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Title	Photosynthesis-dependent Ca <sup>2+</sup> influx and functional diversity between phospholipases in the formation of cell polarity in migrating cells of red algae
Author(s)	Mikami, Koji; Li, Lin; Takahashi, Megumu et al.
Citation	Plant Signaling & Behavior, 4(9), 911-913 <a href="https://doi.org/10.4161/psb.4.9.9534">https://doi.org/10.4161/psb.4.9.9534</a>
Issue Date	2009-09
Doc URL	<a href="https://hdl.handle.net/2115/44171">https://hdl.handle.net/2115/44171</a>
Type	journal article
File Information	Plant_Signal_Behav2009.pdf



**Photosynthesis-dependent Ca<sup>2+</sup> influx and functional diversity  
between phospholipases in the formation of cell polarity in red  
algal migrating cells**

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**Addendum to:**

Li L, Saga N, Mikami K. Ca<sup>2+</sup> influx and phosphoinositide signalling are essential for the establishment and maintenance of cell polarity in monospores from the red alga *Porphyra yezoensis*. J Exp Bot 2009; DOI:10.1093/jxb/erp183.

**Submitted:** 10 July 2009

**Accepted:**

**Key words:** Ca<sup>2+</sup> influx, cell polarity, phospholipase C, phospholipase D, photosynthesis,  
*Porphyra yezoensis*

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**Abbreviations:** PLC, phospholipase C; PLD, phospholipase D; PI3K, phosphatidylinositol 3-kinase; PtdIns(3,4,5)P<sub>3</sub>, phosphatidylinositol-3,4,5-trisphosphate; PtdIns(4,5)P<sub>2</sub>, phosphatidylinositol-4,5-bisphosphate; PA, phosphatidic acid

## **Abstract**

Unicellular spore cells, designated as monospores (also called archeospores), are well known as migrating plant cells, in which establishment of the anterior-posterior axis directs asymmetrical distribution of F-actin. Since the mechanisms of cell polarity formation are not yet fully elucidated in monospores, we investigated the roles of phosphoinositide signaling systems and Ca<sup>2+</sup> mobilization in migration. Although we have already found the critical involvement of phosphatidylinositol 3-kinase in the establishment of cell polarity, we recently demonstrated the important roles of extracellular Ca<sup>2+</sup> influx, phospholipase C (PLC) and phospholipase D (PLD). The remarkable characteristics of these factors are that Ca<sup>2+</sup> influx depends on photosynthetic activity and that PLC and PLD play roles in the establishment and maintenance of cell polarity, respectively. These findings could provide new insight into the regulation of migration in eukaryotic cells.

## **TEXT**

Monospores are responsible for asexual and clonal propagation of the marine multicellular red algae *Porphyra* and have an exceptional characteristic as migrating plant cells.<sup>1-5</sup> Monospores possess a round shape just after release from gametophytic blades (Figs. 1A and 1B), then undergo morphological change during migration. The establishment of cell polarity leads to the determination of anterior-posterior axis and

asymmetrical localization of F-actin (Fig. 1C). After migration, monospores adhere to the substratum in which the apical-basal axis has been established for further development (Fig. 1D). Asymmetrical distribution of F-actin is also found in chemotaxic migration of *Dictyostelium* cells and leukocytes.<sup>6,7</sup> In these cells, reciprocal local accumulation of phosphoinositides, such as phosphatidylinositol-3,4,5-trisphosphate (PtdIns(3,4,5)P<sub>3</sub>) at the leading edge and phosphatidylinositol-4,5-bisphosphate (PtdIns(4,5)P<sub>2</sub>) at the trailing side, is critical for the establishment of cell polarity. Phosphatidylinositol 3-kinase (PI3K) and PtdIns(3,4,5)P<sub>3</sub>-specific D-3-phosphatase PTEN have been identified as key modulators in the establishment of cell polarity, bringing asymmetrical distribution of these two phosphoinositides in plasma membranes.<sup>6,8</sup> Similarly, we found the involvement of PI3K activity in the establishment of cell polarity in migrating monospores,<sup>3</sup> suggesting the evolutionary conservation of the function of PI3K in migrating eukaryotic cells. On the other hand, the importance of cell wall synthesis has been found in the maintenance of the cell polarity during monospore migration<sup>4</sup> as reported in *Fucus* zygotes.<sup>9,10</sup> Therefore, the establishment and maintenance of cell polarity are thought to be regulated separately in monospores of *P. yezoensis*. In this addendum, further evidence of differential regulation of cell polarity formation by extracellular Ca<sup>2+</sup> influx and phospholipases in migrating monospores of red algae is documented according to our recent report.<sup>5</sup>

### **Photosynthesis-dependent extracellular Ca<sup>2+</sup> influx in cell polarity establishment**

Since migration and early development of monospores are completely inhibited in the dark, it is clear that monospore migration requires light illumination. There are two possibilities how light promotes monospore migration: first is sensing of the light direction by photoreceptors as found in Fucooid zygotes,<sup>11</sup> and the other is the energy

supply via photosynthesis. Here, since unilateral light did not influence the direction of migration, and DCMU, a photosynthesis inhibitor, completely inhibited migration, the establishment of cell polarity required for migration depends on photosynthetic activity in red algal monospores.

In animal cells, it is well known that  $\text{Ca}^{2+}$  influx triggers the establishment of cell polarity.<sup>12,13</sup> Moreover,  $\text{Ca}^{2+}$  influx is also critical for polarized tip growth of root hairs and pollen tubes in plants.<sup>14,15</sup> In *P. yezoensis*, monospore migration was completely inhibited by treatment with the  $\text{Ca}^{2+}$  chelator EGTA and  $\text{Ca}^{2+}$  channel blocker  $\text{LaCl}_3$ , whereas  $\text{Ca}^{2+}$  ionophore A23187 accelerated monospore migration. Thus, channel-mediated extracellular  $\text{Ca}^{2+}$  influx promotes the establishment of cell polarity following monospore migration. It is noteworthy that monospores treated with ionophore A23187 were able to migrate in the dark, indicating that artificial  $\text{Ca}^{2+}$  influx can mimic the photosynthesis-dependent monospore migration.

Taken together with the above findings, it was concluded that photosynthetic activity regulates extracellular  $\text{Ca}^{2+}$  influx to promote the establishment of cell polarity in monospores. Photosynthesis-mediated  $\text{Ca}^{2+}$  influx is a novel system for increasing cytosolic  $\text{Ca}^{2+}$  concentration in plants, which has been reported only in blue light-stimulated  $\text{Ca}^{2+}$  uptake via the  $\text{Ca}^{2+}$  channel in guard cells.<sup>16</sup>

### **Functional diversity between phospholipases C and D in monospore migration**

Recently, phosphatidic acid (PA) has been recognized as a second messenger involved in diverse cellular processes such as tip growth, stress response and acclimation, and auxin

signal transduction.<sup>17,18</sup> Phospholipase C (PLC) catalyzes the generation of two second messengers, namely, inositol-1,4,5-trisphosphate and diacylglycerol from PtdIns(4,5)P<sub>2</sub>, the latter of which is immediately converted to PA via phosphorylation by diacylglycerol kinase, although phospholipase D (PLD) directly produces PA from phosphatidylserine.<sup>17,18</sup> Thus, PLC and PLD play different roles in the production of PA.

There is evidence indicating the functional involvement of PLC and PLD in polarity establishment of mammalian cells and *Fucus* zygotes.<sup>19,20</sup> We also found the critical involvement of PLC in the establishment of cell polarity, based on complete inhibition of monospore migration and asymmetrical distribution of F-actin by treatment with a PLC inhibitor, U73122. In contrast, the inhibition of PLD by 1-butanol prevents migration but not asymmetrical distribution of F-actin, although further incubation resulted in the loss of the F-actin asymmetry. These findings demonstrate the functional diversity between PLC and PLD; that is, PLC is required for the establishment of cell polarity as PI3K, whereas PLD maintains cell polarity. Such diversity resembles the differential involvement of PI3K and cell wall synthesis in the establishment and maintenance of cell polarity in monospores.<sup>3,4</sup>

## **Perspectives**

The involvement of photosynthesis-dependent extracellular Ca<sup>2+</sup> influx and phospholipases in cell migration has been shown in the process of cell polarity formation in monospores from *P. yezoensis* (Fig. 1E). Since little is known how photosynthesis regulates Ca<sup>2+</sup> influx, further studies should focus on photosynthesis-dependent activation of the Ca<sup>2+</sup> channel and targets for increased Ca<sup>2+</sup> influx during monospore migration. For instance, the catalytic activity of PLC depends on Ca<sup>2+</sup> concentration, which in turn is mediated by

the Ca<sup>2+</sup>-binding module EF hand motif.<sup>21</sup> On the other hand, since PI3K activity also depends on Ca<sup>2+</sup>,<sup>12</sup> PLC and PI3K are thought to be candidates of the Ca<sup>2+</sup> target. Moreover, functional diversity of PLC and PLD in the formation of cell polarity was also presented. Such a differential involvement of these two phospholipases has also been observed in PtdIns(4,5)P<sub>2</sub>-dependent activation of the tobacco outward-rectifying K channel and the development of the brown alga *Silvetia compressa*.<sup>20,22</sup> Thus, the interrelationship between PLC and PLD in physiological regulations appears to be conserved in eukaryotic cells. In the future, elucidation of the relationship between photosynthesis-dependent Ca<sup>2+</sup> influx and functional diversity among phospholipases and PI3K must be addressed to understand how monospores migrate through a photosynthesis- and phosphoinositide signaling-dependent manner.

### **Acknowledgement**

This study was supported by Grants-in-Aid for Scientific Research (C; no. 21580213) from the Ministry of Education, Culture, Sports, Science, and Technology of Japan.

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## Figure Legend

### Figure 1

**Establishment and maintenance of cell polarity in monospores from the red alga *P. yezoensis*.**

(A) Discharge of unicellular monospores from a multicellular gametophytic blade of *P. yezoensis* strain TU-1. Scale bar = 20  $\mu\text{m}$ . (B-D) Asymmetrical distribution of F-actin during early development of monospores. F-actin was stained with Alexa Fluor 488 phalloidin. (B) Discharged monospore. (C) Migrating monospore. (D) Adhering monospore. Upper and lower photos in each panel show bright-field and fluorescent images, respectively. Arrow in (C) indicates the direction of migration. Scale bars = 5  $\mu\text{m}$ . (E) Schematic representation of our working hypothesis about the formation of cell polarity required for monospore migration. Photosynthesis-dependent  $[\text{Ca}^{2+}]_{\text{cyt}}$  increase regulates PLC and PI3K for the establishment of cell polarity, while PLD is required for the maintenance of the established cell polarity. DG, diacylglycerol; IP3, inositol-1,4,5-trisphosphate; IP3R, IP3 receptor; PC, phosphatidylcholine.

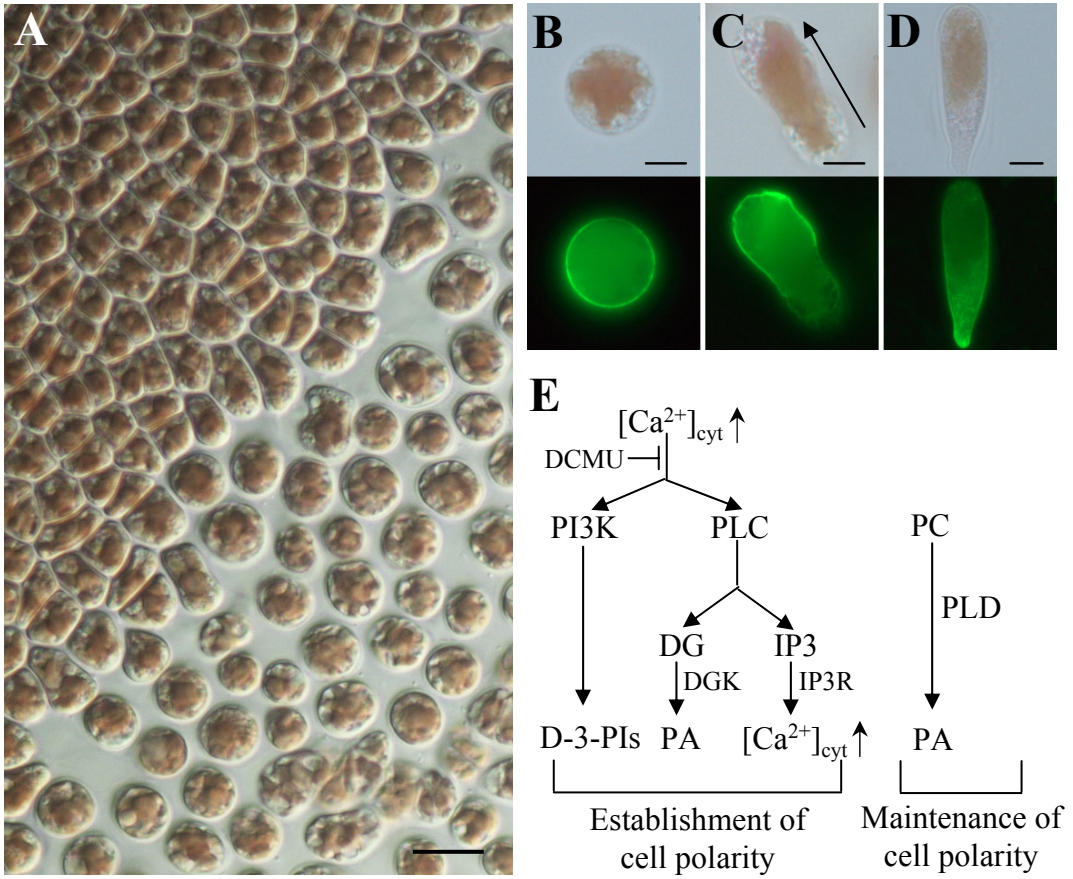


Figure 1