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Picosecond Time-Resolved Fluorescence Spectroscopy of Hematoporphyrin Derivative

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(Invited Paper)

Abstract—The picosecond time-resolved fluorescence decays $I(t)$ and spectra $I(\lambda, t)$ for hematoporphyrin derivative (HPD) in a phosphate buffer saline aqueous solution at different concentrations ($8.4 \times 10^{-6} \sim 8.4 \times 10^{-3}$ M) are measured by a two-dimensional synchroscan streak camera with a mode-locked CW dye laser, and a new emission band (which we call the Y-band) is found at high concentration. It is shown that the fluorescence decays composed of fast and slow components at high concentration are due to the Y-band (120 ps lifetime) from head-to-tail aggregates including equilibrium dimer and stable dimer, and the usual band (3.6 ns lifetime) from monomer, respectively, and the latter band is dynamically quenched by the Förster type resonance energy transfer from the monomer to the aggregate. Furthermore, the measurement of static fluorescence spectra from human gastric cancers and the surrounding *in vivo* after HPD injection shows that a band corresponding to the Y-band from the aggregate appears at only the cancerous cells.

INTRODUCTION

RECENTLY, it has been demonstrated that photoradiation therapy with the use of hematoporphyrin derivative (HPD) activated by laser light at a wavelength of 620-640 nm is an effective treatment of various types of cancers [1]-[3]. There has been an extensive investigation of the behavior of HPD from physical, photochemical, and biological aspects [4]. Many of the mechanisms of energy relaxations and photochemical reactions in the course of the treatment are not yet made clear. In order to reveal the mechanisms, there are the following four subjects to be investigated: 1) elucidation for mechanisms of decay kinetics and energy transfer from the excited state after absorbing the laser light, 2) identification of intermediate states and products in the processes of photochemical reactions and determination of their kinetic rates, 3) identification of target biomolecules which are finally chemically changed and subsequently do not enable cancerous cells to live, and 4) elucidation of the reason why HPD accumulates selectively in cancerous cells.

According to an analysis by Dougherty's group, HPD consists of a mixture of hematoporphyrin IX (HPIX), hydroxyethylvinyl-deuteroporphyrin IX (HVDPIX), protoporphyrin IX (PPIX), and their aggregates, including ether-bonded dimer of HVDPIX and HPIX [4], [5]. On the other hand, it is already known from static spectroscopic studies that HPIX, DPIX, and PPIX are easily dimerized in the equilibrium with monomer in a neutral aqueous solution [6]-[10]. Therefore, it is deduced that in the HPD solution two types of dimers exist, one is chemically bonded and stable, and the other is kinetically in the equilibrium with the monomer and the concentration dependent. For prevention of confusion we define aggregates in this paper as follows: the aggregates mean ether-bonded dimers (composite molecules, S dimers) and dissociable aggregates (D aggregates). The latter are weak coupling dimers (E dimers), timers, and oligomers such as van der Waals coupling aggregates, which are in the equilibrium with molecular units and the concentration dependent. More recently from clinical studies on the relation between HPD components and treatment efficacy, it has been pointed out that the aggregates are essential to the effective treatment of cancer by photoradiation therapy [4], [5], [11], [12]. This is thought to be due to the point that the selective accumulation of HPD involves the properties of aggregation of HPD components.

Recent pulse-laser techniques have made it possible to resolve excited state decay kinetics for a HPD solution. The fluorescence decay kinetics from the first excited singlet state has been investigated in the time region of nanoseconds by Milan's group [13] and in picoseconds by the present authors [14] and Rochester's group [15]. Consequently, it has been found that a fluorescence decaying curve of HPD in a phosphate buffer saline aqueous solution (PBS solution) shows two components of fast (less than a few hundred picoseconds) and slow (more than several nanoseconds) decays [14], [15]. The fluorescence decay increases when the HPD concentration decreases. It is generally thought that the fast and slow decay components are due to aggregates and monomers, respectively [15]. However, no direct evidence for the existence of a fast emission band from aggregates, which is clearly distinguished from a slow fluorescence band from monomers, has yet been obtained. It is generally known that the fluorescence from aggregates of organic molecules rapidly decays in the picosecond time-region [16], [17]. In this paper, we report the first investigation of the picosecond time-dependent fluorescence spectra $I(\lambda, t)$ from HPD in a PBS solution by using a two-dimensional synchroscan streak camera method. A new

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emission band (which we call the *Y*-band) due to the aggregates is found. In addition, mechanisms of the fluorescence relaxation for the HPD solution are quantitatively discussed, including the effect of resonance energy transfer from monomers to aggregates. The result of the direct measurement of the static fluorescence spectra for human gastric cancers after HPD injection *in vivo* is compared with the dynamic fluorescence spectra, and it is shown that a third emission band corresponding to the *Y*-band appears only at the cancerous cells after injection with HPD.

Experimental

The HPD solution of 5 mg/ml (8.4×10^{-3} M) concentration was offered from Dougherty's group [12] and diluted to desired concentrations with a PBS solution (0.02 M, pH = 7.2). The diluted HPD solution in a quartz cell was circulated to avoid any effects from photodegradation. The experimental apparatus for measuring picosecond time-dependent fluorescence spectra from the HPD solution is improved in comparison with the one we described in previous papers [14], [18]. A Jovin Yvon HR320 polychromator was attached between a sample optics and a Hamamatsu C1587 synchroscan streak camera. For data recording and processing, a Princeton Applied Research OMA II optical multichannel analyzer (two-dimensional) with a SIT visicon of high dynamic range (1.6×10^4) and sensitivity (20 photons/count) was used. The sensitivity of the synchroscan streak camera was also improved by ~50 times as high as the previous one by replacing a microchannel plate incorporated in a streak tube. A tunable synchronously mode-locked CW dye (R6G) laser which was constructed by the present authors generates continuous trains of pulses at 82 MHz with an average power of ~30 mW and a duration of less than 5 ps at 570 nm. A 570 nm laser light was employed as an excitation source to avoid photodegradation due to irradiation around 400 nm of the absorption peak. The overall time-resolution of the system was examined by measuring the duration of pulses from the dye laser. The recorded pulse duration at the present operational conditions was 19 ps at the accumulation of $\sim 10^9$ pulses. The reabsorption effect of the fluorescence at high concentration was carefully avoided by measuring the front fluorescence near the surface of the sample cell pumped at some angle in respect to its surface.

All absorption spectra for HPD in the PBS solution were carefully measured on a Hitachi Model 340 recording spectrophotometer over a wide range of concentration (2.5×10^{-8}-8.4×10^{-3} M). This was made possible by the use of cuvettes of different path lengths (0.1, 1.0, 10.0, and 100 mm) and by calibrations of background spectra of their cuvettes containing the PBS solution without HPD. Measurements of fluorescence spectra and fluorescence excitation spectra were made on a Hitachi Model 850 fluorescence spectrophotometer. The fluorescence spectra were taken at the excitation wavelengths of 390, 515, or 570 nm. The excitation spectra were measured at 615 or 680 nm.

Results and Discussion

A. Steady-State Absorption and Fluorescence Excitation Spectra

Absorption spectra of HPD in the PBS solution measured over the concentration range from 2.5×10^{-8}-8.4×10^{-3} M are shown in Fig. 1. It is seen from Fig. 1 that with the increase of the concentration the molar extinction coefficient decreases, the spectral shape of the Soret band changes from two peaks at 372 and 390 nm to a single peak at 363 nm, and a red-shift of four peaks in the *Q*-band occurs. This behavior is very similar to the change from the absorption spectra of the monomer to those of the *E* dimer of HPIX, DPIX, and PPIX in an aqueous solution [7]-[10], except that the peak at the shorter wavelength of the Soret band still remains at the most diluted concentration. This suggests that in the HPD solution at high concentration the *D* aggregate including mainly the *E* dimer exists, while at the diluted concentration, in addition to the monomer, another molecular species independent of the concentration exists. In order to confirm the latter part of this suggestion, we measured how the relative intensity of the fluorescence peaks at 615 or 680 nm depends on the wavelength of the excitation light. A typical excitation spectrum independent of the concentration is shown in Fig. 2(a).

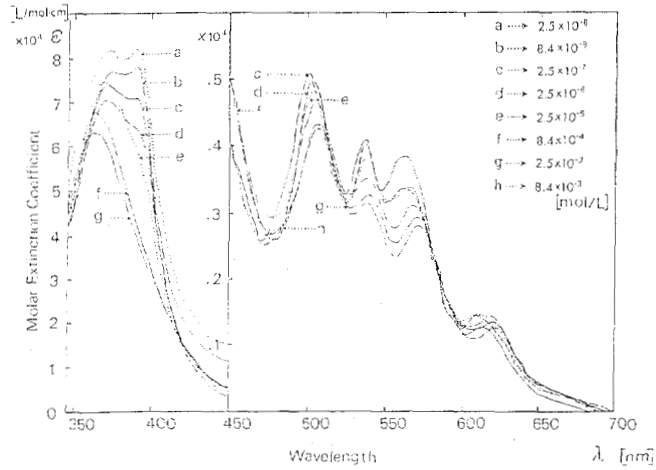


Fig. 1. Absorption spectra of different concentrations of HPD in the PBS solution: (a) 2.5×10^{-8}, (b) 8.4×10^{-8}, (c) 2.5×10^{-7}, (d) 2.5×10^{-6}, (e) 2.5×10^{-5}, (f) 8.4×10^{-4}, (g) 2.5×10^{-3}, and (h) 8.4×10^{-3} M.

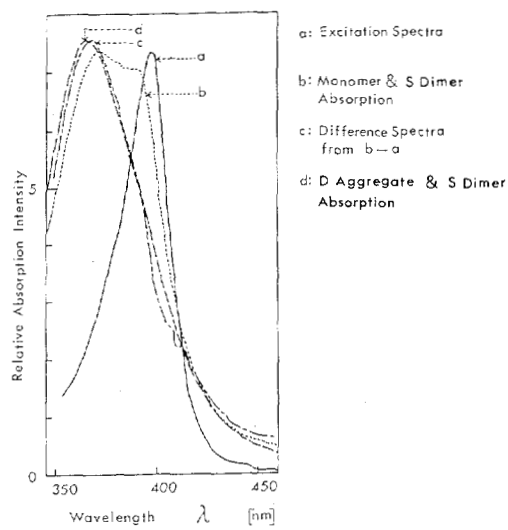


Fig. 2. (a) (——) and (b) (- - - - -) are fluorescence excitation (corresponding to monomer absorption) spectrum and absorption spectrum at the diluted concentration (corresponding to absorption spectrum of monomer and *S* dimer), respectively. (c) (- — - — -) is the difference spectrum subtracted from (b) by (a), corresponding to the absorption spectra of *S* dimer. (d) (— — —) is the absorption spectrum at high concentration corresponding to absorption spectra of *D* aggregate and *S* dimer.

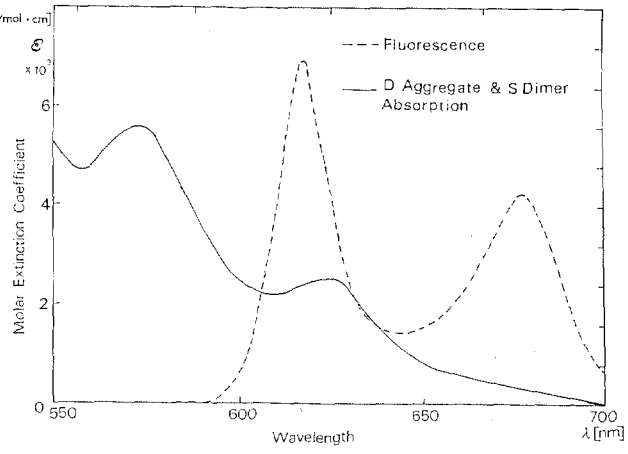


Fig. 3. Overlap of fluorescence spectra (· · · · ·) and absorption spectra of *D* aggregate and *S* dimer (——).

Favorable comparison of this spectrum with the absorption spectrum of the monomer of HPIX and DPIX already known [7]-[9] indicates that both the spectra are almost the same. This means that the monomer of HPD emits the fluorescence having two peaks of 615 and 680 nm like the monomer of HPIX, DPIX, and PPIX. Fig 2(c) is the difference spectrum, which was subtracted from the absorption spectrum at the diluted concentration [Fig. 2(b)] by the excitation spectrum so as not to become negative. This difference spectrum coincides with the absorption spectrum at high concentration [Fig. 2(d)], as shown in Fig. 2(c) and (d). This fact gives the evidence that the HPD solution contains not only the *D* aggregate, but also another stable species in addition to the monomer which is independent of the concentration. It is deduced that this corresponds to the recent analytical result that the HPD solution contains ether-bonded dimer (*S* dimer) of HVDPIX and HPIX [4]-[5]. Therefore, it seems that the absorption spectrum of the *S* dimer is similar to that of the *D* aggregate containing mainly the *E* dimer.

Furthermore, the spectral overlap between the fluorescence and the absorption was examined as shown in Fig. 3. The fluorescence spectrum from the monomer overlaps the absorption spectrum at the high concentration due to the *D* aggregate and *S* dimer over the range between 590 and 700 nm. Therefore, the dynamic fluorescence quenching due to the resonance energy transfer can occur in the HPD solution such as a rhodamine *B* monomer-dimer system in an aqueous solution [19]-[20]. This dynamic phenomenon will be discussed later.

B. Picosecond Time-Resolved Fluorescence Measurement

The measured picosecond decays $I(t)$ of the total fluorescence spectra at different concentrations (8.4×10^{-3}, 8.4×10^{-4}, 8.4×10^{-5}, and 8.4×10^{-6} M) are shown in Fig. 4. From Fig. 4 it is seen that the decays have fast and slow components at high concentration, which agrees with the experimental result of [15], and the amplitude of the fast component decreases with the decrease of the concentration. In order to clarify the mechanism of the fluorescence relaxation, the picosecond time-dependent fluorescence spectra $I(\lambda, t)$ were also measured at the same concentrations. In Fig. 5, the typical time-dependent fluorescence spectra with the corresponding total fluorescence decays at the highest and lowest (1/100) concentrations are shown. It is seen from Fig. 5 that a new

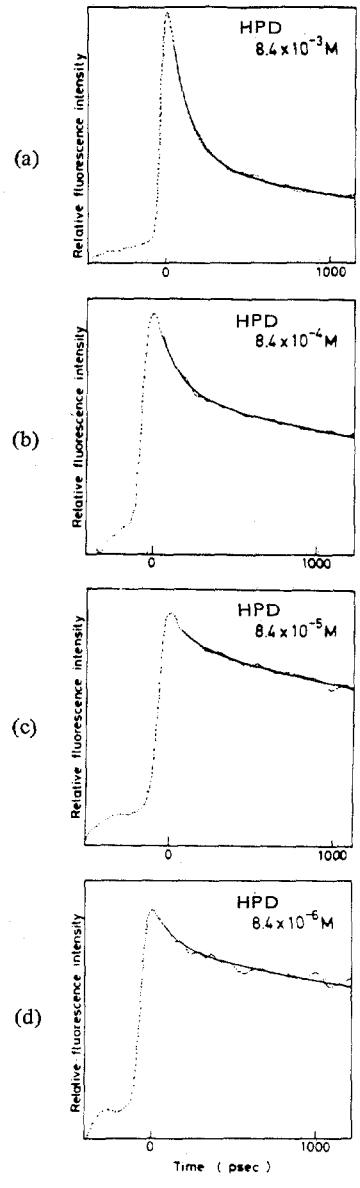


Fig. 4. Picosecond decay $I(t)$ of the total fluorescence for different concentrations of HPD in the PBS solution: (a) 8.4×10^{-3}, (b) 8.4×10^{-4}, (c) 8.4×10^{-5}, and (d) 8.4×10^{-6} M. The solid lines are numerical fitting curves to the data, as discussed in the text.

emission band (the *Y*-band) around 640 nm between the usual two bands around 615 and 680 nm appears in the fast time region for the solution at the highest concentration. The *Y*-band decays faster than a few hundred picoseconds while the usual bands decay slower than several nanoseconds. With decreasing the concentration the *Y*-band is rapidly buried in the usual band. From the comparison between the time-dependent spectra and the total fluorescence decays, it is found that the time and concentration variations for the fast component in the decays definitely correspond to the behavior of the *Y*-band. Therefore, it can be concluded that the fast component appearing in the total fluorescence decay is assigned to the *Y*-band emitted from the at least *D* aggregate. This was also confirmed by measuring the wavelength-resolved fluorescence decays $I(\Delta\lambda, t)$ for the peak of each band, as shown in Fig. 6.

The same picosecond measurement was also done for the HPD II solution (photofrin II, Oncology Research and Develop-

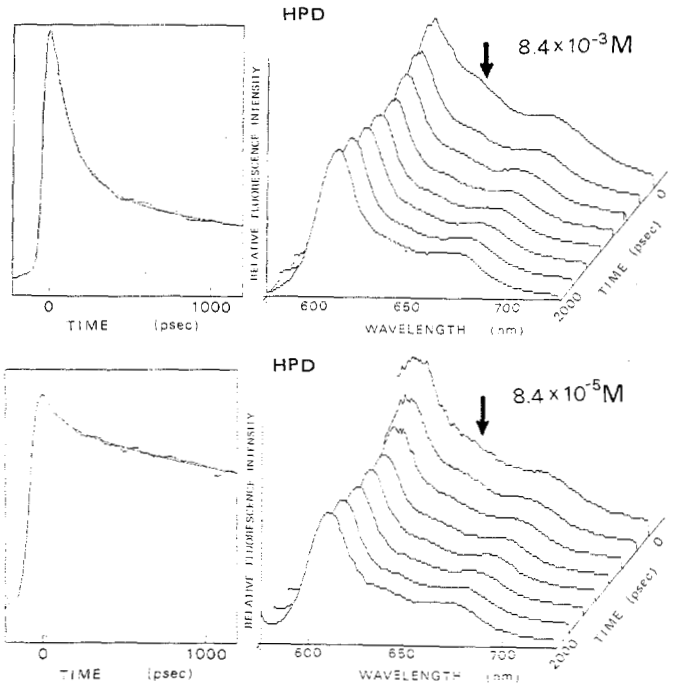


Fig. 5. Picosecond time-dependent fluorescence spectra $I(\lambda, t)$ and corresponding total fluorescence decays $I(t)$ from the HPD solutions at high (8.4×10^{-3} M) and low (8.4×10^{-5} M) concentrations.

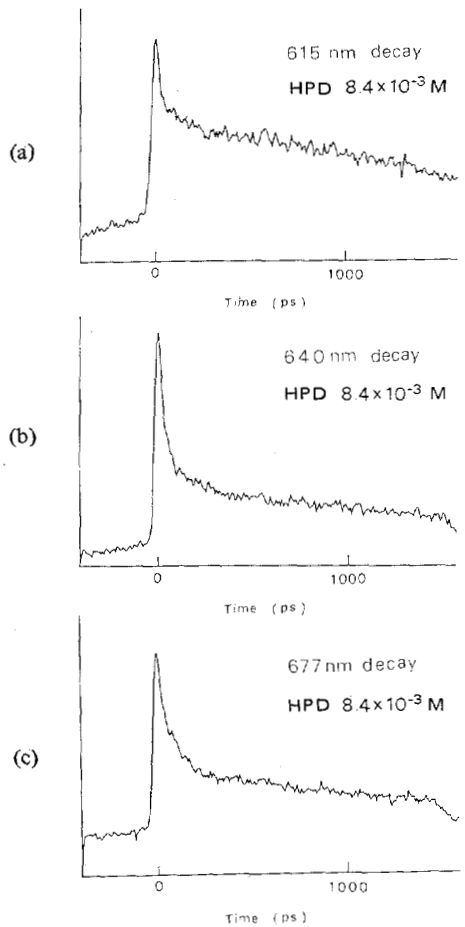


Fig. 6. Wavelength-resolved fluorescence decays $I(\Delta\lambda, t)$ at (a) 615, (b) 640, and (c) 677 nm for the HPD solution at high concentration (8.4×10^{-3} M).

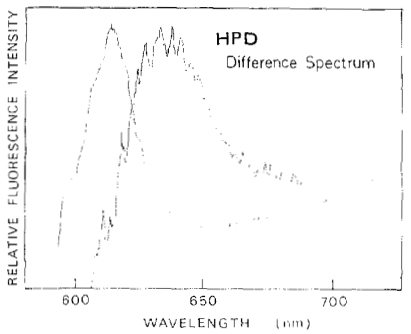


Fig. 7. Difference spectra subtracted from the fluorescence spectra right after pumping (within 300 ps) by the fluorescence spectra long after pumping (6.0 ns passing) for the HPD solution at the high concentrations (8.4×10^{-3} M).

ment) [12] in order to examine the effect of the S dimer. The HPD II solution is the effective aggregate fraction separated by gel filtration from the HPD solution and contains much more of the S dimer. It is known from recent clinical studies [3], [4], [11], [12] that the HPD II solution is superior to the HPD solution concerned with the properties of the selective accumulation and the side-effect. The experimental result showed that the above mentioned behaviors of the Y-band are more remarkable for the HPD II solution than for the HPD solution. Therefore, it can be said that the Y-band is due to not only the D aggregate containing mainly the E dimer, but also the S dimer.

Fig. 7 is the difference spectrum which was subtracted from the fluorescence spectrum right after the excitation pulse (t = 0 ~ 300 ps) by that long after the excitation pulse (t = 6.0 ~ 6.3 ns) so as not to become negative. From Fig. 7 it is found that the Y-band shifts toward the red wavelength in respect to the usual band. It seems that this fact corresponds to the red-shift of the Q-band in the absorption spectra at high concentration. According to Kasha's theoretical analysis on the energy relaxation from the excited states of the molecular aggregates [16], the face-to-face aggregate (sandwich type) shows the blue-shift relative to the monomer excited state level while the head-to-tail aggregate shows the red-shift. This suggests that both the D aggregate and the S dimer emitting the Y-band are the head-to-tail aggregates. It should be noted that the formation of the S dimer necessitates the preformed non-covalent dimer properly aligned [12].

On the basis of the above consideration, we try to describe quantitatively the total fluorescence decay $I(t)$. The decay is the sum of the fast decay of the Y-band from the D aggregate and S dimer, and the slow decay of the usual band from the monomer. The latter can be dynamically quenched through the Förster-type resonance energy transfer [20] from the monomer as donor to the D aggregate and S dimer as acceptor since the fluorescence spectrum from the monomer partially overlaps the absorption spectrum at the highest concentration due to the D aggregate and S dimer. Therefore, the total decay $I(t)$ is expressed by the following equation:

$$I(t) = A_1 \exp\left[-t/\tau_1\right] + A_2 \exp\left[-\sqrt{\pi}\gamma\sqrt{t/\tau_2} - t/\tau_2\right] \qquad (1)$$

TABLE I
FLUORESCENCE RELAXATION PARAMETERS OBTAINED FROM NUMERICAL FITTINGS FOR HPD SOLUTIONS: AMPLITUDES OF FAST A_1 AND SLOW A_2 DECAYING COMPONENTS, RATIO γ OF TOTAL DIMER-CONCENTRATION TO CRITICAL TRANSFER CONCENTRATION, CONCENTRATIONS OF MONOMER C_m AND TOTAL DIMER C_d, AND APPARENT CRITICAL DISTANCE R_o OF ENERGY TRANSFER

Total Concentration C [M]	A1	A2	Monomer Concentration C_m [M]	Total Dimer Concentration C_d [M]	$\gamma = C_d/C_o$	Apparent Critical Distance Cubed R_o^3 [Å^3]	Apparent Critical Distance R_o [Å]
8.4×10^{-3}	0.48	0.52	2.0×10^{-4}	4.1×10^{-3}	0.38	3.7×10^{4}	33
8.4×10^{-4}	0.26	0.74	5.2×10^{-5}	3.9×10^{-4}	0.12	1.2×10^{5}	50
8.4×10^{-5}	0.07	0.93	2.0×10^{-5}	3.2×10^{-5}	0.03	4.2×10^{5}	75
8.4×10^{-6}	0.13	0.87	1.1×10^{-6}	3.6×10^{-6}	0.00	—	—

where the first and second terms describe the fast fluorescence decay of the Y-band, and the slow and dynamically quenched fluorescence decay of the usual band, respectively. As a result of numerical fitting of (1) to the measured total fluorescence decays at different concentrations in Fig. 4, τ_1, τ_2, A_1, A_2, and γ are determined and given in Table I. The solid lines in Fig. 4 represent the numerical fitting curves. The fluorescence lifetimes of the Y-band from the D aggregate and S dimer, and of the usual band from the monomer are 120 ps and 3.6 ns, respectively. As the HPD concentration decreases, A_1 proportional to the sum concentration of the D aggregate and the S dimer decreases, while A_2 proportional to the concentration of the monomer increases.

For simplification of the discussion, we assume as follows: 1) all the D aggregates are only the E dimer, 2) the molar extinction coefficients ϵ_1 at the wavelength (570 nm) of the excitation pulse, the fluorescence quantum yields ϕ_1, and the fluorescence lifetimes τ_1 of the E dimer and those of the S dimer are the same. The assumptions allow us to express the ratio of A_2/A_1 as

$$\frac{A_2}{A_1} = \frac{C_m \epsilon_2 \phi_2 / \tau_2}{C_d \epsilon_1 \phi_1 / \tau_1} \tag{2}$$

where C_m, ϵ_2 and ϕ_2 are the concentration, the molar extinction coefficient at 570 nm, and the fluorescence quantum yield of the monomer respectively, and C_d is the total dimer concentration of the E and S dimers. By using the obtained values of τ_1, τ_2, A_1, and A_2, and the ratio values of $\epsilon_2/\epsilon_1 = 0.65$ (from the absorption spectra in the previous section), and $\phi_2/\phi_1 = 1000$ (the assumed value) in (2), the monomer concentration and the total dimer concentration for different HPD concentrations are calculated as given in Table I. According to a theory of the Förster-type energy transfer between donor and acceptor molecules [20]-[21], the γ in (1) is equal to the ratio C_d/C_o of the acceptor concentration to the critical transfer concentration. For the HPD solution, the acceptor concentration is equal to the total dimer concentration. The critical transfer concentration is expressed by $(7.35/R_o)^3$ with the critical transfer distance R_o units of Å [21]. The R_o is used to indicate the distance at which energy transfer has the same probability as the sum of all other excited state deactivation processes of the monomer. The distance is given by

$$R_o^6 = \frac{(9000 \ln 10)\, k^2 \phi_2}{128 \pi^5 n^4 N} \int_0^\infty \frac{f_2(\nu)\, \epsilon_1(\nu)}{\nu^4}\, d\nu \tag{3}$$

where k^2 is an orientation factor, ($k^2 = \frac{2}{3}$ for HPD with a random direction distribution in the aqueous solution [22]), ϕ_2 is the fluorescence quantum yield of the monomer in the absence of the E and S dimers, n is the refractive index of the solvent, N is Avogadro number, $f_2(\nu)$ and $\epsilon_1(\nu)$ are the spectral distribution of the monomer fluorescence and the molar extinction coefficient of the E and S dimers respectively, and ν is the wavenumber [20]-[21].

The combination of the toal dimer concentration C_d with the γ enables us to determine the apparent critical distance R_o, and the result is also given in Table I. The dependence of the apparent critical distance on the total dimer concentration is shown in Fig. 8. The distance is rapidly decreasing with the increase of the total dimer concentration, like the experimental results of the resonance transfer from rhodamine B monomer to dimer [19] and from DODCI to malachite green and to DQOCI [23]. The true critical distance R_{ot} is determined by extrapolating the total dimer concentration toward the higher

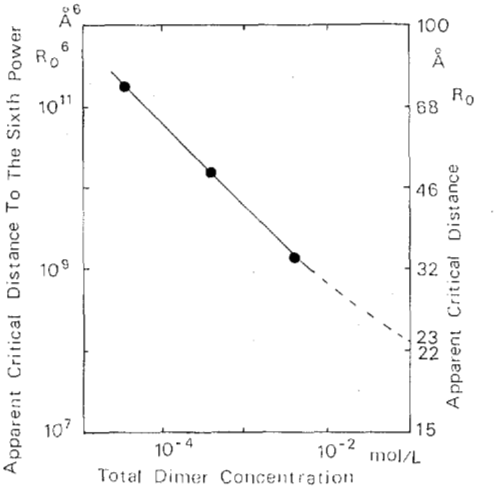


Fig. 8. Total dimer-concentration dependence of the apparent critical distance R_o of energy transfer for the HPD solution.

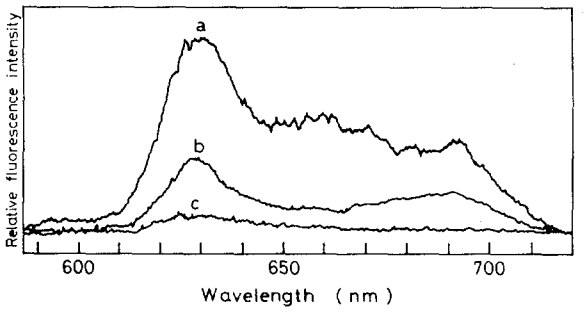


Fig. 9. Static fluorescence spectra from (a) human gastric cancers, (b) the erosion of the cancers, and (c) the surrounding normal cells at the early stage *in vivo* 48 h after HPD injection.

concentration and is estimated to be 23 Å. This value is almost equal to the value of 24 Å which is evaluated by using (3) on the basis of data of the spectral overlap in Fig. 3.

Finally, we describe the correlation between the picosecond time-resolved fluorescence spectra from the HPD solution and the static fluorescence spectra from HPD in cancerous cells *in vivo*. The static fluorescence spectra from the human gastric cancers, the erosion of the cancers and the surrounding normal cells at the early stage (72 years old male) after injection of the HPD solution (5 mg/kg weight) were simultaneously measured by using a fiber endoscope with a Jobin Yvon HR320 polychromator and a Princeton Applied Research OMA I under 405 nm Kr-ion laser pumping. The results measured in 48 h after injection showed that only the static fluorescence spectra from the gastric cancers have a third emission band which corresponds to the *Y*-band as shown in Fig. 9. It was also observed that the third emission band gradually appears at only the gastric cancers as time passes after injection. Therefore, it seems that the head-to-tail aggregates emitting the *Y*-band play a role important for the selective accumulation of HPD in cancerous cells, and the fluorescence lifetime of the aggregates is remarkably increased by binding to some biomolecule *in vivo*. As for this point, further investigation is needed.

Conclusion

From the detailed measurement of absorption spectra and excitation spectra for the HPD solution, it has been found that the HPD solution at high concentration contains the *D* aggregate including mainly the *E* dimer and the *S* dimer, and both the absorption spectra are similar. The measurement of the picosecond time-resolved fluorescence spectra by means of the two-dimensional synchroscan streak camera has shown that a new fast emission band due to the head-to-tail *D* aggregate and *S* dimer exists. The mechanism of the fluorescence relaxation in the HPD solution has been made clear. That is, the fluorescence decays are due to the fast emission (τ_1 = 120 ps) from the head-to-tail *D* aggregate and *S* dimer and the slow emission (τ_2 = 3.6 ns) from the monomer. The latter emission is quenched by Förster-type resonance energy transfer from the monomer to the *D* aggregate and *S* dimer. Furthermore, from the observation of an emission band corresponding to the fast emission band only at the gastric cancers *in vivo*, it has been suggested that the head-to-tail *D* aggregate and *S* dimer has a important role for the selective accumulation properties of the HPD solution.

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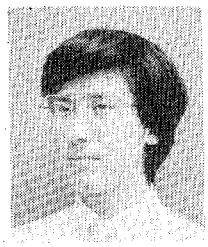
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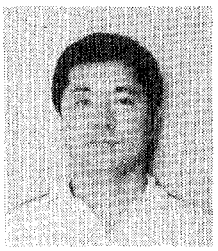
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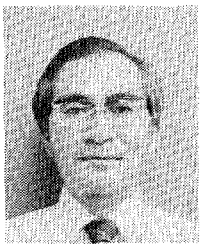
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