



Title	A Novel Epimerase Catalyzing Multiple Isomerization of Amino Acid Residues of Ribosomal Peptide [an abstract of dissertation and a summary of dissertation review]
Author(s)	肖, 宛璐
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学 位 論 文 内 容 の 要 旨

博士の専攻分野の名称 博士（工学） 氏名 ショウ エンロ

学 位 論 文 題 名

A Novel Epimerase Catalyzing Multiple Isomerization of Amino Acid Residues of Ribosomal Peptide

(リボソームペプチドの複数アミノ酸残基を異性化する新規エピメラーゼ)

Salinipeptins, grisemycin, and cypemycin are ribosomally synthesized and post-translationally modified peptides (RiPPs). Among these, salinipeptins was reported to comprise 22 amino acid residues with multiple D-amino acids and its biosynthetic gene cluster was identified. However, no genes homologous to known isomerases such as epimerases and racemases existed in the cluster, but a gene, *salL*, which showed no similarities to function known enzymes, located in the cluster, suggesting that *SalL* might be a novel epimerase. Actually, biosynthetic gene clusters of grisemycin and cypemycin also possess orthologs of *salL* although chirality of amino acids composing grisemycin and cypemycin, have not been reported. Therefore, I first examined grisemycin also contains D-amino acid residues. By heterologous expression of grisemycin biosynthetic gene cluster (*grm*) in *Streptomyces lividans*, grisemycin was confirmed to contain multiple D-amino acids, in the similar manner to salinipeptins. The heterologous expression experiments also confirmed the involvement of a novel peptide epimerase in grisemycin biosynthesis. Gene-deletion experiments indicated that *grmL*, an ortholog of *salL*, was indispensable for grisemycin production and that the epimerization preceded decarboxylation and methylation, which are other modifications installed into the precursor peptides of grisemycin (*GriA*).

To obtain further evidence that *GriL* encodes the novel epimerase, recombinant precursor peptide (*GriA*) and *GriL* were prepared and used for in vitro analysis. However, no isomerase activities were observed under various conditions. Considering that grisemycin contains dehydroamino acid and its biosynthetic gene, *griH*, exists in the gene cluster, the dehydration reaction might occur before isomerization. To examine the possibility, recombinant *GriH* was prepared and incubated with *GriA*, but no dehydration activity was detected. Because *GriL* is the novel enzyme found in *Streptomyces* strain, I consider a possibility that *GriL* requires a co-factor specifically utilized in *Streptomyces* strain. I therefore again utilized the above-mentioned heterologous expression system. When *griA*, *griH*, and *griL* were co-expressed, a dehydrated and isomerized *GriA* was produced, but *griA* and either of the two resulted in the production of no modified *GriA*. The results suggested that an interaction among three enzymes would be essential for dehydration and isomerization of *GriA*.