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Author(s)	Nakajima, Souta; Suzuki, Hiroaki; Nakatsugawa, Makoto; Matsuo, Ayumi; Hirota, Shun K.; Suyama, Yoshihisa; Nakamura, Futoshi	
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Instructions for use

1	Inferring future changes in gene flow under climate change in riverscapes: a pilot case		
2	study in fluvial sculpin		
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4	Souta Nakajima ^{1,2,*} , Hiroaki Suzuki ^{3,4} , Makoto Nakatsugawa ³ , Ayumi Matsuo ⁵ , Shun K. Hirota ^{5,6} ,		
5	Yoshihisa Suyama ⁵ , Futoshi Nakamura ¹		
6			
7	¹ Graduate School of Agriculture, Hokkaido University, Kita-ku N9W9, Sapporo, Hokkaido 060-8589,		
8	Japan		
9	² Present address: Water Environment Research Group, Public Works Research Institute, Minamihara		
10	1-6, Tsukuba, Ibaraki 305-8516, Japan		
11	³ Graduate School of Engineering, Muroran Institute of Technology, Mizumoto-cho 27-1, Muroran,		
12	Hokkaido 050-8585, Japan		
13	⁴ Research Institute of Energy, Environment and Geology, Hokkaido Research Organization, Kita-ku		
14	N19W12, Sapporo, Hokkaido 060-0819, Japan		
15	⁵ Graduate School of Agricultural Science, Tohoku University, Yomogida 232-3, Naruko-onsen,		
16	Osaki, Miyagi 989-6711, Japan		
17	⁶ Present address: Botanical Gardens, Osaka Metropolitan University, Kisaichi 2000, Katano, Osaka		
18	576-0004, Japan		
19			
20	* Corresponding author		
21	Souta Nakajima: n.souta891@gmail.com		
22	Watershed Restoration Team, Water Environment Research Group, Public Works Research Institute,		
23	Minamihara 1-6, Tsukuba, Ibaraki 305-8516, Japan		
24	Phone: +81-29-879-6775, Fax: +81-29-869-5082		
25			
26	ORCID		
27	Souta Nakajima: https://orcid.org/0000-0003-3701-5428		

- 28 Shun K. Hirota: https://orcid.org/0000-0002-6104-1119
- 29 Yoshihisa Suyama: https://orcid.org/0000-0002-3136-5489
- 30 Futoshi Nakamura: https://orcid.org/0000-0003-4351-2578
- 31

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- 36

37 Abstract

38 *Context*: Global climate change poses a significant threat to the habitat connectivity of cold-water-

39 adapted organisms, leading to species extinctions. If gene flow can be modeled by landscape variables,

40 changes in connectivity among populations could be predicted. However, in dendritic and

41 heterogeneous stream ecosystems, few studies have estimated the changes in gene flow from genetic

42 data, in part due to the difficulty in applying landscape genetics methods and accessing water

43 temperature information.

44 *Objectives*: Inferring the determinants and future changes of the gene flow in the cold-water adapted

45 fluvial sculpin *Cottus nozawae* using a recently developed model-based riverscape genetics technique

46 and a hydrological model for estimating water temperature.

47 *Methods*: The strength of gene flow on each stream section was modeled by watershed-wide

48 riverscape variables and genome-wide SNP data for *C. nozawae* in the upper reaches of the Sorachi

49 River, Hokkaido, Japan. Future changes in gene flow were inferred by this model and hydrologically

50 estimated water temperatures under the high greenhouse gas concentration scenario (IPCC RCP8.5).

51 *Results*: Stream order, water temperature, slope, and distance were selected as riverscape variables

52 affecting the strength of gene flow in each stream section. In particular, the trend of greater gene flow

53 in sections with higher stream order and lower temperature fluctuations or summer water temperatures

54 was pronounced. The map from the model showed that gene flow is overall prevented in small

55 tributaries in the southern area, where spring-fed environments are less prevalent. Estimating future

56 changes, gene flow was predicted to decrease dramatically at the end of the 21st century.

57 *Conclusions*: Our results demonstrated that the connectivity of cold-water sculpin populations is

58 expected to decline dramatically in a changing climate. Riverscape genetic modeling is useful for

59 gaining information on population connectivity that does not fully coincide with habitat suitability.

60

61 Keywords

62 model-based riverscape genetics; cold-water fish; *Cottus*; water temperature; global warming63

64 Introduction

65 Global climate change modifies water temperatures and flow regimes, the two key habitat factors 66 affecting freshwater species, posing a critical threat to stream ecosystems (Barbarossa et al. 2021). The 67 spatial distribution of species' suitable habitats shifts with environmental changes, and population 68 fragmentation due to impassable environments may eventually result in local and/or species 69 extinctions (Woodward et al. 2010). Numerous studies have predicted changes in species distributions 70 and suitable habitats of stream organisms (Elith and Leathwick 2009; Comte et al. 2012; Ishiyama et 71 al. 2023; Rahel et al. 1996), but how will the actual population connectivity and migration potential 72 change?

73 Gene flow represents the functional connectivity among wild populations and is critical in 74 species viability (Kottler et al. 2021; Manel and Holderegger 2013). The strength of gene flow is 75 usually discussed individually from the observed genetic structure, but if gene flow could be modeled 76 by landscape variables, the gained knowledge regarding gene flow could be generalized and used to 77 predict its future changes (McRae and Beier 2007). The relationships between gene flow and 78 landscape variables have been investigated in the field of landscape genetics (Balkenhol et al. 2015). 79 However, most analytical techniques developed in landscape genetics exert only poor power in linear 80 and dendritic stream ecosystems (Davis et al. 2018; Chafin et al. 2021), making it difficult to predict 81 future changes in gene flow in riverscapes. Even in streams, regression models can be created by 82 contrasting a genetic distance matrix against pairwise differences in local conditions (Grummer et al. 83 2019), but this approach fails to account for the network architecture and for the attributes in all the 84 spaces that individuals must pass through when traveling between sampling sites (Davis et al. 2018; 85 White et al. 2020; Escalante et al. 2020). Another versatile approach to investigating the effects of 86 landscape elements on gene flow is defining "landscape resistance" surfaces and assessing the 87 relationship between genetic distance and cumulative resistance between populations (isolation by 88 resistance; IBR (McRae 2006)). Although this idea has been applied to studies on stream ecosystems 89 in several cases (e.g., Inoue and Berg 2017; Oliveira et al. 2019; Landguth et al. 2016; Escalante et al. 90 2018), the landscape resistance must be parametrized *a priori* through expert opinion or other 91 empirical methods (e.g., using the inverse of species distribution model estimates) (Spear et al. 2015;

Zeller et al. 2012). To understand the gene flow itself, its determinants should be identified directly
from genetic data (Sartor et al. 2022; Wasserman et al. 2010). Fortunately, alternative methods for
modeling gene flow from genetic data within a spatially explicit graph-theoretic framework have been
developed rapidly in recent years (White et al. 2020; Chafin et al. 2021). Although not yet practically
applied to predictions under environmental changes, we thought that these "riverscape genetics"dedicated methods are the key to determining landscape resistance and modeling current and future
gene flow.

99 Another theme that makes riverscape genetics challenging is the data availability of key 100 environmental elements such as water temperatures. For terrestrial organisms, globally available 101 climate data such as WorldClim (Hijmans et al. 2005) are commonly used to estimate the effects of 102 climate change. However, data on current and future water temperatures that are critical for stream 103 organisms are difficult to obtain as are data on flow rate. Although some studies have used air 104 temperature data as a surrogate of water temperatures (Almodóvar et al. 2012), water temperatures do 105 not actually coincide with air temperatures. In particular, local spatial heterogeneity in water 106 temperatures caused by groundwater discharge and other factors is truly a source of ecosystem 107 diversity and resilience to climate change that cannot be ignored (Koizumi and Maekawa 2004; 108 Nakajima et al. 2021; Ishiyama et al. 2023; Nakamura 2022). Therefore, it is critical in riverscapes to 109 utilize water temperature information considering the spatial heterogeneity generated by 110 hydrogeological factors.

111 Cottus nozawae is a cold-water-adapted sculpin inhabiting northern Japan. Since the 112 distribution and ecology of this species are highly influenced by summer water temperatures (Yagami 113 and Goto 2000), available habitats are expected to decrease significantly under climate change (Suzuki 114 et al. 2021). At a local scale, streams with low summer water temperatures characterized by spring-fed 115 environments have been shown to display high population densities and to be the source of individuals 116 in a watershed (Suzuki et al. 2021; Nakajima et al. 2021). Under ongoing climate change, the 117 migration of this species is expected to be frequently blocked by unsuitable habitats, resulting in 118 population fragmentation and shrinkage. To sustain the species into the future, it is critical to 119 accurately predict the relationship between population connectivity and climate-related variables. Such

predictions will contribute to the advancement of climate change adaptation measures for this species,
such as by identifying sections where stream continuity should be ensured.

122Considering the challenges of data availability and analysis in riverscape genetics, we thought123that the recently developed model-based riverscape genetics approaches and physics-based124hydrological model to estimate water temperatures would enable the modeling and future prediction of125gene flow in cold-water fish. The aims of this study are (i) to identify the factors determining the gene126flow of *C. nozawae* in the stream network, (ii) to model the strength of gene flow using riverscape127variables and predict its future changes, and (iii) to discuss the applicability of riverscape genetic128modeling in conservation ecology.

129

130 Material and Methods

131 Study sites and sampling

132 In 2019, small pieces of fin tissue were sampled from 376 individuals of *C. nozawae* caught by 133 electrofishing (model 12-B Backpack Electrofisher; Smith-Root Inc.) at 13 sites located in the 134 upstream section of the Sorachi River, Hokkaido, Japan (Fig. 1; Table S1). Because no river-crossing 135 structures that would obviously prevent fish migration are present between sampling sites, this area is 136 considered suitable for evaluating the effects of riverscape variables. Regarding the environmental 137 conditions, the tributaries in the northern volcanic watersheds have spring-fed environments with 138 stable water temperatures and flow regimes (García Molinos et al. 2022; Ishiyama et al. 2023). For 139 riverscape genetic modeling, the stream network among sampling sites was viewed as a graph consisting of 24 "nodes" and 23 "edges" (Fig. 2a). We defined "nodes" as the sampling sites and 140 141 major tributary confluences between them, and analysis was conducted with "edges", the stream 142 segments between adjacent nodes, as units.

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144 Genetic data

Genomic DNA was extracted using the QIAGEN DNeasy Blood and Tissue Kit (QIAGEN
Inc.). In this study, we used the multiplexed ISSR genotyping by sequencing (MIG-seq) method

147 (Suyama and Matsuki 2015; Suyama et al. 2022), a technique in which loci between two microsatellite

148 regions are amplified and neutral genome-wide single nucleotide polymorphisms (SNPs) are detected. 149 A MIG-seq library preparation and read quality filtering were performed according to the protocol 150 described in Suyama et al. (2022), with the modification that two runs were conducted and the 151 obtained data were combined after quality filtering. In addition, quality filtering was performed on 71 152 bases with six 5'-end bases and three 3'-end bases removed. After quality filtering, SNP selection was 153 performed using STACKS 2.41 (Catchen et al. 2013). First, the reads were grouped to each locus 154 using the *ustacks*, *cstacks*, *sstacks*, *tsv2bam*, and *gstacks* commands with the following parameters 155 recommended by Paris et al. (2017): minimum depth option creating a stack (m) = 3, maximum 156 distance between stacks (M) = 2, maximum mismatches between loci when building the catalog 157 (n) = 2, and number of mismatches allowed to align secondary reads (N) = 4. From the derived dataset 158of assembled loci, SNPs were detected using the *populations* commands under the following criteria: 159 only loci present at a rate of more than 80% of individuals within all populations were extracted (-p 13 160 -r 0.8); the minimum minor allele frequency was 5% (--min-maf 0.05); sites showing excess 161 heterozygosity were removed (--max-obs-het 0.6); and the output was limited to one SNP per locus (--162 write-single-snp). After filtering, 212 SNPs were obtained.

163 For populations in each sampling site, the expected heterozygosity (H_E) and fixation index 164 (F_{IS}) were calculated using the *populations* command in STACKS. Significant deviations from Hardy– 165 Weinberg equilibrium, as indicated by F_{IS} deviating from zero, were tested by 1000 randomizations 166 using FSTAT 2.9.4 (Goudet 1995). Genetic differentiation among populations was assessed by G_{ST} 167 (Nei 1973) and D_{PS} (Bowcock et al. 1994). D_{PS} is the genetic distance based on the dissimilarities of 168 population allele pools and reflects gene flow over a shorter timescale (approximately 10 generations; 169 Landguth et al. 2010; Leroy et al. 2018), whereas $G_{\rm ST}$ is assumed to reflect long-term gene flow 170 (Holsinger and Weir 2009). G_{ST} was calculated using GenAlEx 6.51 (Peakall and Smouse 2012), and 171D_{PS} was calculated using the package graph4lg (Savary et al. 2021) in R 3.6.0 (R Core Team 2019). 172 To understand the general patterns of population structure, STRUCTURE 2.3.4 (Pritchard et al. 2000) 173 was performed in the setting of the admixture and allele frequency correlated model with previous 174 sampling location information (LOCPRIOR; Hubisz et al. 2009). The algorithm was run 10 times for 175 each K from 1 to 10 with a burn-in of 20,000 followed by 30,000 MCMC replicates. The program

176 CLUMPAK (Kopelman et al. 2015) was then used to summarize the results for each K. STRUCTURE 177 HARVESTER (Earl and vonHoldt 2012) was employed to calculate the probability of the data for 178 each K (LnP(D); Pritchard et al. 2000), the corresponding standard deviation, and the Δ K (Evanno et 179 al. 2005).

180

181 Riverscape data

182 Riverscape variables were collected as a unit of edges. Edge length, slope, stream orders, and 183 catchment area were calculated in ArcGIS 10.7.1 (ESRI Inc.) using National Land Numerical 184 Information (nlftp.mlit.go.jp) from the Ministry of Land, Infrastructure, Transport and Tourism 185 (MLIT) of Japan. Flow rate and water temperature were estimated by a hydrological model based on 186 Suzuki et al. (2022), which considers differences in groundwater discharge depending on catchment 187 geology (see Appendix 1 for details). Briefly, the daily flow rate on the stream in each 1 km mesh was 188 reproduced by four-layered tank models (Sugawara 1979), and the flow and heat flux were tracked 189 along the streamflow. Importantly, different tank parameters were given for the volcanic areas and 190 other areas, based on validation using measured water temperature data from field surveys at multiple 191 sites in the study area. Flow rate and water temperature from September 2018 to August 2019 were 192 reproduced and used to calculate the riverscape variables. In future predictions, 1 km-downscaled data 193 on meteorological elements (Ueda et al. 2020), calculated under the climate data projected in the 194 representative concentration pathway scenario 8.5 (RCP8.5) in the IPCC 5th Assessment (IPCC 2014), 195 was used to derive the input water amount to the tank model. We used the mean of the predicted 196 variables for the years 2081 to 2100 (all from September to next August) as future riverscape 197 variables.

198

199 Gene flow analysis

We probabilistically modeled the relative migration rate (edge passability) of each edge as a function of riverscape variables using the "BGR model" (White et al. 2020). This is a novel method that can model bidirectional gene flow in stream networks using genetic distance matrices as input data and riverscape variables as explanatory variables, rigorously accounting for the spatial autocorrelation

structure of stream networks using a graph-theoretical framework and a spatial autoregressive model. Specifically, the nearly homogeneous stream segments (delimited by nodes that are sampling sites or major tributary confluences) were defined as edges, and the relative migration rate (edge passability; w_{ij}) of each edge linking nodes *i* and *j* was estimated as a function of *k* riverscape variables ($x_{ij1}, x_{ij2}, \dots, x_{ijk}$) and the corresponding parameters ($\beta_1, \beta_2, \dots, \beta_k$) as:

$$w_{ij} = \exp \left(\beta_0 + \sum_k \beta_k x_{ijk}\right)$$

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where β_0 is the intercept term. Here, all riverscape variables were normalized from 0 to 1. The posterior distribution of parameters $\beta_0, \beta_1, ..., \beta_k$ was estimated by a Markov Chain Monte Carlo (MCMC) sampler, to fit the input genetic data. The mathematics linking w_{ij} to genetic distance are described in Peterson et al. (2019).

214 The BGR model was run in R. We used G_{ST} and D_{PS} as genetic distances and 10 possible 215 riverscape variables (Table 1; Fig. S1) as covariates x_{iik} . All variables except direction are symmetric. 216 For each summary statistic, forward selections were conducted based on the deviance information 217 criterion (DIC). Variables were added until the DIC no longer decreased by 7 or more (Cain and 218 Zhang 2019). At each step of the forward selection, the variables that were highly correlated 219 (Pearson's r > 0.7) with other variables already included in the model were not added to the model. 220 Models with fewer than four variables were run for 50,000 MCMC iterations and parameters were 221 estimated after 25,000 burn-in. Models with four or more variables were run for 100,000 iterations 222 including 50,000 burn-in. After the final model was identified, we conducted a long run with 500,000 223 iterations including 200,000 burn-in, to accurately estimate the β values and 95% credible intervals. 224 Landscape resistance, calculated as the inverse of w_{ii} of each edge, was estimated and mapped from 225 the selected models. To evaluate the estimates, the correlations between genetic distances and 226 estimated landscape resistance (sum of edges between populations) were calculated by Mantel tests 227 with 9999 permutations, and compared to the correlations between genetic distances and waterway 228 geographical distance. The Mantel tests were conducted using the package VEGAN 2.5.6 (Oksanen et 229 al. 2019) in R. Future landscape resistance was inferred by substituting the future water temperature 230 variable into the final BGR model. At this stage, we used the model derived from D_{PS} because G_{ST}

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231

233 **Results**

234 The level of H_E was similar across the watershed (ranged from 0.241–0.272), and F_{IS} ranged from -235 0.012–0.023 with no populations deviating significantly from zero (Table S1). The average $G_{\rm ST}$ was 236 0.029 (ranged from 0.000–0.047; Table S2) and D_{PS} was 0.087 (ranged from 0.050–0.135). In the 237 STRUCTURE, while LnP(D) for each K increased progressively, ΔK was highest at K = 2, and locally 238 maximum at K = 6 (Fig. S2). Populations in the southern area were grouped into distinct clusters from 239 low K, and as K increased, populations in other tributaries were also mixed with geographically 240 uneven clusters. The strength of population structure differed geographically (Fig. 2a), but the factors 241 determining this difference are not known by the STRUCTURE. From the forward selection of the 242 model explaining the strength of gene flow, Shreve's stream order, water temperature fluctuation, 243 slope, and edge length were selected for G_{ST} , and Strahler's stream order and summer water 244 temperature were selected for D_{PS} , in this order (Tables 2 and S3). In both cases, the first and second 245 variables added to the model were the stream order and water temperature, respectively. While 246 different types of variables were selected for G_{ST} and D_{PS} (Shreve's or Strahler's; summer water 247 temperature or water temperature fluctuation), these results show the importance of the stream order 248 and water temperature on the strength of gene flow. The stream orders had a positive effect on gene 249 flow, while the water temperature fluctuation or summer water temperature had a negative effect. The 250 effect (β) of the water temperature on gene flow was higher in D_{PS} than in G_{ST} . In G_{ST} , the slope and 251 edge length were also selected and had negative effects, indicating their relevance to long-term gene 252flow. Geographically, the southern upstream area had generally higher landscape resistance (lower 253 gene flow) than the main stream, while in the northern volcanic area, landscape resistance was not so 254 high even upstream (Fig. 2). The Mantel tests between the genetic distances and estimated landscape 255 resistance suggested significant relationships (r = 0.46, p < 0.05 for G_{ST} ; r = 0.60, p < 0.01 for D_{PS}), 256 and the correlations were much higher than those between the genetic and geographic distances (Fig. 257 3). The future prediction indicated that the landscape resistance would increase overall from the 258 current levels. Some sections in the main stream and in the upper reaches in the volcanic area were

displays long-term patterns and D_{PS} is more likely to reflect current changes in gene flow.

estimated to exhibit as high landscape resistance levels as the present southern upstream area. Thesouthern upstream area was projected to display very high resistance.

261

262 **Discussion**

In this study, we succeeded in modeling and future predicting of gene flow of *C. nozawae* in the stream network. Although there are still challenges in the modeling technique (e.g., simplicity of model assumptions, difficulty of model evaluation, etc.), the modeled landscape resistance explained the genetic distances well (Fig. 3); the strength of gene flow could be largely explained by riverscape variables.

268 It was a somewhat unexpected result that the stream order was identified as the variable with 269 the strongest effect on gene flow. Previous studies of cold-water fish have reported both higher and 270 lower gene flow in streams with higher stream orders (Aunins et al. 2015; Escalante et al. 2018; White 271 et al. 2020). Within the arbitrary study areas, main streams tend to have higher water temperatures and 272 are often unsuitable environments for cold-water fish. On the other hand, in dendritic stream 273 structures, confluences are often known to be stable gene accumulation and source points for stream 274 organisms (Grant et al. 2007; Paz-Vinas and Blanchet 2015), and the downstream passage of those 275 organisms may result in higher gene flow in higher-order streams. In addition, as the main stream 276 tends to be more severely affected by flooding (Han et al. 2007; Koizumi et al. 2013), individuals, 277 especially those of low-mobility species, may have a greater chance of being flushed. There may be 278 differences regarding which processes are predominant depending on the studied species or areas. 279 Summer water temperature (or water temperature fluctuation in G_{ST}) negatively affected gene flow. 280 This is probably because streams with high summer water temperatures and large fluctuations are not 281 suitable environments for C. nozawae (Suzuki et al. 2021), making successful dispersal difficult. 282 While it is not uncommon for studies of cold-water fishes to implicate an association between gene 283 flow and water temperature-related variables (Kanno et al. 2011; Escalante et al. 2018; Hand et al. 284 2016), the present study was able to represent this pattern using more realistic water temperature 285 information. The model from G_{ST} also included the slope and edge length, but the model from D_{PS} did not. We found that topography and distance affected the formation of the long-term population 286

structure as in many other systems (Kanno et al. 2011; Caldera and Bolnick 2008), but that most of the
current gene flow can be explained by the stream order and water temperature. The upstreamdownstream direction did not affect gene flow, probably because environmental conditions influence
the direction of gene flow (Nakajima et al. 2021).

291 Maps displaying modeled landscape resistance from G_{ST} and D_{PS} were visually similar (Fig. 292 2), indicating that the pattern has probably been maintained for a long time. Overall, gene flow is 293 prevented in small tributaries in the southern area. This area displays higher water temperature 294 fluctuations than the northern volcanic area where spring-fed environments are more prevalent 295 (Ishiyama et al. 2023); gene flow in the southern area is probably suppressed by the effects of water 296 temperature. When comparing this geographical pattern with the STRUCTURE barplots, the upper 297 reaches in the non-volcanic area where gene flow is prevented roughly corresponded to the areas 298 where a strong population structure was observed. While the reason for the heterogeneity in the 299 strength of population structure could not be known by the STRUCTURE, a possible explanation was 300 explicitly presented in the gene flow analysis.

301 Under the RCP8.5 scenario, reduced gene flow and increased landscape resistance across the 302 watershed were predicted (Fig. 2d). Since the studied species exhibited a clear genetic structure only 303 in the southern area, the prediction that the northern area will have the same level of gene flow as the 304 present southern area indicates that each tributary within the watershed may experience genetic 305 fragmentation in the future. Nevertheless, gene flow in the northern area was expected to be 306 maintained spatially continuously to some extent, indicating that streams with volcanic watersheds are 307 important for ensuring population connectivity under climate change. A previous study suggested that 308 streams with low summer temperatures behave as source habitats in the watershed (Nakajima et al. 309 2021). Our study showed that these streams may serve not only as source habitats but also as 310 migration pathways in the watershed. As a scenario analysis, Inoue and Berg (2017) considered 311 landscape resistance to be the inverse of the species distribution model (SDM) estimates and predicted 312 that an increased landscape resistance would reduce the gene flow of freshwater bivalves in the future. 313 This is a valuable study that attempts to predict future changes in gene flow; however, it is known that 314 the habitat suitability maps created by SDMs provide poor estimates of genetic resistance, because of

the conceptual differences between habitat selection and entire gene flow (Wasserman et al. 2010,
2012; Sator et al. 2022; Mateo-Sánchez 2015). Actually, in *C. nozawae*, the SDM created in Suzuki et
al. (2021) indicated that the catchment area, analogous to the stream order, had a negative effect on the

318 occurrence of this species, in contrast to the gene flow characteristics estimated in our study.

319 Therefore, genetic population connectivity should be considered separately from habitat suitability.

320 The present study is novel in that gene flow was modeled using riverscape variables identified 321 from genetic data and including water temperature. Our results showed that gene flow in the cold-322 water sculpin is expected to decrease dramatically in response to a changing climate. Therefore, under 323 ongoing climate change, it is important to maintain habitat continuity within the distribution ranges. In 324 particular, it is necessary to consider that the risk is high in sections where water temperature 325 fluctuations are large (such as non-volcanic watersheds). Additionally, while main streams may be less 326 suitable as habitats, they are important as migration corridors. No structures such as weirs should be 327 installed so that drifted individuals can quickly enter suitable habitats.

To obtain more robust results, it would be desirable to increase the number of sampling populations. This study has the potential for further development. For example, demography simulations using inferred landscape resistance (Landguth et al. 2010, 2016) could reveal population viability. Also, combined with habitat quality analyses such as SDMs, population connectivity could be quantified for more detailed predictions from the viewpoint of habitat availability (Saura and Pascual-Hortal 2007). We hope that riverscape genetic modeling will be applied to predict the consequences of environmental changes on a variety of freshwater organisms.

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- 538 Data Availability: Genetic and environmental data generated in this study were deposited at Figshare

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Table 1 Riverscape variables considered in the present study.

Variable	Description	Hypothesis / Ecological importance	Ranges
Summer water temperature ^A	Mean water temperature from July to August (July 2019 to August 2019) [°C]	Streams with low summer water temperatures are suitable for <i>C. nozawae</i> occupancy/survival (Yagami and Goto 2000; Suzuki et al. 2021) and therefore migration may also occur frequently in these streams.	8.32–14.2
Water temperature fluctuation ^A	Standard deviation of the water temperature in one year (September 2018 to August 2019)	Thermally stable streams can be suitable for migration.	1.97–5.09
Drought water discharge ^{BC}	Flow rate on the day when the flow is 355th highest in one year [m ³ /s] (September 2018 to August 2019)	Drought water discharges, which particularly reflect the environmental heterogeneity created by groundwater (Nagasaka and Sugiyama 2010), ensure opportunities to colonize throughout the year.	0.07–6.24
Flow fluctuation ^{AB}	Coefficient of variation in the daily flow rate in one year (September 2018 to August 2019)	Hydrologically stable streams can be suitable for migration.	0.31–0.65
Edge length	Length of edges [km]	Isolation by distance (Wright 1943)	0.05-7.19
Slope	Mean gradient of the edge, i.e., the elevation range divided by edge length	Fish movement and migration are often impeded on steep slopes (Kanno et al. 2011).	4.15–56.5
Strahler's stream order c	Strahler's stream order of the edge	Even in cold-water fish, the mainstem may function as a corridor that facilitates connectivity among populations (White et al. 2020).	1–4
Shreve's stream order BC	Shreve's stream order (link magnitude), i.e., the numbers of confluence points	Given the dendritic arrangement and asymmetry of stream networks, sections with more confluence points upstream may increase the number of migrants passing through (Paz Vinas and Blanchet 2015)	1–62
Catchment area ^{BC}	Cumulative area of the catchment calculated at the midpoint of the edge [km ²]	Sections with larger catchment areas may have more migrants passing through, based on the same principle as that of the stream order. Or, conversely, streams with larger catchment areas have been identified to have lower <i>C. nozawae</i> occupancies (Suzuki et al. 2021) and therefore it is also possible that less migration occurs in sections with larger catchment areas.	6.4–296.3
Direction	Whether the gene flow is toward the upstream (0) or downstream (1) direction	Most stream organisms have higher migration rates in the downstream direction than in the upstream direction (Lamphere and Blum 2012).	0 or 1

542 Variables with the same letters (A, B, C) have high correlations (|r| > 0.7); these variables were not included in the same model.

543 **Table 2** Selected models explaining the strength of gene flow on the edges. Estimated β values

Variables	β	95% CI
(A) $G_{\rm ST}$		
(Intercept)	5.84	5.65, 5.98
Shreve's stream order	4.51	3.96, 5.19
Water temperature fluctuation	-0.48	-0.61, -0.35
Slope	-0.94	-1.15, -0.67
Edge length	-0.42	-0.54, -0.32
(B) $D_{\rm PS}$		
(Intercept)	6.06	4.94, 6.85
Strahler's stream order	2.50	1.72, 3.15
Summer water temperature	-1.42	-2.49, -0.38

544 (median) and their 95% credible intervals (95% CI) are displayed.

545

546 **Figure Legends**

547 Fig. 1 Location of the study area. The blue network indicates the rivers belonging to the Ishikari River

548 system, which has the second largest watershed in Japan and includes the Sorachi River.

549

550 Fig. 2 Maps of the Sorachi River watershed showing the study area (a) and landscape resistance

551 estimated by BGR models (b–d). In panel (a), sampling nodes (sampling sites) and unsampled nodes

552 (major confluences between them), which are delimitations of edges (stream sections), are denoted.

553 Barplots with each sampling node indicate the population structure inferred by STRUCTURE (K = 6).

Landscape resistance is shown in three patterns: long-term gene flow modeled by G_{ST} (b), recent gene

flow modeled by D_{PS} (c), predicted gene flow at the end of the 21st century derived by substituting

556 future water temperatures into the model derived by D_{PS} (d).

557

558 Fig. 3 Isolation by distance and isolation by resistance. The relationship between pairwise genetic

559 distance and cumulative landscape resistance between populations (b, d) is compared to the

relationship with simple waterway geographic distance (a, c). The cases of G_{ST} (a, b) and D_{PS} (c, d) are

shown.





(a) G_{ST} - Isolation by distance







(b) G_{ST} - Isolation by resistance



(d) D_{PS} - Isolation by resistance

