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Rose Bengal-decorated Rice Husk-Derived Silica Nanoparticles Enhanced Singlet Oxygen Generation for Antimicrobial Photodynamic Inactivation

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ABSTRACT

Rice husks (RHs) are well known for their high silica content, and the RH-derived silica nanoparticles (RH NPs) are amorphous and biocompatible; therefore, they are suitable raw materials for biomedical applications. In this study, rose bengal-impregnated rice husk nanoparticles (RB-RH NPs) were prepared for their potential photosensitization and ¹O₂ generation as antimicrobial photodynamic inactivation. RB is a halogen-xanthene type's PS showing high singlet oxygen efficiency, and the superior photophysical properties are desirable for RB in the antimicrobial photodynamic inactivation of bacteria. To enhance the binding of anionic RB to RH NPs, we conducted cationization for the RH NPs using polyethyleneimine (PEI). The control of the RB adsorption state on cationic PEI-modified RH NPs was essential for RB RH-NP photosensitizers to obtain efficient ¹O₂ generation. Minimizing RB

aggregation allowed highly efficient ${}^{1}O_{2}$ production from RB-RH NPs at the molar ratio of RB with the PEI, $X_{RB/PEL}$ =0.1. The RB-RH NPs have significant antimicrobial activity against S. *mutants* compared to free RB after white light irradiation. The RB-RH NP-based antimicrobial photodynamic inactivation can be employed effectively in treating S. *mutants* for dental applications.

Key words: rice husk; silica; rose bengal; photosensitizer; singlet oxygen; photodynamic therapy

Introduction

Diseases caused by bacteria and viruses are called "infectious diseases. Among these infectious diseases, antimicrobial agents are effective against diseases caused by bacteria because they kill or inhibit the growth of the causative bacteria and other organisms, and they treat various infectious diseases [1,2]. However, the number of "drug-resistant" bacteria that do not respond to conventional antimicrobial agents has been increasing worldwide, and various bacteria resistant to antimicrobial agents have already been identified [3]. Antimicrobial resistance (AMR) has made it increasingly difficult to prevent and treat infections [4]. As the number of resistant bacteria increases, antimicrobial drugs become ineffective, making it more difficult to treat infections that could previously be treated with appropriate therapy and recover from minor illnesses, increasing the likelihood of serious illness and even death. With the COVID-19 coronavirus pandemic, humanity has drawn concern about new antimicrobial treatments for infectious diseases.

As an alternative treatment for infectious diseases, antimicrobial photodynamic therapy (aPDT) provides no selective inactivation of pathogenic microorganisms in response to the present AMR issue [5,6]. The inactivation mechanism of aPDT is based on a photoactivatable photosensitizer (PS) to generate cytotoxic reactive oxygen species under light irradiation of appropriate wavelengths, such as

singlet oxygen (¹O₂) and other reactive oxygen species (ROS). Besides the ability of the non-selective inactivation, the microorganism resistance of aPDT is rarely reported, indicating the possibility of escaping AMR issues [7]. Many efforts have been focused on developing more effective organic PSs for aPDT treatment. Still, PSs have several disadvantages, such as water-solubility, photostability, biocompatibility, enzymatic degradation, and chemical modulation for specific functions.

The composite of PS with nanoparticles (NPs) can improve water-solubility, overcome aggregation issues, and protect PSs from enzymatic degradation [8-11]. Among the variety of NPs, silica-based nanomaterials have been attracted as PS support for PDT applications [12]. Silica NPs (SiOx NPS) are chemically inert and have high water dispersibility and surface modification capability. Various type's silica supports have been proposed for PDT applications, such as mesoporous SiOx NPs with the pore-integrated PSs [13-15], aggregated SiOx NPs with the pore-integrated PSs [16,17], and covalently or physically bound complexes of PSs and SiOx NPs.[18-23]. Most of these SiOx -based nanomaterials are synthetic amorphous silica via thermal (pyrogenic) or wet routes (colloidal, precipitated, and gel) since their particle size, shape, and porosity can be controlled during their synthetic methods. However, precursor solutions such as tetramethylorthosilicate (TMOS) and tetraethylorthosilicate (TEOS) for producing SiOx NPs are not economically and environment-friendly because of their high cost and toxicity.

Rice husks (RHs) are well known for their high silica content, and they contain organic compounds (70–80%) (including cellulose or lignin) and about 20-30% silica [24,25]. Many rice husks are generated yearly as a byproduct of rice cultivation in a cost-effective and eco-friendly process. In recent years, the shift from fossil resources to biomass has attracted attention with the growing awareness of environmental issues such as global warming. Thus, the natural resources-based RHs have gained considerable interest in energy/ecological fields since various chemical products can be derived from

RH biomass, including active carbon, silica, syngas, and biofuel [26-28]. RH-derived SiOx NPs are amorphous and biocompatible; therefore, they are suitable raw materials for biomedical applications [29-31]. However, to our knowledge, there is no study on the encapsulation of photosensitizers into RHderived SiOx NPs for a-PDT.

The present work aimed to develop the composite of Rose Bengal (RB) with RH-derived SiOx NPs for aPDT applications. RB is a halogen-xanthene type's PS showing intense absorption bands in the visible region of 480–550 nm, a high triplet quantum yield of 0.76, a long-lived triplet state ($t_{1/2} = 0.1-0.3$ ms), and a high singlet oxygen quantum yield of 0.75 under 540 nm light irradiation [32]. These superior photophysical properties are desirable for RB in aPDT [13, 15, 18, 20, 22, 33]. We prepared amino-functionalized RH-derived SiOx NPs to conjugate RB through the electrostatic interaction to yield RB-modified RH NPs (RB-RH NPs) for aPDT, and the ¹O₂ generation efficiency of RB-RH NPs depending on the RB loading amount was investigated. Finally, we demonstrate that the RB-RH NPs can reduce the bacterial proliferation of *Streptococcus mutans* after white light-emitting diode irradiation, commonly found in the human oral cavity.

Materials and methods

Materials

N, N-Dimethylformamide (DMF) (assay[HCON(CH₃)₂](GC)min.99.5%(mass/mass)) and Rose Bengal (RB) were purchased from FUJIFILM Wako Pure Chemicals Co. 9,10-Antracenediyl-bis(methylene) dimalonic acid (ABDA, \geq 90.0%), poly(ethylene imine) 800 (PEI 800) Polyethylenimine,branched (average Mw ~800 by LS, average Mn ~600 by GPC) was used. Methanol (99.8%) was purchased from Tokyo Kasei Kogyo Co. Pure water was purified by a water distillation apparatus (aquarius RFD250, ADVANTEC). The ultrasonic cleaner was SANSYO's D-SONiC.

Instrumentation and characterization

UV-visible (UV-Vis) absorption spectra and Fourier transform infrared (FT-IR) spectra were measured using the V-670 spectrometer (JASCO, Japan) and the FTIR 4200 spectrometer (JASCO, Japan) with an attenuated total reflection (ATR) instrument (ATR PRO ONE, JASCO, Japan)., respectively. The zeta potential was evaluated using the Zetasizer Nano ZS (Malvern Instruments Ltd., GB). Before the zeta potential measurements, all samples were dispersed by ultrasonication for 10 minutes. Scanning electron microscopy (SEM) and transmission electron microscopy (TEM) images were captured using a field-emission SEM apparatus (JSM-6700, JEOL, Japan) operated at an acceleration voltage of 5.0 kV and using a JEOL 1400 microscope at 120 kV. Thermogravimetric analysis (TGA) was performed using a TGA system (Thermo Plus EVO, Rigaku, Japan) at a heating rate of 5 °C/min under an airflow. To determine surface area, pore size, and pore volume, N₂ adsorption and desorption isotherm measurements were conducted using a Microtrac BELSORP-Mini instrument at 77 K under continuous adsorption conditions. Brunauer–Emmett–Teller (BET) and Barrett–Joyner–Halenda (BJH) analyses were used. The crystal nature of RH NP powders was analyzed by X-ray diffraction (XRD, D2 PhaseRBruker AXS GmbH, Karlsruhe, Germany).

Preparation of amorphous silica NPS from rice husks (RH-NPs)

Rice husks harvested in Shiga, Japan were used as input raw materials to produce high-purity amorphous silica materials, according to the literature, where the citric acid solution leaching treatment and air combustion of rice husks are performed to remove the metallic impurities without the use of strong acids such as sulfuric acid [34, 35]. About 30 g of husks were immersed in 5 wt% citric acid solution of 500 mL at 50 °C for 30 minutes. The water rinsing treatment on acid-leached husks was carried out to remove the acid solution from rice husks and followed by water washing three times. The washed rice husks were dried at 100 °C for 1 hour in the drying oven under an air atmosphere and then

combusted at 800 °C for 30 minutes in the muffle furnace. The airflow rate in the combustion was 0.42 ml/s using a small air compressor.

Synthesis of PEI-modified RH NPs

To enhance the binding of rose bengal to RH NPs, we performed cationization of RH NPs using polyethyleneimine (PEI). 0.4 g of RH NPs dispersed in 1mL methanol were added into 5 mL methanol solution with 7.7g PEI, and the mixture was stirred at 650 rpm all night. The solid products were isolated by centrifugation. After removing the supernatant, we added 10 mL methanol for washing, and then did the centrifugation again and repeated this procedure three times. Finally, we obtained PEI-modified RH NPs by drying the washed product under reduced pressure for 24 hours.

Preparation of rose bengal modified RH NPs (RB-RH NPs)

The RB-RH NPs are schematically summarized in Figure 1. RB-RH NPs were prepared as follows:

20 mg of PEI-modified RH NPs was added to aqueous RB solutions (2 mL) of various concentrations $(3.5 \times 10^{-2} \text{ mg/mL}, 7.0 \times 10^{-2} \text{ mg/mL}, 1.75 \times 10^{-1} \text{ mg/mL}, \text{ and } 7.0 \times 10^{-1} \text{ mg/mL})$ [i.e., molar ratio of RB/PEI polymer, $X_{RB/PEI} = 0$, 0.1, 0.25, and 1.0, respectively], and the mixture was stirred at 500 rpm for 1 h. Here, we regard the molecular weight of PEI as ~ 800. The anionic RB binds to the cationic PEI-modified RH NPS, producing RB-RH NPs. The RB-RH NPs were obtained as a precipitate after centrifugation at 600 rpm for 5 min, followed by washing with 25 mL water, and further centrifugation multiple times until the absorbance of the washingolution was measured to be less than 0.01. Finally, dried powdered RB-RH NPs were obtained by vacuum drying.



Figure 1 Preparation procedures of rose bengal modified RH NPs (RB-RH NPs)

Detection of ¹O₂

ABDA was used as a probe to detect ${}^{1}O_{2}$ under excitation by a white light-emitting diode (LED) (SPF-D2, Shodensha, Japan). ABDA can react irreversibly with ${}^{1}O_{2}$, which causes a decrease in the ABDA absorption at 378 nm. An aqueous suspension of RB-RH NPs (1.8 mL, ~ RB absorbance of 0.1 at 665 nm) was mixed with ABDA in DMF (0.5 mM, 0.2 mL), and the final concentration of ABDA was adjusted to be 50 μ M according to the references. The RB absorbance of around 0.05 at 570 nm was selected to obtain dispersed RB-RH NP solution. The ABDA absorbance of the solution at 378 nm was monitored at 5 min intervals by UV–Vis spectra. The possible absorption from the RB-RH NP scattering backgrounds was subtracted from the UV–Vis spectra of RH NPs.

Antimicrobial effects of RB-RH NPs under white LED irradiation

RB-RH NPs were dispersed in a suspension of Streptococcus mutans (ATCC 35668) and dispensed into 48-well plates. Before incubation, the suspension was irradiated with white LED for 1 min. After incubation at 37 °C under anaerobic conditions for 24 h, the bacterial turbidity was determined using a turbidimeter (CO7500 Colorwave, Funakoshi Co., Ltd., Tokyo, Japan) at 590 nm. Scheffe's test performed the statistical analysis, and P values <0.05 were considered statistically significant. All

statistical analyses were performed using a software package (SPSS 11.0, IBM Corporation, Armonk, NY). For comparison, we also examined antibacterial tests on RB only. We adjusted the RB concentration to equal the RB concentration in RB-RH NPs ($X_{RB}/_{PEI} = 1.0$), using the RB adsorption mass per mg of RH NPs: 6.8×10^{-2} mg/mg.

Results and discussion

Characterization of RH NPs

The XRD pattern of RH NPs is shown in **Figure 2a**. There are no peaks derived from crystalline silica structure. The pattern is characteristic of amorphous silica with a broad diffraction peak near $2\theta = 22^{\circ}$, which is consistent with typical amorphous silica [16, 34, 35]. The SEM image of RH NPs shows the size and shape were not controlled during their preparation, but the RH NPs consisted of fused NPs with small sizes of ca. 30–50 nm (**Figure 2b**). The nitrogen adsorption and desorption isotherms of RH NPs had a BET surface area of 196 m²·g⁻¹, a pore volume of 0.32 cm³·g⁻¹, and an average pore size of 6.47 nm (**Figure 2c**). Herein, we suppose the RH NPs with a diameter of ca. 30 nm are non-porous solid. In that case, the specific surface area can be ca. 84 m²/g, assuming the spherical particles without any surface area of RH NPs than the calculated value of non-porous solid suggests porous structures for RH NPS. The TEM image of RH NPs supports the porous nature of nano-aggregates of NPs consisting of single-nanosized NPs (<10 nm) (**Figure 2d**).

PEI-modified RH NPs

The PEI amount adsorbed on RH NPs was examined using TGA analysis. The TG curve of the PEImodified RH-NPs showed weight loss from 0-150°C from the water evaporation and further weight decrease from the PEI degradation above 150 °C. The amount of PEI adsorbed on RH NPs is defined as a weight loss of around 150-500°C in the TG curves (**Figure S1**). The amount of PEI adsorbed on RH-NPs was 7.0×10^{-8} mol (PEI polymer unit) /mg (RH NPs) for the PEI-modified RH-NPs. We used the PEI-modified RH-NPs for further RB modification.

Figure 3a shows FT-IR spectra of PEI-modified RH-NPs, confirming the presence of PEI on the RH-NPs from the observation of the N-H (1500 cm⁻¹) and C-H bands (2800-3000 cm⁻¹) of PEI (**Figure 3a**). The zeta potential value of unmodified RH NPs was -32 mV, and the negative value originates from the dissociation of the silanol group on the silica surface. The PEI-modified RH-NPs showed a positive value of +32 mV (**Figure S2**), and the charge reversal indicates the cationic PEI polymer adsorbed on negatively charged RH NPs.

The N₂ adsorption/desorption isotherms of RH NPs and PEI-modified RH NPs are shown in **Figure 2c**. After the PEI adsorption on the RH NPs, the specific surface area decreases from 196 m²g⁻¹ to 146 m²g⁻¹. The pore volume of 0.31 cm³·g⁻¹ and average pore size of 8.38 nm were evaluated for PEI-modified RH-NPs, similar to unmodified RH-NPs. Thus, the PEI-modified RH-NPs maintain a high specific surface area even after the adsorption of PEI on the RH NPs.



Figure 2 (a) XRD pattern of RH NPs. (b) SEM image of RH NPs. (c) Nitrogen adsorption and desorption isotherms of RH NPs (red circles: adsorption and red circles: desorption) and PEI-RH NPs (green circles: adsorption and blue circles: desorption).

RB-modified RH NPs (RB RH-NPs)

The control of the RB adsorption state on silica is essential for RB RH-NP photosensitizers to obtain efficient ${}^{1}O_{2}$ generation because the aggregation of RB quenches ${}^{1}O_{2}$ production. Thus, we prepared RB-RH NPs with different loading amounts of RB. We varied the molar ratio of RB with the PEI polymer unit (Mw ~800), X_{RB/PEI} as follows: X_{RB/PEI} =0.1, 0.25, and 1.0. **Figure 3b** shows normalized absorption spectra of free RB and suspensions of RB-RH NPs with different values of X_{RB/PEI}. The free RB shows well-defined adsorption at 549 nm with a shoulder absorption at 514 nm, as expected for RB dyes. The two absorption bands of free RB in water are consistent with the reported ones: monomeric RB (549 nm) and dimeric (or aggregation) RB (515nm) bands [36-38]. The absorption ratio of monomeric and dimeric (or aggregation) RB, A_m/A_d can be regarded as a measure of the RB aggregation. The values of (A_m/A_d) were the followings: 2.97 for RB only, 2.24 for $X_{RB/PEI} = 0.1$, 1.79 for $X_{RB/PEI} = 0.25$, and 1.65 for X_{RB/PEI} =1.0. The decreasing ratio with the RB loading amount suggests the aggregation of RB molecules due to the dye-dye interactions in the RB-RH NPs, which are particularly dominant in the case of RB-RH NPs ($X_{RB/PEI} = 0.25$ and 1.0). The relatively higher value of ($A_{m/Ad}$) for RB-RH NPs (X_{RB/PEI} =0.1) suggests the suppression of the aggregates. We also observe that the RB adsorption into the surface of PEI-modified RH NPs causes the red shift of the monomeric RB absorption from 549 nm (free RB) to 564 nm (RB-RH NPs, $X_{RB/PEI} = 0.1$). This red-shifted absorption at 564 nm was consistent with the absorption of monomeric RB on the amine-modified silica surface, which has been reported to be the molecular deformation of RB induced by the interaction of silica surfaces [13]. The FT-IR spectra of RB-RH NPs ($X_{RB/PEI} = 0.1$) support the physical adoption of RB on RH NPs without the chemical structural change, where the RB absorption bands are similar between RB and RB-RH NPs (Figure 3c). However, we observed that the C=O vibration band of carboxyl groups in RB molecules shifted from 1610 cm⁻¹ to 1630-1660 cm⁻¹ by the RB adsorption into PEI-modified RH NPs. The result supports the electrostatic interaction between the carboxylate groups of RB and PEI-modified silica surface, as schematically shown in Figure 3d. Thus, we can say that the optimal RB loading in monomeric form is $X_{RB/PEI} = 0.1$ for RB-RH NPs.



Figure 3 (a) FT-IR spectra of PEI, PEI RH-NPs, and RH-NPs. (b) Normalized absorption spectra of free RB and suspensions of RB-RH NPs with different values of $X_{RB/PEI}$ ($X_{RB/PEI} = 0.1, 0.25$, and 1). (c) FT-IR spectra of RB and RB- RH NPs. (d) Schematic picture of possible interaction between the electrostatic interaction between the carboxylate groups of RB and PEI-modified silica surface.

Evaluation of ${}^{1}O_{2}$ generation by RB-RH NPs

The ¹O₂ generation efficiency can be influenced mainly by the concentration of RB molecules bound on the RH NPs. If too few RB molecules are deposited on the RH matrix, the resultant NPs may be few ¹O₂ generations. On the other hand, if too many RB molecules are densely deposited on the RH NPs, the ¹O₂ generation can be reduced through the ¹O₂ self-quenching due to the RB aggregation. Thus, we have investigated the relationship between the loading amount of RB onto PEI-modified RH NPs and their ${}^{1}O_{2}$ generation rate of RB-RH NPs. We evaluated the ${}^{1}O_{2}$ generation efficiency of RB-RH NPs under white light irradiation using the ${}^{1}O_{2}$ detection probe ABDA. We can monitor the ${}^{1}O_{2}$ generation from RB-RH NPs from the absorbance decrease of ABDA by preferential reaction with ${}^{1}O_{2}$.

Figures 4a-4c show the UV–Vis absorption spectra of (a) RB, (b)RB-RH NPs ($X_{RB/PEI} = 0.1$), and (c) RB-RH NPs ($X_{RB/PEI} = 1.0$) in the presence of ABDA under white light irradiation. The ABDA absorbance at around 300–400 nm decreased during the light irradiation, indicating ¹O₂ generation from RB-RH NPs. We evaluated the decreasing of ABDA absorbance by reacting ¹O₂ as the measure of ¹O₂ generation as follows:

$$Q = \ln \left[A_{ABDA(t=0)} / A_{ABDA(t)}\right] / A_{RB}$$

An _{ABDA(t=0)} and A _{ABDA(t)} are the ABDA absorbance at 379 nm before and after the light irradiation, respectively. A_{RB} is the peak absorbance of RB at 550-570 nm. **Figure 4d** shows the Q values of RB-RH NPs as a function of light irradiation time. A linear increase in Q values was observed in all cases. The slope of the line (= S) can be regarded as the ¹O₂ generation rate of RB-RH NPs, as shown in **Figure 4d**. The S values for RB and RB-RH NPs (X_{RB/PEI} =0.1, 0.25, and 1.0) and RB only are summarized in **Figure 5a**. The S values of RB-RH NPs showed the maxima at X_{RB/PEI} =0.1, which is more prominent than that of RB. This indicates that RB-RH NPs have maximum ¹O₂ generation efficiency at X_{RB/PEI} =0.1, corresponding to the optimal RB loading in the monomeric form on RB-RH NPs, as described above. We also examined the light intensity-dependent behavior of the ¹O₂ generation rate for the RB-RH NPs (X_{RB/PEI} =0.1). With a higher intensity of the incident LED, a faster generation of ¹O₂ was accomplished, confirming the photo-mediated process (**Figure 5b**). Thus, it is easy to control the ¹O₂ generation rate by changing the incident LED intensity. In contrast, the S value of RB-RH NPs (X_{RB/PEI} =0.25) is smaller than that of free RB (i.e., 0.25 times). The lower ¹O₂ generation rate of the RB-RH NPs (X_{RB/PEI} =0.25 and 1.0) can be attributed to the RB aggregation within the RB-RH NPs. Although

the enhanced mechanism of adsorbed RB on PEI-modified RH NPs at $X_{RB/PEI} = 0.1$ is unclear at present, the coordinating bonds between the carboxylate groups of RB and the amino groups of PEI as shown in **Figure 3d** could restrict the relaxation pathway of the excited RB-RH NPs by locking the RB in place, leading to fewer nonradiative decay pathways. The inhibition of nonradiative decay pathways in RB-RH NPs may enable enhanced ${}^{1}O_{2}$ generation.



Figure 4 UV–Vis spectra of ABDA during white LED irradiation in the presence of (a) RB, (b) RB-RH NPs ($X_{RB/PEI} = 0.1$, and (c) RB-RH NPs ($X_{RB/PEI} = 1.0$). (d) Normalized ABDA absorbance at 378 nm (*Q*) as a function of irradiation time in the presence of RB or RB-RH NPs ($X_{RB/PEI} = 0.1$, 0.25, and 1.0).



Figure 5 (a) ${}^{1}O_{2}$ generation efficiency (*S*) of RB and RB-RH NPs(($X_{RB/PEI} = 0.1, 0.25, and 1.0$). (b) The values of *S* as a function of the white light irradiation intensity.

aPDT Using RB-RH NPs under white LED irradiation

The RB-RH NPs have the ${}^{1}O_{2}$ -generation capability under white LED irradiation. Therefore, we applied the RB-RH NPs (X_{RB/PEI} =0.1) for a-PDT against gram positive bacteria, *S. mutans*. **Figure 6a** shows turbidities of S. mutans cultures exposed to increasing doses of RB-RH NPs (X_{RB/PEI} =0.1) with LED irradiation at a range of concentrations (0 (absence), 1, 5, and 10 mg/mL). The turbidity directly reflects the growth over time of the bacterial population in the suspension. Low turbidity indicates a small number of bacteria in the suspension. With LED irradiation, exposure to RB-RH NPs resulted in the decrease of turbidities. Thus, photoexcited 1 mg/mL RB-RH NPs significantly reduced the turbidity of S. mutans cultures (Figure 6a). These results suggested that Lys-Au NCs/RB at concentrations of 1 mg/mL was effective for aPDT in this experimental system. To examine the conjugating effect of RB and RH NPs, we compared aPDT activity of RB-RH NPs and RB alone. The RB-RH NPs (X_{RB/PEI} =1.0) were estimated to include 6.8×10^{-2} mg RB /RH NP mg from the RB adsorption experiment. Even

without LED irradiation, the decrease in turbidity was observed after 24 h in the presence of the RB-RH NPs (the concentration of RB, $6.8\mu g$ /mL), indicating the antimicrobial effect of RB-RH NPs themselves without the LED light. However, under white LED irradiation in the presence of the RB-RH NPs, a further decrease in the turbidity of the bacterial suspensions occurred, indicating the suppression of bacterial growth (i.e., a-PDT activity). The ${}^{1}O_{2}$ generated by photoexcited RB-RH NPs likely suppressed S. mutans growth. For comparison, we also examined antibacterial tests on RB only. We adjusted the RB concentration to equal the RB concentration in RB-RH NPs (X_{RB/PEI} =1.0), using the RB adsorption mass per mg of RH NPs: 6.8×10^{-2} mg/mg. The aPDT activity of RB-RH NPs was higher than that of free RB by the combined effect of RB with RH NPs (**Figure 6B**). The result demonstrates that RB-RH NPs are promising photosensitizers for aPDT applications. Thus, our RB-RH NPs appear to possess greater antibacterial potency against S. mutans than that obtained with RB alone.



Figure 6 (a) Dose-dependent antimicrobial effects of RB-RH NPs ($X_{RB/PEI} = 0.1$) on Streptococcus mutans (ATCC 35668) under white LED irradiation for 1 min after 24 h incubation (n = 5, mean ± standard deviation). *P < 0.05 vs other groups. (b) Antimicrobial effects of RB ($6.8 \times 10^{-2} \text{ mg/mL}$) and RB-RH NPs ($X_{RB/PEI} = 1.0$) (1 mg/mL) on Streptococcus mutans (ATCC 35668) under white LED irradiation for 1 min after 24 h incubation (n = 5, mean ± standard deviation). *P < 0.05 vs other groups.

Conclusion

We fabricated a novel photosensitizer based on rose Bengal (RB)-decorated rice husk-derived silica nanoparticles (RH NPs) to enhance ${}^{1}O_{2}$ generation for antimicrobial photodynamic inactivation. The polyethyleneimine (PEI)-modified RH NPs to conjugate RB through the electrostatic interaction were prepared to yield RB-modified RH NPs (RB-RH NPs). We have investigated the relationship between the loading amount of RB onto PEI-modified RH NPs and their ${}^{1}O_{2}$ generation rate of RB-RH NPs (i.e., ${}^{1}O_{2}$ generation efficiency). The RB-RH NPs have maximum ${}^{1}O_{2}$ generation efficiency at the molar ratio of RB/PEI polymer, X_{RB/PEI} =0.1, corresponding to the optimal RB loading in the monomeric form on RB-RH NPs. In contrast, high loading RB with more than of X_{RB/PEI} =0.25, the overall ${}^{1}O_{2}$ generation efficiency is reduced through the ${}^{1}O_{2}$ self-quenching due to the RB aggregation. The RB-RH NPs have significant antimicrobial activity on *Streptococcus mutans* when compared to free RB after white light irradiation. The RB-RH NP based aPDT can be employed effectively in the treatment of *Streptococcus mutans*.

Declarations

Conflict of interest: The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Supplementary Information: The online version contains supplementary material available at https://doi.org/10.1007/xxxxxxx.

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