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Evidence for instantaneous e-vector detection in the honeybee using an associative learning
paradigm

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List of abbreviations

CS+: e-vector orientation that will be or has been paired with sucrose reward

CS-: e-vector orientation that will be or has been specifically unpaired with sucrose reward

DRA: dorsal rim area of the compound eye

exDRA: area of the compound eye except for the DRA

PER: proboscis extension reflex

SUMMARY

Many insects use the polarization pattern of the sky for obtaining compass information during orientation or navigation. E-vector information is collected by a specialized area in the dorsal-most part of the compound eye, the dorsal rim area (DRA). We tested honeybees' capability of learning certain e-vector orientations by using a classical conditioning paradigm with the proboscis extension reflex. When one e-vector orientation (CS+) was associated with sugar water (US), while another orientation (CS-) was not rewarded, the honeybees could discriminate CS+ from CS-. Bees, whose DRA was inactivated by painting, did not learn CS+. When ultraviolet polarized light (350 nm) was used for CS, the bees discriminated CS+ from CS-, but no discrimination was observed in blue (442 nm) or green light (546 nm). Our data indicate that honeybees can learn and discriminate between different e-vector orientations, sensed by the UV receptors of the DRA, suggesting that bees can determine their flight direction from polarized UV skylight during foraging. Fixing the bees' heads during the experiments did not prevent learning indicating that they use an "instantaneous" algorithm of e-vector detection, i.e. the bees do not need to actively scan the sky with their DRAs ("sequential" method) to determine e-vector orientation.

INTRODUCTION

Many animals show striking capabilities in spatial orientation and navigation over short-range and long-range distances. For example, sea turtles, birds and butterflies travel thousands of kilometers for seasonal habitats or a nest [1-3]. Insects such as honeybees [4] and desert ants [5] are capable of navigating, up to several kilometers, in straight lines back to their nest after complicated feeding journeys. To solve navigational tasks, animals are forced to choose the appropriate traveling direction [6]. Spatial orientation should be encoded as internal representations of the spatial relationship between the animal's body and its surrounding space. In rats and other mammals, place cells and head direction cells are well-studied examples of neurons encoding spatial properties in the brain [7-9]. However, the brain mechanisms of mammals for spatial orientation and calculating distances are not fully understood. The smaller brain of an insect allows investigation of basic brain mechanisms for spatial navigation at a single neuron level [10].

It is well known that insects use a skylight compass to obtain directional information during navigation by path integration, i.e. they continually monitor their net distance and direction from the starting point [11-15]. A major source of skylight compass information is the polarization pattern of blue sky. As a result of sunlight scattering, light from blue sky is partially polarized with the celestial e-vectors arranged along concentric circles around the sun [16]. Behavioral studies have shown that desert ants use celestial e-vector patterns during travel for finding the correct direction to the nest [12,15]. The finding that honeybees failed to transfer the direction to the food source by a waggle dance under an unpolarized light condition [11,17] suggests the usage of e-vectors of polarized light for deducing the direction.

The e-vector detection in insects is mediated by a group of specialized ommatidia located in the most dorsal part of the compound eye, the dorsal rim area (DRA). The sensory function of DRA ommatidia as polarization sensors have been studied broadly across species

anatomically, electrophysiologically and behaviorally (for reviews, see [13,18]). In honeybees, the DRA consists of 4-5 rows of ommatidia at the dorsal margin of the compound eye [19,20]. Electron microscopic studies have shown that in each DRA ommatidium there are two sets of the photoreceptors with the microvilli oriented 90° to each other, i.e. they are tuned to mutually orthogonal e-vector orientations [19,21]. These two receptor types have been suggested to connect antagonistically to second-order neurons, effectively enhancing e-vector contrast [22]. Electrophysiological and behavioral studies indicate that UV-sensitive photoreceptors in the DRA ommatidia with high polarization sensitivities and wide visual fields are the primary information carriers for polarization vision in honeybees [23,24]. Wavelength dependence for e-vector-guided waggle dances was previously found to be strictly UV-sensitive [25].

In contrast to the peripheral sensory system, neural mechanisms of polarization vision in the brain have been studied in only few species, mainly in crickets and locusts. In crickets, e-vector information from the DRA is processed by a group of polarization-opponent interneurons in the optic lobe (POL1-neurons; [26-28]). There are only three types of POL1-neurons, which are tuned to different e-vectors oriented 10° , 60° and 130° to the body axis, suggesting that e-vector orientation is coded as signals of three differently tuned information channels by the so-called *instantaneous method* of e-vector detection [13]. In the higher center of the brain, the central complex has been suggested to be the location of the internal compass [29]. Electrophysiological studies have revealed a number of different types of polarization-sensitive neurons in the central complex [29-32]. In the locust, zenithal e-vector orientations are topographically represented in columnar organization in the protocerebral bridge of the central complex [29]. In the cricket, Sakura *et al.* [32] proposed that polarization information is represented by e-vector orientation-selective neurons (so-called “compass neuron”) in the central body, as a result of integration of combinatory activations by the three types of POL1-neurons. The compass neurons have been discussed as analogues of head-direction cells in mammals

[13,32,33].

Since no electrophysiological data are available for honeybees beyond the retinal level, it is unknown if the neural basis of e-vector detection in honeybees is similar to the one of crickets and locusts. In principle, every organism has two ways of measuring celestial e-vector orientation, the *instantaneous method* described above, and the so-called *sequential method* [13]. With the instantaneous method e-vector orientation can be recognized at a glance, since each e-vector is unambiguously defined by a certain neural activity pattern in the brain. In contrast, with the sequential method an animal has to make rotatory movements around its vertical body axis thereby scanning the sky with their DRA. So far, two previous studies with honeybees came to opposite conclusions. Based on the idea that the insects perceive polarization not as a separate modality of light but, as a modulation of light intensity when the bees made rotational movements, Rossel & Wehner [11,22, 34] were able to elicit directed waggle-dances when the light intensity of an unpolarized dorsal stimulus was modulated as a function of body orientation. Edrich & von Helversen [35], on the other hand, were unable to disturb the oriented dances of bees when the degree of polarization of a dorsal polarized stimulus was temporally modulated at different rates. This suggested instantaneous processing because the animal should have been severely confused if it had made sequential comparisons. These hypotheses for the polarized light perception have been still in argument because of poor amount of direct evidence.

To study the cognitive functions of polarization vision, monitoring of behavioral outputs and also the establishment of a reliably reproducible experimental paradigm are required. In honeybees, the Pavlovian classical conditioning with the proboscis extension reflex (PER) has been widely used as a tool for determining cognitive capabilities. A honeybee will extend its proboscis when sugar water (unconditioned stimulus, US) is applied to its antenna, proboscis, or leg [36]. When sugar solution is applied to the bee shortly after presentation of a conditioned sensory stimulus (CS), which is not originally related to the PER, the bee will show PER when

CS alone is presented without sugar water. By using this associative learning paradigm, the capabilities of bees for sensory discrimination and cognition have been investigated [37-39].

In the present study, we established a new behavioral paradigm for studying polarization vision in honeybees using PER, allowing us to examine two controversial mechanisms for polarization recognition because the head of the bee can be easily fixed in a place under this condition. We demonstrate (1) that honeybees can learn e-vector orientations by using the classical conditioning paradigm with PER, and (2) that bees discriminate certain e-vector orientations under these conditions. We show (3) that the DRA is crucial for the discrimination of e-vector orientations, and (4) that polarization vision is restricted to the UV range of light. Most importantly, our data indicate (5) that honeybees can use an instantaneous mechanism to code e-vector orientation in the brain.

1. MATERIALS AND METHODS

(a) Experimental animals

The honeybees, *Apis mellifera ligustica*, used in this study were reared in a normal ten-frame hive at the campus of Tokushima Bunri University. All experimental bees were collected from the same colony. Forager honeybees with pollen loads were collected at the hive entrance in the evening (mostly between 4 and 6 p.m.), at least one day before the experiment. The bees were anesthetized on ice and mounted in a plastic tube as described previously [36,40]. Contrary to the previous conditioning experiments using visual stimulus [37,38], the antennae of the bees were kept intact during the whole experiments. The bees were fed 4 or 5 drops of 30% sucrose solution and kept at room temperature until the next day in a dark environment. In the morning, each bee was fed with one drop of the sucrose solution. Although it was generally difficult for the bees to move their heads under these conditions, the heads were further immobilized at the neck with

wax in the experiments for which results are shown in Fig. 4.

(b) Experimental setup

The experiments were performed using a custom-made experimental box (electronic supplementary material, Fig.S1a) in a dark room. The box consisted of two parts divided by a horizontal partition with a circular opening containing a polarizer: the upper part was for generating unpolarized light while the lower part was for manipulating an experimental animal. Inside of the box was covered with black non-glossy paper to avoid reflection of the light. Only the frontal plane of the box was open for manipulating. Light from a xenon lamp (LC8, Hamamatsu Photonics, Hamamatsu, Japan) was applied from the top of the upper part by a light guide. In addition to white light, we used UV, blue and green monochromatic light to examine the wavelength dependency of polarization vision. For these monochromatic stimuli, interference filters with λ_{max} of 350 nm, 441.6 nm or 546 nm and half-widths of 10 nm (VPF, Sigma-Koki, Tokyo, Japan) was mounted at the end of the light guide. Within the box, the light passed through a hollow tube the wall of which was diffusing paper for removing any specific polarized light components. After passing through the diffusing paper, unpolarized light passed through the opening in the center of the partition between the upper and the lower part of the box. For the polarized light stimulus, a slider with a linear polarizer (HN42HE, Polaroid Company, Cambridge, MA) was inserted manually into a slit below the diffusing paper. The e-vector orientation of the stimulus could be changed by changing the inserting direction of the slider. Two bees were placed on the black platform in the center of the lower part so that the DRA would face the light stimulus. The intensity of the polarized white light at the animal level was 8700 lx (measured by LM-331, AS ONE, Osaka, Japan), and the powers of the polarized UV, blue, and green lights at the animal level were 5.0, 17.1, and 13.4 $\mu\text{W}/\text{cm}^2$, respectively (measured by PM100, ThorLabs, Newton, NJ for blue and green light and YK-34UV, Lutron Electronic Enterprise, Taipei, Taiwan for the UV light). The size of the stimulus was 62° as seen

by the bees.

(c) Visual stimulation and conditioning

The procedures of the conditioning paradigm were adapted from the color-conditioning paradigm [37]. Differential conditioning was applied with two polarized light stimuli with different e-vector orientations (electronic supplementary material, Fig. S1b). The angular difference between the e-vector orientations presented during conditioning was 90° . Here, we refer to the e-vector parallel to the body axis of the bee as 0° and that at a right angle to the body axis as 90° , for convenience. At least 5 minutes before conditioning began, the bees were placed in the center of the lower part below the unpolarized light stimulus to become familiar with the environment. Unpolarized light illumination was kept on throughout the experiments and the polarizer was inserted manually only during polarized light stimulation. One e-vector orientation was selected as the conditioned stimulus (CS+) and was applied for 4 s followed by manual application of 30% sucrose solution (unconditioned stimulus, US) to the right antenna or to the proboscis for 3 s together with the CS+. US application was performed by the standard procedure [36,40]. The other e-vector orientation was presented for 7 s without sucrose as CS-. CS+ and CS- were balanced among animals, i.e. one of the bees in the box was conditioned with 0° of the e-vector as CS+, while the other was with 90° at the same time. The interval between CS+ and CS- was 2 min. This differential conditioning trial was repeated 15 times (inter-trial interval = 4 min). In the test phase, the animal was exposed to polarized light once without sucrose 4 min after the last conditioning trial. Since bees could freely extend their proboscis and since the CS+ light during the conditioning phase preceded the US by 4 s, the response to the CS+ could be monitored visually and expressed as an acquisition function. For the experiments under UV light, a small spotlight of dim red light was focused on the bee's head to observe the bee's behavior.

(d) Selective painting of the compound eye

The dorsal rim area (DRA) of the compound eye is visually identifiable because its cornea appears slightly grey and cloudy [19]. Either both DRAs or the whole compound eyes except for the DRAs (exDRA) were painted with black acrylic emulsion paint (Herbol, Cologne, Germany) under a dissecting microscope at least 1 hour before experiments. Because it was technically not possible to cover the DRA alone, which consists of only 4-5 horizontal rows of ommatidia (see [19,24]), a small area of the unspecialized dorsal part next to the DRA was covered together with the DRA. *Vice versa*, a small area of the dorsal eye part near the DRA was not covered when the DRA was left open because it was impossible to leave the DRA alone open. We estimated that a 3-fold area of the DRA was covered in DRA-painted animals and a less than 2-fold area of the DRA was not covered in exDRA animals. After the experiments, the paint cover was checked in all animals under the dissecting microscope. Data for cases in which even a small area of paint was missing, were excluded from further analysis. The three ocelli, which are not involved in polarization vision [41], were not painted in the experiments.

(e) Analysis and statistics

Animals that extended their proboscis during the 4-s polarized light stimulation period prior to US application were counted as showing PER. Data for animals that failed to respond to US were excluded from analysis.

Comparisons of the learning effects across the experimental parameters were performed using GLMs calculated by R statistical software (version 2.11.0, R Development Core Team, <http://www.R-project.org>). Based on the data of number of the bees that exhibited PER in each trial, we constructed four models for binomial distributions with different parameter sets, trial number, CS, both trial number and CS, and null, i.e., neither trial number nor CS. Akaike Information Criterion (AIC) was calculated for each model, and the best model was selected as

the model providing the smallest AIC value among the four models.

To determine the number of the trials to establish the e-vector orientation learning, we performed the change point analysis using GLMs. Sixteen trials were divided into two groups, the former and later groups. For the set 1, the former group included the trial 1 and the later group included trial 2-16. We applied the null model to the former group and the CS model to the later group. AIC for this set was obtained by summing AICs of both former and later group. Next, for the set 2, trial 1-2 was in the former group and trial 3-16 was in the later group, and then calculating AIC for the set 2. We repeated this procedure until the set 15, trial 1-15 for the former group and trial 16 for the later group. Finally, the number of trial for learning establishment was determined by the set which provided the smallest AIC value among 15 sets.

2. RESULTS

(a) General features of polarized light learning

First, we examined the capability of a honeybee to discriminate between different e-vector orientations. The differential conditioning paradigm with two mutually orthogonal e-vector orientations revealed that honeybees could discriminate those e-vectors (Fig. 1). Model selection using AIC values of GLMs indicated that the best model was the model including both the trial and CS as the parameters (Table 1). In the first acquisition trial, only one (2.4%) and four (9.5%) of 42 bees exhibited PER to CS+ and CS-, respectively. After 15 acquisition trials, 14 (33.3%) and 2 (4.8%) of the 42 bees exhibited PER to CS+ and CS-, respectively. The change point analysis indicated that seven trials was needed to discriminate CS+ and CS- (Fig. 1). At about the seventh trial, the number of bees showing PER to CS+ became large and then slightly decreased until the 10th trial. The number of bees showing PER to CS+ increased again at about 12th trial. In the test trial (16th trial from the beginning), the number of animals showing PER was a

slightly smaller than that in the 12th trial. In most cases, the proportion of animals showing PER did not exceed 50%. On the other hand, the number of animals showing PER to CS- remained small (less than 10%) throughout the experiment. When 15 acquisition trial was divided into two parts (early part: 1-8, late part: 9-15) and the number of PERs to CS+ in each part was counted, there were only three bees among 42 bees in which the number of PER in the early part was higher than that in the late part. This indicates that the most bees learned CS+ although the learning score was only about 35%. The learning properties described above were generally observed in all experiments in this study.

We further analyzed the behavioral responses to the two e-vector orientations (0° and 90°) when each e-vector was used as CS+ separately using GLMs. No bias was found for either of the two e-vectors as CS+ (GLM including CS as a parameter was not selected as the best model, meaning that 0° and 90° for CS+ was not different). Any clear interactive effects were also not observed in terms of PER responses between two simultaneously conditioned bees (only 6 cases were potentially interactive cases, data not shown). Therefore, we pooled the data for these two e-vectors according to their functions.

From the comparison of bee learning performance, we found that the effect of conditioning depended on the season although the bees were from the same colony. The maximum ratios of animals responding to CS+ were 50.0% and 27.8% in spring and autumn, respectively. On the other hand, the maximum ratios of animals responding to CS- were 8.3% and 5.6% in spring and autumn, respectively (with PER probability in the first trial excluded). These results suggest that spring bees learned e-vector orientation better than autumn bees, but we cannot exclude other possibilities such as aging, experience etc, except for the seasonal effect. Although we do not know any causal factors, we used only spring bees for further experiments because of this difference in the learning score.

(b) Selective stimulation of eye regions

To confirm the sensory input area for e-vector learning in the eye, we covered part of each compound eye restricting the area receiving light stimulation to the DRA or exDRA (electronic supplementary material, Fig. S2).

We found that the exDRA-painted animals exhibited the PER to CS⁺ but not to CS⁻, whereas the DRA-painted animals could not learn CS⁺ (Fig. 2, Table 1). The number of PER animals increased gradually with increase in acquisition trials. In the 5th trial, CS⁺/CS⁻ discrimination was established (Fig. 2b). In the test trial, 17 (65.4%) of 26 exDRA-painted animals showed PER for CS⁺. The best model, “trial and CS”, indicates that bees learned CS⁺ along the acquisition trials (Table 1). In general, the probability of DRA-painted animals showing PER to CS⁻ was slightly higher than that of exDRA-painted animals throughout the experiment (mean values of 14.6% and 6.7% for DRA-painted and exDRA-painted animals, respectively).

(c) Wavelength-dependence of e-vector discrimination

The compound eye of the honeybee contains three types of photoreceptors with different spectral sensitivities, peaking in the ultraviolet (UV), blue, and green, respectively [42]. Here, we addressed to determine the wavelength selectivity for e-vector discrimination learning. The bees exhibited e-vector discrimination by conditioning with UV polarized light only (Table 1, Fig. 3a). The maximum ratios of animals that responded were 44.8% and 17.2% for CS⁺ and CS⁻, respectively. In the case of blue or green polarized light, the bees did not learn CS⁺ because the best model was “null” (Table 1, Fig. 3b,c). The probabilities for PER during CS⁺ or CS⁻ were both in the same low range. These results indicate that a honeybee perceives polarized light information from UV photoreceptors.

(d) Instantaneous or sequential system for e-vector detection?

Next, we fixed the bees' heads with wax, preventing any scanning movements necessary for a

sequential approach, and trained them with UV polarized light. If the bees depended on spatial scanning, they would not be able to discriminate CS+ from CS- under this condition. On the other hand, if the bees could discriminate the two e-vectors, this suggested that e-vector discrimination “at a glance” was possible by the instantaneous mechanism. Fixation of the head revealed that the honeybee’s ability for significant discrimination between CS+ and CS- because the “CS” model was selected (Table 1, Fig. 4) same as the result for animals without fixation. The PER probability reached a plateau with a maximum value at 43.3%. As in the case of DRA-painted animals, head-fixed animals showed slightly higher PER responses to CS- in general than that for the animals shown in Fig. 1 (10.0% and 8.3% of the bees showed PER for CS- in the test phase in head-fixed and normal animals, respectively.).

4. DISCUSSION

(a) General properties of e-vector learning

We have established a new appetitive-learning paradigm in which an e-vector orientation was associated with sugar solution and demonstrated that honeybees can discriminate between two different e-vectors. In this study, we used 0° and 90° e-vector orientations for CS+ and could not find any differences in learning performances between these two e-vectors. If we use e-vector orientations for CS that coded by POL-neurons in the medulla (e.g. 10°, 60° and 130° in the crickets), bees might show better discrimination ability. Nevertheless we first have to find orientations coded by polarization-sensitive neurons because neither selective orientations nor such brain neurons have been studied in the honeybee yet. Learning of e-vector orientation was slower than olfactory learning, i.e. the bees required seven acquisition trials for polarized light learning, whereas only one sole trial was sufficient for olfactory learning [36,43]. Furthermore, PER probability after 15 trials was much lower than in olfactory learning (about 80% in the third

trial) [36]. These differences are probably based on the different sensory modality for learning. PER probability of harnessed honeybees was 40% after 20 trials in color learning [37]. This is comparable with our results, although color learning required more acquisition trials, i.e. 14 trials for color learning and seven trials for polarized light learning. Rather than color learning in honeybees, the properties of polarized light learning might be similar to those of motion learning. In motion learning [38], harnessed honeybees were presented with black-white gratings moving from front to back or *vice versa*, mimicking optical flow during foraging, and the bees were conditioned to associate the movement direction. In these experiments, eight acquisition trials were required to reach a plateau of learning performance at approximately 40%, i.e. the same as in the present study. Since information on optical flow and e-vector orientation are both closely related to foraging behavior, i.e., navigation and orientation [15,44], and since the learning performance for these two sensory cues are similar, these information may have similar biological meaning for bees.

The comparatively low performance of polarized light learning was not improved by either a longer (20 min) inter-trial interval (ITI) or a larger number of trials (data not shown). In olfactory learning, spaced training (ITI > 3 min) induces higher learning performance than massed training (ITI = 30 s), and the memory established by spaced training consolidated better than that by massed training [45]. It would be interesting to examine how long polarized light memory is maintained under natural conditions. This memory is probably short-term memory, like working memory in mammals, because it is unlikely that a foraging bee needs to retain e-vector memory for path integration during flight. Our preliminary experiments in which bees were tested three times (4, 8 and 12 min after the last conditioning trial) by CS+ without any rewards suggested that the memory extinction curve for e-vector orientation was comparable to that of olfactory learning with the short ITI (data not shown, [46]). The 4-min ITI in our experiments might be long considering their normal flight speed, 210-350 m/min [47]. Assuming

that polarized light learning is related to navigation, we might have obtained a better performance score if we had selected a massed training paradigm (ITI < 4 min) because each type of learning paradigm has a specifically optimal ITI in general [45,48].

We found a seasonal difference in the polarized light learning score. Honeybees are well known to have an annual life cycle in their colony [49]. Both olfactory and tactile learning performances by using PER were affected by the foraging season [50]. Our finding that spring bees showed better performance in polarized light learning than autumn bees together with the seasonality previously found in three different sensory modalities, i.e. olfactory, tactile and visual, suggests that the seasonal difference in learning ability is a general feature for honeybees. We still need additional experiments to reveal seasonal difference in learning capability for polarized light.

(b) Polarized light information processing mechanism in the sensory system

Bees learned e-vector orientation only when UV light was used for a CS, but no significant learning performance was observed for blue or green stimulus even though the intensity of UV light was about a third of other wavelength intensities (Fig. 3). This finding supports a previous study in which the direction of waggle dances could be controlled by e-vector orientation provided that the stimulus contained UV [25]. These data indicate that honeybees rely on polarized UV light of skylight to detect e-vector information. In addition, our covering experiments demonstrate that the DRA is the eye region responsible for e-vector learning (Fig. 2). This result is in agreement with a previous study showing that blocking visual inputs to the DRA abolished polarized-light guided orientation in the honeybee [24]. Their data perfectly agree with the high polarization sensitivity in the UV receptors of the DRA found in an electrophysiological study [23]. Thus, all available data strongly indicate that the UV receptors of the DRA are the input elements for polarized light perception in honeybees.

(c) Instantaneous system of polarization vision

In the *sequential method*, successive readings of an insect's POL-neurons are compared while the insect scans a dorsal polarized stimulus (e.g. the sky) by rotating about its vertical axis. Whenever one of the POL-neurons exhibits maximal activity, this indicates that the e-vector of the stimulus is aligned with the e-vector tuning axis of the POL-neuron. With (for instance) three tuning types of POL-neuron, just three body orientations can be determined directly by the polarization vision system. Based on these reference orientations, all other orientations must be determined either by proprioception or by evaluating the optic displacement of the retinal image of the world perceived on making a turn. In the *instantaneous method*, the brain compares the simultaneously available signals of differently tuned POL-neurons. Theory shows that the signals of three independent polarization-sensitive channels suffice to encode e-vector orientation unambiguously [51-53]. In other words, each body orientation is defined by a unique combination of activity levels in the three POL-neurons. Using the instantaneous approach, an insect can perceive e-vector orientation (or body orientation relative to it) at a glance without performing any movements.

Which of these alternative algorithms is implemented in honeybees? This question has been addressed by behavioral experiments. Rossel & Wehner [11,22] support the idea of the sequential mechanism by directing waggle-dance experiments with an unpolarized dorsal stimulus. Edrich & von Helversen [35], on the other hand, have shown that bees kept the oriented dances even under temporarily modulating the degree of polarization of a dorsal polarized stimulus, suggesting the instantaneous mechanism. In the present study, fixing the bees' heads to prevent scanning movements during the conditioning experiments did not prevent the insects to discriminate between two sequentially presented orthogonal e-vector orientations (Fig. 4). This finding can only be explained by an instantaneous algorithm of e-vector detection. Tethered flying locusts and walking crickets responded spontaneously to the e-vector of a dorsally presented polarized stimulus even with their heads fixed suggesting instantaneous e-vector

detection in orthopteran insects ([54]; T. Labhart, unpublished observations).

Recent electrophysiological studies in the central complex of orthopteran insect brains have provided data strongly consistent with instantaneous processing. The central complex has been suggested to be involved in spatial orientation in *Drosophila* [55], and many types of polarization-sensitive neurons have been found electrophysiologically in the central complex of crickets and locusts [29-32]. A group of neurons in the central complex of crickets, which have been suggested to code e-vector orientation [32] supporting the idea of an internal compass. In the locust, the protocerebral bridge of the central complex consists of 16 columns with each column containing polarization-sensitive neurons that respond to specific e-vector orientation [29]. Thus, information on e-vector orientation seems to be represented in a topographical manner in the protocerebral bridge of the central complex. These findings imply that actual e-vector orientation is finely encoded in the orthopteran brain. On the other hand, because the polarized light stimuli were applied by a slowly rotating polarizer in all these studies, there is still a possibility that the animals enable to analyze the e-vector orientation “sequentially” even under the fixation of the heads.

Our present conclusion that honeybees use the instantaneous mechanism for polarization vision does not exclude the possibility that the sequential mechanism is also used, as proposed by the analysis of dance behavior under unpolarized light [11,22]. Another learning paradigm using PER in which bees can scan polarized stimuli will allow us to examine whether or not they use sequential mechanisms. Possibly, these two essentially different mechanisms are not exclusive but function complementary depending on the circumstantial conditions the animal is facing, e.g., light intensity, degree of polarization, and visible size of the blue sky. If so, it would be interesting to reveal situations in which one mechanism dominates. It remains unknown what angular view of the sky and what intensity of polarized light are needed for honeybees to discriminate two e-vectors. Elucidation of these factors will facilitate an understanding of the manner of selection of those two mechanisms. Since directional discrimination capability is

necessary for all navigating animals [6], similar neural mechanisms probably underlie navigation in both invertebrates and vertebrates.

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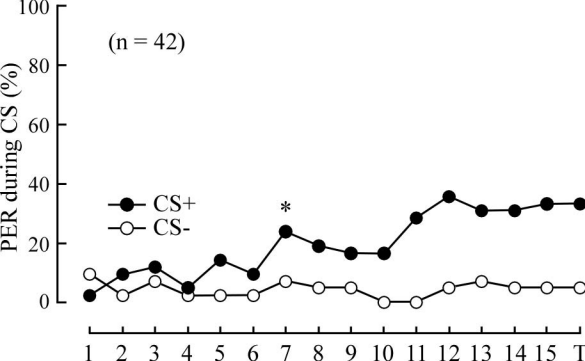
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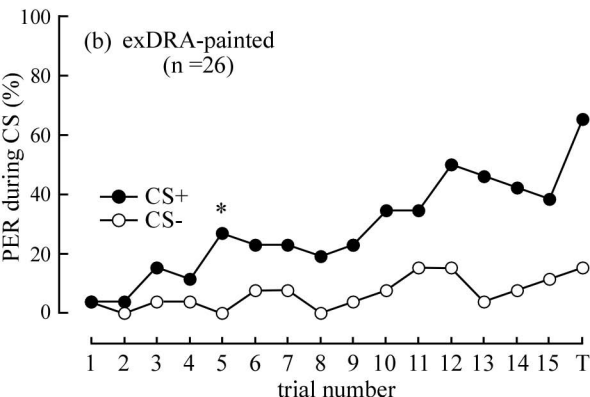
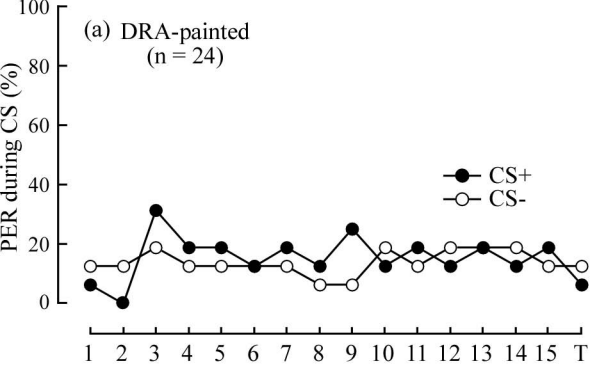
Fig. 1. Associative learning of e-vector orientation. Proboscis extension response probabilities in each trial are shown. Bees exhibited learning performance after the 7th conditioning trial. The asterisk indicates the point when the effect of conditioning was found by the change point analysis. 1-15: Number of each conditioning trial. T: Test trial.

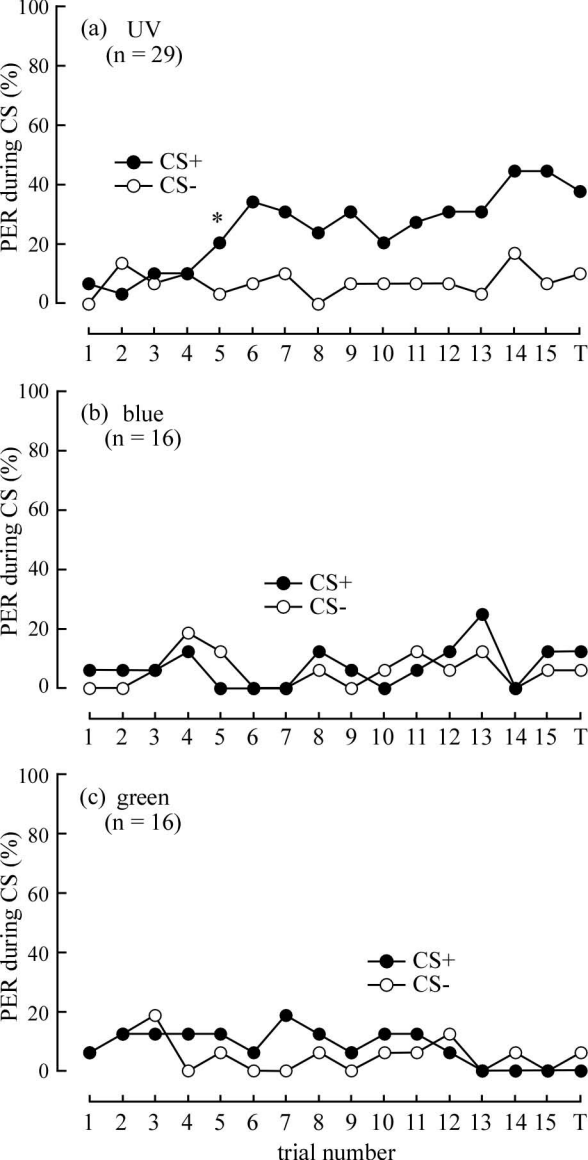
Fig.2. Learning curves of DRA-painted (a) and exDRA-painted (b) bees. **(a)** The DRA-painted bees did not learn CS+ throughout the trials. **(b)** The exDRA-painted bees exhibited leaning performance after the 5th conditioning trial. The asterisk indicates the point when the effect of conditioning was found by the change point analysis. 1-15: Number of each conditioning trial. T: Test trial.

Fig. 3. Learning curves for UV (a), blue (b), and green (c) polarized light. **(a)** The UV-trained bees exhibited leaning performance after the 5th conditioning trial. The asterisk indicates the point when the effect of conditioning was found by the change point analysis. **(b,c)** The blue- and green-trained bees did not learn CS+ throughout the trials. 1-15: Number of each conditioning trial. T: Test trial.

Fig. 4. Learning curves of head-fixed bees for UV polarized light. The head-fixed bees exhibited learning performance after the 2nd conditioning trial. The asterisk indicates the point when the effect of conditioning was found by the change point analysis. 1-15: Number of each conditioning trial. T: Test trial.







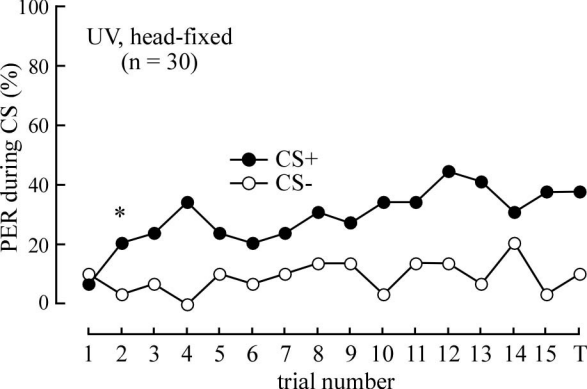


Table 1. AICs of GLMs with different parameter sets in each experimental group

parameters	normal (Fig. 1)	DRA (Fig. 2a)	exDRA (Fig. 2b)	UV (Fig. 3a)	blue (Fig. 3b)	green (Fig. 3c)	neck-fixed (Fig. 4)
null	243.66	116.21 *	240.84	210.21	86.84 *	84.88 *	201.92
only trial	216.90	138.13	207.63	205.33	90.44	96.37	214.72
only CS	161.80	118.17	168.83	152.86	88.33	85.36	139.86 *
Trial + CS	133.22 *	140.08	129.69 *	145.75 *	91.14	96.81	151.51

(*: selected model with minimum AIC)