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学位論文内容の要旨 (Summary of dissertation)

博士の専攻分野の名称 博士 (医学) (Degree conferred: Doctor of Philosophy) 氏名 スン シン (Name of recipient: Sun Xin

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学位論文題名 (Title of dissertation)

Studies of roles of NLRC5 expression in cancers on antigen presentation and host

anti-cancer immunity

(癌の抗原提示とホストの抗癌免疫における NLRC5 発現の役割の研究)

[Background and Objectives] NLRC5 (NLR Family CARD Domain Containing 5), also known as CITA (Class I transactivator), is identified as transcription activator for MHC class I and related genes. CD8+ T cells recognize antigens presented by MHC class I and consist of a critical part of anti-cancer immunity. Although a data mining study has identified *NLRC5* as a target for immune evasion in cancer, the molecular mechanisms are not well defined. In this study, we investigated the role of *NLRC5* expression in cancer in modulating host anti-cancer immunity, and efficacy of anti-PD-1 treatment. We sought to enhance our understanding of the interactions involving NLRC5, cancer, and the immune system, which eventually aimed to help the development of novel cancer therapy.

[Materials and methods] To investigate the role of NLRC5 in host anti-cancer immunity, we induced genomic deletion of *Nlrc5* in mouse melanoma B16F10 and mouse breast cancer E0771 cell lines. Furthermore, to investigate whether *NLRC5* is a viable target for development of novel immunotherapy, we developed a novel CRISPR/dCas9-TET1cd based targeted demethylation and activation (TDMa) system to enhance *Nlrc5/NLRC5* expression in mouse melanoma B16F10 and human breast cancer MCF7 cell lines. QRT-PCR and RNA-Seq were performed to determine the mRNA expression levels after knockout of *Nlrc5* or *Nlrc5/NLRC5* activation. Bisulfite sequencing was performed to determine promoter methylation levels. Flow cytometry were performed to determine cancer cell immunogenicity. Subsequently, mouse cell lines were subcutaneously implanted *in vivo* to observe tumor growth. Flow cytometry and immunohistochemistry were performed using *in vivo* tumors to characterize tumor infiltrating lymphocytes (TILs) and tumor cells.

[Results] We found that genomic deletion of *Nlrc5* specifically downregulated MHC class I gene expression in B16F10 and E0771 cells. *Nlrc5* deletion promoted E0771 tumor growth *in vivo* by inhibiting CD8+ T cell infiltration and activation through MHC class I. On the other hand, *Nlrc5* deletion did not affect B16F10 tumor growth without stimulation. This is because B16F10 has low basal MHC class I expression and genomic deletion of *Nlrc5* did not have significant effect on B16F10 basal MHC class I levels. However, we found that *Nlrc5* is required for response to anti-PD-1 therapy in B16F10 tumor transplantation model. Based on these results, we performed transcriptome screening and generated the optimal prediction model for prediction of *Nlrc5* promoter in B16F10 and MCF7 cell lines using CRISPR/dCas9 based system (TDM and TDMa systm). Although targeted demethylation of *Nlrc5* (TDM

system) alone was insufficient to significantly augment MHC class I expression and tumor immunogenicity, we found that the expression of MHC class I is robustly augmented by combination of targeted demethylation activation of *Nlrc5* (TDMa system). RNA-Seq analysis showed that TDMa system induced specific augmentation of expression of *Nlrc5* and dependent MHC class I expression. OT-I co-culture analysis showed that the immunogenicity of B16F10 were augmented by targeted demethylation and activation of *Nlrc5*. *In vivo* implantation of these cells showed that tumor growth is impaired by targeted demethylation and activation and activation of *Nlrc5* in C57BL/6 mice, but not in NUDE mice. CD8+ T cell infiltration and activation was augmented by targeted demethylation and activation of *Nlrc5* in B16F10 not only increased the number of TILs, but it also promoted concentration of *CD8+* T cells into the tumor center. Furthermore, we showed that targeted demethylation and activation of *Nlrc5* sensitized the response to anti-PD-1 treatment in B16F10. Finally, we showed that CD8+ T cell.

[Discussion] Although numerous studies have identified the role of NLRC5 as class I transactivator, most of the investigations were focused its role in the immune system. Few studies investigated the role of NLRC5 in cancers and cancer derived NLRC5 in modulating host anti-cancer immunity. MHC class I is of high priority as target for cancer treatment, there, our research is importance. Here, using both genomic deletion, and targeted demethylation and activation approach, we systemically illustrated the role of NIrc5 in modulating immunogenicity through MHC class I and host anti-cancer immunity in melanoma and breast cancer. For genomic deletion of Nlrc5, consistent with the previous investigations using Nlrc5 knockout mice, we found that Nlrc5 deletion downregulated MHC class I expression levels with or without mIFN- γ stimulation. Our study is the first to characterize the effect of *Nlrc5* in MHC class I antigen presentation pathway to our knowledge. On the contrary, we also induced targeted demethylation and activation of Nlrc5/NLRC5 in B16F10 and MCF7 using the TDMa system that we developed. A previous study characterized the phenotype of NIrc5 overexpression in B16F10 in vitro and in vivo. Our results further confirmed that activation of Nlrc5 enhanced the expression of MHC class I and host anti-cancer immunity. In addition, we found that targeted demethylation and activation of Nlrc5 promoted concentration of CD8+ T cell into the tumor center. Previous studies observed that aggravation of CD8+ T cell into tumor center is correlated with a better prognosis, while the mechanism is left unexplored. Here, we further confirmed that expression of surface MHC class I in cancer is one of the mechanisms that promote CD8+ T cell infiltration into tumor center. Furthermore, we found that targeted demethylation and activation of NIrc5 sensitized response of B16F10 to anti-PD-1 treatment. Previous studies have identified that surface MHC class I levels and NLRC5 expression levels correlates with a good response to immune checkpoint blockade therapy. However, the underlying mechanisms are not well studied. Our results indicate that CD8+ T cell were exclusively activated by anti-PD-1 treatment and targeted demethylation and activation of NIrc5 synergistically enhanced CD8+ T cell mediated anti-cancer immunity, highlighting *Nlrc5* as a therapeutic target to synergistically enhance the efficacy of anti-PD-1 treatment.

[Conclusion] In conclusion, our research deepened our understanding of the role of *Nlrc5* in host anti-cancer immunity. Our strategy to induce and genotyping genomic deletion is solid and can be applied to other genes and cell lines. We show that the CRISPR/dCas9 mediated targeted demethylation and activation strategy of *Nlrc5* is a viable and novel immunotherapy approach to enhance cancer immunogenicity. For future study, optimization of delivery system to effectively induce targeted demethylation and activation *in vivo* could contribute to development of novel immunotherapy.