



HOKKAIDO UNIVERSITY

Title	Larval Diapause in <i>Chymomyza costata</i> (Diptera: Drosophilidae) I. Effects of Temperature and Photoperiod on the Development
Author(s)	ENOMOTO, Osamu; 榎本, 修
Citation	Low temperature science. Ser. B, Biological sciences, 39, 21-29
Issue Date	1982-03-25
Doc URL	https://hdl.handle.net/2115/19115
Type	departmental bulletin paper
File Information	39_p21-29.pdf



Larval Diapause in *Chymomyza costata* (Diptera : Drosophilidae) I. Effects of Temperature and Photoperiod on the Development¹

Osamu ENOMOTO²

榎本 修

Abstract *Chymomyza costata* is one of few drosophilid species known to commit larval diapause. The thermo-photoperiodic induction of diapause is released by rearing conditions of 18°C/LD 14:10 or shorter photoperiods. At a high temperature, 23°C, diapause is not induced even in a short photoperiod, LD 10:14. The stage sensitive to photoperiodic stimuli for induction of diapause lies at 18°C between 10 to 15~20 days after being oviposited.

Introduction

According to the phenological records of Drosophilidae hitherto accumulated, adult diapause is prevailing in the Holarctic region. Pupal diapause is known only in *Drosophila alpina* (11, 12), and larval diapause in a few species, *Chymomyza amoena* (1), *Chy. distincta* (5), *Chy. costata* (7), and *Scaptodrosophila deflexa* (2). Consequently, the studies on larval diapause have less advanced than those on adult diapause. In Finnish strains the mechanisms controlling larval diapause of *Chy. costata* was reported as entirely temperature-dependent involving some geographical variation but with no reference to the role of photoperiodism (7). Recently the positive effect of photoperiod on larval diapause was confirmed in some Finnish strains (15). The present paper deals with the role of photoperiod and temperature on the development of *Chy. costata* Zetterstedt in Hokkaido, northern Japan.

Materials and Methods

Chy. costata is known from northern, central and western Europe, Hungary, western Russia, Japan (7, 14) as well as Arctic America (17). The stock used for experiments are offsprings of several females collected on July 27, 1978, by net-sweeping or direct aspirating at stumps in recently cut-over areas and timberyards nearby Jozankei, a mountaneous suburb of Sapporo. The parental flies were allowed to oviposit on the culture medium (sugar 50, agar 14, yeast 80, water 1000) in a milk bottle kept at room temperature. Then eggs were daily collected and sterilized with 70%

¹ Received for publication November 30, 1981. Contribution No. 2407 from the Inst. Low Temp. Sci.

² The Institute of Low Temperature Science, Hokkaido University, Sapporo, 060

alcohol for about five minutes to prevent mold. Sixty eggs were put in each glass vial (3 cm diameter, 10 cm height) with the culture medium (10 ml), cotton-plugged, and kept in eight different thermo-photoperiodic conditions; $18\pm 1^\circ\text{C}/\text{LD } 8:16$, $10:14$, $12:12$, $14:10$, $16:8$ and $18:6$, $23\pm 1^\circ\text{C}/\text{LD } 10:14$ and $16:8$. The light cycles were administered by white fluorescent tubes. In each condition, their developmental stage was identified every two days under a dissecting microscope. The eggs remained unhatched on 2nd day (23°C) and 4th day (18°C) were excluded from the count as considered unfertilized. The identification of larval instar was made by measuring the length of mandibular hooks. In each measurement all individuals (30~60; variable by survival ratio) in one culture vial were examined and abandoned afterward to avoid the invasion of mold, which was the main cause making successive observations of larval growth difficult.

Another series of experiment was set up in order to elucidate the period sensitive to the photoperiodic stimuli inducing diapause. Under 18°C the immatures were transferred from longer (LL) to shorter (LD 10:14) photophase and *vice versa* at 5, 10, 15, 20, 25 and 30 (only from shorter to longer) days since being oviposited. The number of pupated individuals was daily counted in each condition. The count was continued at least for 50 days.

Further, the ovarian development of flies in various photocycles, the criterion hitherto employed to judge the incidence or not of reproductive diapause, was checked as follows: Until emergence the immatures were reared at $18\pm 1^\circ\text{C}/\text{LD } 16:8$. On the first day of adult emergence, the flies were separated into eight groups and each incubated under the following

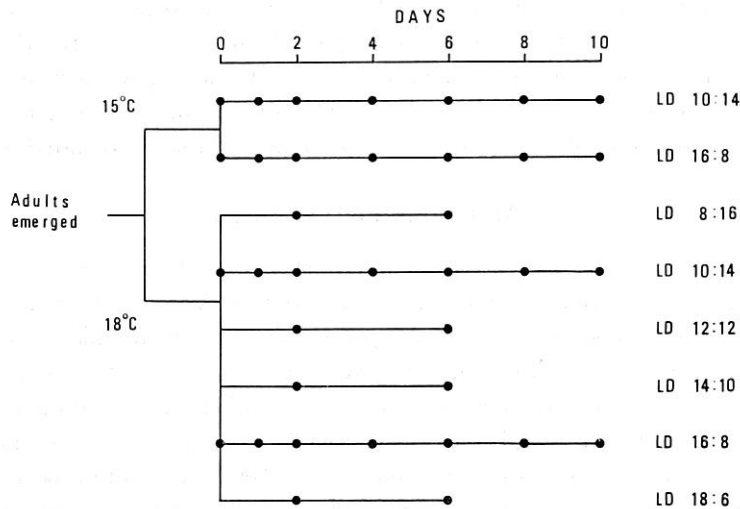


Fig. 1. The dissection schedule of adult females in various thermo-photoperiodic conditions. Adults were transferred from the stock condition, $18\pm 1^\circ\text{C}/\text{LD } 16:8$, soon after emergence. About 30 females were dissected on each examining date (black circle).

conditions; $15 \pm 1^\circ\text{C}/\text{LD } 10:14$ and $16:8$, $18 \pm 1^\circ\text{C}/\text{LD } 8:16$, $10:14$, $12:12$, $14:10$, $16:8$ and $18:6$. About 30 individuals were reared in each glass vial (3 cm diameter, 10 cm height) containing culture medium (5 ml) and females were dissected to classify their ovarian conditions (20) according to the schedule shown in Fig. 1. About 30 females were examined at each dissection.

Results and Discussion

Figure 2 shows the effects of photoperiod on the development of immatures at 18°C . Eggs, the first instar and the second instar larvae showed no or little difference in their growth rate among the six photoperiodical cycles adopted. All eggs hatched by 4 days, almost all first instar larvae moulted by 8 days and nearly all second instar larvae moulted by 16 days after being oviposited. The third instar larvae, however, showed a significant prolongation of larval stage at LD $14:10$ and shorter photoperiodic regimes. Larvae, having committed the developmental suppress in shorter photophases, did not begin the pupation at least by 50 days under the same photoperiodic condition. Especially in LD $10:14$ the developmental suppress succeeded at least for 120 days. At longer photophases, LD $16:8$ and $18:6$, the third instar larvae began to pupate by 18 days and 26 days, respectively. But the onset and frequency of pupation were quite variable among culture vials even at the same photoperiodic condition. Adult emergence began by about 40 days in both photophases, though neither the onset nor the frequency showed synchronization among culture vials and individuals within a culture vial.

Figure 3 shows the development of the immatures both in longer (LD $16:8$) and shorter (LD $10:14$) photophases at 23°C . No interruption of growth was detected in either longer or shorter photophases, though the frequency of the pupation was lower in the latter.

Thus the experimental induction of developmental suppress or diapause at the third larval instar was easily realized by the artificial photoperiodic cycles LD $14:10$ or shorter under 18°C . Under longer photophases, though larvae did not inevitably interrupt further development, timing of the initiation of pupation was quite irregular. This was probably due to other environmental factors such as humidity or undetectable invasion in culture medium of microorganisms, either harmful or nutritive for larval growth. Although the malt medium was reported to serve good nutrition for rearing this species in Finland (10), the preliminary experiment by the Japanese strain did not yield better result. Thus the other factors, which were not considered in this study also seemed to affect the diapause incidence. Such an integral influence for the induction is a characteristic feature of diapause in many insects (6, 13, 18). Nevertheless, photoperiod is considered to be in the primary factor forecasting the approach of adverse conditions in the

favourable season (16). It is well known that temperature affects the photoperiodical response of diapause or alters the critical daylength (3, 4, 8, 16). The larvae reared at high temperatures seem not to enter diapause even in short photophases. The retardation of diapause by high temperatures is also reported in several adult diapausing drosophilid species (8, 9). The

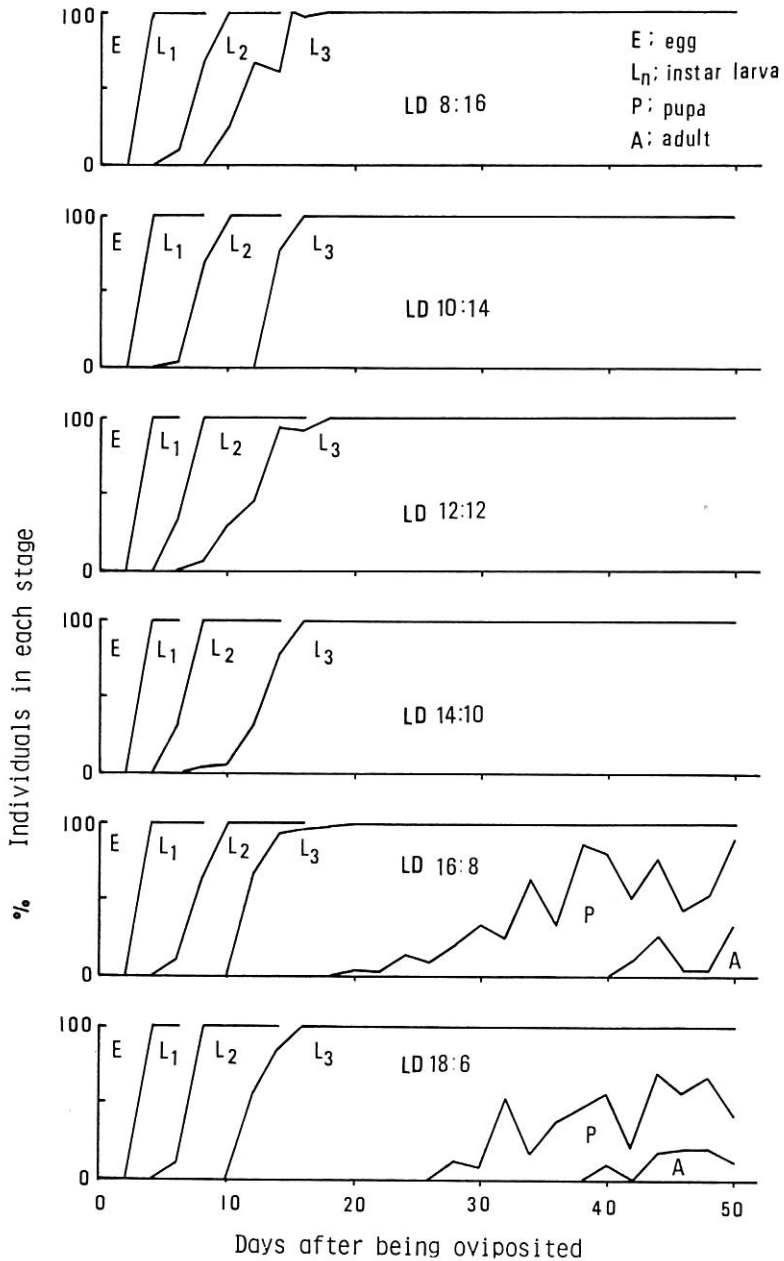


Fig. 2. Development of immatures at different photophases under $18 \pm 1^\circ\text{C}$

critical daylength lying approximately at LD 15:9 under 18°C and its disappearance under a high temperature, 23°C, is ecologically interpreted as follows: The natural daylength in Sapporo exceeds 15 hours a day from early June to mid July. This period seems too short to explain the adult collection records because the present data on natural population, though limited, show the production of newly emerged flies in May, June, July, August and even in early September.

However, the flies emerged in May and June are considered to be overwintered individuals and therefore free from the photoperiodic condition inducing diapause. Further, the production of newly emerged flies in August and early September in spite of the shorter daylength is considered to be resulted by high temperature during July and August (monthly mean temperature; 21.2 and 21.7°C, respectively recorded by the Sapporo Meteorological Observatory). Thus the temperature compensation of the photoperiodic response seems to be significant in natural population.

Figure 4 shows the effects of transference from a long (LL) photocycle to a short one (LD 10:14) or *vice versa* on the duration spent for pupation at 18°C. When larvae transferred from a shorter to a longer photoperiodic condition at 5 and 10 days after being oviposited, the mean duration of entire egg and larval stages did not significantly differ from that of LL. The larvae, transferred from a shorter regime to longer one at 15, 20, 25 and 30 days, differed significantly in their larval duration from nondiapausing individuals. While individuals transferred from LL to LD 10:14 at 5 and 10 days after being oviposited seemed to enter the larval diapause, the larvae transferred at 15, 20 and 25 days seemed not to be committed the developmental suppress. The transference experiment shows that first ten days, including egg, the first instar and a part of the second instar, are insensitive both to long and short photoperiods. After 15 days exposure to longer photoperiods, larvae became also insensitive to shorter photoperiods, mostly completing pupation, though with a slight delay. Further, in the larvae spent their first life span in a shorter photoperiod for more than 15 days, the pupation after exposure to a longer photoperiod began with gradual delays according to the duration kept in the shorter photoperiod. Thus it is concluded that even after 15 days the larvae are sensitive to the shorter photoperiod for diapause maintenance or increasing the diapause intensity.

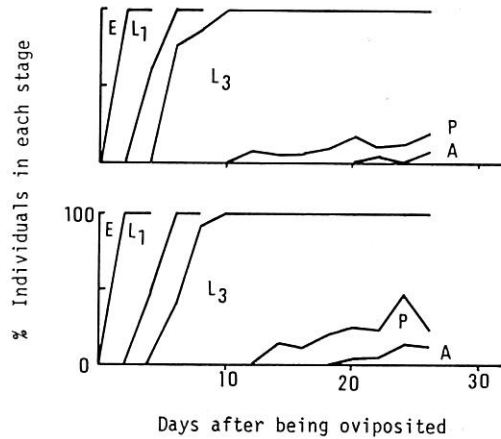


Fig. 3. Development of immatures at two different photoperiods under $23 \pm 1^\circ\text{C}$ LD 10:14 (top) and 16:8 (bottom)

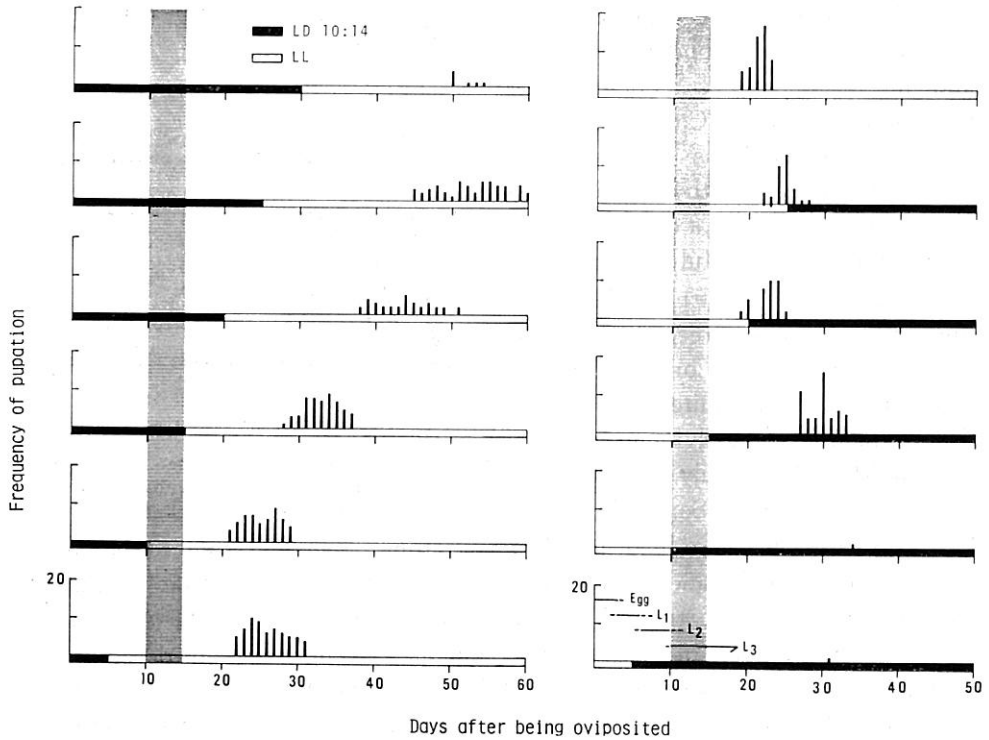


Fig. 4. Effects of transference from short (LD 10:14) to long (LL) photoperiod or *vice versa* on the duration by the pupation. Vertical bars shows the number of pupation on each day. Shaded area indicates the photo-sensitive stage.

In other words, the main sensitive stage to short photoperiods inducing diapause is laid after 10 days, which corresponds to the latter part of the second and the early one of the third instar. After 15 days the larvae are still sensitive to the cumulated effect of the photoperiods that enhances the maintenance or intensity of diapause, though they are no more capable to enter in diapause.

Figure 5 shows the percentage of undeveloped ovaries in females dissected about 30 for each experiment. Under 18°C and 15°C the ovarian development proceeded without interruption both in two photoperiodic regimes LD 10:14 and 16:8. Most ovaries matured at least within 4 days after emergence at 18°C and 10 days at 15°C. The duration spent for ovarian maturation was identical between two photoperiodic regimes. Fig. 6 shows the frequency of undeveloped ovaries under various photoperiods at 18°C. All photoperiodic regimes studied here were ineffective to interrupt the ovarian development. The similar inability to respond the photoperiodic stimuli in controlling the ovarian maturation was reported in several domestic *Drosophila* species in Hokkaido, hence they are regarded as incapable of outdoor overwintering (19). *Chy. costata*, therefore, should be considered

not to overwinter in the adult stage. Although precise information on the evolution of photoperiodism and diapause is not yet available for this species, the manifestation of photoperiodism in a unique developmental stage seems reasonable compared to other adult diapausing drosophilids, for it enhances the synchronization of life cycle in a population.

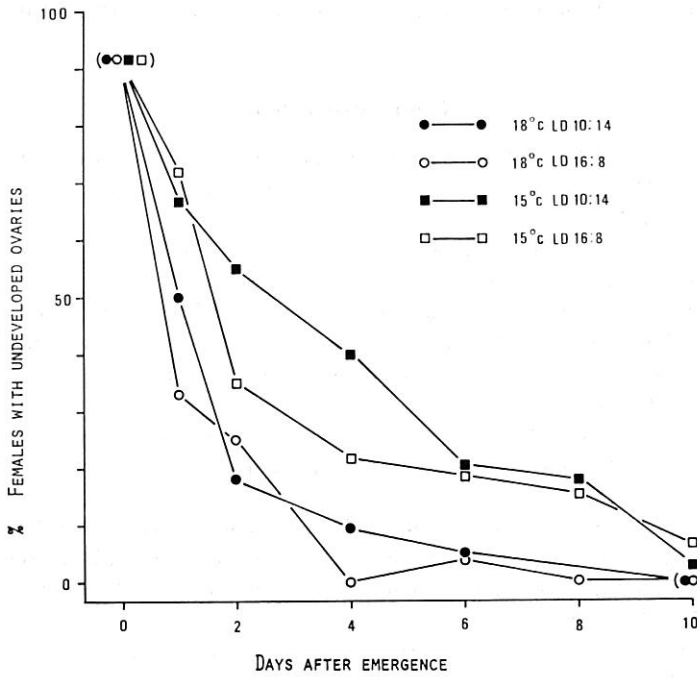


Fig. 5. Effects of temperature and photoperiod on the ovarian development. Percentage of females with undeveloped ovaries plotted as a function of time

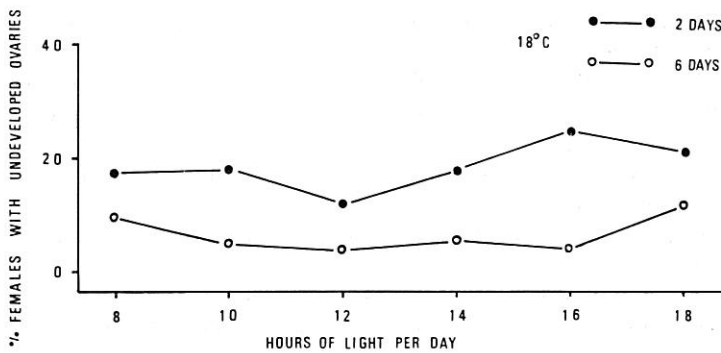


Fig. 6. Effects of photoperiod on the ovarian development in females under different photocycles at $18 \pm 1^\circ\text{C}$

Acknowledgments

I wish to express my sincere thanks to Prof. Shôichi F. Sakagami for his reading of the manuscript. Cordinal thanks are also due to Mr. Masanori J. Toda and Dr. Masahito T. Kimura for their valuable suggestions and helps and to Mrs. Ikuko Numasa for her preparation of the manuscript. Dr. Ari Riihimaa kindly informed me the knowledge on the Finnish populations.

Literature Cited

- 1) Band, H. T. and R. N. Band 1980 Overwintering of *Chymomyza amoena* larvae in apples in Michigan and preliminary studies on the mechanism of cold hardiness. *Experientia*, 36(10): 1182-1183.
- 2) Basden, E. B. 1954 Diapause in *Drosophila* (Diptera: Drosophilidae). *Proc. R. ent. Soc. Lond. (A)*, 29: 114-118.
- 3) Beck, S. D. 1968 Insect Photoperiodism. Academic Press, New York, 288 pp.
- 4) Danilevskii, A. S. 1961 Photoperiodism and seasonal development of insects. Japanese translation, 1966. Tokyo Univ. Press, Tokyo, 293 pp.
- 5) Enomoto, O. unpublished.
- 6) Frankos, V. H. and A. P. Platt 1976 Glycerol accumulation and water content in larvae of *Limenitis archippus*: their importance to winter survival. *J. Insect Physiol.*, 22: 623-628.
- 7) Hackman, W., S. Lakovaara, A. Saura, M. Sorsa and K. Vepsäläinen 1970 On the biology and karyology of *Chymomyza costata* Zetterstedt, with reference to the taxonomy and distribution of various species of *Chymomyza* (Dipt., Drosophilidae). *Ann. Ent. Fenn.* 36: 1-9.
- 8) Ichijô, N., M. T. Kimura and N. Minami 1980 Eco-physiological aspects of reproductive diapause in *Drosophila sordidula* and *D. lacertosa* (Diptera: Drosophilidae). *Jap. J. Ecol.*, 30: 221-228.
- 9) Kimura, M. T. and N. Minami 1980 Reproductive diapause in *Drosophila auraria*: Sensitivity of different developmental stages to photoperiod and temperature. *Kontyû*, 48: 458-463.
- 10) Lakovaara, S. 1969 Malt as a culture medium for *Drosophila* species. *Drosophila Inform. Serv.*, 44: 128.
- 11) Lumme, J. 1978 Phenology and photoperiodic diapause in northern populations of *Drosophila*. 145-170. In "Evolution of insect migration and diapause" (H. Dingle ed.), Springer-Verlag. New York.
- 12) ———, O. Muona and M. Orell 1978 Phenology of boreal drosophilids (Diptera). *Ann. Entmol. Fenn.*, 44: 73-85.
- 13) Mansingh, A. 1971 Physiological classification of dormancies in insects. *Can. Ent.*, 103: 983-1009.
- 14) Okada, T. 1976 Subdivision of the genus *Chymomyza* Czerny (Diptera, Drosophilidae), with description of three new species. *Kontyû*, 44: 496-511.
- 15) Riihimaa, A. 1981 Personal communication.
- 16) Saunders, D. S. 1976 Insect clocks. Pergamon Press, Oxford, 280 pp.
- 17) Takada, H. and M. J. Toda 1981 Notes on Arctic Canadian Diastatidae and Drosophilidae (Diptera), with the description of a new species. *J. Fac. Gen. Edu. Sapporo Univ.*, No. 18-A: 1-8.

- 18) Tauber, M. J. and C. A. Tauber 1976 Insect seasonality: diapause maintenance, termination, and postdiapause development. *Ann. Rev. Ent.*, 21: 81-107.
- 19) Watabe, H. 1979 *Drosophila* survey of Hokkaido, XXXVI. Seasonal changes in the reproductive condition of wild and domestic species of *Drosophila*. *J. Fac. Sci. Hokkaido Univ.* VI, Zool., 21: 365-372.
- 20) ——— and K. Beppu 1977 *Drosophila* survey of Hokkaido, XXXIII. Ovarian development of *Drosophila* in relation to wild population. *J. Fac. Sci. Hokkaido Univ.* VI, Zool., 20: 611-620.