



# HOKKAIDO UNIVERSITY

Title	Identification and molecular variations of CAN-SINEs from the ZFY gene final intron of the Eurasian badgers (genus Meles)
Author(s)	Tashima, Sara; Kaneko, Yayoi; Anezaki, Tomoko et al.
Citation	Mammal Study, 36(1), 41-48 <a href="https://doi.org/10.3106/041.036.0105">https://doi.org/10.3106/041.036.0105</a>
Issue Date	2011-03
Doc URL	<a href="https://hdl.handle.net/2115/48490">https://hdl.handle.net/2115/48490</a>
Type	journal article
File Information	MS36-1_41-48.pdf



## Identification and molecular variations of CAN-SINEs from the *ZFY* gene final intron of the Eurasian badgers (genus *Meles*)

Sara Tashima<sup>1</sup>, Yayoi Kaneko<sup>2</sup>, Tomoko Anezaki<sup>3</sup>, Minoru Baba<sup>4</sup>, Shuuji Yachimori<sup>5</sup>, Alexei V. Abramov<sup>6</sup>, Alexander P. Saveljev<sup>7</sup> and Ryuichi Masuda<sup>1,\*</sup>

<sup>1</sup> Department of Natural History Sciences, Graduate School of Science, Hokkaido University, Sapporo 060-0810, Japan

<sup>2</sup> Tokyo University of Agriculture and Technology, Fuchu 183-8538, Japan

<sup>3</sup> Gunma Natural History Museum, Tomioka 370-2345, Japan

<sup>4</sup> Kitakyushu Museum of Natural History and Human History, Kitakyushu 805-0071, Japan

<sup>5</sup> Shikoku Institute of Natural History, Susaki 785-0023, Japan

<sup>6</sup> Zoological Institute, Russian Academy of Sciences, Saint-Petersburg 199034, Russia

<sup>7</sup> Russian Research Institute of Game Management and Fur Farming, Russian Academy of Agricultural Sciences, Kirov 610000, Russia

**Abstract.** The short interspersed nucleotide elements (SINEs) are specific to the taxa and thought to be one of powerful phylogenetic gene markers. Especially, the SINE sequences, which exist uniquely in genome of order Carnivora, are named CAN-SINEs. Among the Eurasian badgers (genus *Meles*), a member of the family Mustelidae in order Carnivora, the Japanese badger (*M. anakuma*) was previously reported to have an insertion of CAN-SINE in the final intron of the zinc finger protein gene on Y chromosome (*ZFY*). In the present study, we examined occurrence of the CAN-SINE of the *ZFY* final intron in the Eurasian badgers, and three continental and four Japanese haplotypes were identified from a total of 40 male badgers. Among the Eurasian badger CAN-SINEs, a 12-bp deletion specific to the Japanese haplotypes was found, whereas the 12-bp region (non-deletion) in the continental haplotypes consisted of one 6-bp direct repeat and 6-bp microsatellite-like sequences. Moreover, the continental haplotypes were phylogenetically divided into three lineages: eastern Eurasia, Caucasus and western Eurasia. These genetic differentiations supported the classification recently proposing that genus *Meles* are grouped into the European badger (*M. meles*), the Southwest Asian badger (*M. canescens*), the Northwest & Central Asian badger (*M. leucurus*) and the Japanese badger (*M. anakuma*). In addition, the number of adenines in the poly A/T rich tails was polymorphic among all lineages of Eurasian badgers, and geographically variable within the Japanese badgers.

**Key words:** CAN-SINE, Eurasian badger, *Meles*, molecular variation, *ZFY* gene.

The short interspersed nuclear elements (SINEs), which are the 80–400 base-pair (bp) long nonretroviral retrotransposons, disperse in nuclear genome via RNA with the help of endonuclease or the complex of reverse transcriptase, and they do not encode the reverse transcriptase in their own sequences (Alberts et al. 2001). They acquire the retropositional activity from the corresponding long interspersed nuclear elements (LINEs), which encode reverse transcriptase and which are thought to be the origins of the SINEs (Okada et al. 1997). More than 10<sup>4</sup> copies of SINEs exist in eukaryotic genome (Shedlock and Okada 2000). The majority of SINEs are

reported to have derived from transfer RNAs (tRNAs) with the exceptions of primate Alu and rodent B1, which derived from 7SL RNA (Ullu and Tschudi 1984; Daniels and Deininger 1985; Labuda et al. 1991), and consist of three regions: a tRNA-related region, a tRNA-unrelated region and an A/T-rich region (Kramerov and Vassetzky 2005). The tRNA-related region contains an internal promoter of the RNA polymerase III, and the tRNA-unrelated region includes the LINE derived region, and the A/T-rich region relates to generate microsatellite sequences in Mustelidae (López-Giráldez et al. 2006).

The SINE is thought to be one of powerful phylo-

\*To whom correspondence should be addressed. E-mail: masudary@ees.hokudai.ac.jp

genetic markers because of its wide dispersion in genome and no elimination after insertion to specific sites (Shedlock and Okada 2000). In addition, they are specific to the taxa (Van der Vlugt and Lenstra 1995), as reported in the primate Alu (Weiner et al. 1986; Batzer et al. 1990), cetacean CHR1 and CHR2 (Shimamura et al. 1997) and carnivore-specific SINE (CAN-SINE) (Minnick et al. 1992). Alberts et al. (2001) showed a possibility that the activities of transposable elements are related to speciation. The phylogenetic relationships by SINE markers were reconstructed in many mammalian taxa such as cetaceans (Shimamura et al. 1997), vespertilionids (Kawai et al. 2002) and felids (Pecon-Slattery et al. 2004).

The CAN-SINEs exist in representatives of all carnivore families (Vassetzky and Kramerov 2002). Pecon-Slattery et al. (2000) reported the SINE insertions within the nonrecombining region on the Y chromosome in the six carnivore families: Felidae, Canidae, Mustelidae, Ursidae, Procyonidae and Phocidae. For Mustelidae, Yamada and Masuda (2010) reported the independent insertion of CAN-SINE into the final intron of the zinc finger protein on Y chromosome (*ZFY*) in the ermine (*Mustela ermine*) and the Japanese badger (*Meles anakuma*).

The Eurasian badgers were once thought to comprise three groups of subspecies in the single species *Meles meles*: subspecies “*meles*” in Europe westward to the Volga River, subspecies group “*arenarius-leptorhynchus*” in the east of the Volga River to Siberia, and another subspecies group “*amurensis-anakuma*” in Far East and Japan (Heptner et al. 1967). Recently, however, based on morphological features such as the skull, color pattern, baculum structure (Lynch 1994; Abramov 2001, 2002; Abramov and Puzachenko 2006), they are classified into three independent species: the European badger (*Meles meles*), the Asian badger (*M. leucurus*) and the Japanese badger (*M. anakuma*) (Lynch 1994; Abramov 2001; Wozencraft 2005). Based on the mitochondrial DNA (mtDNA) cytochrome *b* phylogenetic analysis, Kurose et al. (2001) reported the clear genetic distinction between the Japanese badgers and the Eurasian continental badgers. In addition, Marmi et al. (2006) showed the four mtDNA lineages in the Eurasian badgers: Europe, Southwest Asia, North & East Asia, and Japan. Abramov and Puzachenko (2005, 2006) already mentioned a remote position of badgers of Middle East from the others within the European badger *Meles meles*. Del Cerro et al. (2010) examined mtDNA and nuclear

DNA sequences and proposed a new distinct species *M. canescens* for the Southwest Asian badger, in addition to the previous three species. Tashima et al. (2011) also supported the four lineages using new data of the mtDNA control region and the sex-determining region on Y chromosome (*SRY*), which is paternally inherited.

In the present study, we identified CAN-SINEs from the *ZFY* final intron of not only the Japanese badgers but also continental Eurasian badgers. Based on a comparison of the haplotype sequences, we presented the molecular structural variations and phylogenetic features of the paternally inherited CAN-SINEs, and then discuss molecular evolution of CAN-SINEs related to speciation of the Eurasian badgers.

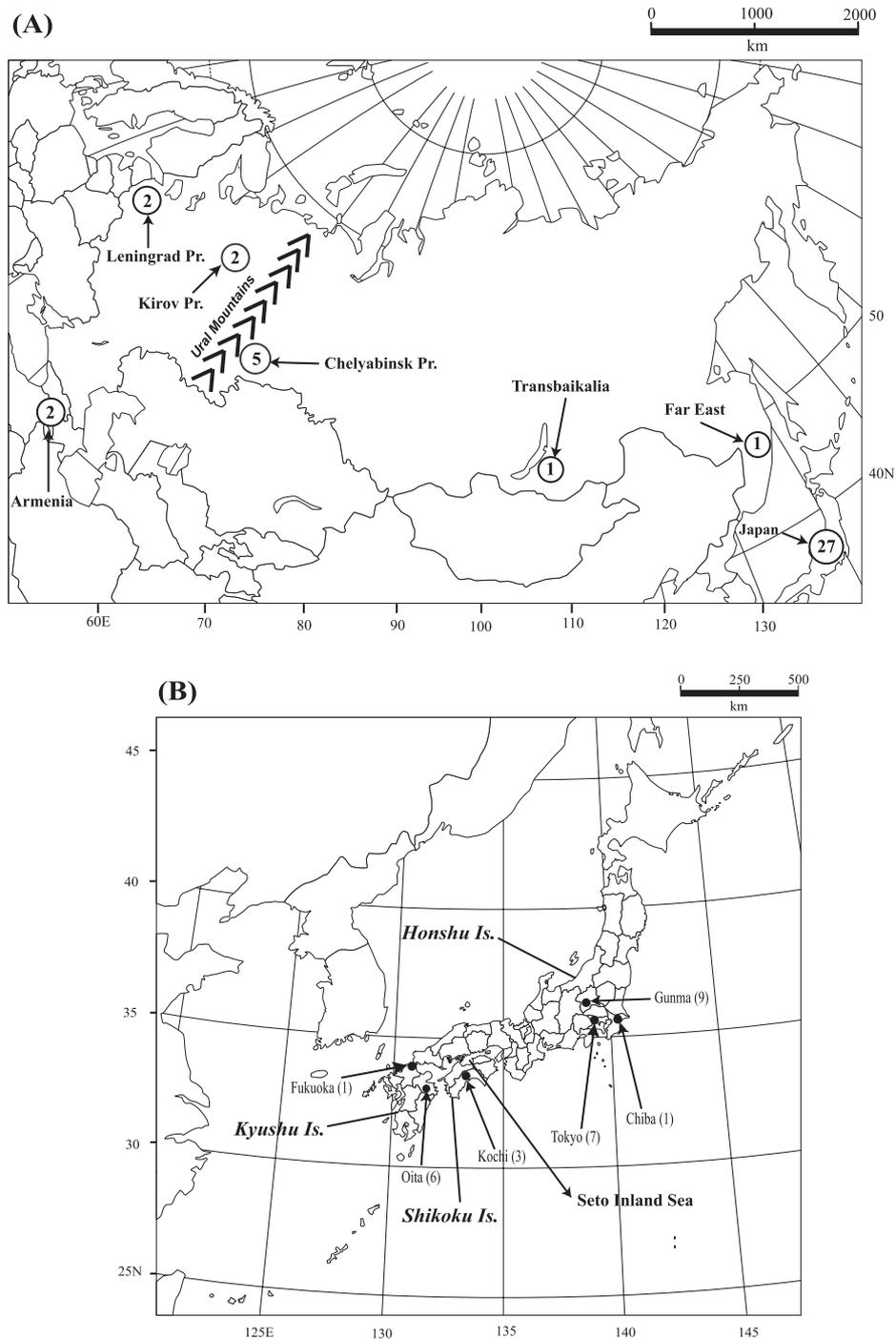
## Materials and methods

### *Samples and DNA extraction*

Tissue samples (muscle, hair, skin and blood) were obtained from 40 male Eurasian badgers. Sampling locations are shown in Fig. 1. The DNA extraction was performed by using the QiAamp DNA Micro Kit (Qiagen) for hairs and the DNeasy Blood & Tissue Kit (Qiagen) for the other tissues. Extracted DNAs (about 1–100 ng/μl) were dissolved in 200 μl of TE buffer and preserved at 4°C until use.

### *Amplification and sequencing of SINE in the ZFY final intron*

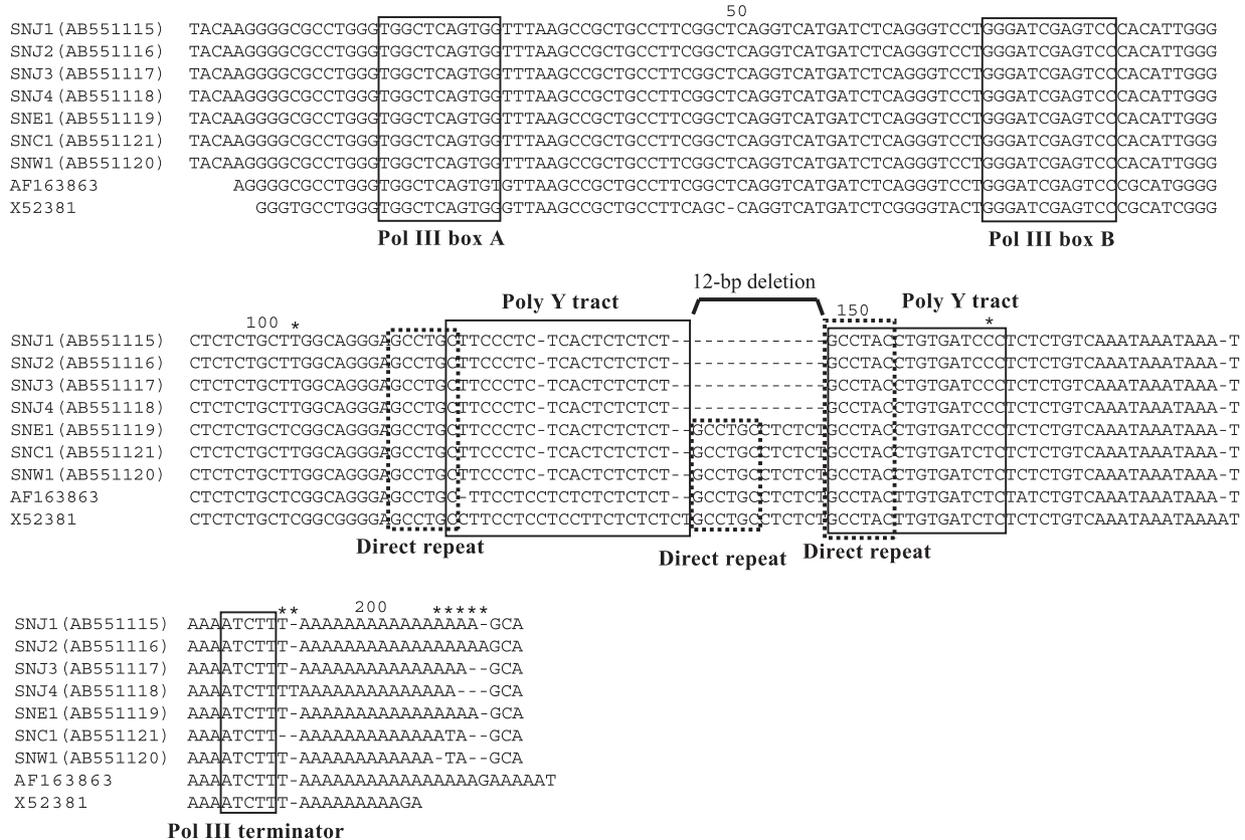
Two PCR primers to amplify the SINE inserted in the *ZFY* final intron were newly designed using the published sequence data of the Japanese badger (accession no. AB491600: Yamada and Masuda 2010): MELSN-F2 (5'-GGAGCAAGACAAATAATTCAG-3') and MELSN-R2 (5'-CATTTAAAGCCACATGTATTTGG-3'). A total of 50 μl of the PCR reaction solution consisted of 5 μl of 10 × PCR buffer (Takara), 4 μl of 2.5 mM dNTP mixture, 0.25 μl of *rTaq* DNA polymerase (5 units/μl, Takara), 0.5 μl of each of the above two primers and 5 μl of the DNA extract (about 1–100 ng/μl). The PCR reactions were carried out in a PCR thermal cycler (TP600, Takara) with the following reaction conditions: one cycle of 94°C for 3 min; 35 cycles of 94°C for 1 min, 57°C for 1 min, 72°C for 1 min; and one cycle of 72°C for 10 min. The PCR products were purified with the QIAquick PCR Purification Kit (Qiagen) and subjected to the sequencing reaction. In the present study, two sequencing primers were newly designed based on the published *ZFY* final intron sequence of the Japanese badger (acces-



**Fig. 1.** (A) The sampling locations in the Eurasian Continent. Numerals in circles are the numbers of individuals from those locations. (B) The precise sampling locations on the Japanese islands. Numerals in parentheses are the numbers of individuals from those locations. The boundaries of prefectures are shown by lines within three main islands of the Japanese archipelago.

sion number AB491600; Yamada and Masuda 2010): MELSN-SQ1 (5'-GCCACATGTATTTGGATATCTA-3') and MELSN-SQ2 (5'-AGACAAATAATTCAGTTGTCAT-3'), and 3'-labeled with Texas Red. The cycle PCR condition was one cycle of 94°C for 3 min; 30 cycles of

94°C for 30 sec; 55°C for 30 sec; 72°C for 1 min; 1 cycle of 72°C, 7 min, using the Thermo Sequence pre-mixed cycle sequencing kit (Amersham). The PCR products were run using an automated DNA sequencer (SQ5500, Hitachi).



**Fig. 2.** The components of the seven CAN-SINE haplotypes (SNJ1 to SNJ4, SNE1, SNC1, and SNW1) identified from the Eurasian badger. Their accession numbers are shown together with the seven CAN-SINE haplotypes. RNA polymerase III boxes A and B and terminator, and the polypyrimidine (C/T shown as Y) tracts are enclosed by solids lines. Direct repeat motifs are enclosed by broken lines. Numerals (end characters) and asterisks above the SNJ1 sequence indicate nucleotide site numbers and polymorphic sites of haplotypes, respectively. AF163863 (Leib et al. 2005) and X52381 (Lavrentieva et al. 1991) are accession numbers of the published CAN-SINE sequences of the American mink (*Neovison vison*).

*Phylogenetic analysis of nucleotide sequences*

A sequence alignment was performed by software CLUSTAL W (Thompson et al. 1994). The published CAN-SINE sequence of the tyrosine aminotransferase gene (AF163863: Leib et al. 2005) and that of the X-chromosome (X52381: Lavrentieva et al. 1991) of the American mink (*Neovison vison*) were used as references. A median-joining network was constructed by using software NETWORK (fluxus-engineering.com <http://www.fluxus-engineering.com/sharenet.htm>: Bandelt et al. 1999), where a gap was counted as one nucleotide substitution.

**Results**

*Sequence variations and structures of the SINE insertion in the ZFY final intron*

The two PCR primers newly designed for targetting the CAN-SINE sequence of the Japanese badger *ZFY* final intron successfully amplified the highly homologous

CAN-SINE sequences from a total of 40 males of the Japanese and continental populations of the Eurasian badgers (Fig. 2). Of the determined SINE sequences (197–210 bp), 21 nucleotide sites were polymorphic including 12-bp insertions/deletions (indels), and four haplotypes from Japan (SNJ1, SNJ2, SNJ3 and SNJ4) and three haplotypes from the Eurasian Continent (SNE1, SNC1 and SNW1) were identified (Fig. 2). The substitutions among the haplotypes were transitions between T and C at nucleotide sites 102 and 161, except the A/T rich region. A 12-bp deletion occurred at nucleotide sites 135–146 in the Japanese haplotypes, compared from the continental ones (Fig. 2). The other nucleotide differences between the haplotypes were polymorphisms of the numbers (14–17 nucleotides) of the A/T repeats at the A/T rich tail (Fig. 2). Even within the Japanese badgers, the geographical variation in the numbers of the A/T repeats led to classification of the four Japanese haplotypes.

An alignment and components of SINE sequences

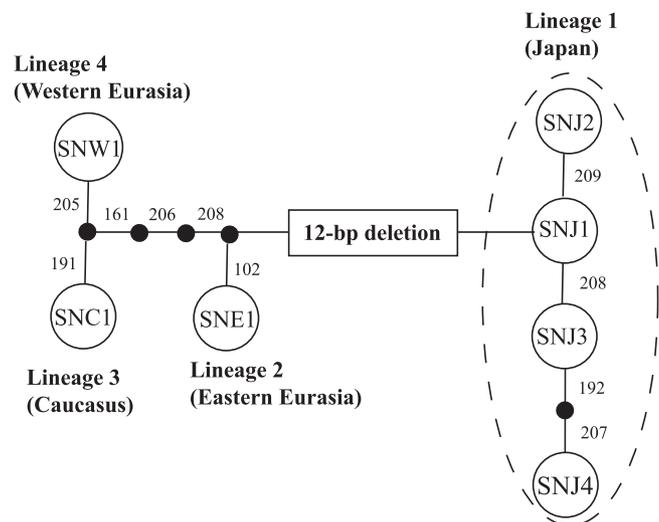
obtained in the present study showed the high similarity with the previously reported CAN-SINEs (AF163863 and X52381) from the American mink (Fig. 2). The CAN-SINE haplotype sequences included RNA polymerase III boxes A and B and terminator, two polypyrimidine tracts (C/T shown as Y in Fig. 2), and two direct repeats (GCCTGC and GCCTAC), each of which adhered to the 5' ends of the two polypyrimidine tracts, respectively. In addition, the 12-bp deletion between the two polypyrimidine tracts was found to be specific to the Japanese haplotypes (SNJ1 to SNJ4), whereas the 12-bp region (non-deletion) in the continental haplotypes (SNE1, SNC1 and SNW1) consisted of one direct repeat (GCCTGC) at the 5' end and 6-bp microsatellite-like sequences (TCTCTC) at the 3' end (Fig. 2). Consequently, in the entire sequence of these CAN-SINEs, the four Japanese haplotypes contained two direct repeats, whereas the three continental haplotypes had three.

The CAN-SINE sequences determined in the present study were deposited to DDBJ/GenBank/EMBL databases with the following accession numbers: AB551115–AB551121.

#### Molecular phylogeny of CAN-SINE in the badger *ZFY* final intron

A median-joining network (Fig. 3) showed that the SINE haplotypes were grouped into four lineages (Lineages 1–4). Lineage 1 consisting of the four Japanese haplotypes (SNJ1, SNJ2, SNJ3 and SNJ4) was apparently separated from the other three lineages, all of which consisted of the continental haplotypes with the 12-bp region (non-deletion at nucleotide sites 135–146) (Figs. 2 and 3).

Table 1 showed the distribution and frequencies of the badger CAN-SINE haplotypes in Eurasia. SNE1 (Lin-



**Fig. 3.** A median joining network of CAN-SINE haplotypes detected from the badger *ZFY* final intron. The 12-bp deletion found in the Japanese badger is shown as a square between Lineages 1 and 2. Open circles and filled circles represent identified CAN-SINE haplotypes and hypothesized haplotypes, respectively. Numerals near lines indicate numbers of polymorphic nucleotide sites.

age 2) was identified from eastern Eurasian badgers of Far East, Transbaikalia and Chelyabinsk Province. SNW1 (Lineage 4) was found in western Eurasian badgers of Kirov and Leningrad Provinces. SNC1 (Lineage 3) was from Armenia (Table 1). The distribution pattern shows the genetic differentiation among the sampling localities in the Eurasian Continent.

Among the four Japanese haplotypes, SNJ1 was most closely related to the continental haplotypes (Fig. 3), and shared by 15 badgers from Honshu Island and three from Shikoku Island (Table 1). SNJ2 was specific to Honshu. SNJ3 and SNJ4, both of which were identified from Kyushu Island, were connected with each other (Fig. 3; see Fig. 1 for geographical locations of the continent and Japanese islands).

**Table 1.** Distribution and frequencies of CAN-SINEs of the *ZFY* final intron among regional populations of Eurasian badgers (genus *Meles*)

Haplotype	Number of animals	Eurasian Continent								
		Japan ( <i>M. anakuma</i> )			Eastern Eurasia ( <i>M. leucurus</i> )			Western Eurasia ( <i>M. meles</i> )		Caucasus ( <i>M. meles</i> )
		Honshu	Shikoku	Kyushu	Far East	Transbaikalia	Chelyabinsk	Kirov Pr.	Leningrad Pr.	Armenia
SNJ1	18	15	3							
SNJ2	2	2								
SNJ3	5			5						
SNJ4	2			2						
SNE1	7				1	1	5			
SNW1	4							2	2	
SNC1	2									2
Total	40	17	3	7	1	1	5	2	2	2

## Discussion

### *Components of CAN-SINEs in the Eurasian badger ZFY final intron*

The majority of SINEs consist of three parts (tRNA-related and -unrelated regions and A/T rich regions), and the most common source of SINEs is tRNA of lysine (Shedlock and Okada 2000). Furthermore, the typical CAN-SINEs have the two promoter regions of RNA polymerase III (Pol III boxes A and B) in the tRNA-related region, polypyrimidine tracts, and the A/T rich region overlapping with a polyadenylation signal (AATAAA), and direct repeats in the tRNA unrelated region (Vassetzky and Kramerov 2002; López-Giráldez et al. 2005). The SINEs of the badger *ZFY* final intron obtained in the present study possessed these characteristic structures. Thus, the structural homologies clearly show that the badger SINEs can be classified in CAN-SINEs.

In both Pol III boxes A and B, there were no substitutions among the haplotypes obtained in the present study, whereas one substitution (at nucleotide site 161) in the second polypyrimidine tract was found between haplotypes of the Japanese/eastern Eurasian badgers and those of the western Eurasian/Caucasian badgers. The polypyrimidine tract was reported in not only the most families of order Carnivora (Vassetzky and Kramerov 2002), but also the hedgehog (*Mesochinus dauuricus*: Borodulina and Kramerov 2001) and tree shrew (*Tupaia glis*: Ten et al. 2006). Moreover, because the sequences and length of the polypyrimidine tract vary even between phylogenetically related species such as the wolf (*Canis lupus*) and red fox (*Vulpes vulpes*), it is assumed that the generation of this tract is associated with expansion of microsatellite-like regions (Vassetzky and Kramerov 2002). Especially, we found that the direct repeats (GCCTGC and GCCTAC) adhere to the 5' end of the first and second polypyrimidine tracts, respectively (Fig. 2). Vassetzky et al. (2003) and Ten et al. (2006) reported that such direct repeats consisting of 3–6 bp could generate internal duplications and deletions in mammal SINEs. They also mentioned that one of the direct repeats was present within the deletions. Actually in the present study, within the 12-bp indels of all the three continental badgers' CAN-SINEs, one of the direct repeats (GCCTGC) was found to adhere to the 5' end of 6-bp microsatellite-like sequences (CTCTCT) (Fig. 2). These suggest that the formation of two polypyrimidine tracts and the 12-bp indels found in the badger CAN-

SINEs has been mediated by the 6-bp direct repeats.

In addition, the repeat number of A/T in the downstream of the Pol III terminator was variable (Fig. 2). The poly A/T rich tail was known to be microsatellite-like sequences in the human (Shriver et al. 1992) and pig (Ellegren 1993), and it was thought to be associated with generation of microsatellites in mustelids (López-Giráldez et al. 2005, 2006). Polymorphisms of the nucleotide repeat number in the A/T rich tail within the species were reported in the European wild cat (*Felis silvestris*: Pecon-Slattey et al. 2004). The present study first shows the intraspecific and interspecific variations of length in the A/T rich region of the Eurasian badgers.

### *Phylogenetic features of the CAN-SINE sequences for the Eurasian badgers*

The four lineages identified in the present study are separately localized following geographical distributions of the Eurasian badgers. It means that the molecular differentiation of the CAN-SINE in the *ZFY* final intron corresponds with the morphological differentiation of the Eurasian badgers classified into the four groups: the classification recently proposed by Del Cerro et al. (2010) using an agreement with molecular phylogeny. In addition, the SINE lineages were in agreement with those of mtDNA (Marmi et al. 2006; Del Cerro et al. 2010; Tashima et al. 2011). Yamada and Masuda (2010) reported that the CAN-SINE sequence was inserted to the *ZFY* final intron independently to the Japanese badger and the ermine, after diversification of at least nine Asian mustelid species. The present study revealed that the CAN-SINE could be inserted once prior to speciation of the Eurasian badgers, because it occurs commonly at the same nucleotide position in the *ZFY* final intron of genus *Meles*.

It is likely that the 12-bp region was deleted from the CAN-SINE of the ancestor of the Japanese badgers, rather than this region was inserted into the *ZFY* final intron of the continental badgers. That is, the 12-bp deletion could have caused secondarily after the SINE insertion into the badger *ZFY* final intron. The first reason is that the previously reported mustelid CAN-SINEs in addition to the haplotypes identified from the present study also contain the homologous 12-bp sequences (6-bp direct repeat and 6-bp microsatellite-like sequences) as shown in Fig. 2. The second is that the ancestors of the Japanese badgers are thought to have immigrated from the Eurasian continent via land bridges during Pleistocene (Kawamura et al. 1989; Kawamura 1991).

Thus, the finding of the 12-bp deletion within the CAN-SINEs of the Japanese badgers (Fig. 2) suggests their geographical and genetic separation from the continental badgers.

It is interesting that single haplotypes of CAN-SINE were identified from three continental lineages, whereas four haplotypes were found in the Japanese lineage. Tashima et al. (2011) reported the similar frequencies of identified haplotypes of the *SRY* gene as another paternally inherited gene: 6 types in 33 Japanese badgers; 2 types in 8 badgers of Chelyabinsk Province; 1 type in both 2 badgers of Kirov Province and 2 badgers of Leningrad Province; 1 type in 2 badgers of Armenia. Although this could be attributed to a small number of samples and sampling localities in the continent, the findings suggest a higher genetic variability in the Japanese badgers. The variation of mtDNA haplotypes in the Japanese badgers was higher than that in the continental badgers (Tashima et al. 2011). Thus, the geographic isolation within islands could have resulted in the relatively higher genetic variability in the Japanese badgers.

The four CAN-SINE haplotypes identified from the Japanese badgers were distributed region-specifically: the number of single adenine repeats in the tail part was lower (15 in SNJ3 and 14 in SNJ4) in individuals from Kyushu Island than those from Honshu (16 in SNJ1 or 17 in SNJ2) and Shikoku (16 in SNJ1). Geographical isolation of Honshu, Shikoku and Kyushu Islands by the Seto Inland Sea, which started 7,000–5,000 years ago (Ohshima 1990), would have genetically differentiated the island populations within Japan. Tashima et al. (2010) examined biparentally inherited microsatellite loci of the Japanese badgers and showed the genetic differentiation among Honshu, Shikoku, and Kyushu Islands, suggesting the geographic isolation of the Japanese populations via the Seto Inland Sea. Long uninterrupted mono-nucleotide repeats as seen in the 3' end of the CAN-SINEs are likely related to the time since its mutation, that is, the ages of longer repeats are thought to be younger (Batzer et al. 1990; Ellegren 1993). The variabilities in the number of nucleotides in the poly A/T tail of the badger CAN-SINEs could have been generated by nucleotide slippages after the geographical isolation between the continent and the Japanese islands and even among the Japanese islands.

**Acknowledgments:** We thank T. Tsujimoto, H. Tanaka, S. Dakemoto, V. Sotnikov, O. Kruglova, and I. Khorozyan for supplying badger samples. This study was supported

in part by Grants-in-Aid for Scientific Research (Nos. 17405012 and 22405003) from the Japan Society for the Promotion of Science and the Russian Foundation for the Basic Research (No. 09-09-00073).

## References

- Abramov, A. V. 2001. Notes on the taxonomy of the Siberian badgers (Mustelidae: *Meles*). Proceedings of Zoological Institute Russian Academy of Sciences 288: 221–233.
- Abramov, A. V. 2002. Variation of the baculum structure of the Palearctic badger (Carnivora, Mustelidae, *Meles*). Russian Journal of Theriology 1: 57–60.
- Abramov, A. V. and Puzachenko, A. Yu. 2005. Sexual dimorphism of craniological characters in Eurasian badgers, *Meles* spp. (Carnivora, Mustelidae). Zoologischer Anzeiger 244: 11–29.
- Abramov, A. V. and Puzachenko, A. Yu. 2006. Geographical variability of skull and taxonomy of Eurasian badgers (Mustelidae, *Meles*). Zoologicheskii Zhurnal 85: 641–655.
- Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K. and Walter, P. 2001. Molecular Biology of the Cell. 4th ed. Garland, New York.
- Bandelt, H. J., Forster, P. and Röhl, A. 1999. Median-joining networks for inferring intraspecific phylogenies. Molecular Biology and Evolution 16: 37–48.
- Batzer, M., Kilroy, G. E., Richard, P. E., Shaikh, T. H., Desselle, T. D., Hoppens, C. L. and Deininger, P. L. 1990. Structure and variability of recently inserted Alu family members. Nucleic Acids Research 18: 6793–6798.
- Borodulina, O. R. and Kramerov, D. A. 2001. Short interspersed elements (SINEs) from insectivores. Two classes of mammalian SINEs distinguished by A-rich tail structure. Mammalian Genome 12: 779–786.
- Daniels, G. R. and Deininger, P. L. 1985. Repeat sequence families derived from mammalian tRNA genes. Nature 317: 819–822.
- Del Cerro, I., Marmi, J., Ferrando, A., Chashchin, P., Taberlet, P. and Bosch, M. 2010. Nuclear and mitochondrial phylogenies provide evidence for four species of Eurasian badgers (Carnivora). Zoologica Scripta 39: 415–425.
- Ellegren, H. 1993. Variable SINE 3' poly (A) sequences: an abundant class of genetic markers in the pig genome. Mammalian Genome 4: 429–434.
- Heptner, V. G., Naumov, N. P., Yutgenon, P. B., Sludskii, A. A., Chirkova, A. F. and Bannikov, A. G. 1967. Subfamily Melinae, Genus *Meles* Brisson, 1762, *Meles meles* (Linnaeus, 1758). In (V. G. Heptner and N. P. Naumov, eds.) Mammals of the Soviet Union, Vol. II, Part 1 Sirenia and Carnivora, pp. 1228–1282. Vysshaya Shkola, Moscow (Scientific Publisher, Enfield, for the English-translated version).
- Kawai, K., Nikaido, M., Harada, M., Matsuura, S., Lin, L.-K., Wu, Y., Hasegawa, M. and Okada, N. 2002. Intra- and interfamily relationships of Vespertilionidae inferred by various molecular markers including SINE insertion data. Journal of Molecular Evolution 55: 284–301.
- Kawamura, Y. 1991. Quaternary mammalian fauna in the Japanese islands. Quaternary Research 30: 213–220.
- Kawamura, Y., Kamei, T. and Taruno, H. 1989. Middle and late Pleistocene mammalian fauna in Japan. Quaternary Research 28: 317–326 (in Japanese with English abstract).
- Kramerov, D. A. and Vassetzky, N. S. 2005. Short Retrotransposons in Eukaryotic Genomes. In (K. W. Jeon, ed.) International

- Review of Cytology: A Survey of Cell Biology, pp. 165–221. Elsevier Academic Press, London.
- Kurose, N., Kaneko, Y., Abramov, A. V., Siritroonrat, B. and Masuda, R. 2001. Low genetic diversity in Japanese populations of the Eurasian badger *Meles meles* (Mustelidae, Carnivora) revealed by mitochondrial cytochrome *b* gene sequences. *Zoological Science* 18: 1145–1151.
- Labuda, D., Sinnet, F., Richar, C., Deragon, J. M. and Striker, A. 1991. Evolution of mouse B1 repeats: 7SL RNA falling pattern conserved. *Journal of Molecular Evolution* 32: 405–514.
- Lavrentieva, M. V., Rivkin, M. I., Shilov, A. G., Kobetz, M. L., Rogozin, I. B. and Serov, O. L. 1991. B2-like repetitive sequence from the X chromosome of the American mink (*Mustela vison*). *Mammalian Genome* 1: 161–170.
- Leib, S. R., McGuire, T. C. and Prieur, D. J. 2005. Comparison of the tyrosine aminotransferase cDNA and genomic DNA sequences of normal mink and mink affected with tyrosinemia type II. *Journal of Heredity* 96: 302–309.
- López-Giráldez, F., Gómez-Moliner, B., Marmi, J. and Domingo-Roura, X. 2005. Genetic distinction of American and European mink (*Mustela vison* and *M. luticola*) and European polecat (*M. putorius*) hair samples by detection of a species-specific SINE and a RFLP assay. *Journal of Zoology* 265: 405–410.
- López-Giráldez, F., Andr ez, O., Domingo-Roura, X. and Bosch, M. 2006. Analysis of carnivore microsatellites and their intimate association with tRNA-derived SINE. *BMC Genetics* 7: 269–280.
- Lynch, J. M. 1994. Morphometric variation in the badger (*Meles meles*): clinical variation in cranial size and shape across Eurasia. *Small Carnivore Conservation* 10: 6–7.
- Marmi, J., L pez-Gir ldez, F., Macdonald, D. W., Calafell, F., Zholnerovskaya, E. and Domingo-Roura, X. 2006. Mitochondrial DNA reveals a strong phylogeographic structure in the badger across Eurasia. *Molecular Ecology* 15: 1007–1020.
- Minnick, M. F., Stillwell, L. C., Heineman, J. M. and Stiegler, G. L. 1992. A highly repetitive DNA sequence possibly unique to canids. *Gene* 110: 235–238.
- Ohshima, K. 1990. The history of straits around the Japanese Islands in the late-Quaternary. *Quaternary Research* 29: 193–208 (in Japanese with English abstract).
- Okada, N., Hamada, M., Ogiwara, I. and Ohsima, K. 1997. SINES and LINES share common 3' sequences: a review. *Gene* 205: 229–243.
- Pecon-Slattery, J., Murphy, W. J. and O'Brien, S. J. 2000. Patterns of diversity among SINE elements isolated from three Y-chromosome genes in carnivores. *Molecular Biology and Evolution* 17: 825–829.
- Pecon-Slattery, J., Wilkerson, A. J. P., Murphy, W. J. and O'Brien, S. J. 2004. Phylogenetic assessment of introns and SINES within the Y chromosome using the cat family Felidae as a species tree. *Molecular Biology and Evolution* 21: 2299–2309.
- Shedlock, A. M. and Okada, N. 2000. SINE insertions: powerful tools for molecular systematics. *BioEssays* 22: 148–160.
- Shimamura, M., Yasue, H., Ohshima, K., Abe, H., Kato, K., Kishiro, T., Goto, M. and Okada, N. 1997. Molecular evidence from retroposons that whales from a clade within even-toed ungulates. *Nature* 388: 666–670.
- Shriver, M. D., Siest, G. and Boerwinkle, E. 1992. Length and sequence variation in the apolipoprotein B intron 20 *Alu* repeat. *Genomics* 14: 449–454.
- Tashima, S., Kaneko, Y., Anezaki, T., Baba, M., Yachimori, S. and Masuda, R. 2010. Genetic diversity within the Japanese badgers (*Meles anakuma*), as revealed by microsatellite analysis. *Mammal Study* 35: 221–226.
- Tashima, S., Kaneko, Y., Anezaki, T., Baba, M., Yachimori, S., Abramov, A. V., Saveljev, A. P. and Masuda, R. 2011. Phylogeographic sympatry and isolation of the Eurasian badgers (*Meles*, Mustelidae, Carnivora): implications for an alternative analysis using maternally as well as paternally inherited genes. *Zoological Science* 28: 293–303.
- Ten, O. A., Borodulina, O. R., Vassetzky, N. S., Oparina, N. Iu. and Kramerov, D. A. 2006. Repetitive sequences of the tree shrew genome (Mammalia, Scandentia). *Molecular Biology* 40: 63–71.
- Thompson, J. D., Higgins, D. G. and Gibson, T. J. 1994. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research* 22: 4673–4680.
- Ullu, E. and Tschudi, C. 1984. *Alu* sequences are processed 7SL RNA genes. *Nature* 312: 171–172.
- Van der Vlugt, H. H. J. and Lenstra, J. A. 1995. SINE elements of carnivores. *Mammalian Genome* 6: 49–51.
- Vassetzky, N. S. and Kramerov, D. A. 2002. CAN-a pan-carnivore SINE family. *Mammalian Genome* 13: 50–57.
- Vassetzky, N. S., Ten, O. A. and Kramerov, D. A. 2003. B1 and related SINES in mammalian genomes. *Gene* 319: 149–160.
- Weiner, A. M., Deninger, P. L. and Efstratiadis, A. 1986. Nonviral retroposons: genes, pseudogenes, and transposable elements generated by the reverse flow of genetic information. *Annual Review of Biochemistry* 55: 631–661.
- Wozencraft, W. C. 2005. Order Carnivora. In (D. E. Wilson and D. M. Reeder, eds.) *Mammals Species of the World: A Taxonomic and Geographic Reference*. 3rd ed., pp. 532–628. Johns Hopkins University Press, Baltimore.
- Yamada, C. and Masuda, R. 2010. Molecular phylogeny and evolution of sex-chromosomal genes and SINE sequences in the family Mustelidae. *Mammal Study* 35: 17–30.

Received 15 September 2010. Accepted 28 December 2010.