



| | |
|------------------|--|
| Title | Influence of acute beetroot juice supplementation on cold-induced vasodilation and fingertip rewarming |
| Author(s) | Wakabayashi, Hitoshi; Sugiyama, Koji; Suzuki, Shinichi; Sakihama, Yasuko; Hashimoto, Makoto; Barwood, Martin J. |
| Citation | European journal of applied physiology, 123(3), 495-507 https://doi.org/10.1007/s00421-022-05071-6 |
| Issue Date | 2023-03 |
| Doc URL | http://hdl.handle.net/2115/90609 |
| Rights | This version of the article has been accepted for publication, after peer review (when applicable) and is subject to Springer Nature 's AM terms of use, but is not the Version of Record and does not reflect post-acceptance improvements, or any corrections. The Version of Record is available online at: http://dx.doi.org/10.1007/s00421-022-05071-6 |
| Type | article (author version) |
| File Information | Wakabayashi2022EJAP_beetroot.pdf |



[Instructions for use](#)

1 **Title**

2 Influence of acute beetroot juice supplementation on cold-induced vasodilation and fingertip rewarming

3

4 **Authors**

5 Hitoshi Wakabayashi^{1*}, Koji Sugiyama¹, Shinichi Suzuki¹, Yasuko Sakihama², Makoto Hashimoto²,

6 Martin J Barwood³

7

8 **Affiliations**

9 1 Laboratory of Environmental Ergonomics, Faculty of Engineering, Hokkaido University, Sapporo,

10 Japan

11 2 Laboratory of Molecular and Ecological Chemistry, Faculty of Agriculture, Hokkaido University,

12 Sapporo, Japan

13 3 Faculty of Social and Health Sciences, Leeds Trinity University, Leeds, UK

14

15 * Correspondence: Hitoshi Wakabayashi

16 e-mail: wakabayashi@eng.hokudai.ac.jp

17 ORCID: 0000-0003-0451-8758

18

19 **Abstract**

20 **Purpose:** Vasoactive ingredients in beetroot (BR) such as nitrate are known to induce vasodilation in
21 temperate conditions. This study investigated the effect of BR ingestion on cold induced vasodilation
22 (CIVD) and rewarming of finger skin temperature (T_{fing}) during and after hand immersion in cold water.

23 **Methods:** Twenty healthy males (mean \pm SD; age 22.2 ± 0.7 yrs, height 172.6 ± 6.0 cm, body mass 61.3 ± 11.7
24 kg) repeated a hand cold water immersion test twice with prior BR or water beverage ingestion (randomised
25 order). They rested for two hours in thermoneutral conditions (27°C , 40% relative humidity) after
26 consuming the beverage, then immersed their non-dominant hand in 8°C water for 30 min. They then
27 rewarmed their hand in the ambient air for 20 min. Skin temperature at seven body sites, T_{fing} , finger skin
28 blood flow ($SkBF_{\text{fing}}$), and blood pressure were measured.

29 **Results:** During hand immersion parameters of CIVD (T_{fing} and $SkBF_{\text{fing}}$) were not different between BR
30 and water conditions although skin temperature gradient from proximal to distal body sites was significantly
31 smaller with BR ($P < 0.05$). During rewarming, $SkBF_{\text{fing}}$ and cutaneous vascular conductance were
32 significantly higher with BR than with water ($P < 0.05$). The rewarming speed in T_{fing} and $SkBF_{\text{fing}}$ was
33 significantly faster with BR at 15- (BR 1.24 ± 0.22 vs water $1.11 \pm 0.26^{\circ}\text{C}/\text{min}$) and 20-min rewarming
34 ($P < 0.05$). Additionally, individuals with slower rewarming speed with water demonstrated accelerated
35 rewarming with BR supplementation.

36 **Conclusion:** BR accelerated rewarming in T_{fing} and $SkBF_{\text{fing}}$ after local cold stimulus, whereas, CIVD
37 response during hand cold immersion was not affected by BR ingestion.

38

39 **Keywords**

40 Nitrate, nitric oxide, rewarming speed, skin blood flow, red beet

41

42 **Abbreviations**

| | | |
|----|-------------------|--------------------------------|
| 43 | AVA | Arteriovenous anastomoses |
| 44 | CVC | Cutaneous vascular conductance |
| 45 | CIVD | Cold-induced vasodilation |
| 46 | MAP | Mean arterial blood pressure |
| 47 | NFCI | Non-freezing cold injury |
| 48 | NO_3^- | Nitrate |
| 49 | NO_2^- | Nitrite |
| 50 | NO | Nitric oxide |
| 51 | NOS | Nitric oxide synthase |
| 52 | ROS | Reactive oxygen species |
| 53 | $SkBF$ | Skin blood flow |
| 54 | T_{fing} | Finger skin temperature |

| | | |
|----|-----------------------|--|
| 55 | $T_{\text{pro-dis}}$ | Proximal to distal skin temperature difference |
| 56 | T_{subl} | Sublingual temperature |
| 57 | \bar{T}_{sk} | Mean skin temperature |
| 58 | | |

59 **Introduction**

60 In cold environments, cutaneous vasoconstriction is induced for maintaining homeostasis of core body
61 temperature. This vasomotor response for body temperature regulation results in the reduction of skin
62 temperature especially at distal extremities thereby widening the gradient between skin and core and
63 resulting in cooler extremities. Accordingly, the risk of frostbite and non-freezing cold injury in the distal
64 extremities have been reported in the workers in cold environments (e.g. fishery, military, and cold storage
65 workers), especially when they are continuously and longitudinally exposed to cold (Imray et al. 2009;
66 Makinen and Hassi 2009). Additionally, it is well known that performance of manual dexterity using fingers
67 and hands is impaired by the reduction of skin and subcutaneous tissue temperature resulting in impaired
68 motor coordination (Heus et al. 1995; Castellani and Tipton 2015; Wakabayashi et al. 2015), which could
69 be an additional injury risk factor for accidents in the workplace. Thus, wearing thermal protective clothing
70 and gloves are generally recommended for the workers in cold (Castellani et al. 2006; Holmer 2009).
71 However, wearing thick gloves can impair the manual dexterity performance especially when workers
72 conduct technical operations which need fine motor control of their fingers and hands (Brajkovic et al.
73 2001; Dianat et al. 2012). Therefore, some practically available alternative solutions for keeping warm
74 distal extremities are required for delicate manual work in cold environments in otherwise healthy
75 individuals.

76
77 Recently, supplementation with nitrate (NO_3^-) rich beetroot (BR) drink has been considered as an ergogenic
78 aid for enhancing blood circulation and exercise performance, especially in athletes, with promising results
79 (Wylie et al. 2013; Hoon et al. 2013; Dominguez et al. 2017). These studies have concurrently investigated
80 the effect of BR on the blood pressure and vasodilation response but only in thermoneutral or hot
81 environments (Wylie et al. 2013; Hobbs et al. 2013; Amano et al. 2018). Wylie et al. (2013) reported a dose
82 response relationship in reducing the oxygen cost of exercise with no improvement in exercise tolerance
83 above a dose of 140 mL (~ 8.4 mmol NO_3^-) in a thermoneutral environment. Hobbs et al. (2013), also in a
84 thermoneutral environment, demonstrated increase in the endothelium-independent vasodilation and
85 decreased diastolic blood pressure following beetroot infused bread ingestion. These effects were
86 concurrent with increased plasma and urinary nitrate. Amano et al. (2018) conducted their study in hot
87 conditions (30°C, 50% relative humidity) but only showed changes in mean arterial pressure but not skin
88 blood flow or cutaneous vascular conductance. Compared to studies in warm or thermoneutral
89 environments, fewer studies have investigated the effect of BR on peripheral circulation and distal skin
90 temperature during and after local cold exposure (Eglin et al. 2017; Shepherd et al. 2019; Wickham et al.
91 2021). This is surprising given that an endothelial-independent NO donor is known to increase vasodilation
92 as demonstrated following glyceryl trinitrate (GTN) ingestion in cold-sensitive individuals (Hope et al.
93 2014) and topical GTN application in patients with Raynaud's phenomenon (Anderson et al. 2002)
94 suggesting a plausible role for NO in improving peripheral re-warming. Considering the evidence of

95 cutaneous vasodilation with dietary ingested BR primarily in thermoneutral environments and in persons
96 with cold injury, this intervention could also be applied for maintaining warm finger skin temperature in a
97 cold environment which may also translate to an occupational benefit in improving manual dexterity.

98

99 Wickham et al. (2021) investigated the effect of acute BR supplementation on the cold-induced vasodilation
100 (CIVD) in finger skin temperature and blood flow *during* hand cold-water (8°C) immersion in ten healthy
101 males. They found no difference in any CIVD parameters between conditions with BR and NO₃⁻ depleted
102 placebo drink. Accordingly, they suggested only a minor contribution of nitric oxide (NO) as a mechanism
103 for the CIVD response (Wickham et al. 2021). Shephard et al. (2019) examined the effect of acute and
104 chronic BR supplementation on vasomotor regulation in a cohort of cold-sensitive elderly people
105 (64.3±15.3 yrs) with Raynaud's syndrome (i.e., recurrent transient vasospasm of the fingers and/or toes in
106 response to a cold or stressful stimulus (Wigley 2002)). They reported that both chronic BR
107 supplementation and chronic nitrate depleted BR juice supplementation enhanced skin blood flow (SkBF)
108 during 10-min rewarming phase but in the thumb only following local transient cold stimulus (2-min cold
109 water immersion, 15°C) compared to the baseline no supplementation trial. While, noteworthy there were
110 no other differences between BR and NO₃⁻ depleted placebo drink. Based on the results, they suggested
111 that some of the vasoactive ingredients in BR, other than NO₃⁻, like betanin, quercetin, and chlorogenic
112 acid (Wootton-Beard et al. 2011) might be a factor for enhancing vasodilation after cold stimulus (Shepherd
113 et al. 2019). Importantly these vasoactive substances could be common to both the depleted NO₃⁻ test
114 supplementation conditions (i.e., the placebo controls) and the BR supplementation conditions used in most
115 studies. Hence, tests using a control condition to effectively separate and distinguish the effects of BR
116 supplementation is warranted. Consistent with this idea, Thompson et al. (2018) compared the physiological
117 effects of BR juice with potassium nitrate (KNO₃) supplementation containing similar amounts of NO₃⁻
118 (Thompson et al. 2018). They found lower resting blood pressure with ingestion of BR compared to KNO₃,
119 which suggested ingredients other than NO₃⁻ in BR might be responsible for improving the bioavailability
120 of NO; yet the study of Thompson et al (2018) was focussed on enhancing sprint interval training
121 performance. Whilst theoretically sound the evidence for the efficacy of BR supplementation in expediting
122 the rewarming of the extremities during and following cold exposure in a healthy population is equivocal
123 although further protocol manipulations are required to explore the putative effects.

124

125 With the controls used and findings revealed from previous studies, it remains possible that BR
126 supplementation as a sole ingredient might enhance the CIVD response during local cold exposure and
127 accelerate the subsequent rewarming, in comparison to a control absent of NO₃⁻ and other vasoactive
128 ingredients (i.e., water ingestion). Accordingly, the purpose of this study is to investigate the effect of acute
129 BR ingestion on CIVD response during hand cold immersion and subsequent rewarming, compared to a
130 control condition (water) in healthy young individuals. It was hypothesized that acute BR supplementation

131 would enhance the CIVD response during hand cold immersion, and finger skin blood flow and skin
132 temperature rewarming after the immersion. With occupational cold exposure often far longer than the short
133 exposure windows used in prior studies (e.g. Shephard et al. 2019 used 2-min immersion), we also sought
134 to examine an extended period of cold exposure (matching that of Wickham et al. 2021; 30-min) coupled
135 with an extended period of re-warming (i.e., 20-min). Lastly and from a practical perspective, the individual
136 variation in the response to BR ingestion was quantified to enable targeted future intervention toward cold
137 sensitive individuals.

138

139 **Methods**

140 ***Participants***

141 The experimental protocol was approved by the IRB of Hokkaido University. All participants were
142 informed of the experimental protocols and gave their written informed consent before participation.
143 Twenty healthy Japanese males living in Sapporo (mean \pm standard deviation age: 22.2 \pm 0.7 yrs, height:
144 172.6 \pm 6.0 cm, body mass: 61.3 \pm 11.7 kg, % body fat: 15.2 \pm 5.0%) participated in the experiment. Their
145 percentages of body fat were estimated using bioelectrical impedance (RD-800, TANITA, Japan). All
146 experimental protocols in this study were designed according to the principle of the Helsinki Declaration.
147 Participants were asked to prohibit eating nitrate (NO₃⁻) rich foods, e.g. processed meats, green leaf
148 vegetable like Spinach, Chin gin cai, Seaweed, Sayaingen beans (Sobko et al. 2010) on the test day and the
149 day before. Additionally, they fasted for 2 hours before arriving the laboratory and were asked to refrain
150 using mouth rinse on the test day, since the oral bacteria are involved in the reduction of NO₃⁻ to NO₂⁻
151 (Govoni et al. 2008).

152

153 ***Experimental Design***

154 Participants completed a total of two test conditions separated by a minimum of 7 days to enable washing
155 out of the BR effect (Amano et al. 2018; Shepherd et al. 2019). On each occasion hand immersion in to
156 cold water and a rewarming test was completed with prior ingestion of beetroot (BR) or water as a
157 representative control. The order of the test conditions was randomised using crossover design.

158

159 ***Protocol***

160 Participants arrived at the laboratory and changed their clothes to half sleeve shirts, long pants and socks
161 (insulation \sim 0.6 clo). They then rested in upright sitting position on a chair with their arms on a table in a
162 climatic chamber controlled to thermoneutral conditions (27°C and 40% relative humidity) for 30 min
163 before drinking 140 mL of water or 140 mL of BR (Beet It Sport Pro-Elite Shot, James White Drinks,
164 Ipswich, UK), which were maintained at room temperature (\sim 27°C). The BR drink contains \sim 12.9 mmol
165 (800 mg) NO₃⁻ and 0.28 mmol (154 mg) betanin. The concentration of betanin was calculated from
166 absorbance measured with a UV/vis spectrophotometer (U-3310, Hitachi, Japan) using the molar extinction

167 coefficient $\epsilon_{538}=60,000 \text{ M}^{-1}\text{cm}^{-1}$ (Wylter and Meuer 1979; Kugler et al. 2004). After the beverage ingestion,
168 they continued resting for two hours, which has previously shown to be a sufficient time course to increase
169 plasma NO_2^- after drinking BR containing 4.2 to 16.8 mmol NO_3^- (Wylie et al. 2013). Then, after measuring
170 pre-immersion baseline for the measurement items described below, they immersed their non-dominant
171 hand covered by a waterproof polyethylene glove (12 μm thickness) in 8°C water up to their wrist for 30
172 min. The water in the tank was stirred and temperature controlled using a thermostat water circulation
173 device (LV-200, Advantec, Japan); water temperature was monitored at the start and end of each immersion.
174 After the 30-min hand water immersion, they removed the glove and rewarmed their hand in lateral position
175 on the table for 20 min.

176

177 *Measurements*

178 Sublingual temperature (T_{subl}) was measured at baseline, at the end of the hand immersion and rewarming
179 phases using a thermometer (MC-172L, Omron, Japan). Participants were asked to place the tip of the
180 thermometer below the tongue and to close their mouth for 5 minutes until the measurement stabilized.
181 Skin temperature was measured using thermistor probes (ITP082-24, Nikkiso-Therm, Japan) at seven body
182 sites (forehead, chest, forearm, thigh, foot, non-immersed hand and immersed fingertip). The skin
183 temperatures were monitored every second using data loggers (NR543R, Nikkiso-Therm, Japan), and
184 averaged every minute for subsequent data analyses. Mean skin temperature (\bar{T}_{sk}) was estimated using a
185 modified Hardy and DuBois' equation (Hardy and Du Bois 1938), as follow:

186

187 Equation 1
$$\bar{T}_{\text{sk}} = 0.07T_{\text{head}} + 0.35T_{\text{chest}} + 0.14T_{\text{forearm}} + 0.05T_{\text{hand}} + 0.32T_{\text{thigh}} + 0.07T_{\text{foot}}$$

188

189 Where: T_{chest} was the selected site for trunk temperature and T_{thigh} included the additional 0.13 weighting
190 ordinarily allocated to T_{legs} (not recorded in the current study) from the original \bar{T}_{sk} formula.

191

192 The difference between proximal (average of forehead and chest) and distal (average of hand and foot) skin
193 temperatures ($T_{\text{pro-dis}}$) were calculated as surrogate measure for assessing peripheral blood flow (Rubinstein
194 and Sessler 1990). Since vasoconstriction occurs remarkably in the distal part relative to the proximal part,
195 $T_{\text{pro-dis}}$ well reflects the peripheral vasomotor tone, although there is a limitation that $T_{\text{pro-dis}}$ takes longer
196 time to reach steady state compared to the vasomotor response (Rubinstein and Sessler 1990; House and
197 Tipton 2002). This method has been verified during cooling and rewarming (House and Tipton 2002) and
198 also been used in studies on circadian rhythm as a parameter of distal heat loss (Krauchi et al. 1999). Skin
199 blood flow in the volar side of index finger ($SkBF_{\text{fing}}$) was measured by laser Doppler flowmetry (ALF21,
200 ADVANCE, Japan) and sampled using an analogue to digital data converter (Powerlab/16SP, AD
201 Instruments, Australia) and recorded every 1 sec interval using a laptop computer. Arbitrary units (AU)
202 were used for the data of $SkBF_{\text{fing}}$. Systolic (SBP) and diastolic blood pressure (DBP) was measured at the

203 upper (contralateral) arm using an inflatable cuff and an automated blood pressure monitor (HEM-7430,
204 Omron, Japan) every 5 min during the rewarming phase. Mean arterial blood pressure (*MAP*) was calculated
205 using the following formula:

206

207 Equation 2 $MAP \text{ [mmHg]} = (SBP - DBP) / 3 + DBP$

208 Cutaneous vascular conductance in finger (*CVC_{finger}*) was calculated from *SkBF_{finger}* and *MAP* as follows:

209

210 Equation 3 $CVC_{\text{finger}} \text{ [AU/mmHg]} = SkBF_{\text{finger}} / MAP$

211

212 Parameters of CIVD were analysed in accordance with (Cheung 2015) where a minimum increase of 0.5°C
213 in *T_{finger}* was required and the associated change in *SkBF_{finger}* during the hand cold water immersion was then
214 considered. The onset time of the CIVD, the minimal (*T_{min}*), the first peak (*T_{peak}*), and maximal (*T_{max}*) *T_{finger}*
215 was detected, then the amplitude from *T_{min}* to *T_{peak}* (*T_{peak}* - *T_{min}*) and mean value of *T_{finger}* after 5 min to the
216 end of the immersion (mean *T_{finger}*) were calculated. Additionally, the numbers of CIVD oscillations were
217 counted. These CIVD parameters for *SkBF_{finger}* were similarly analysed.

218

219 Rewarming speed in *T_{finger}* and *SkBF_{finger}* was calculated from 1 min to 5, 10, 15, and 20 min after hand
220 immersion, respectively, e.g. *T_{finger}* rewarming speed in 10 min was calculated as follows:

221

222 Equation 4 $T_{\text{finger}} \text{ rewarming speed in 10 min [}^\circ\text{C/min]} = (T_{\text{finger}} \text{ at 10 min} - T_{\text{finger}} \text{ at 1 min}) / (10 - 1)$

223

224 Thermal sensation of whole-body and immersed hand was assessed using a 7-points categorical scale (-3:
225 cold, -2: cool, -1: slightly cool, 0: neither, +1: slightly warm, +2: warm, +3: hot) every 5 min during the
226 experiment. Thermal comfort was assessed using a 7-points scale (-3: very uncomfortable, -2:
227 uncomfortable, -1: slightly uncomfortable, 0: neither, +1: slightly comfortable, +2: comfortable, +3: very
228 comfortable) every 5 min. Pain sensation of the immersed hand was assessed using 4-points scale (0: no
229 pain, 1: slightly painful, 2: painful, 3: very painful) every 5 min.

230

231 **Statistics**

232 Comparisons of datasets from time-course measurements every 5 min were performed using repeated two-
233 way (time × condition) analysis of variance (ANOVA) for each phase of hand cold immersion and
234 rewarming. If Mauchly's sphericity test was not satisfied, the degrees of freedom were adjusted by
235 Greenhouse-Geisser's ε. Partial η^2 (η_p^2) was calculated for assessing effect size for ANOVA where 0.01,
236 0.06 and 0.14 indicate small, medium and large effect sizes, respectively. Post-hoc test was conducted using
237 a paired Student's *t*-test with multiple comparisons adjustment using Benjamin-Hochberg's false discovery
238 rate (FDR) at time points between water and beetroot conditions. CIVD parameters of *T_{finger}* and *SkBF_{finger}* in

239 two conditions were compared using a paired Student's *t*-test. Pearson's correlation coefficients and 95%
240 confidence interval (CI) for the slope of the regression line were calculated to examine the relationships
241 between rewarming speeds in the two conditions. Wilcoxon signed-rank test was conducted for comparing
242 subjective sensation between conditions. Statistical significance was set at $P<0.05$. Analyses were
243 conducted using a statistical software (IBM SPSS Statistics version 20, IBM). All data are presented as
244 mean values and standard deviation (SD).

245

246 **Results**

247 *Hand cold immersion phase*

248 Time course of T_{fing} and $SkBF_{\text{fing}}$ in the immersed hand are shown in **Fig. 1**. A significant main effect of
249 time was detected in T_{fing} ($F_{3,2, 61.1}=3803.2, \eta_p^2=0.995, P<0.001$) and $SkBF_{\text{fing}}$ ($F_{2,1, 40.0}=102.2, \eta_p^2=0.843,$
250 $P<0.001$). Parameters of CIVD in T_{fing} and $SkBF_{\text{fing}}$ are presented in **Table 1**. There was no significant
251 difference between conditions in any parameters of CIVD.

252 Time course of $\bar{T}_{\text{sk}}, T_{\text{hand}}$ on non-immersed body region and difference between proximal and distal skin
253 temperatures ($T_{\text{pro-dis}}$) are shown in **Fig. 2**. During the hand immersion phase, a significant main effect of
254 time was detected in \bar{T}_{sk} ($F_{2,5, 47.5}=19.6, \eta_p^2=0.507, P<0.001$), T_{hand} ($F_{2,1, 39.6}=65.4, \eta_p^2=0.775, P<0.001$), and
255 $T_{\text{pro-dis}}$ ($F_{1,9, 36.6}=133.3, \eta_p^2=0.875, P<0.001$). A significant main effect of condition was detected in T_{hand} ($F_{1,$
256 $19}=6.0, \eta_p^2=0.239, P<0.05$) and $T_{\text{pro-dis}}$ ($F_{1, 19}=6.4, \eta_p^2=0.253, P<0.05$). $T_{\text{pro-dis}}$ in BR was significantly smaller
257 than in water condition at 20- and 25-min hand immersion (both $P<0.05$) primarily due to higher
258 temperatures at the distal site. No difference in T_{subl} between water and BR conditions was observed at the
259 baseline (36.60 ± 0.30 and $36.66\pm 0.34^\circ\text{C}$) and at the end of hand immersion (36.58 ± 0.34 and $36.66\pm 0.33^\circ\text{C}$).
260 Time course of thermal sensation of the whole-body and immersed hand, thermal comfort and pain
261 sensation of the immersed hand are shown in **Fig. 3**. No difference in these subjective sensations was
262 observed between conditions during hand immersion.

263

264 *Rewarming phase*

265 Time course of T_{fing} and $SkBF_{\text{fing}}, CVC_{\text{fing}}$ in the immersed hand, and MAP during the rewarming phase is
266 shown in **Fig. 4**. A significant main effect of time was detected in T_{fing} ($F_{1,8, 34.0}=379.3, \eta_p^2=0.952, P<0.001$),
267 $SkBF_{\text{fing}}$ ($F_{2,3, 43.4}=55.1, \eta_p^2=0.744, P<0.001$), CVC_{fing} ($F_{2,5, 47.9}=53.8, \eta_p^2=0.739, P<0.001$), and MAP ($F_{2,9,$
268 $54.7}=4.8, \eta_p^2=0.202, P<0.01$). A significant main effect for condition was detected in CVC_{fing} ($F_{1, 19}=4.4,$
269 $\eta_p^2=0.189, P<0.05$). A significant interaction between time and condition were detected in T_{fing} ($F_{2,3, 44.6}=5.5,$
270 $\eta_p^2=0.224, P<0.01$), $SkBF_{\text{fing}}$ ($F_{2,8, 53.3}=4.9, \eta_p^2=0.204, P<0.01$), CVC_{fing} ($F_{2,8, 53.1}=4.2, \eta_p^2=0.182, P<0.05$),
271 and MAP ($F_{2,5, 48.1}=3.3, \eta_p^2=0.148, P<0.05$). $SkBF_{\text{fing}}$ in BR condition was significantly higher than in the
272 water condition at 45- and 50-min rewarming ($P<0.05$). CVC_{fing} in BR condition was significantly higher
273 than in water condition at 40- to 50-min rewarming ($P<0.05$).

274 Time course of \bar{T}_{sk} , T_{hand} and $T_{pro-dis}$ on non-immersed body region are shown in **Fig. 2**. During the
275 rewarming phase, a significant main effect of time was detected in \bar{T}_{sk} ($F_{2,1, 40.1}=27.6, \eta_p^2=0.592, P<0.001$)
276 and $T_{pro-dis}$ ($F_{1.5, 28.6}=3.7, \eta_p^2=0.163, P<0.05$). A significant main effect of condition was detected in $T_{pro-dis}$
277 ($F_{1, 19}=5.8, \eta_p^2=0.235, P<0.05$). $T_{pro-dis}$ in BR condition was significantly smaller than the water condition
278 at 30- to 50-min rewarming ($P<0.05$). No difference in T_{subl} between water and BR conditions (36.52 ± 0.33
279 and $36.60\pm 0.35^\circ\text{C}$) was observed at the end of rewarming phase.

280

281 The rewarming speeds in T_{fing} from 1 min to 5, 10, 15, and 20 min after hand immersion are presented in
282 **Table 2**. Significantly faster rewarming was observed in beet condition compared to water at 15- and 20-
283 min rewarming (both $P<0.05$). Similarly, the recovery speed in $SkBF_{fing}$ was significantly faster in BR
284 condition at 15- and 20-min rewarming (both $P<0.05$).

285

286 The individual values of T_{fing} rewarming speeds in water and BR conditions are plotted in **Fig. 5**. Significant
287 correlations were evident between conditions in 10, 15, and 20 min rewarming ($r=0.69, P<0.01$; $r=0.68,$
288 $P<0.01$; $r=0.58, P<0.01$; respectively). The slope of the regression lines (95% CI) are 0.60 (0.28-0.91), 0.57
289 (0.27-0.88), and 0.43 (0.13-0.73) at 10, 15, and 20 min, respectively. The slopes of the regression lines (and
290 95% CI) were gentler compared to the $y = x$ (reference line), which represents identical rewarming speed
291 in water and BR conditions. Thus, the individuals with slower rewarming speed with water showed greater
292 improvement in rewarming with BR supplementation.

293

294 Time course of thermal sensation of the whole-body and immersed hand, thermal comfort and pain
295 sensation of the immersed hand are shown in **Fig. 3**. No difference in these subjective sensations was
296 observed between conditions during the rewarming phase.

297

298 **Discussion**

299 This study investigated the effect of BR ingestion on CIVD response and rewarming of T_{fing} during and
300 after hand immersion in cold water, by comparing with drinking water as a control. In contrast to our
301 hypothesis for the immersion phase of the experiment, CIVD in T_{fing} and skin blood flow was not affected
302 by BR. On the other hand, as a major finding of this study, rewarming of T_{fing} and skin blood flow was
303 accelerated by drinking BR 2-hours before hand immersion. Hence, the hypothesis is only partially
304 supported.

305

306 ***Cold-induced vasodilation***

307 We originally hypothesised that CIVD would be enhanced by ingestion of BR, since it contains vasoactive
308 ingredients such as NO_3^- , betanin, and chlorogenic acid (Wootton-Beard et al. 2011). However, the results
309 showed no significant difference in all parameters of CIVD for T_{fing} and $SkBF_{fing}$ between BR and water

310 conditions during immersion. Additionally, thermal sensation and pain sensation of the immersed hand was
311 not different between conditions.

312

313 Starting from the original observation of the hunting reaction of finger temperature to cold (Lewis 1930),
314 the CIVD response has been studied in humans (Daanen and Ducharme 1999) and animals including
315 adrenergic neural mechanism in isolated vascular smooth muscle (Rusch et al. 1981). Based on the current
316 knowledge of the CIVD, NO-dependent active vasodilation and/or sympathetic withdrawal has been
317 suggested as potential mechanisms of CIVD response (Daanen 2003; Cheung 2015). In this study,
318 considering the significantly smaller $T_{\text{pro-dis}}$ in BR condition during the latter half of the hand immersion,
319 whole-body vasodilation appears to be enhanced with BR supplementation during a prolonged (compared
320 to other studies; e.g., Eglin et al., 2017; Hope et al., 2014) cold stimulus of the extremity. However, finger
321 skin temperature and blood flow in the immersed hand did not show any effect of BR supplementation.
322 This result was in line with the previous finding of no difference in CIVD response during hand immersion
323 to 8°C water between BR and NO₃⁻-depleted placebo drink (Wickham et al. 2021). NO-mediated active
324 vasodilation probably has a minor contribution to enabling the CIVD response when cold stimulus from
325 the extremities is significant whilst also coupled with the effect of hydrostatic squeeze on the surface blood
326 vessels of the immersed hand in healthy individuals. In persons with cold injury such as Raynaud's
327 syndrome or those who are cold-sensitive with an abnormal endothelial function, NO supplementation is
328 more likely to influence the vasomotor responses during cold challenge but not by a large magnitude (e.g.,
329 Shepherd et al. 2019; Hope et al. 2014). Hope et al. (2014) suggested that GTN, acting experimentally as
330 the NO donor, bypasses the endothelium dependent NO pathway to re-establish the vascular response in
331 cold-sensitive individuals. Hope et al. (2014) only observed facilitated post-immersion re-warming in their
332 cold-sensitive group but not in their healthy controls. Shepherd et al. (2019) showed some transient
333 evidence of increased CVC with chronic supplementation with BR and nitrate depleted BR but not acute
334 supplementation indicating a greater dose of BR than used here might be needed to induce CIVD during
335 cold immersion. Collectively, there is now a growing body of evidence that suggests acute BR
336 supplementation doesn't have the potency to evoke vasodilation during local cold challenge in healthy
337 (Wickham et al. 2021) and cold-sensitive/injured individuals (Eglin et al. 2013; Shepherd et al. 2019). Thus,
338 it is speculated that release of vasoconstrictor tone might be one of the major mechanisms for the CIVD
339 response rather than NO-dependent active vasodilation. On the other hand, systemic effects of BR
340 supplementation on blood pressure (Shepherd et al. 2019; Wickham et al. 2021) are plausible, as shown in
341 the observation of higher distal skin temperature during prolonged hand immersion to this body of evidence
342 (see Fig. 2). Therefore, BR could also evoke vasodilatory effects in nonglaborous regions where
343 noradrenergic vasoconstrictor nerves and cholinergic active vasodilator nerves are active (Kellogg 2006).
344 Nevertheless, our observations of accelerated rewarming following cold exposure are novel and require
345 further exploration.

346

347 *Acceleration of rewarming*

348 The major finding of this study was that BR ingestion accelerated the rewarming speed of T_{fing} and finger
349 skin blood flow after 30-min hand cold immersion. During the rewarming phase, after terminating the cold
350 stimulus, the vasoactive ingredients in BR enhanced the whole-body and local vasodilation but only after
351 10 ($SkBF_{\text{fing}}$) to 15 minutes (T_{fing}) of rewarming (Table 2). The duration of rewarming studied here might
352 also be another reason why prior studies have not revealed this difference having primarily measured for
353 up to 10-min of rewarming (Eglin et al. 2017; Shepherd et al. 2019). It is evident from the present data that
354 the experimental effects of BR probably extend beyond the 20-min rewarming period where these
355 significant differences remained. Although rewarming speed in $SkBF_{\text{fing}}$ and T_{fing} was accelerated with BR,
356 it was not associated with perception of local thermal sensation or whole-body thermal comfort.

357

358 One of the most potent vasoactive ingredients in BR is NO_3^- , with many studies reporting NO_3^- - NO_2^- - NO
359 pathway induced vasodilation and consequently lowered blood pressure through exogenous NO mediated
360 relaxation of endothelial cells to relax vascular smooth muscle (Wylie et al. 2013; Lara et al. 2016; Richards
361 et al. 2018). Unlike the present study, previous studies reported no effect of acute BR ingestion on
362 peripheral vasodilatory response following 2-min cold immersion of extremities in comparison with NO_3^- -
363 depleted placebo (Eglin et al. 2017; Shepherd et al. 2019). The short duration (2 min) cold immersion in
364 the previous studies and prolonged (30 min) cold exposure in the present study are categorized into early-
365 phase (i.e., primarily skin cooling) and late-phase local cooling (i.e., skin and superficial muscle cooling),
366 respectively (Hodges and Johnson 2009; Alba et al. 2019). At the onset of the local cold stimulus, early
367 vasoconstriction is induced mostly via adrenergic and neural mechanisms (Ekenvall et al. 1988; Stephens
368 et al. 2004), whereas later vasoconstriction during prolonged cooling is mediated via combination of
369 continued vasoconstrictor nerve excitation and inhibition of NO vasodilator pathway (Hodges et al. 2006;
370 Hodges and Johnson 2009; Alba et al. 2019). It is likely that there is a more substantial reduction in
371 bioavailable NO at the end of 30-min hand immersion in the present study due to the decreased activity of
372 endothelial NOS and downstream of NOS (Hodges et al. 2006) and hence this is the reason for the efficacy
373 of increasing NO in the present study. The likely reduction of NO bioavailability during longer cold
374 exposure enabled us to find a significant vasodilatory effect of BR, as external NO donor via NO_3^- - NO_2^- -
375 NO pathway, accelerating T_{fing} rewarming following the cold exposure. Yet, the time course of our $SkBF_{\text{fing}}$
376 and T_{fing} (**Table 2**) data during rewarming along with the magnitude of T_{fing} change (**Fig 4**) suggest plausible
377 successive relief to both mechanisms (i.e., relief of vasoconstrictor nerve excitation and restoration of the
378 NO vasodilator pathway; Hodges et al. 2006; Hodges and Johnson 2009; Alba et al. 2019) that are
379 associated with prolonged cooling. $SkBF_{\text{fing}}$ laser doppler flowmetry data from Hodges et al. (2018) during
380 30-min hand immersion indicate the onset of the CIVD response precedes increases in T_{fing} (i.e., the same
381 as our data) but closely matched the measured neurogenic activity in the finger but not the endothelial nitric

382 oxide dependent or independent activity (Hodges et al. 2018). The changes seen in T_{fing} in the study of
383 Hodges et al. (2018) were small and transient due to the ongoing cold-water immersion. During rewarming
384 in the present study, we suggest the BR supplementation facilitated the earlier onset of vasoconstrictor
385 nerve relief, facilitating an increase in $SkBF_{\text{fing}}$ at 10-min of rewarming and accelerating the increase in T_{fing}
386 from 15-minutes. This increase in $SkBF_{\text{fing}}$ raises T_{fing} towards the vasomotor range (i.e., 26°C, (Folkow
387 and Neil 1971); see **Fig 4**) where further active vasodilation is plausible (Kellogg et al. 1998; Kellogg 2006).
388

389 The initial reduction of NO bioavailability, including the inhibition of NOS activity, is partly induced by
390 the elevation of reactive oxygen species (ROS) from vascular smooth muscle mitochondria (Bailey et al.
391 2005; Holowatz and Kenney 2007; Hodges and Johnson 2009; Johnson et al. 2014). In addition, the ROS
392 generated by local cooling also enhances Rho-kinase activity to increase vascular tone to the neural
393 noradrenalin release, which is mainly explained by the translocation of α_{2c} -adrenoreceptors to the surface
394 of vascular smooth muscle cell (Bailey et al. 2004; Bailey et al. 2005; Hodges and Johnson 2009). The Rho-
395 kinase pathway is relatively slow event as shown in time course of Rho activity to cooling in human cultured
396 dermal arteriolar vascular smooth muscle (Bailey et al. 2004). Thus, the enhanced vasodilatory response
397 with BR ingestion after prolonged cold exposure might also be related to slow reversal of the ROS and
398 Rho-kinase pathway which fits with the time course of responses shown in the present study. Previous
399 research investigating the vasomotor effect of BR supplementation suggested the nitrate-independent
400 vasodilatory response due to other bioactive ingredients in BR (Bahadoran et al. 2017; Thompson et al.
401 2018; Shepherd et al. 2019). Among them betanin was focused on as one of the phytochemical antioxidants
402 in BR (Esatbeyoglu et al. 2015; Hadipour et al. 2020). A study reported that local administration of
403 ascorbate antioxidant inhibited the vasoconstriction during local skin cooling (Yamazaki 2010). Thus, it
404 was suggested that quenching of ROS by the antioxidants might decrease efficacy of adrenoreceptors and
405 influence the vascular response to cooling. The antioxidant, betanin contained in BR, inhibits the diffusion-
406 controlled reaction of NO with superoxide (i.e., an ROS) by scavenging super-oxide radicals that create
407 peroxynitrite thereby slowing the appearance of this ROS and improving the bio-availability of NO
408 (Sakihama et al. 2012). Hence, consumption of betanin is a plausible means to increase bioavailable NO
409 for stimulating vasodilation (Esatbeyoglu et al. 2015). In the present study, 30-min hand immersion in 8°C
410 was probably sufficient oxidative stress to decrease bioavailable NO (Christmas et al. 2016). Following this
411 cold stress, the combined effect of nitrate (NO production) and betanin (antioxidation) in BR could increase
412 bioavailable vasodilatory NO, which could plausibly enhance T_{fing} rewarming compared to the water
413 ingestion condition following initial relief to vasoconstrictor nerve activity. It is a limitation of the present
414 study that we cannot discern the separate contribution of the bioactive ingredients (e.g., nitrate and betanin)
415 in BR that contribute to the responses we report and that we could not blind the treatment conditions;
416 although it did not evoke significant change in thermal sensation and comfort, both scenarios require further
417 research. Lastly, our observations are restricted to males only.

418

419 ***Individual variation in the effect of beetroot supplementation***

420 There are individual variations in vasomotor response to local cooling. Cold sensitive individuals such as
421 with non-freezing cold injury (NFCI) and Raynaud's syndrome present with colder hands and foot skin
422 temperature, greater vasoconstriction to cold and slower rewarming rate following cold exposure compared
423 to normal individuals (Eglin et al. 2013; O'Reilly et al. 1992). Patients with Raynaud's phenomenon show
424 deficiency of NO in response to cold stimulus (Tanaka et al. 2012), which could be due to their greater
425 oxidative damage resulting from higher serum ROS level (Biondi et al. 2008). It was also reported that
426 increased ROS in response to cooling might be one of the mediators for tissue damage in NFCI (Geng et
427 al. 2015). Moreover, even in healthy people with cold constitution ("hi-e-sho" in Japanese), greater
428 vasoconstriction was provoked to local skin cooling and iontophoretic noradrenaline application (Yamazaki
429 2015). These findings indicated that cold sensitive individuals would have an increased sensitivity of
430 adrenoceptor on vascular smooth muscle and/or decreased bioavailability of NO, that has been shown to
431 evoke oxidative stress (Biondi et al. 2008).

432

433 In the present study, individual variation in healthy participants was observed in the rewarming speed after
434 30-min cold exposure. The slopes of the regression lines between the rewarming speed in water and BR
435 conditions were gentler compared to the $y = x$, which represents identical speed in both conditions. This
436 result indicated that cold sensitive individuals with slower rewarming in the water condition showed more
437 remarkable improvement in the rewarming speed with BR ingestion. We speculate that cold sensitive
438 individuals, who potentially have more oxidative stress and less bioavailable NO, could benefit more so
439 following BR supplementation containing antioxidants like betanin and NO_3^- as NO donor; a focus of future
440 research.

441

442 **Conclusions**

443 This study investigated the effect of beetroot supplementation on vasomotor responses during and after
444 hand cold immersion in human. CIVD in finger temperature and skin blood flow was not affected by BR
445 ingestion, but a significantly smaller $T_{\text{pro-dis}}$ in BR condition, primarily due to higher distal temperature,
446 would indicate that whole-body vasodilation was enhanced during the local cold stimulus. The major
447 finding of this study was that BR accelerated rewarming in finger skin temperature and skin blood flow
448 after the local cold stimulus. Additionally, cold sensitive individuals with a slower rewarming rate with
449 water demonstrated more remarkable acceleration in rewarming with BR supplementation.

450

451 **Declarations**

452 **Funding** This study was supported by Hokkaido University Research and Education Center for Robust
453 Agriculture, Forestry and Fisheries Industry.

454

455 **Conflicts of interest** None declared.

456

457 **Ethics approval** All experimental protocols in this study were approved by the IRB of Hokkaido
458 University.

459

460 **Author contributions** HW and MB conceived and designed research. KS and SS conducted the
461 experiments. KS and HW analyzed data. HW and MB wrote the draft of manuscript. All authors read and
462 approved the manuscript.

463

464 **Acknowledgments** The authors wish to thank all those who participated in this study.

465

466 **References**

467 Alba BK, Castellani JW, Charkoudian N (2019) Cold-induced cutaneous vasoconstriction in humans:
468 Function, dysfunction and the distinctly counterproductive. *Exp Physiol* 104 (8):1202-1214.
469 doi:10.1113/EP087718

470 Amano T, Okushima D, Breese BC, Bailey SJ, Koga S, Kondo N (2018) Influence of dietary nitrate
471 supplementation on local sweating and cutaneous vascular responses during exercise in a hot
472 environment. *Eur J Appl Physiol* 118 (8):1579-1588. doi:10.1007/s00421-018-3889-9

473 Anderson ME, Moore TL, Hollis S, Jayson MIV, King TA, Herrick AL (2002) Digital vascular response to
474 topical glyceryl trinitrate, as measured by laser Doppler imaging, in primary Raynauds
475 phenomenon and systemic sclerosis. *Rheumatology* 41 (3):324-328. doi:DOI
476 10.1093/rheumatology/41.3.324

477 Bahadoran Z, Mirmiran P, Kabir A, Azizi F, Ghasemi A (2017) The Nitrate-Independent Blood Pressure-
478 Lowering Effect of Beetroot Juice: A Systematic Review and Meta-Analysis. *Adv Nutr* 8 (6):830-
479 838. doi:10.3945/an.117.016717

480 Bailey SR, Eid AH, Mitra S, Flavahan S, Flavahan NA (2004) Rho kinase mediates cold-induced
481 constriction of cutaneous arteries: role of alpha2C-adrenoceptor translocation. *Circ Res* 94
482 (10):1367-1374. doi:10.1161/01.RES.0000128407.45014.58

483 Bailey SR, Mitra S, Flavahan S, Flavahan NA (2005) Reactive oxygen species from smooth muscle
484 mitochondria initiate cold-induced constriction of cutaneous arteries. *Am J Physiol Heart Circ*
485 *Physiol* 289 (1):H243-250. doi:10.1152/ajpheart.01305.2004

486 Biondi R, Coaccioli S, Lattanzi S, Puxeddu A, Papini M (2008) Oxidant/antioxidant status in patients with
487 Raynauds disease. *Clin Ter* 159 (2):77-81

488 Brajkovic D, Ducharme MB, Frim J (2001) Relationship between body heat content and finger temperature
489 during cold exposure. *J Appl Physiol* 90 (6):2445-2452

490 Castellani JW, Tipton MJ (2015) Cold stress effects on exposure tolerance and exercise performance.
491 *Compr Physiol* 6 (1):443-469. doi:10.1002/cphy.c140081

492 Castellani JW, Young AJ, Ducharme MB, Giesbrecht GG, Glickman E, Sallis RE, American College of
493 Sports M (2006) American College of Sports Medicine position stand: prevention of cold injuries
494 during exercise. *Med Sci Sports Exerc* 38 (11):2012-2029.
495 doi:10.1249/01.mss.0000241641.75101.64

496 Cheung SS (2015) Responses of the hands and feet to cold exposure. *Temperature (Austin)* 2 (1):105-120.
497 doi:10.1080/23328940.2015.1008890

498 Christmas KM, Patik JC, Khoshnevis S, Diller KR, Brothers RM (2016) Sustained cutaneous
499 vasoconstriction during and following cryotherapy treatment: Role of oxidative stress and Rho
500 kinase. *Microvasc Res* 106:96-100. doi:10.1016/j.mvr.2016.04.005

501 Daanen HA (2003) Finger cold-induced vasodilation: a review. *Eur J Appl Physiol* 89 (5):411-426.
502 doi:10.1007/s00421-003-0818-2

503 Daanen HAM, Ducharme MB (1999) Finger cold-induced vasodilation during mild hypothermia,
504 hyperthermia and at thermoneutrality. *Aviat Space Environ Med* 70 (12):1206-1210

505 Dianat I, Haslegrave CM, Stedmon AW (2012) Methodology for evaluating gloves in relation to the effects
506 on hand performance capabilities: a literature review. *Ergonomics* 55 (11):1429-1451.
507 doi:10.1080/00140139.2012.708058

508 Dominguez R, Cuenca E, Mate-Munoz JL, Garcia-Fernandez P, Serra-Paya N, Estevan MC, Herreros PV,
509 Garnacho-Castano MV (2017) Effects of Beetroot Juice Supplementation on Cardiorespiratory
510 Endurance in Athletes. A Systematic Review. *Nutrients* 9 (1). doi:10.3390/nu9010043

511 Eglin CM, Costello JT, Bailey SJ, Gilchrist M, Massey H, Shepherd AI (2017) Effects of dietary nitrate
512 supplementation on the response to extremity cooling and endothelial function in individuals with
513 cold sensitivity. A double blind, placebo controlled, crossover, randomised control trial. *Nitric
514 Oxide* 70:76-85. doi:10.1016/j.niox.2017.09.005

515 Eglin CM, Golden FS, Tipton MJ (2013) Cold sensitivity test for individuals with non-freezing cold injury:
516 the effect of prior exercise. *Extrem Physiol Med* 2 (1):16. doi:10.1186/2046-7648-2-16

517 Ekenvall L, Lindblad LE, Norbeck O, Ezzell BM (1988) alpha-Adrenoceptors and cold-induced
518 vasoconstriction in human finger skin. *Am J Physiol* 255 (5 Pt 2):H1000-1003.
519 doi:10.1152/ajpheart.1988.255.5.H1000

520 Esatbeyoglu T, Wagner AE, Schini-Kerth VB, Rimbach G (2015) Betanin--a food colorant with biological
521 activity. *Mol Nutr Food Res* 59 (1):36-47. doi:10.1002/mnfr.201400484

522 Folkow B, Neil E (1971) *Circulation*. Oxford University Press, London-Toronto.

523 Geng Z, Tong X, Jia H (2015) Reactive oxygen species (ROS) mediates non-freezing cold injury of rat
524 sciatic nerve. *Int J Clin Exp Med* 8 (9):15700-15707

525 Govoni M, Jansson EA, Weitzberg E, Lundberg JO (2008) The increase in plasma nitrite after a dietary

526 nitrate load is markedly attenuated by an antibacterial mouthwash. *Nitric Oxide-Biol Ch* 19
527 (4):333-337. doi:10.1016/j.niox.2008.08.003

528 Hadipour E, Taleghani A, Tayarani-Najaran N, Tayarani-Najaran Z (2020) Biological effects of red beetroot
529 and betalains: A review. *Phytother Res* 34 (8):1847-1867. doi:10.1002/ptr.6653

530 Hardy JD, Du Bois EF (1938) The technic of measuring radiation and convection. *J Nutr* 15 (5):461-475

531 Heus R, Daanen HA, Havenith G (1995) Physiological criteria for functioning of hands in the cold: a review.
532 *Appl Ergon* 26 (1):5-13

533 Hobbs DA, Goulding MG, Nguyen A, Malaver T, Walker CF, George TW, Methven L, Lovegrove JA (2013)
534 Acute ingestion of beetroot bread increases endothelium-independent vasodilation and lowers
535 diastolic blood pressure in healthy men: a randomized controlled trial. *J Nutr* 143 (9):1399-1405.
536 doi:10.3945/jn.113.175778

537 Hodges GJ, Johnson JM (2009) Adrenergic control of the human cutaneous circulation. *Appl Physiol Nutr*
538 *Metab* 34 (5):829-839. doi:10.1139/H09-076

539 Hodges GJ, Mallette MM, Cheung SS (2018) Cutaneous neural activity and endothelial involvement in
540 cold-induced vasodilatation. *Eur J Appl Physiol* 118 (5):971-978. doi:10.1007/s00421-018-3832-
541 0

542 Hodges GJ, Zhao K, Kosiba WA, Johnson JM (2006) The involvement of nitric oxide in the cutaneous
543 vasoconstrictor response to local cooling in humans. *J Physiol* 574 (Pt 3):849-857.
544 doi:10.1113/jphysiol.2006.109884

545 Holmer I (2009) Evaluation of cold workplaces: an overview of standards for assessment of cold stress. *Ind*
546 *Health* 47 (3):228-234. doi:10.2486/indhealth.47.228

547 Holowatz LA, Kenney WL (2007) Local ascorbate administration augments NO- and non-NO-dependent
548 reflex cutaneous vasodilation in hypertensive humans. *Am J Physiol-Heart C* 293 (2):H1090-
549 H1096. doi:10.1152/ajpheart.00295.2007

550 Hoon MW, Johnson NA, Chapman PG, Burke LM (2013) The effect of nitrate supplementation on exercise
551 performance in healthy individuals: a systematic review and meta-analysis. *Int J Sport Nutr Exerc*
552 *Metab* 23 (5):522-532. doi:10.1123/ijsnem.23.5.522

553 Hope K, Eglin C, Golden F, Tipton M (2014) Sublingual glyceryl trinitrate and the peripheral thermal
554 responses in normal and cold-sensitive individuals. *Microvasc Res* 91:84-89.
555 doi:10.1016/j.mvr.2013.11.002

556 House JR, Tipton MJ (2002) Using skin temperature gradients or skin heat flux measurements to determine
557 thresholds of vasoconstriction and vasodilatation. *Eur J Appl Physiol* 88 (1-2):141-145.
558 doi:10.1007/s00421-002-0692-3

559 Imray C, Grieve A, Dhillon S, Caudwell Xtreme Everest Research G (2009) Cold damage to the extremities:
560 frostbite and non-freezing cold injuries. *Postgrad Med J* 85 (1007):481-488.
561 doi:10.1136/pgmj.2008.068635

562 Johnson JM, Minson CT, Kellogg DL (2014) Cutaneous Vasodilator and Vasoconstrictor Mechanisms in
563 Temperature Regulation. *Comprehensive Physiology* 4 (1):33-89. doi:10.1002/cphy.c130015

564 Kellogg DL, Jr. (2006) In vivo mechanisms of cutaneous vasodilation and vasoconstriction in humans
565 during thermoregulatory challenges. *J Appl Physiol* (1985) 100 (5):1709-1718.
566 doi:10.1152/jappphysiol.01071.2005

567 Kellogg DL, Jr., Crandall CG, Liu Y, Charkoudian N, Johnson JM (1998) Nitric oxide and cutaneous active
568 vasodilation during heat stress in humans. *J Appl Physiol* (1985) 85 (3):824-829.
569 doi:10.1152/jappl.1998.85.3.824

570 Krauchi K, Cajochen C, Werth E, Wirz-Justice A (1999) Warm feet promote the rapid onset of sleep. *Nature*
571 401 (6748):36-37. doi:10.1038/43366

572 Kugler F, Stintzing FC, Carle R (2004) Identification of betalains from petioles of differently colored Swiss
573 chard (*Beta vulgaris* L. ssp. *ciela* [L.] Alef. Cv. Bright Lights) by high-performance liquid
574 chromatography-electrospray ionization mass spectrometry. *J Agric Food Chem* 52 (10):2975-
575 2981. doi:10.1021/jf035491w

576 Lara J, Ashor AW, Oggioni C, Ahluwalia A, Mathers JC, Siervo M (2016) Effects of inorganic nitrate and
577 beetroot supplementation on endothelial function: a systematic review and meta-analysis. *Eur J*
578 *Nutr* 55 (2):451-459. doi:10.1007/s00394-015-0872-7

579 Lewis T (1930) Observations upon the reactions of the vessels of the human skin to cold. *Heart-J Stud Circ*
580 15 (2):177-208

581 Makinen TM, Hassi J (2009) Health problems in cold work. *Ind Health* 47 (3):207-220.
582 doi:10.2486/indhealth.47.207

583 OReilly D, Taylor L, el-Hadidy K, Jayson MI (1992) Measurement of cold challenge responses in primary
584 Raynauds phenomenon and Raynauds phenomenon associated with systemic sclerosis. *Ann*
585 *Rheum Dis* 51 (11):1193-1196. doi:10.1136/ard.51.11.1193

586 Richards JC, Racine ML, Hearon CM, Jr., Kunkel M, Luckasen GJ, Larson DG, Allen JD, Dinunno FA
587 (2018) Acute ingestion of dietary nitrate increases muscle blood flow via local vasodilation during
588 handgrip exercise in young adults. *Physiol Rep* 6 (2). doi:10.14814/phy2.13572

589 Rubinstein EH, Sessler DI (1990) Skin-Surface Temperature-Gradients Correlate with Fingertip Blood-
590 Flow in Humans. *Anesthesiology* 73 (3):541-545. doi:Doi 10.1097/00000542-199009000-00027

591 Rusch NJ, Shepherd JT, Vanhoutte PM (1981) The effect of profound cooling on adrenergic
592 neurotransmission in canine cutaneous veins. *J Physiol* 311:57-65.
593 doi:10.1113/jphysiol.1981.sp013572

594 Sakihama Y, Maeda M, Hashimoto M, Tahara S, Hashidoko Y (2012) Beetroot betalain inhibits
595 peroxynitrite-mediated tyrosine nitration and DNA strand cleavage. *Free Radic Res* 46 (1):93-99.
596 doi:10.3109/10715762.2011.641157

597 Shepherd AI, Costello JT, Bailey SJ, Bishop N, Wadley AJ, Young-Min S, Gilchrist M, Mayes H, White D,

598 Gorczynski P, Saynor ZL, Massey H, Eglin CM (2019) "Beet" the cold: beetroot juice
599 supplementation improves peripheral blood flow, endothelial function, and anti-inflammatory
600 status in individuals with Raynauds phenomenon. *J Appl Physiol* (1985) 127 (5):1478-1490.
601 doi:10.1152/jappphysiol.00292.2019

602 Sobko T, Marcus C, Govoni M, Kamiya S (2010) Dietary nitrate in Japanese traditional foods lowers
603 diastolic blood pressure in healthy volunteers. *Nitric Oxide* 22 (2):136-140.
604 doi:10.1016/j.niox.2009.10.007

605 Stephens DP, Saad AR, Bennett LA, Kosiba WA, Johnson JM (2004) Neuropeptide Y antagonism reduces
606 reflex cutaneous vasoconstriction in humans. *Am J Physiol Heart Circ Physiol* 287 (3):H1404-
607 1409. doi:10.1152/ajpheart.00061.2004

608 Tanaka A, Yamazaki M, Saito M, Oh IT, Watanabe Y, Tsuboi R (2012) Highly sensitive high-pressure liquid
609 chromatography with ultraviolet light method detected the reduction of serum nitrite/nitrate levels
610 after cold exposure in patients with Raynauds phenomenon. *J Dermatol* 39 (10):889-890.
611 doi:10.1111/j.1346-8138.2011.01433.x

612 Thompson C, Vanhatalo A, Kadach S, Wylie LJ, Fulford J, Ferguson SK, Blackwell JR, Bailey SJ, Jones
613 AM (2018) Discrete physiological effects of beetroot juice and potassium nitrate supplementation
614 following 4-wk sprint interval training. *J Appl Physiol* (1985) 124 (6):1519-1528.
615 doi:10.1152/jappphysiol.00047.2018

616 Wakabayashi H, Oksa J, Tipton MJ (2015) Exercise performance in acute and chronic cold exposure. *The*
617 *Journal of Physical Fitness and Sports Medicine* 4 (2):177-185. doi:10.7600/jpfsm.4.177

618 Wickham KA, Steele SW, Cheung SS (2021) Effects of acute dietary nitrate supplementation on cold-
619 induced vasodilation in healthy males. *Eur J Appl Physiol* 121 (5):1431-1439.
620 doi:10.1007/s00421-021-04621-8

621 Wigley FM (2002) Clinical practice. Raynauds Phenomenon. *N Engl J Med* 347 (13):1001-1008.
622 doi:10.1056/NEJMcp013013

623 Wootton-Beard PC, Moran A, Ryan L (2011) Stability of the total antioxidant capacity and total polyphenol
624 content of 23 commercially available vegetable juices before and after in vitro digestion measured
625 by FRAP, DPPH, ABTS and Folin-Ciocalteu methods. *Food Research International* 44 (1):217-
626 224. doi:10.1016/j.foodres.2010.10.033

627 Wyler H, Meuer U (1979) Zur Biogenese der Betacyane: Versuche mit [2-14C]-Dopaxanthin. *Helv Chim*
628 *Acta* 62 (4):1330-1339. doi:10.1002/hlca.19790620447

629 Wylie LJ, Kelly J, Bailey SJ, Blackwell JR, Skiba PF, Winyard PG, Jeukendrup AE, Vanhatalo A, Jones
630 AM (2013) Beetroot juice and exercise: pharmacodynamic and dose-response relationships. *J*
631 *Appl Physiol* (1985) 115 (3):325-336. doi:10.1152/jappphysiol.00372.2013

632 Yamazaki F (2010) Local ascorbate administration inhibits the adrenergic vasoconstrictor response to local
633 cooling in the human skin. *J Appl Physiol* (1985) 108 (2):328-333.

634 doi:10.1152/jappphysiol.00814.2009
635 Yamazaki F (2015) The cutaneous vasoconstrictor response in lower extremities during whole-body and
636 local skin cooling in young women with a cold constitution. *J Physiol Sci* 65 (5):397-405.
637 doi:10.1007/s12576-015-0378-3
638

Table 1 Parameters of cold induced vasodilation during hand cold immersion (n = 20).

| | Water | | Beet | | T-test |
|---|-------|-------|------|--------|---------|
| | mean | (SD) | mean | (SD) | p value |
| <i>T_{fing}</i> | | | | | |
| Baseline (°C) | 32.6 | (1.6) | 32.8 | (1.5) | 0.41 |
| Onset time (sec) | 521 | (78) | 535 | (145) | 0.68 |
| <i>T_{min}</i> (°C) | 9.3 | (0.7) | 9.4 | (0.7) | 0.36 |
| <i>T_{peak}</i> (°C) | 10.6 | (1.1) | 11.0 | (1.3) | 0.12 |
| <i>T_{max}</i> (°C) | 11.0 | (1.2) | 11.2 | (1.5) | 0.48 |
| <i>T_{peak}</i> - <i>T_{min}</i> (°C) | 1.2 | (0.9) | 1.6 | (1.1) | 0.21 |
| Mean <i>T_{fing}</i> (°C) | 10.1 | (0.9) | 10.3 | (0.9) | 0.27 |
| Number of CIVD | 1.6 | (0.7) | 1.9 | (1.0) | 0.21 |
| <i>SkBF_{fing}</i> | | | | | |
| Baseline (AU) | 26.8 | (5.7) | 30.2 | (10.6) | 0.14 |
| Onset time (sec) | 83 | (58) | 78 | (64) | 0.80 |
| <i>SkBF_{min}</i> (AU) | 2.7 | (1.9) | 3.3 | (2.1) | 0.30 |
| <i>SkBF_{peak}</i> (AU) | 15.6 | (4.7) | 15.8 | (5.2) | 0.90 |
| <i>SkBF_{max}</i> (AU) | 17.4 | (6.3) | 17.5 | (6.1) | 0.98 |
| <i>SkBF_{peak}</i> - <i>SkBF_{min}</i> (AU) | 12.9 | (3.8) | 12.5 | (3.9) | 0.66 |
| Mean <i>SkBF_{fing}</i> (AU) | 11.8 | (4.7) | 12.2 | (5.0) | 0.74 |
| Number of CIVD | 2.1 | (0.8) | 2.4 | (1.0) | 0.18 |

T_{fing}: finger skin temperature, *T_{min}*: minimal *T_{fing}*, *T_{peak}*: the first peak *T_{fing}*, *T_{max}*: maximal *T_{fing}* during hand cold immersion, Mean *T_{fing}*: mean value of *T_{fing}* after 5 min to the end of immersion, CIVD: cold induced vasodilation, *SkBF*: skin blood flow, AU: arbitrary unit.

Table 2 Rewarming speed in finger skin temperature and blood flow after hand cold immersion (n = 20)

| Rewarming speed | Water | | Beet | | T-test | FDR |
|--|-------|--------|------|--------|---------|---------|
| | mean | SD | mean | SD | p value | p value |
| <i>T</i> _{finger} (°C/min) | | | | | | |
| in 5 min (31-35 min) | 1.92 | (0.63) | 1.86 | (0.57) | 0.74 | 0.74 |
| in 10 min (31-40 min) | 1.47 | (0.43) | 1.61 | (0.38) | 0.09 | 0.11 |
| in 15 min (31-45 min) | 1.11 | (0.26) | 1.24 | (0.22) | 0.01 | 0.02 |
| in 20 min (31-50 min) | 0.86 | (0.19) | 0.97 | (0.14) | 0.01 | 0.02 |
| <i>SkBF</i> _{finger} (AU/min) | | | | | | |
| in 5 min (31-35 min) | 2.33 | (1.74) | 2.75 | (1.98) | 0.40 | 0.40 |
| in 10 min (31-40 min) | 1.50 | (0.91) | 1.88 | (0.91) | 0.04 | 0.05 |
| in 15 min (31-45 min) | 0.88 | (0.59) | 1.29 | (0.61) | 0.01 | 0.02 |
| in 20 min (31-50 min) | 0.67 | (0.45) | 0.99 | (0.44) | 0.01 | 0.02 |

Rewarming speed is calculated from 1 min to 5, 10, 15, and 20 min after hand immersion.

*T*_{finger}: finger skin temperature, *SkBF*_{finger}: finger skin blood flow. P values of multiple paired T-tests are corrected with false discovery rate (FDR).

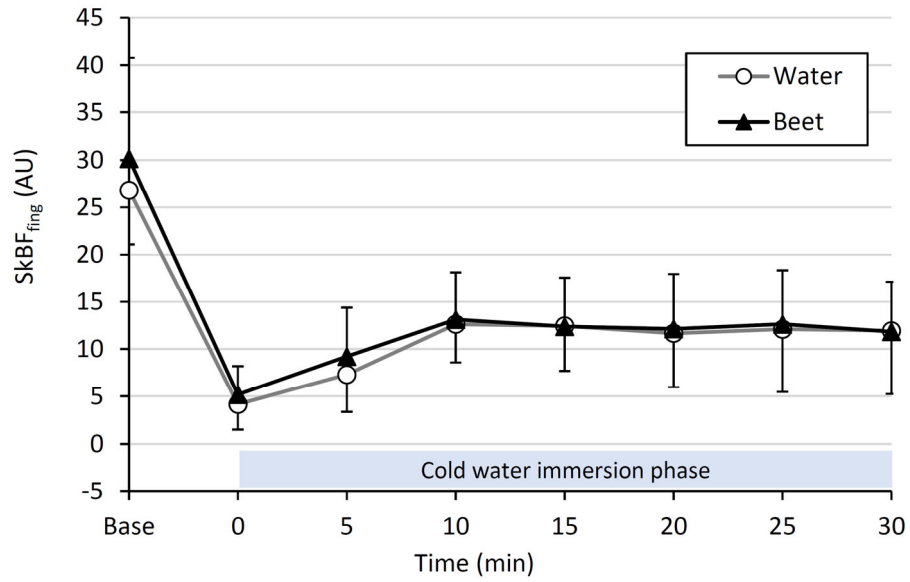
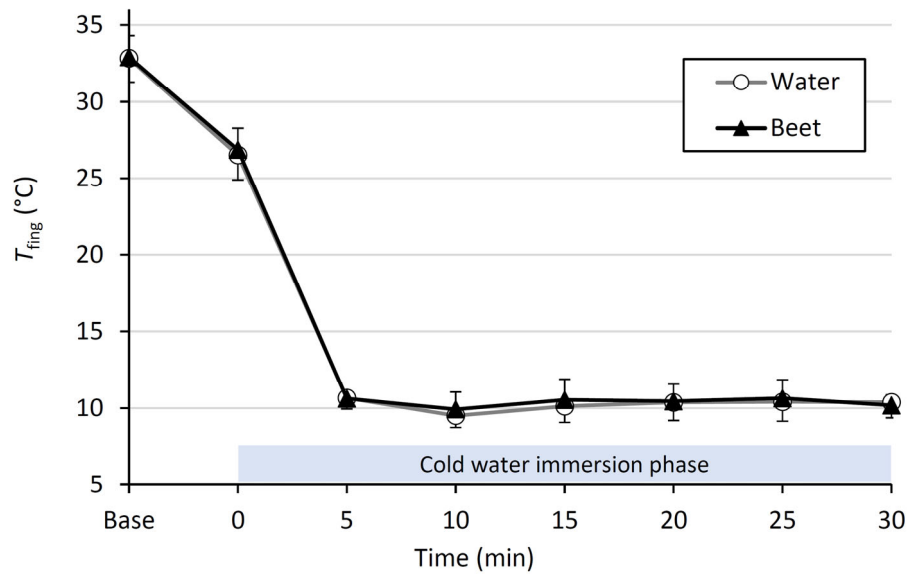


Figure 1

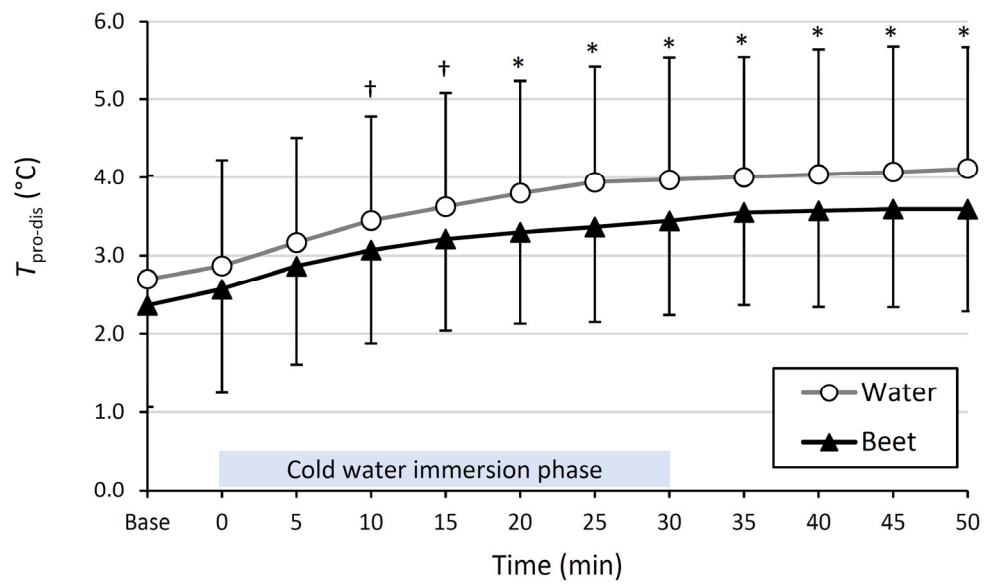
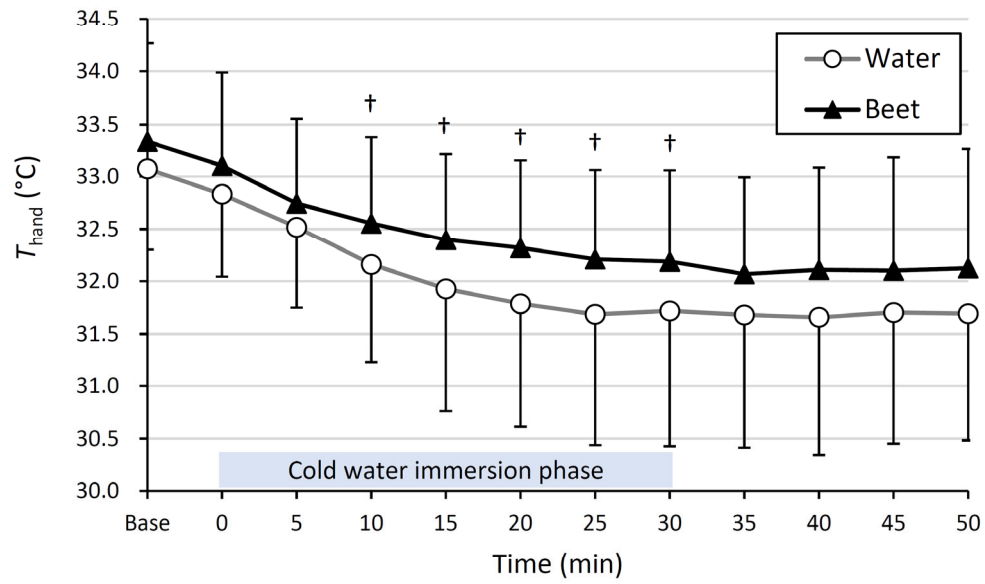
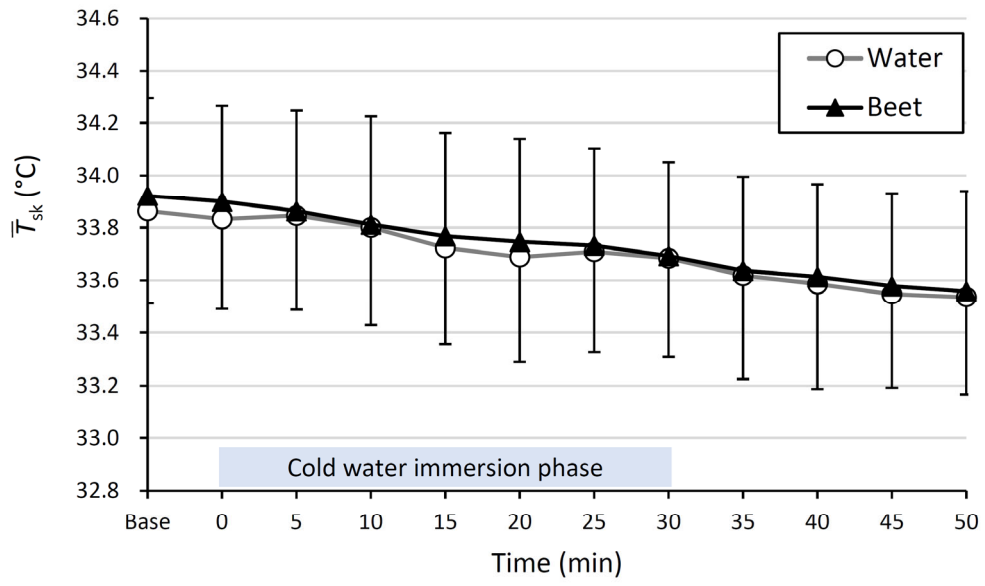


Figure 2

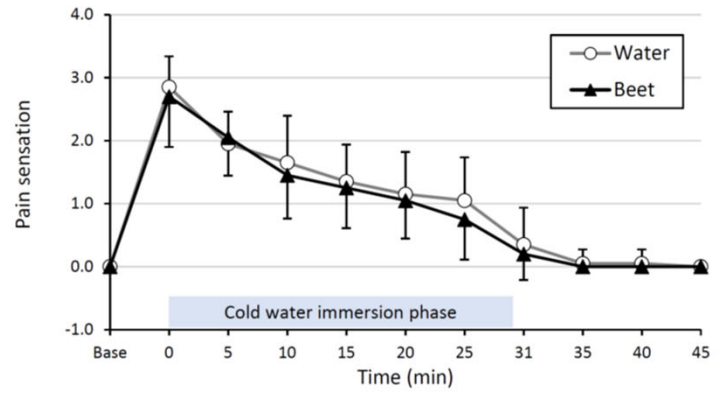
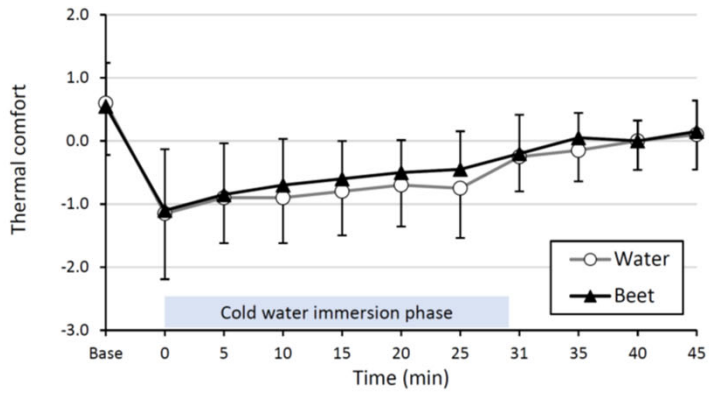
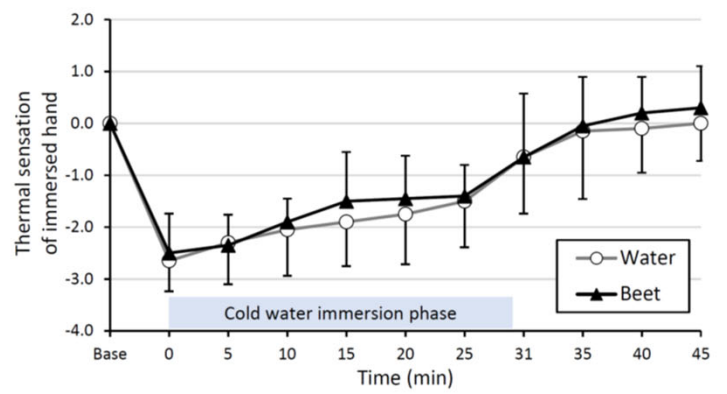
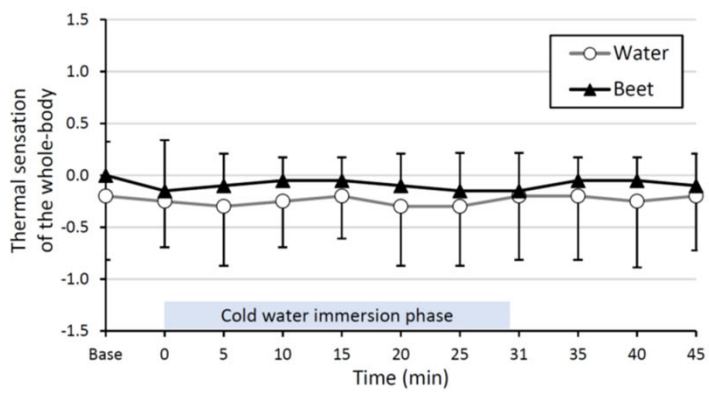


Figure 3

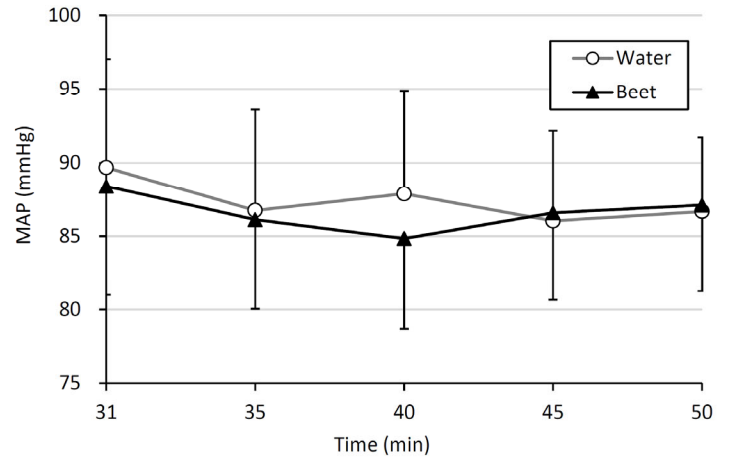
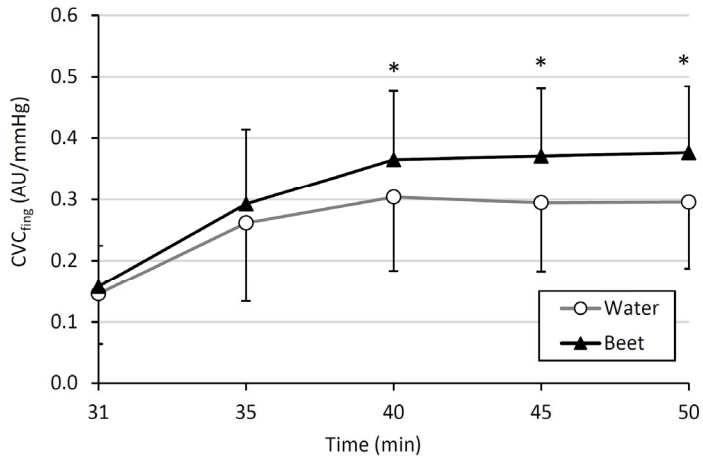
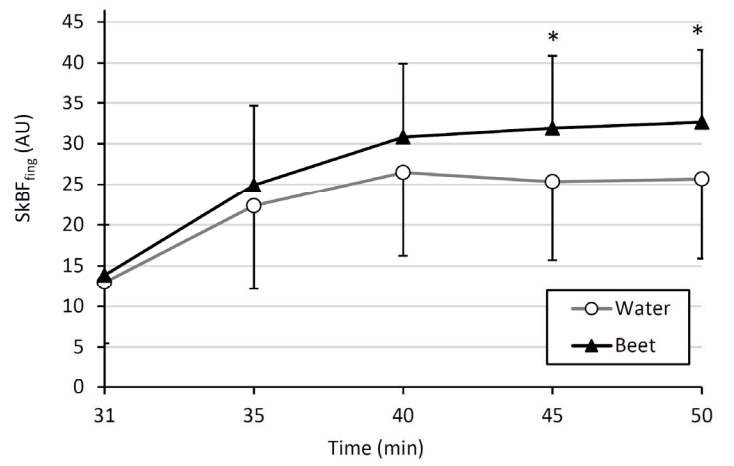
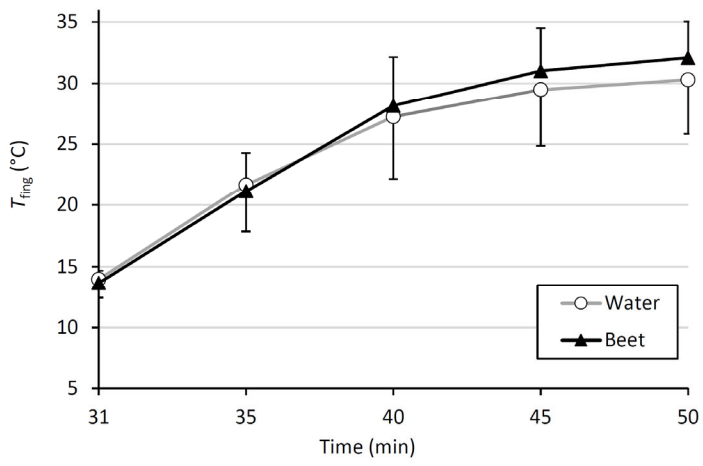


Figure 4

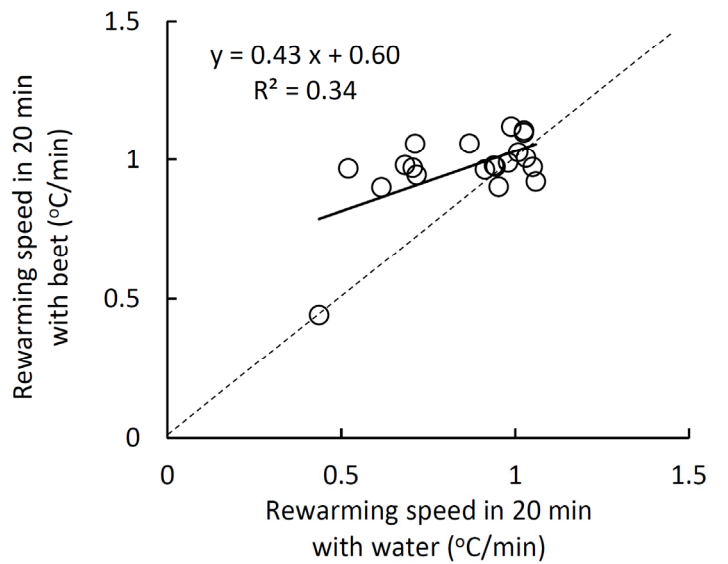
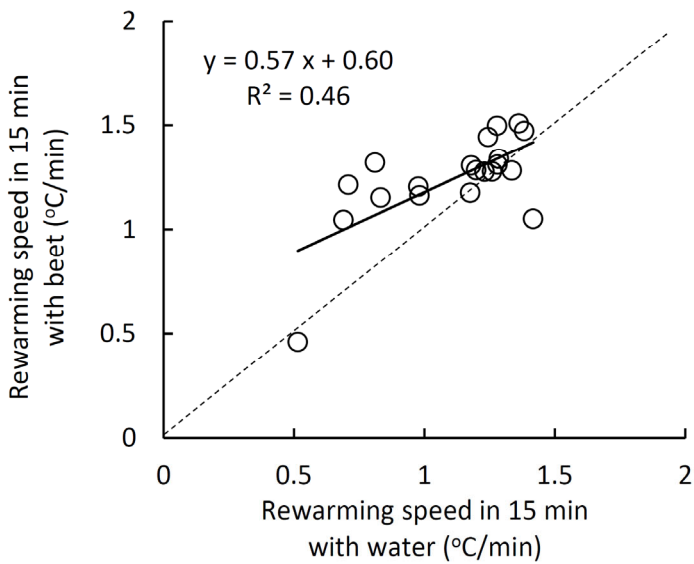
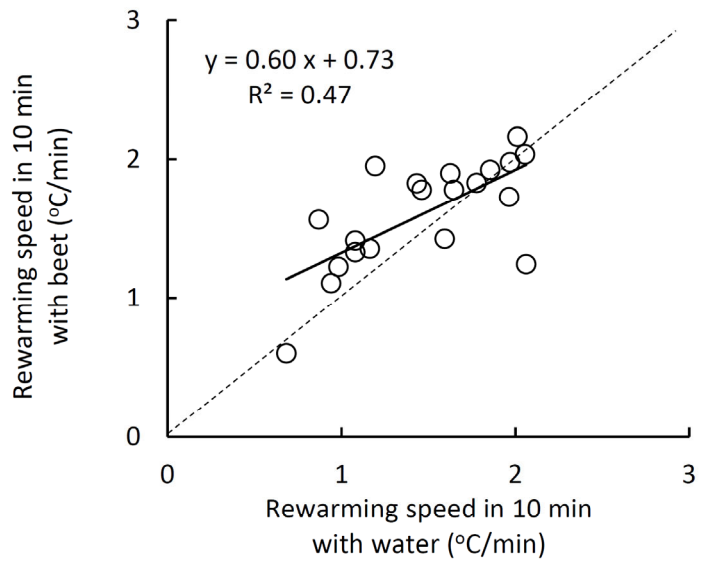
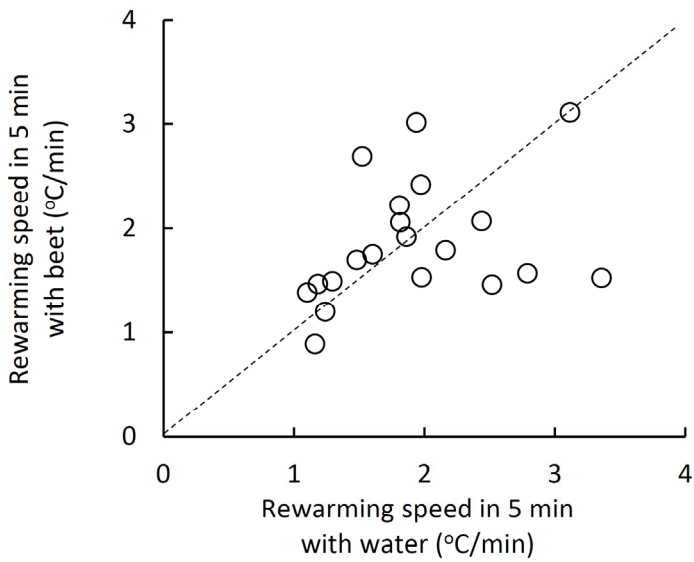


Figure 5