a region containing ca. 650 bases starting from 5'end, produced non-immunoreactive polypeptide.

This result suggests that epitope area located in the region from ca. 650 to ca. 800 position. Furthermore, these immunoreactive product from each deleted fragments reacted only with homologous antiserum to P–1041 strain, and not with heterologous antiser to 9 C. psittaci strains and 2 C. trachomatis strains (11).
Table 1 Percent homology of nucleotide and deduced amino acid sequence among chlamydial hypB genes and other bacterial and higher plant chaperonin genes.

<table>
<thead>
<tr>
<th></th>
<th>C. psittaci P-1041 DNA</th>
<th>C. psittaci GPIC hypB gene</th>
<th>C. trachomatis A hypB gene</th>
<th>Cox. brunetii hsp60 gene</th>
<th>E. coli groES gene</th>
<th>T. aesticum Rubisco binding protein gene</th>
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<tr>
<td>C. psittaci</td>
<td>91.5*</td>
<td>81.2</td>
<td>61.1</td>
<td>59.4</td>
<td>56.5</td>
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<tr>
<td>P-1041 DNA</td>
<td>(96.4)</td>
<td>(91.3)</td>
<td>(59.1)</td>
<td>(56.9)</td>
<td>(49.2)</td>
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<tr>
<td>C. psittaci</td>
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<td>61.2</td>
<td>60.0</td>
<td>57.2</td>
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<tr>
<td>GPIC hypB gene</td>
<td>(93.0)</td>
<td>(61.4)</td>
<td>(60.8)</td>
<td>(51.0)</td>
<td></td>
<td></td>
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<tr>
<td>C. trachomatis</td>
<td></td>
<td></td>
<td>62.1</td>
<td>61.1</td>
<td>57.4</td>
<td></td>
</tr>
<tr>
<td>A hypB gene</td>
<td></td>
<td></td>
<td>(61.3)</td>
<td>(60.6)</td>
<td>(50.8)</td>
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<tr>
<td>Cox. brunetii</td>
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<td></td>
<td></td>
<td>68.5</td>
<td>54.7</td>
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<tr>
<td>hsp60 gene</td>
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<td>(75.5)</td>
<td>(49.5)</td>
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<tr>
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<td>55.8</td>
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<td>(49.5)</td>
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<td>I. aesticum</td>
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<td>Rubisco binding protein gene</td>
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Asterisk indicate nucleotide sequence homology and ( ) indicate amino acid sequence homology.
Homology of nucleotide sequence and the deduced amino acid sequence of the fragment with those of hypB and chaperonin genes: To assess the function of the chlamydial gene, nucleotide and amino acid homology searches were carried out on the published sequence data of organisms. The P-1041 DNA fragment was extensively and highly homologous to two of the chlamydial hyp genes (Table 1). With one of these, the C. psittaci GPIC hypB gene, the P-1041 DNA showed 91.5% homology in nucleotide sequence and 96.4% homology in amino acid sequence. Another hyp gene isolated from C. trachomatis serovar A showed 81.2% and 91.3% homology in nucleotide and amino acid sequence with the DNA from P-1041, respectively. Variable regions among the amino acid sequence were found at two locations, residues 184 to 229, and residues 340 to 380. It was observed that 5 to 10 residue substitutions occurred among three chlamydial sequences (Fig 3).

Fig. 3 Deduced amino acid sequence alignment of the P-1041 chaperonin-like protein and other chlamydial HypB proteins. Comparison was made among P-1041; C. psittaci strain avian P-1041, GPIC; C. psittaci strain guinea pig inclusion conjunctivitis and, A; C. trachomatis serovar A. Dot indicate sequence identity. 3' end of deletion-fragments are represented by 1 with asterisk for fragment 79124 and 2 for fragment 79110.
Hyp gene of C. psittaci avian strain

In addition, the P-1041 DNA fragment showed a highly homologous sequence with several genes of chaperonin family from other bacteria and higher plants (Table 1).

DISCUSSION

In this report, DNA fragment No. 17 of a C. psittaci avian strain P-1041, was cloned and characterized for nucleotide sequencing and the expression of protein. The results show that the corresponding region of ORF of the P-1041 DNA was extensively and highly homologous to hypB genes of C. trachomatis serovar A and C. psittaci GPIC strain both in nucleotide and amino acid sequence.

Furthermore, it can be supposed that the cloned DNA of P-1041 is contained in chaperonin gene family known to code for the stress-response protein of other bacteria and plants. Chaperonin, protein product of this gene family, has been proposed to play a essential role for folding certain polypeptide and their correct assembling into oligomeric structure (3). Therefore, it may be possible that the cloned DNA fragment of P-1041 as well is related to the assembly of the protein during its unique developmental cycle.

The antigen coding region in the P-1041 DNA fragment was defined to locate from ca. 650 position to ca. 800 position in the nucleotide sequence by fragment deletion analysis. The product of the P-1041 DNA seemed to be type-specific, since the polypeptide had no cross-reactivity against 11 antisera to heterologous chlamydial strains including 9 C. psittaci and 2 C. trachomatis strains (11). On the other hand the HypB polypeptide was reported to be the chlamydial genus-common antigen (6, 7, 8). It may be possible that there are more than two epitopes in the HypB protein; a genus-common one, located in the region further upstream the cloned fragment, and a type-specific one, located in the cloned DNA of P-1041. Another possible explanation for antigenic specificity of the product is that the other common epitopes have been expressed, but are hidden by other proteins sterically. Deduced amino acid comparison showed that the variable region in amino acids was found at the region of 184 to 229 corresponding closely to 3' end of deleted fragment 79124. This region may be related to type-specific epitopes since the product of fragment 79110 was antigenic, while the fragment 79124, lacking the variable region, was not. Additionally, the P-1041 DNA did not have any putative transcriptional termination structure, nor the promoter site in the noncoding region between first and second putative ORFs. Therefore, the hypB-like first ORF and the second ORF may be cotranscribed as an operon in the P-1041 gene.

Since the DNA fragment of C. psittaci pigeon strain cloned in this study has a highly homologous sequence with hypB genes of two other chlamydial strains, it is necessary to examine whether the product is also responsible factor for the hypersensitivity or other pathological events of C. psittaci infection in avian hosts.
ACKNOWLEDGMENTS

We would like to thank Dr. Mitsuru Takanami, Institute for Chemical Reserch, Kyoto University, for his valuable advice and discussion.

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