



Title	Type II NKT Cells Stimulate Diet-Induced Obesity by Mediating Adipose Tissue Inflammation, Steatohepatitis and Insulin Resistance
Author(s)	Satoh, Masashi; Andoh, Yasuhiro; Clingan, Christopher Stuart et al.
Citation	PLoS One, 7(2), e30568 https://doi.org/10.1371/journal.pone.0030568
Issue Date	2012-02-22
Doc URL	https://hdl.handle.net/2115/49209
Rights(URL)	https://creativecommons.org/licenses/by/3.0/
Type	journal article
File Information	PLoS0ne7-2_e30568.pdf



Type II NKT Cells Stimulate Diet-Induced Obesity by Mediating Adipose Tissue Inflammation, Steatohepatitis and Insulin Resistance

Masashi Satoh^{1,2,3}, Yasuhiro Andoh^{2,3,3}, Christopher Stuart Clingan^{2,4}, Hisako Ogura², Satoshi Fujii⁵, Koji Eshima¹, Toshinori Nakayama⁶, Masaru Taniguchi⁷, Noriyuki Hirata^{2,8}, Naoki Ishimori³, Hiroyuki Tsutsui³, Kazunori Ono², Kazuya Iwabuchi^{1,2*}

1 Department of Immunology, Kitasato University School of Medicine, Sagamihar, Japan, **2** Division of Immunobiology, Research Section of Pathophysiology, Institute for Genetic Medicine, Hokkaido University, Sapporo, Japan, **3** Department of Cardiovascular Medicine, Hokkaido University Graduate School of Medicine, Sapporo, Japan, **4** Department of Microbiology and Immunology, Vanderbilt University School of Medicine, Nashville, Tennessee, United States of America, **5** Department of Molecular and Cellular Pathobiology and Therapeutics, Graduate School of Pharmaceutical Sciences, Nagoya City University, Nagoya, Japan, **6** Department of Immunology, Graduate School of Medicine, Chiba University, Chiba, Japan, **7** RIKEN Research Center for Allergy and Immunology, Yokohama, Japan, **8** Division of Cancer Biology, Research Section of Pathophysiology, Institute for Genetic Medicine, Hokkaido University, Sapporo, Japan

Abstract

The progression of obesity is accompanied by a chronic inflammatory process that involves both innate and acquired immunity. Natural killer T (NKT) cells recognize lipid antigens and are also distributed in adipose tissue. To examine the involvement of NKT cells in the development of obesity, C57BL/6 mice (wild type; WT), and two NKT-cell-deficient strains, $J\alpha 18^{-/-}$ mice that lack the type I subset and $CD1d^{-/-}$ mice that lack both the type I and II subsets, were fed a high fat diet (HFD). $CD1d^{-/-}$ mice gained the least body weight with the least weight in perigonadal and brown adipose tissue as well as in the liver, compared to WT or $J\alpha 18^{-/-}$ mice fed an HFD. Histologically, $CD1d^{-/-}$ mice had significantly smaller adipocytes and developed significantly milder hepatosteatosis than WT or $J\alpha 18^{-/-}$ mice. The number of $NK1.1^{+}TCR\beta^{+}$ cells in adipose tissue increased when WT mice were fed an HFD and were mostly invariant $V\alpha 14J\alpha 18$ -negative. $CD11b^{+}$ macrophages (M ϕ) were another major subset of cells in adipose tissue infiltrates, and they were divided into $F4/80^{high}$ and $F4/80^{low}$ cells. The $F4/80^{low}$ -M ϕ subset in adipose tissue was increased in $CD1d^{-/-}$ mice, and this population likely played an anti-inflammatory role. Glucose intolerance and insulin resistance in $CD1d^{-/-}$ mice were not aggravated as in WT or $J\alpha 18^{-/-}$ mice fed an HFD, likely due to a lower grade of inflammation and adiposity. Collectively, our findings provide evidence that type II NKT cells initiate inflammation in the liver and adipose tissue and exacerbate the course of obesity that leads to insulin resistance.

Citation: Satoh M, Andoh Y, Clingan CS, Ogura H, Fujii S, et al. (2012) Type II NKT Cells Stimulate Diet-Induced Obesity by Mediating Adipose Tissue Inflammation, Steatohepatitis and Insulin Resistance. PLoS ONE 7(2): e30568. doi:10.1371/journal.pone.0030568

Editor: Simon Afford, University of Birmingham, United Kingdom

Received: August 9, 2011; **Accepted:** December 19, 2011; **Published:** February 22, 2012

Copyright: © 2012 Satoh et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported in part by Grant-in-Aid for Scientific Research (#B: #20390106) by the Japan Society for the Promotion of Science (JSPS), Global COE Program, Establishment of International Collaboration Center for Zoonosis Control, by the Ministry of Education, Culture, Sports, Science, and Technology (MEXT) of Japan, a Health and Labor Sciences Research Grant on Intractable Diseases (Neuroimmunological Diseases) from the Ministry of Health, Labor and Welfare of Japan, and a Grant by Heisei Ijuku Tomakomai East Hospital. MS is a recipient of a Parent's Association (Keyaki Kai) Grant of Kitasato University School of Medicine. No additional external funding received for this study. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: akimari@kitasato-u.ac.jp

These authors contributed equally to this work.

Introduction

Obesity is thought to progress with caloric excess under a chronic inflammatory process characterized by infiltration of adipose tissue by M ϕ and by cells of the adaptive immune system, such as T cells [1–3]. The inflammation in adipose tissue induces alterations in metabolic and endocrine functions of adipocytes, which leads to insulin resistance and various pathological responses [4,5]. Recent studies by Nishimura et al revealed the active participation of $CD8^{+}$ T cells in chronic inflammation in adipose tissue [6]. Moreover, $CD4^{+}Foxp3^{+}$ T cells with unique specificity have been detected in adipose tissue and were suggested to regulate the development of obesity by suppressing inflammatory responses [7]. Furthermore, additional findings showed that

the transfer of $CD4^{+}$ T cells from WT but not from T-cell receptor transgenic mice ameliorated the metabolic dysregulation in $Rag-1^{-/-}$ mice fed a high fat diet (HFD), which led to the idea that $CD4^{+}$ T cells play a suppressive role in diet-induced obesity (DIO) [8]. These studies have indicated that T cells that infiltrate adipose tissue are not just inert bystanders but are active modifiers of inflammation and thus either aggravate or ameliorate obesity.

Natural killer T (NKT) cells are a unique subset of T-lineage cells that recognize various lipid antigens in the context of $CD1d$ molecules [9]. Among lipid ligands, α -galactosylceramide (α -GalCer) is the prototype ligand [10] that can stimulate NKT cells to promptly produce large amounts of various cytokines and chemokines and also demonstrate cytotoxic activity [11]. Endogenous ligands can also stimulate NKT cells to perform their innate

effector functions [12]. Moreover, NKT cells localize to the liver [13], where lipid metabolism is active, and in adipose tissue [14], another location for lipid metabolism with endocrine functions. These considerations led us to suggest that, NKT cells might play a role in a disease that involves abnormal lipid metabolism or lipid-related inflammation. Indeed, several research groups including our team have demonstrated that NKT cells accelerate atherogenesis in a mouse model of atherosclerosis [15–17]. Furthermore, we have examined the involvement of NKT cells in insulin resistance induced in mice fed an HFD and demonstrated that NKT cells play an important role in adipose-tissue inflammation and glucose intolerance in β_2 -microglobulin knockout ($\beta_2m^{-/-}$) mice with DIO [18]. However, both mainstream $CD8^+$ T cells and various innate lymphocytes other than NKT cells also are absent in $\beta_2m^{-/-}$ mice [19]. Thus, we attempted to examine the involvement of NKT cells in DIO and insulin resistance using NKT cell-deficient mice. To this end, we compared B6 (WT) and two strains of NKT cell-deficient mice, $CD1d^{-/-}$ and $J\alpha 18^{-/-}$ mice on a B6 genetic background. Unlike our previous study in $\beta_2m^{-/-}$ mice [18], DIO was significantly suppressed in $CD1d^{-/-}$ mice compared to WT mice. Moreover, in $J\alpha 18^{-/-}$ mice where type I but not type II NKT cells were deficient, DIO was induced to an equal extent as in WT mice. The possible mechanisms underlying lipid-induced NKT-cell activation and the development of chronic inflammation by type II NKT cells in DIO are discussed.

Materials and Methods

Mice

Male and female 8-week-old C57BL/6 (B6; Nippon SLC, Shizuoka, Japan), $B6.CD1d^{-/-}$ [20], and $B6.J\alpha 18^{-/-}$ [21] mice were used. Mice were maintained on food and water *ad libitum* until they reached the desired weight (20–24 g) or age (8 wk) under specific pathogen-free conditions. All experiments were approved by the Committees of Animal Experimentation at Hokkaido University (permit number: #09-0022) and Kitasato University (permit number: #2011 105).

Diet-induced obesity

Mice were fed either regular chow as a standard fat diet (SFD; Nihon Nosan: fat 4.3%, cholesterol 0.03%, protein 18.3%, carbohydrate 58.3%) or an HFD (CLEA Japan HFD-32: fat (powdered tallow and safflower oil of high oleic type) 32.0%, protein 25.5%, fiber 2.9%, mineral 4.0%, nitrogen 29.4%, water 6.2%) starting from 8 wk of age for 18 wk. Mice were weighed weekly. The ingredients of HFD-32 are listed in detail at <http://www.clea-japan.com/Feed/pdf/clea_hfd32.pdf>. Major fatty acids included oleic acid ($C_{18:1}$; 64.3%), palmitic acid ($C_{16:0}$; 12.6%), linoleic acid ($C_{18:2}$; 10.2%), and stearic acid ($C_{18:0}$; 7.5%). After 18 wk of feeding, mice were sacrificed for analysis. For some experiments, mice were injected intraperitoneally with α -GalCer (0.1 μ g/g body weight, BW) or vehicle as control to examine the change of BW.

Blood chemistry

Total cholesterol (T-chol), high-density lipoprotein (HDL) cholesterol (HDL-chol), triglyceride (TG), and alanine aminotransferase (ALT) concentrations in sera were quantified by colorimetric assays with the Fuji Drychem system (Fujifilm Medical, Osaka, Japan) according to the manufacturer's protocol, as described elsewhere [16].

Histology and quantitative analyses of microscopic images

Perigonadal fat tissue was removed and fixed with buffered formaldehyde solution (10%) followed by ordinary processing for

paraffin-embedded sections and hematoxylin-eosin (HE) staining. Images of the HE-stained adipose tissue were incorporated with a BIOREVO microscope (BZ8100; Keyence Corp., Osaka, Japan), and morphometric analyses were performed with image analysis software (BZ-H2A, -H1C) equipped on the microscope. Liver samples were snap-frozen in OCT compound (Sakura Finetek Co., Tokyo, Japan) with liquid nitrogen, and cryosections were stained with Oil Red O (ORO) (Sigma Chemical Co., St. Louis, MO) and Meyer's hematoxylin (Wako Pure Chemical Co. Ltd., Osaka, Japan). Images of lipid droplets in hepatocytes stained red were quantified by computerized image analysis system (Scion Image software, Scion Corp., Frederick, MD).

Flow cytometry

Splenocytes were prepared by mincing the spleen with a glass homogenizer, and red blood cells were lysed with Tris- NH_4Cl solution. Hepatic mononuclear cells (HMNC) were isolated from liver homogenates by density-gradient centrifugation with 33% PercollTM (GE Healthcare Life Sciences, Piscataway, NJ) as previously reported [22]. Stromal vascular cells were isolated from the digest of perigonadal fat by mincing and incubating with collagenase D solution (2 mg/ml) (Roche Diagnostics, Indianapolis, IN) for 1–1.5 h. The cells were incubated with 2.4G2 monoclonal antibody (mAb) (anti-Fc γ R/III/II) to block non-specific binding of primary mAb and then reacted with $CD1d$:Ig recombinant fusion protein (BD Biosciences Pharmingen, San Diego, CA) loaded with α -GalCer (α -GalCer- $CD1d$ -dimer), followed by detection with phycoerythrin (PE)-conjugated anti-mouse IgG₁ mAb (A85-1; BD) according to the manufacturer's protocol [16]. After washing, cells were stained with a combination of the following mAb conjugates: fluorescein isothiocyanate (FITC)-anti-TCR β chain (H57-597; BD), -anti- $CD3\epsilon$ (145-2C11; BD), -anti- $CD1d$ (1B1; BD), -anti- $CD206$ (MR5D3; BioLegend, San Diego, CA), -rat IgG_{2a} (BD), -rat IgG_{2b} (BD), allophycocyanin (APC)-anti-NK1.1 (PK136; BD, BioLegend), -anti- $CD11c$ (HL3; BD), -Streptavidin (BioLegend), PE-anti- $CD4$ (RM4-5; BD), -anti- $CD8$ (53-6.7; BD), -anti- $CD11b$ (M1/70; BioLegend), and APC-Cy7-Streptavidin (BD). Stained cells were acquired with FACS Calibur or Canto II flow cytometers (BD Bioscience Immunocytometry Systems, San Jose, CA) and analyzed with CellQuest, FACS Diva (BDIS), or FlowJo (Tommy Digital Biology, Tokyo, Japan) software as described elsewhere [16]. Propidium iodide (PI; Sigma) or 7-aminoactinomycin D (7-AAD; BD) positive cells were electronically gated as dead cells from the analysis.

Quantification of serum cytokines and adipokines

The concentration of Th1/Th2 and inflammatory cytokines, including IFN- γ , tumor necrosis factor (TNF)- α , IL-1 α , 2, 4, 5, 6, 10, 17, and GM-CSF in serum was quantified with Mouse Th1/Th2 10plex FlowCytomixTM Multiplex (Bender MedSystems GmbH, Vienna, Austria) according to the manufacturer's protocol with a flow cytometer. Leptin (Morinaga Institute of Biological Sciences, Kanagawa, Japan) in sera was respectively quantified with enzyme-linked immunosorbent assay (ELISA) kit according to the manufacturers' protocol.

IPGTT and insulin tolerance test

Intraperitoneal (*i.p.*) glucose tolerance test (IPGTT) was performed by *i.p.* injection of glucose solution (1 g/kg) after 16 h of fasting [6]. The insulin tolerance test (ITT) was performed by *i.p.* injection of insulin (0.751 U/kg; Humulin R 100 U/ml; Eli Lilly Japan KK, Kobe, Japan) after 3.5 h of fasting. The blood glucose level was serially quantified with a blood glucose monitor (MEDISAFE MINI; Terumo Corp., Tokyo, Japan). The insulin

level was also quantified with the mouse insulin ELISA kit (Morinaga) according to the manufacturer's protocol.

HMNC transfer

HMNC were obtained from either $J\alpha 18^{-/-}$ or $CD1d^{-/-}$ mice and were intravenously transferred to $CD1d^{-/-}$ recipient mice (1×10^6 /mouse) at 8 wk. These mice were fed an HFD for 14 weeks and tested for IPGTT and ITT as above.

Culture of M ϕ from the stromal vascular fraction (SVF) of adipose tissue

Cells (1×10^5 /well) obtained from perigonadal adipose tissue were cultured in RPMI medium (supplemented with 10% heat-inactivated fetal calf serum, 100 U/ml penicillin, 100 μ g/ml streptomycin, and 50 μ M β -mercaptoethanol) with 1 μ g/ml LPS for 20 h. After 20 h, supernatants were collected and frozen at -80°C , and cytokines were later quantified as described above.

Statistical analysis

Results were demonstrated as means $\pm$ standard deviation (s.d.). Statistical analysis between two groups was performed by Student's *t*-test and among three groups was performed using ANOVA followed by Tukey-Kramer tests. Pearson's correlation coefficient test was used to examine the correlation. Values with $P < 0.05$ were considered statistically significant.

Results

Impact of the presence of NKT cells on the weight gain of mice fed an HFD

Female or male B6 (WT), $B6.J\alpha 18^{-/-}$ ($J\alpha 18^{-/-}$), and $B6.CD1d^{-/-}$ ($CD1d^{-/-}$) mice were fed either an SFD or an HFD from 8 wk to 22–26 wk of age (for 14–18 wk), and the BW of each group of mice was plotted over time. Three groups of female (Figure 1A) or male (Figure 1B) mice showed a similar and gradual increase of BW when fed an SFD. On the other hand, all groups of female and male mice gained substantially more BW on HFD than those on SFD. However, $CD1d^{-/-}$ mice showed significantly less gains than those of WT and $J\alpha 18^{-/-}$ mice (Figure 1A, B). Consistently, the least difference was observed in $CD1d^{-/-}$ mice when the net gain ([BW on HFD at *n* wk] – [BW on HFD at 8 wk]; $n \geq 8$ wk) was compared (Figure 1C, D). We also compared BW in WT mice that received either the prototypical type I NKT cell ligand α -GalCer or vehicle when fed either an SFD or an HFD. There was no significant difference in weight gain between mice that received α -GalCer or vehicle and fed either an SFD or an HFD (Figure S1A), suggesting that type I NKT cells might have a minimal role in the development of DIO even when the activating ligand is provided.

When the daily food intake per animal was compared in each group of mice by calculating $\Sigma[(\text{amounts given} - \text{amounts left}) / \text{total periods}]$ throughout the feeding periods, $CD1d^{-/-}$ female

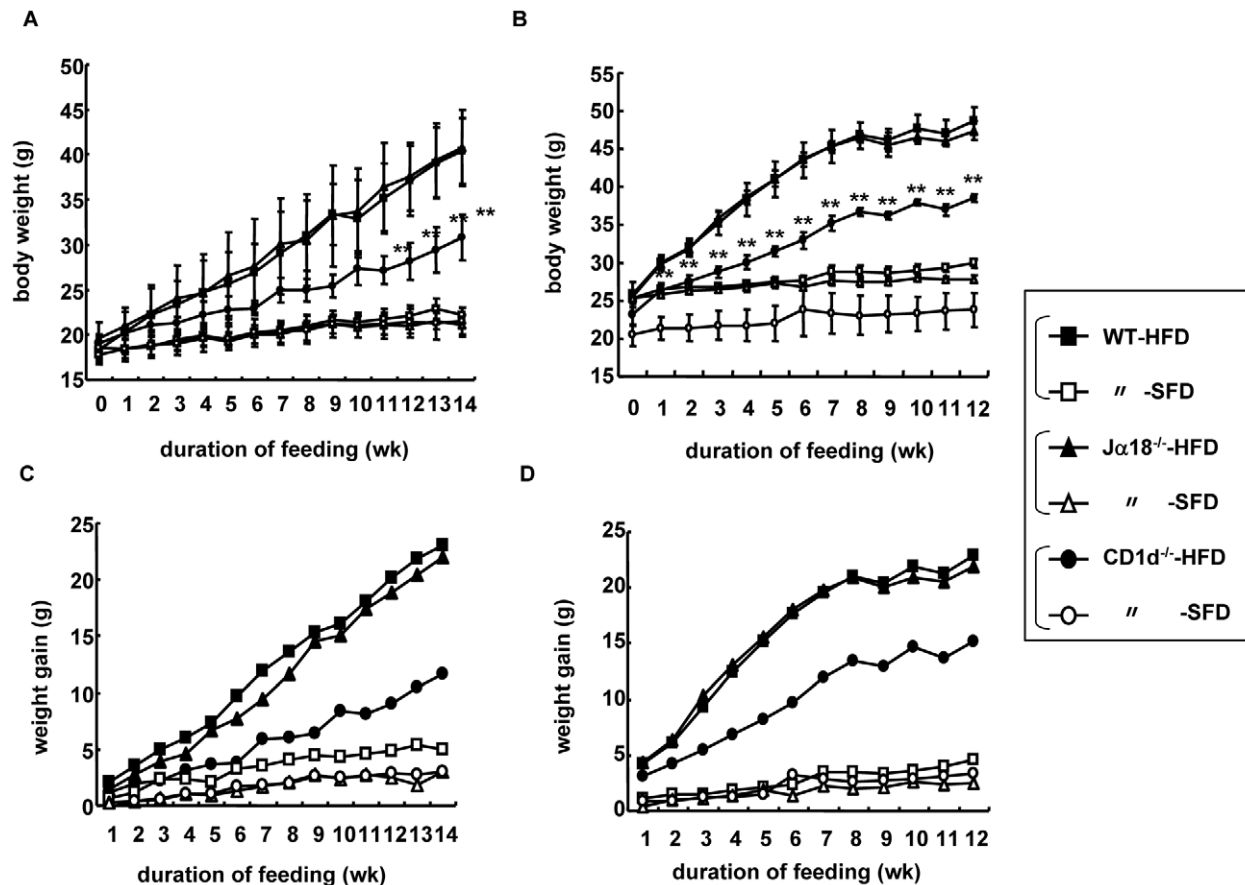


Figure 1. Body weight and weight gain of three strains of mice fed an SFD or an HFD. (A, B) Body weight (BW) of female (A) and male mice (B) fed a high-fat diet (HFD) or a standard-fat diet (SFD) at 8 wk and weighed weekly. ($n = 3-7$ in each group) (C, D) Representative data of three similar experiments are shown. Weight gain ($\Delta BW = BW_{n \text{ wk}} - BW_{8 \text{ wk}}$; $n \geq 9$) of female (C) and male (D) mice. Each point represents mean $\pm$ standard deviation (s.d.). Statistical analysis was performed according to the Tukey-Kramer test. $^{**}p < 0.01$ (WT and $J\alpha 18^{-/-}$ vs $CD1d^{-/-}$). doi:10.1371/journal.pone.0030568.g001

mice took in slightly smaller amounts of both SFD and HFD than the other two strains, WT and $J\alpha 18^{-/-}$ mice (Figure 2A). A similar tendency in food intake was observed in male mice (data not shown).

The BW of $CD1d^{-/-}$ mice was significantly lower than that of WT and $J\alpha 18^{-/-}$ mice after 18 wk of HFD feeding (Figure 2B) ($WT = J\alpha 18^{-/-} > CD1d^{-/-}$; $p < 0.01$ between WT and $CD1d^{-/-}$; $J\alpha 18^{-/-}$ and $CD1d^{-/-}$). The difference in BW in each group of mice was attributable to the sum of the differences in weights of organs and tissues. Thus, the liver, perigonadal fat tissue (white adipose tissue; WAT) and fat tissue between the scapulae (brown adipose tissue; BAT) of each group were obtained and compared. Consistent with the differences in BW, $CD1d^{-/-}$ mice had lower liver weight (Figure 2C) ($WT = J\alpha 18^{-/-} > CD1d^{-/-}$; $p < 0.01$ between WT and $CD1d^{-/-}$; $p < 0.05$ between $J\alpha 18^{-/-}$ and $CD1d^{-/-}$), lower WAT weight (Figure 2D) ($WT \geq J\alpha 18^{-/-} > CD1d^{-/-}$; $p < 0.01$ between WT and $CD1d^{-/-}$; $J\alpha 18^{-/-}$ and $CD1d^{-/-}$) and lower BAT weight (Figure 2E) ($WT \geq J\alpha 18^{-/-} > CD1d^{-/-}$; $p < 0.01$ between WT and $CD1d^{-/-}$), when compared to those of sex-matched WT or $J\alpha 18^{-/-}$ mice fed an HFD (closed bar). In SFD-fed mice, there were no significant differences in the weights of the whole BW, liver, WAT, and BAT among the three groups of mice (Figure 2B–E; open bar).

When serum lipids were quantified in the three groups of mice fed an HFD, WT and $J\alpha 18^{-/-}$ mice exhibited significantly higher levels of T-chole and HDL-chole than $CD1d^{-/-}$ mice (Figure S2A, B). The cholesterol level was consistent with the extent of adiposity, whereas the TG level was not different among the three groups (Figure S2C).

HFD-induced hepatosteatosis and adipocyte hypertrophy are ameliorated in $CD1d^{-/-}$ mice

First, livers were histologically examined for major causes of weight gain among three strains of mice on HFD. WT and $J\alpha 18^{-/-}$ mice on HFD had marked hepatosteatosis, which was revealed by red deposits on ORO staining (Figure 3A). On the other hand, livers of $CD1d^{-/-}$ mice that appeared less fatty exhibited lighter deposits on ORO staining (Figure 3A). The ORO-positive regions in $CD1d^{-/-}$ livers were significantly smaller than those of WT and $J\alpha 18^{-/-}$ mice (Figure 3B). The severity of steatosis appeared consistent with the liver weight and dyslipidemic level in each strain (Figure 2A and B). Furthermore, the ALT level, a marker of liver damage, was significantly suppressed in $CD1d^{-/-}$ mice (Figure 3C). We also quantified pro-inflammatory cytokines that were secreted from HMNC in mice fed an HFD upon stimulation with LPS. Production of IL-6 and TNF- α in culture supernatants was increased in WT and $J\alpha 18^{-/-}$ mice, whereas it was suppressed in HMNC cultures of $CD1d^{-/-}$ mice (Figure 3D, E). Thus, lipid-induced inflammation may be suppressed in the absence of $CD1d$ and $CD1d$ -restricted NKT cells.

Next, we similarly analyzed adipose tissues that also contribute to the difference in BW in the three groups of mice. $CD1d^{-/-}$ mice had apparently smaller adipocytes in adipose tissue even on HFD, whereas enlarged adipocytes occupied the entire tissue in WT and $J\alpha 18^{-/-}$ mice (Figure 4A). Of note, boundaries of adipocytes formed almost straight lines due to the close packing of adipocytes in WT mice (Figure 4A left panel). To compare adipocyte size in WAT of each group of mice, the circumference of

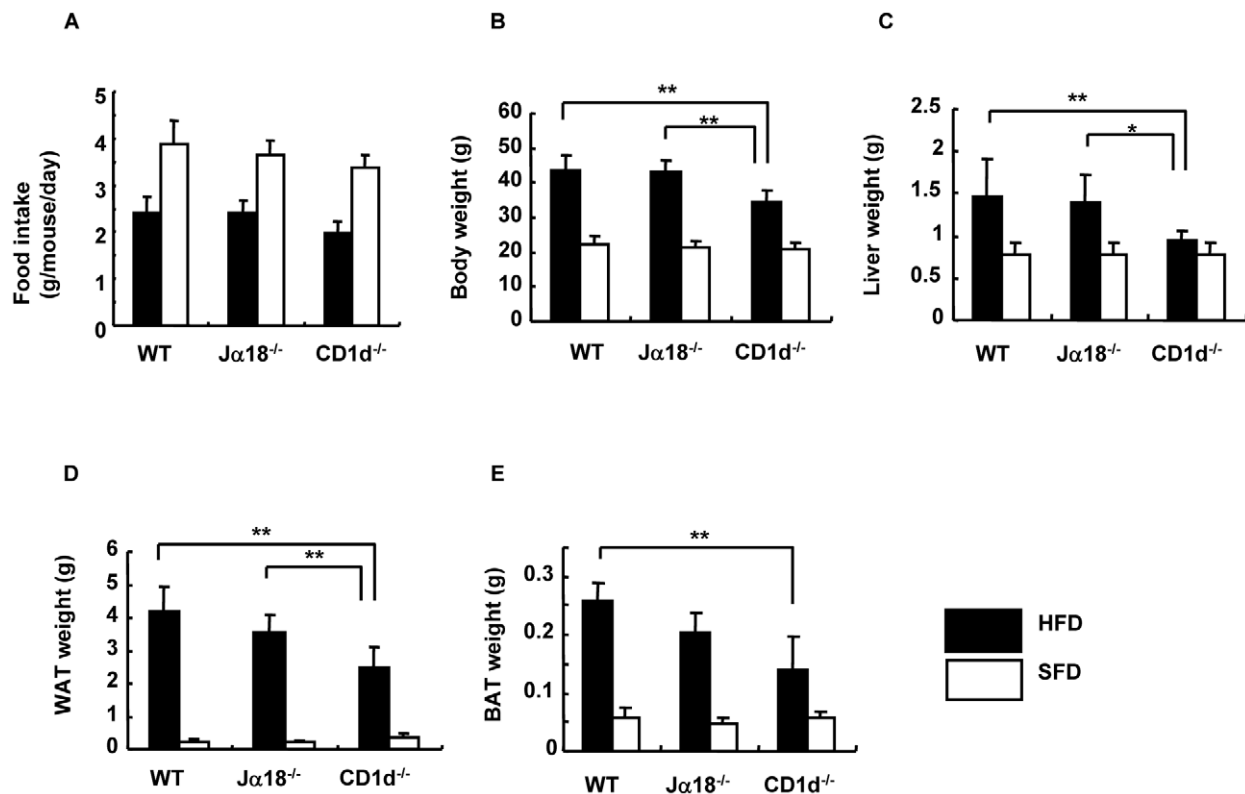


Figure 2. Physiological parameters of three strains of female mice fed an SFD or an HFD. (A) Food intake of mice in each group (g/mouse/day). (B–E) The body, liver, perigonadal adipose tissue (WAT), and brown adipose tissue (BAT) were weighed after 18 wk of feeding at 26 wk of age ($n = 3-7$ in each group). Representative data of three similar experiments are shown. The results are expressed as mean $\pm$ s.d. Statistical analysis was performed according to the Tukey-Kramer test. * $p < 0.05$, ** $p < 0.01$. doi:10.1371/journal.pone.0030568.g002

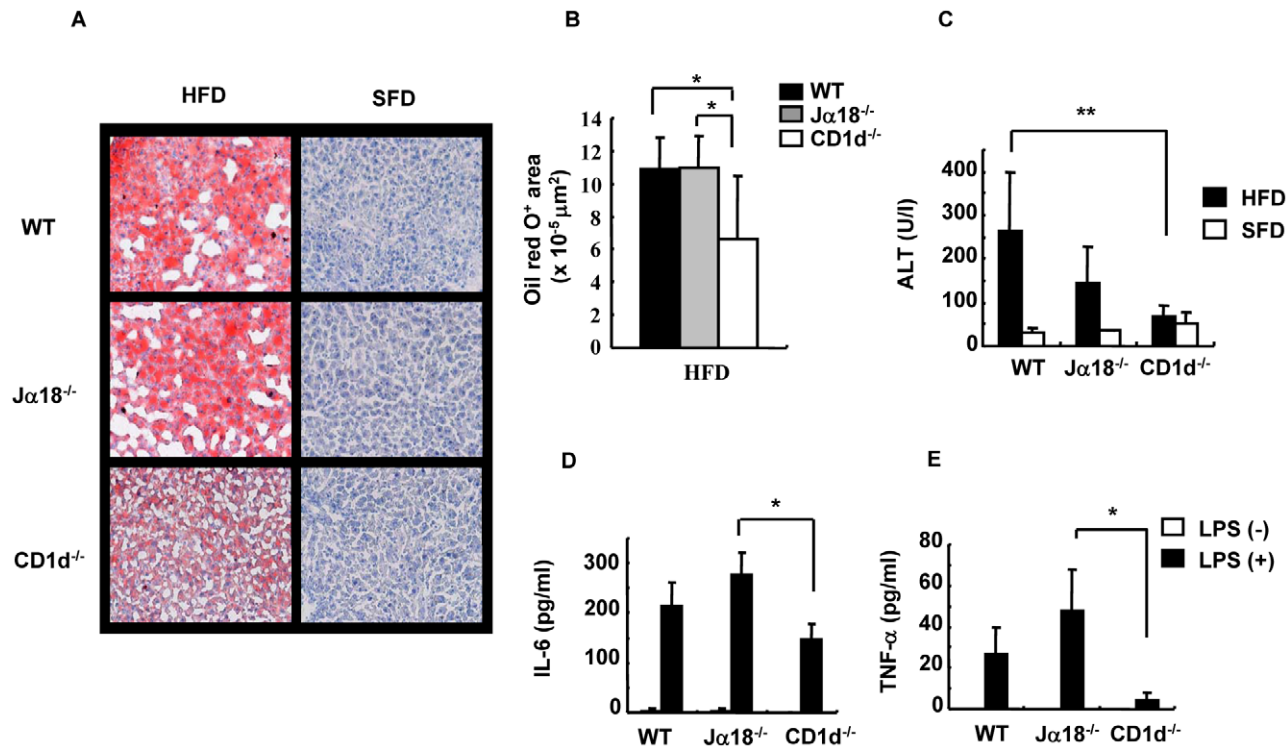


Figure 3. Histology and cytokine production in the liver. (A) A liver section was stained with ORO (frozen section) in SFD- and HFD-fed mice. (B) Red-stained lipid droplets in liver sections of HFD-fed mice were quantified with image analysis software. (C) Serum levels of ALT were quantified with the Drychem system. (D, E) The production of cytokines from HMNC of HFD-fed mice stimulated with LPS for 20 h ($n=3-4$ female mice in each group). The results are expressed as mean $\pm$ s.d. Statistical analysis was performed according to the Tukey-Kramer test. * $p<0.05$, ** $p<0.01$. doi:10.1371/journal.pone.0030568.g003

each adipocyte was morphometrically compared (Figure 4B). The adipocytes from CD1d^{-/-} mice had significantly smaller circumference than those from WT and Jα18^{-/-} mice.

When leptin is overexpressed, mice demonstrate a lean phenotype [23]. To examine whether CD1d^{-/-} expressed a higher level of leptin in sera than WT and Jα18^{-/-} mice, serum leptin levels were quantified. However, CD1d^{-/-} showed the lowest level among the three strains of mice, which was proportional to the volume of WAT (Figure 4C).

Analysis of infiltrated cellular components in liver of mice fed an HFD

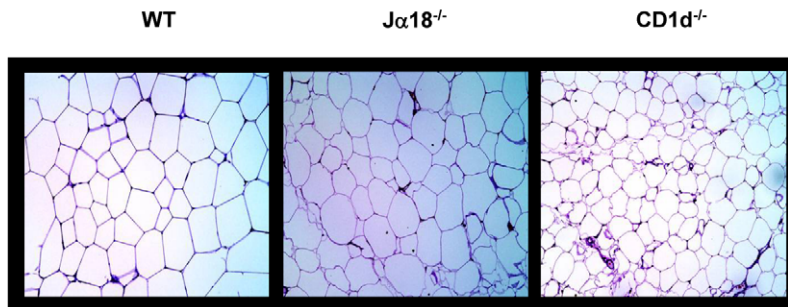
Our findings thus far showed that adiposity and hepatosteatosis were similar between WT and Jα18^{-/-} but were minimal in CD1d^{-/-} mice. The difference between Jα18^{-/-} and CD1d^{-/-} mice is solely the absence of type II NKT cells selected by CD1d in CD1d^{-/-} but not Jα18^{-/-} mice. To study the relationship between type I (iNKT) and type II NKT cells, we analyzed NKT cell subsets in liver and adipose tissue of each strain. As shown in our previous report with the short term feeding of HFD [18], the proportion of NKT cells in liver decreased in WT mice fed an HFD compared to mice on an SFD when the HMNC fraction was stained with a combination of either α-GalCer-loaded CD1d-dimer and anti-TCRβ mAb (iNKT cells) (Figure 5A-a, b; 22.8% → 6.8%) or anti-NK1.1 and TCRβ mAb (total NKT cells, including NKT-like cells) (Figure 5A-c, d; 27.9% → 6.6%). To detect the subset of type II NKT cells and NKT-like cells in the iNKT cell-deficient strains, the latter combination was employed. Although a significant decrease was demonstrated in NKT cells in WT mice (Figure 5B-a), there was no significant difference in the proportion of total NKT

cells in Jα18^{-/-} and CD1d^{-/-} mice fed an HFD (Figure 5B-a), but Jα18^{-/-} mice appeared to exhibit a slight decrease in the prevalence of these cells on an HFD. To examine the residual NKT-like cells in the livers of Jα18^{-/-} and CD1d^{-/-} mice, staining with a combination of anti-NK1.1 and anti-TCRβ mAb was employed. As demonstrated in the FACS profiles, total NKT cells in WT mice were significantly decreased after HFD feeding (Figure 5B-a). As for the subset of residual NKT cells, the subset that expressed neither CD4 nor CD8 (CD4⁻CD8⁻ double negative; DN) did not differ regardless of the feeding regimen used or the mouse strains analyzed. The relative prevalence of the CD4⁺ subset was as follows: WT > Jα18^{-/-} > CD1d^{-/-} mice, and the prevalence of the CD4⁻CD8⁺ subset had an inverse relationship with CD8⁺ cells (WT < Jα18^{-/-} < CD1d^{-/-} mice) (Figure 5B-b, c). Although the total number of liver NKT cells was reduced, the residual population was mainly a CD4⁺ subset in WT mice and the remaining cells were a CD4⁻CD8⁻ subset (Figure 5B-d). The CD8⁺ subset was very minimal in WT mice. On the other hand, CD1d^{-/-} mice had residual numbers of hepatic NK1.1⁺TCRβ⁺, NKT-like cells, and about 40~50% of the population was CD8⁺. There was no significant difference in the proportion of regulatory T (T_{reg}) cells in HMNC among the three strains of mice (data not shown). It should be noted that NKT cells in the liver of WT mice fed with HFD were significantly reduced. This effect of lipid excess was evident as early as day 1 of HFD feeding (data not shown).

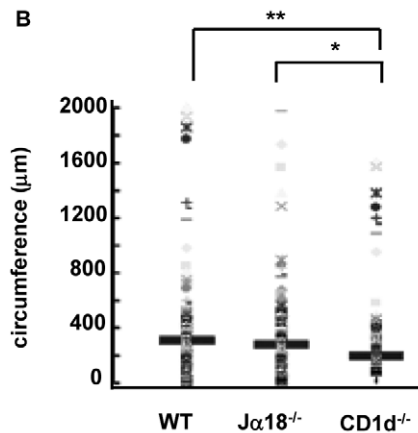
Analysis of infiltrated cellular components in adipose tissue of mice fed an HFD

The SVF in adipose tissue contains innate lymphocytes and Mφ even under normal conditions. In adipose tissue in obese mice, we

A



B



C

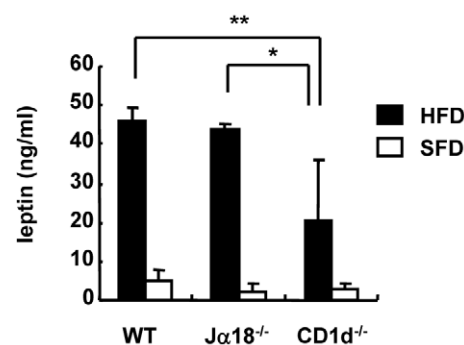


Figure 4. Histology of perigonadal adipose tissue and serum leptin level. (A, B) A paraffin section of perigonadal adipose tissue was stained with HE, and the circumference of adipocytes was morphometrically analyzed (10×10 , 300–400 adipocytes measured) in HFD mice. (C) The serum leptin level was quantified by ELISA ($n=3-4$ female mice in each group). Representative data of two similar experiments are shown. The results are expressed as mean $\pm$ s.d. Statistical analysis was performed according to the Tukey-Kramer test. * $p<0.05$, ** $p<0.01$. doi:10.1371/journal.pone.0030568.g004

readily detected increased numbers of mononuclear cells than in lean mice in spaces surrounded by adipocytes (data not shown). First, we analyzed NK1.1⁺TCR β ⁺ (NKT) cells and the CD4/8 subsets. Since the CD1d-restricted NKT cells are absent in CD1d^{-/-} mice, we used NK1.1⁺TCR β ⁺ staining to detect residual NKT-like cells instead of double staining with a combination of α -GalCer-loaded CD1d-dimers and anti-TCR β mAb. In WT mice, the proportion of iNKT cells in adipose tissue was not significantly different (Figure 6A-a, b; 1.5%–>1.2%), whereas there were significantly more NK1.1⁺TCR β ⁺ cells in mice fed an HFD compared to those on SFD, in the same experimental setting (Figure 6A-c, d; 2.2%–>5.2%). The percentage of NKT cells increased in WT and Jα18^{-/-} cells when the NKT cells of mice on HFD were compared with those on SFD (WT; $p<0.05$; Jα18^{-/-}; $p<0.01$), whereas such an increase was not observed in CD1d^{-/-} mice (Figure 6B-a). Since CD1d^{-/-} mice lack CD4⁺CD1d-restricted NKT cells, there were significant differences in the percentages (Figure 6B-b) (WT; $p<0.05$; Jα18^{-/-}; $p<0.01$) and actual cell numbers (WT, Jα18^{-/-}; $p<0.05$). Of note, the CD8⁺ subset was prominently increased in the adipose tissue of Jα18^{-/-} mice both in percentage (Figure 6B-c) and in number. The CD4⁺ subset of NKT cells appeared to be significantly decreased in Jα18^{-/-} mice fed an HFD, most likely due to the relative abundance of the CD4⁺ subset (Figure 6B-d). Similar results were obtained when analyzing actual cell numbers (WT>Jα18^{-/-}>CD1d^{-/-}) (data not shown).

We analyzed and compared the proportion of NKT cells at early and late phases (18-wk) of HFD feeding. In WT mice, the

number of α -GalCer-CD1d-dimer⁺ cells and NK1.1⁺TCR β ⁺ cells in liver were increased at 1 wk of HFD-feeding, and numbers tended to decrease thereafter (Figure S3A). On the other hand, these cells, especially the NK1.1⁺TCR β ⁺ population, were gradually increased in adipose tissue (Figure S3B). The increase in NK1.1⁺TCR β ⁺ cells (percentage in SVF) in adipose tissue correlated with BW in WT and Jα18^{-/-} mice, whereas no correlation was found in CD1d^{-/-} mice (Figure 6C). A similar correlation was obtained between numbers of NK1.1⁺TCR β ⁺ cells (cell number/g adipose tissue) and BW (data not shown).

Analysis of Mφ in adipose tissue and cytokine production upon LPS stimulation

Mφ are another major cellular subset in SVF and may affect the metabolism of adipose tissue. Therefore, SVF preparations from the three strains of mouse were stained with F4/80 and CD11b (Figure 7A). The three strains had a similar pattern of F4/80⁺/CD11b⁺ staining. Although we anticipated that CD1d^{-/-} mice had fewer Mφ than Jα18^{-/-} mice, the percentage of F4/80⁺/CD11b⁺ cells in CD1d^{-/-} mice was higher than that of Jα18^{-/-} mice (Figure 7B-a). However, when the F4/80⁺ population was divided into two subpopulations according to the MFI of F4/80 staining pattern, i. e., Population 1 (P1: F4/80^{high}) and Population 2 (P2: F4/80^{low}) cells (both CD11b⁺), WT mice had higher frequencies of P1 than Jα18^{-/-} mice (Figure 7B-b), whereas CD1d^{-/-} mice had an increased frequency of P2 than WT and Jα18^{-/-} mice (Figure 7B-c). Analyses of actual cell numbers

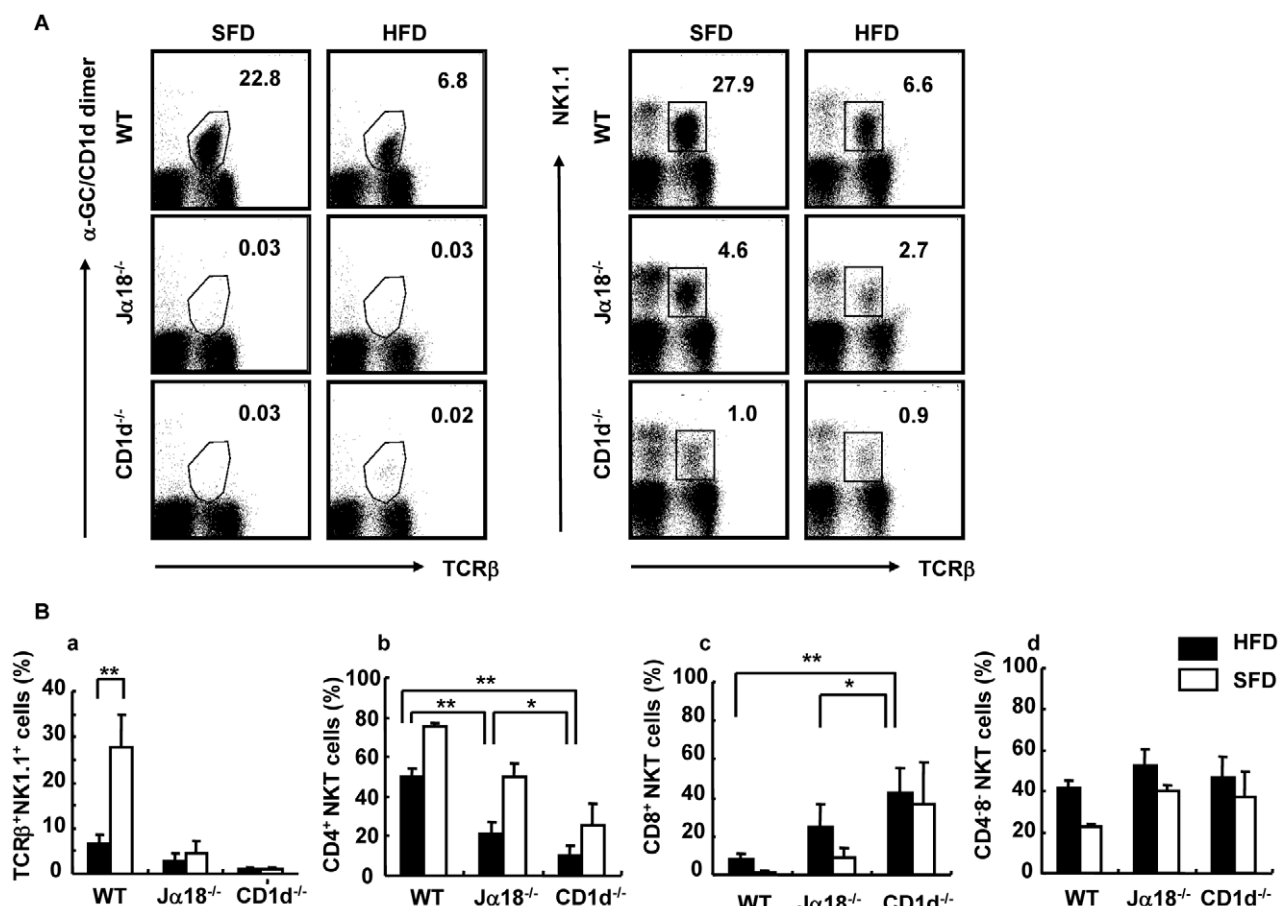


Figure 5. Flow cytometric analyses of HMNC in mice fed an SFD or an HFD. (A) A representative flow cytometric dot-plot defining the population of α -GalCer/CD1d dimer⁺TCR β ⁺ and NK1.1⁺TCR β ⁺ cells in the liver of SFD-fed mice (a, c) and HFD-fed mice (b, d). (B) The proportion of NK1.1⁺TCR β ⁺ cells (a), CD4⁺ (b), CD8⁺ (c), and CD4⁺CD8⁻ (d) NKT cells ($n=3-6$ female mice in each group). Representative data of three similar experiments are shown. The results are expressed as mean $\pm$ s.d. Statistical analysis was performed according to the Tukey-Kramer test. * $p<0.05$, ** $p<0.01$.

doi:10.1371/journal.pone.0030568.g005

demonstrated the same tendency among the three mouse strains (data not shown). Of note, a higher MFI of CD1d expression was detected in the cells of the P1 population compared with the P2 population, both in WT and Jα18^{-/-} mice (Figure 7C-a, b, c).

To examine functional differences in a specific Mφ population, total cells from the SVF fraction were stimulated with LPS for 20 h and cytokines were measured (Figure 8). IL-10 levels were significantly higher in SVF cultures from CD1d^{-/-} mice compared with WT mice (Figure 8A), and GM-CSF was significantly lower in Jα18^{-/-} and CD1d^{-/-} mice (Figure 8B). Notably, the production of TNF-α in the culture supernatant that could affect insulin resistance was not significantly different among the three strains (Figure 8C).

Glucose and insulin tolerance are ameliorated CD1d^{-/-} mice

Since the pattern of adiposity should reflect levels of glucose intolerance, IPGTT was performed in the three mouse strains (Figure 9). No difference was observed in fasting blood sugar levels and in elevation after *i.p.* administration of glucose over time in the three strains fed an SFD (Figure 9A, left). On the other hand, CD1d^{-/-} mice demonstrated the lowest fasting blood sugar level and a suppressed elevation profile after glucose administration among the three strains on an HFD (Figure 9A, right). Insulin

levels were significantly higher in WT and Jα18^{-/-} mice before and at 120 min after infusion than that obtained in the respective groups on an SFD (data not shown) and for CD1d^{-/-} mice on an HFD, suggesting that insulin resistance was present in those strains (Figure 9B). To further examine insulin resistance, an ITT was performed in HFD-fed mice that had been fasted for 3.5 h [6] (Figure 9C). Following injection of insulin, blood glucose levels fell most prominently in CD1d^{-/-} mice at 30 and 60 min after the injection (Figure 9C). The decrease in glucose level in WT mice injected with α -GalCer was less than that of mice injected with vehicle (Figure S1B), suggesting that insulin resistance was reproduced in mice when iNKT cells were activated [18], although there was no net increase in BW with the treatment. Thus, CD1d^{-/-} mice developed the least resistance to insulin in the absence of both iNKT and type II NKT cells.

To examine whether type II NKT cells alone could reproduce the pathophysiological status observed in Jα18^{-/-} mice, HMNC obtained from either Jα18^{-/-} or CD1d^{-/-} mice were intravenously transferred to CD1d^{-/-} recipient mice and weight gain was examined on an HFD. CD1d^{-/-} mice that received HMNC from Jα18^{-/-} mice showed a slight increase in BW and exhibited a less profound decrease in blood glucose levels in the ITT (Figure S4A, C). Although no significant difference was detected with the IPGTT (Figure S4B), CD1d^{-/-} mice that received HMNC from

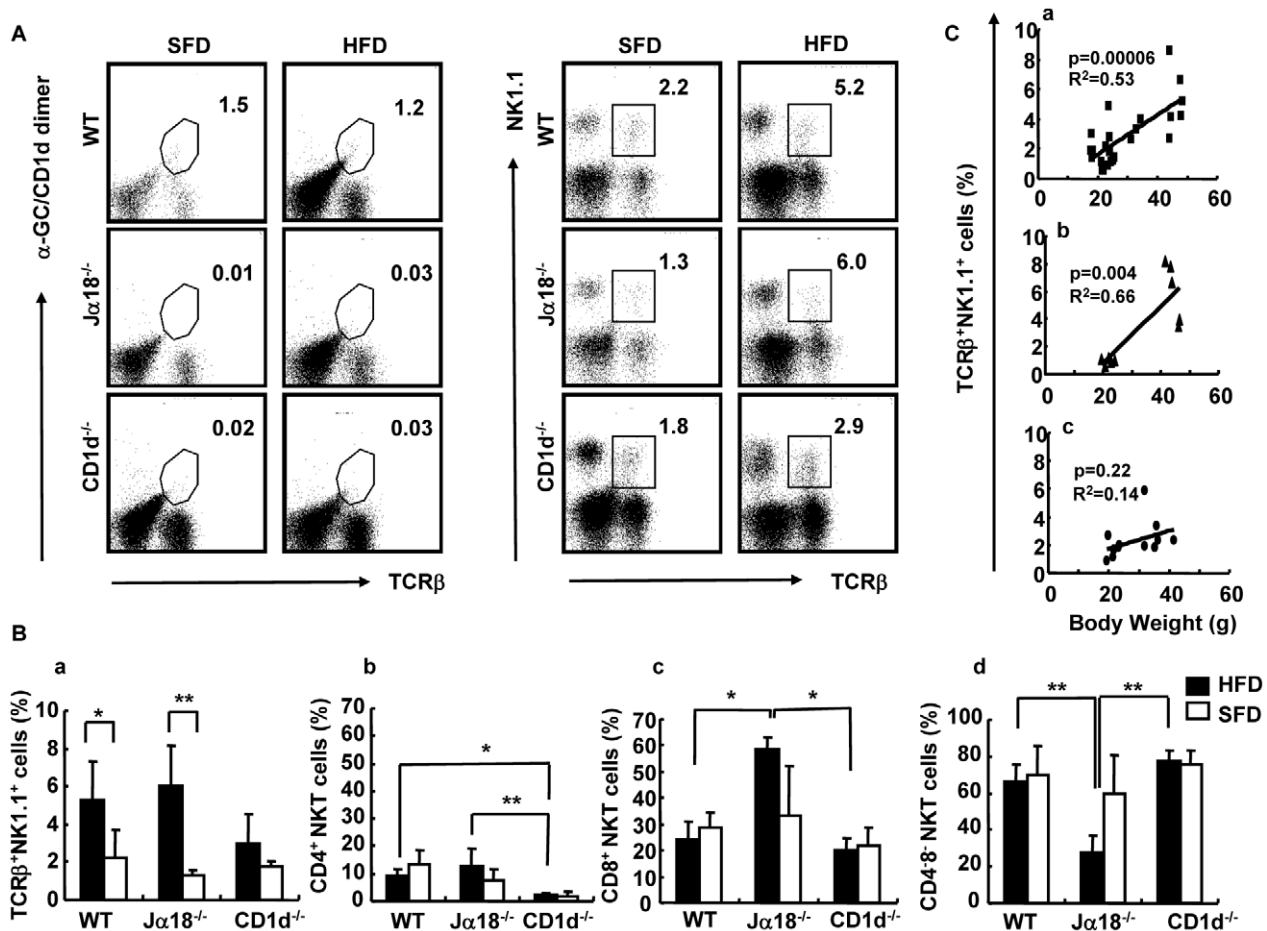


Figure 6. Flow cytometric analyses of NKT cells in adipose tissue in mice fed an SFD or an HFD. (A) A representative flow cytometric dot-plot defining the population of α -GalCer/CD1d dimer⁺TCRβ⁺ and NK1.1⁺TCRβ⁺ cells in adipose tissue of SFD-fed mice (a, c) and HFD-fed mice (b, d). (B) The proportion of each subset in SVF. NK1.1⁺TCRβ⁺ cells (a), CD4⁺ (b), CD8⁺ (c), and CD4⁺CD8⁻ (d) NKT cells (n = 3–6 female mice in each group). Representative data of three similar experiments are shown. The results are expressed as mean $\pm$ s.d. Statistical analysis was performed according to the Tukey-Kramer test. * $p < 0.05$, ** $p < 0.01$. (C) The correlation between BW and the percentage of NKT cells in SVF by Pearson's correlation coefficient (R^2) test. WT (a), $P = 0.00006$; Jα18^{-/-} (b), $P = 0.004$; and CD1d^{-/-} (c), $P = 0.2$. Values with $P < 0.05$ were considered statistically significant. doi:10.1371/journal.pone.0030568.g006

Jα18^{-/-} mice showed higher insulin resistance than CD1d^{-/-} mice that received HMNC from CD1d^{-/-} mice. The percentage of NKT cells (NK1.1⁺TCRβ⁺) and CD4⁺ NKT cells was similar in the liver and adipose tissue of CD1d^{-/-} hosts (Figure S4D-a, b). However, CD8⁺ NK1.1⁺ T cells in adipose tissue were significantly increased in mice transferred with HMNC from Jα18^{-/-} mice (Figure S4D-c), similarly to what we observed in Jα18^{-/-} mice fed an HFD.

Discussion

Adipocytes, Mφ and T cells are now thought to constitute a functional triad to promote obesity and obesity-associated disorders [1–3,6–8]. Recently, the involvement of eosinophils and B cells has also been demonstrated [24,25], implying that many other immune cells participate in the physiologic and pathologic processes of adipocyte hypertrophy. This may be a natural process since adipose tissue is thought to be an ancestor of lymphoid organs [5,14].

In the present study, we demonstrated that an innate lymphocyte of the T cell lineage, the CD1d-restricted NKT cell, also has an active role in the development of obesity. When fed an

HFD (CLEA Japan), CD1d^{-/-} mice gained the least weight compared to Jα18^{-/-} and WT mice. This difference was demonstrated when mainstream CD8⁺ T cells were primarily unaffected. Furthermore, similar results were obtained with different types of HFD (Harlan Teklad [26] and our own original preparation [18]), suggesting that the suppression of weight gain was solely dependent on the deficiency of CD1d-restricted NKT cells but not on the specific source of fat, i.e. fatty acid composition.

Since Jα18^{-/-} mice gained comparable BW as WT mice, the residual CD1d-restricted, type II NKT cells appeared to contribute to the accumulation of visceral fat in the body. In HFD-fed WT and Jα18^{-/-} mice, the number of NK1.1⁺TCRβ⁺ cells increased along with the hypertrophic change of abdominal WAT (Figure 4, 6), whereas the α -GalCer-CD1d dimer⁺ population did not change in adipose tissue of WT mice, suggesting that the type II NKT cells were the major population affecting adipose tissue metabolism. Accordingly, the transfer of NKT cells from Jα18^{-/-} mice to CD1d^{-/-} mice resulted in weight gain and significant increase in insulin resistance (Figure S4). Both the percentage and the cell number of NK1.1⁺TCRβ⁺ cells in adipose tissue correlated with the BW weights in Jα18^{-/-}

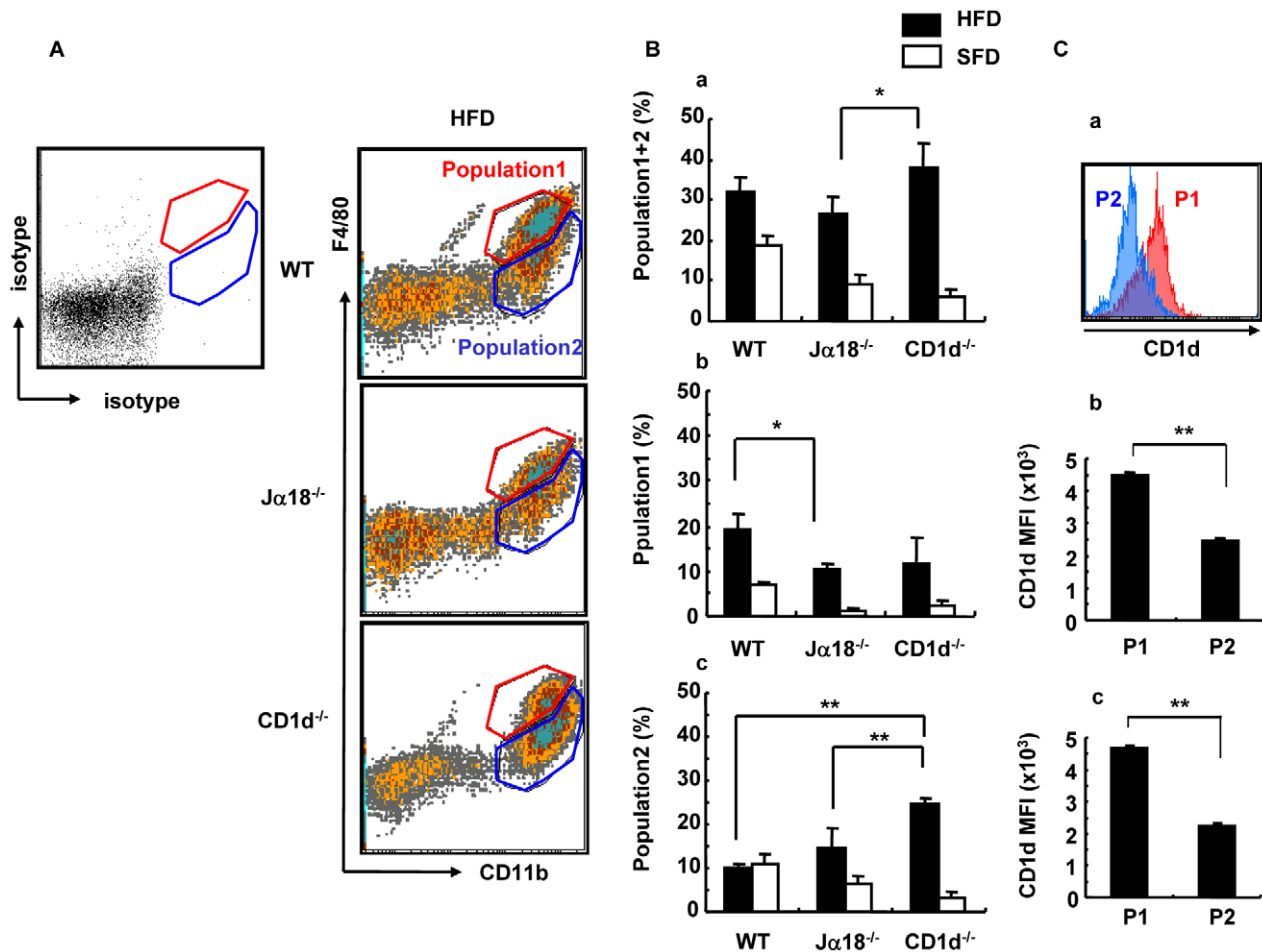


Figure 7. Flow cytometric analyses of infiltrated Mφ in perigonadal adipose tissue. (A) A representative flow cytometric dot-plot defining the F4/80⁺CD11b⁺ population in perigonadal adipose tissue in HFD-fed mice. F4/80^{hi} (Population 1; red gate) and F4/80^{low} (Population 2; blue gate) are set to a staining with isotype control mAb. (B) The percentage of cells of CD11b⁺ (total) (a), Population 1 (b) and Population 2 (c). (C) CD1d expression by the Population 1 and Population 2 (a), MFI in WT (b) and Jα18^{-/-} (c) mice fed an HFD (n = 3–4 male mice in each group). The results are expressed as mean ± s.d. Statistical analysis was performed according to the Tukey-Kramer test. **p* < 0.05, ***p* < 0.01. doi:10.1371/journal.pone.0030568.g007

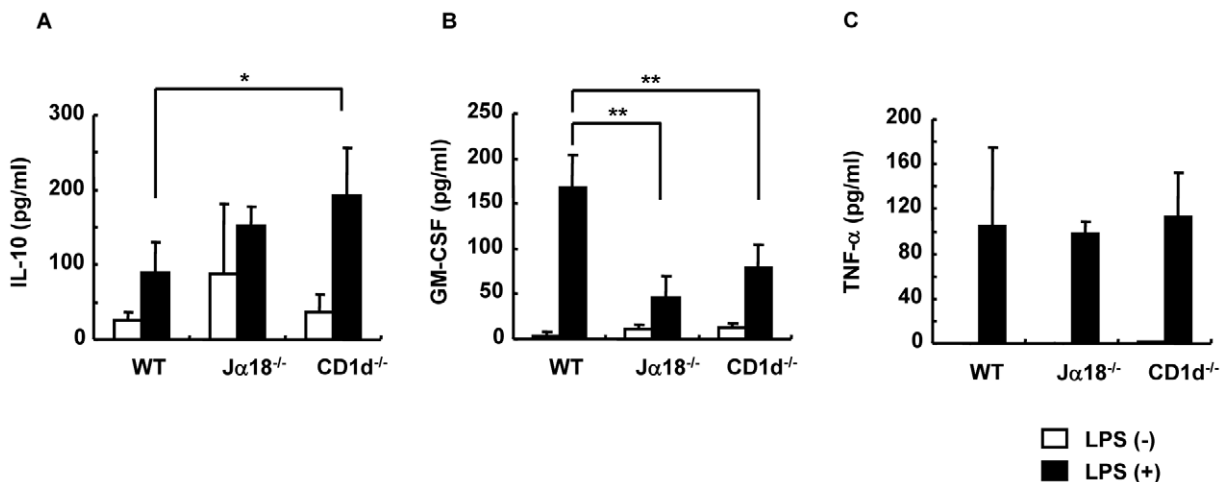


Figure 8. Cytokine production by Mφ in perigonadal adipose tissue upon stimulation with LPS. (A–C) IL-10, GM-CSF and TNF-α concentrations after stimulation of Mφ in perigonadal adipose tissue with LPS (1 μg/ml) for 20 h (n = 3–4 male mice in each group). Representative data of three similar experiments are shown. The results are expressed as mean ± s.d. Statistical analysis was performed according to the Tukey-Kramer test. **p* < 0.05, ***p* < 0.01. doi:10.1371/journal.pone.0030568.g008

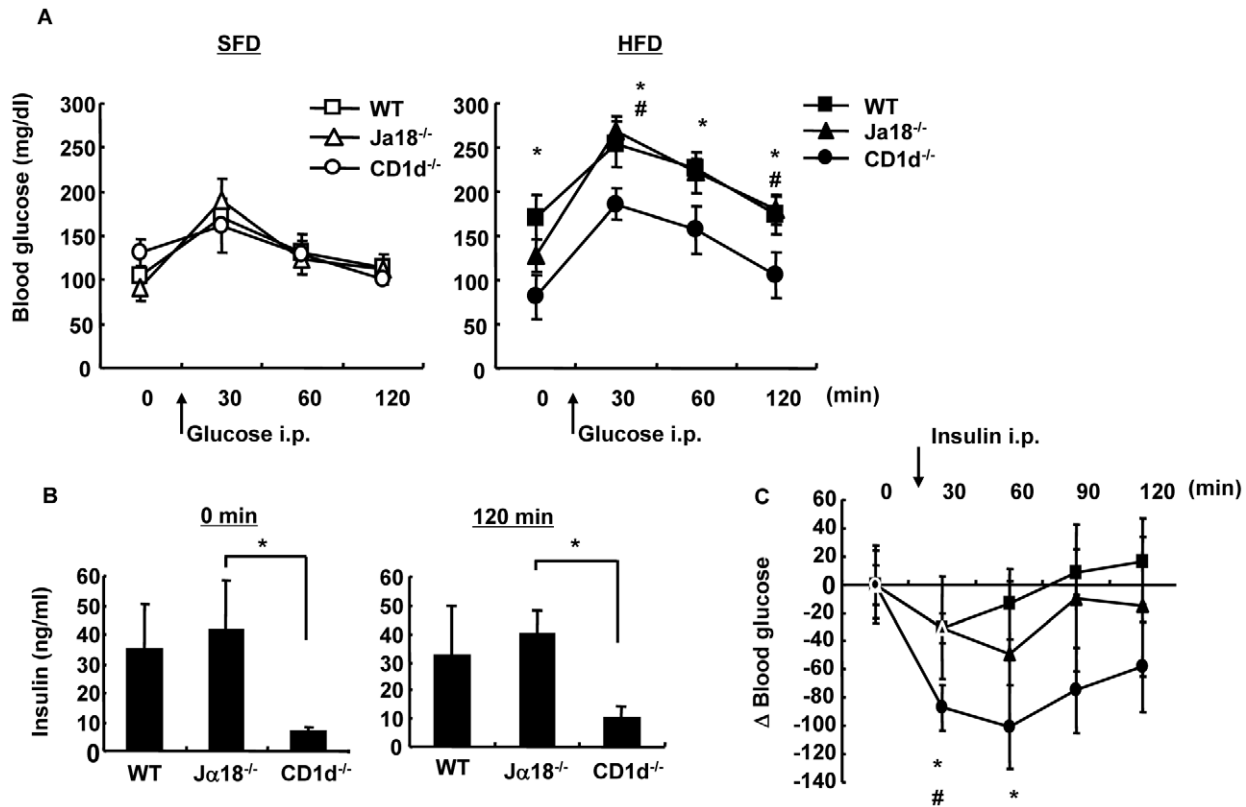


Figure 9. Glucose and insulin tolerance. (A) (IPGTT). Glucose (1 g/kg BW) was administered *i.p.* to female mice fed an SFD or an HFD. (B) The serum level of insulin 0 min and 120 min after glucose administration (*i.p.*) to HFD-fed mice was quantified by ELISA. (C) ITT. Insulin (0.75 U/kg BW) was administered *i.p.* to HFD-fed mice ($n = 3-4$ female mice in each group). Representative data of three similar experiments are shown. The results are expressed as mean $\pm$ s.d. Statistical analysis was performed according to the Tukey-Kramer test. * $p < 0.05$ (WT versus CD1d^{-/-}); # $p < 0.05$ (Ja18^{-/-} versus CD1d^{-/-}). doi:10.1371/journal.pone.0030568.g009

as well as in WT mice, whereas no correlation was found in CD1d^{-/-} mice (Figure 6C). When the NK1.1⁺TCR β ⁺ cells in adipose tissue were analyzed, we were unable to detect these cells with sulfatide-loaded CD1d-multimers that are specific for a subset of type II NKT cells [27] (data not shown). Of note, these cells exhibited a biased repertoire in specific V β s in HFD-fed WT mice (data not shown), although the significance of this finding remains unclear. It should also be noted that α -GalCer administration did not boost weight gain in WT mice fed an HFD, although insulin action was blunted with α -GalCer administration. This result indicated that insulin resistance but not adiposity could be reproduced through the activation of type I NKT cells (Figure S1A).

Nishimura et al demonstrated that CD8⁺ T cells are important for recruitment of M ϕ and chronic inflammation in adipose tissue, which was proposed to propagate inflammation [6]. In our experiments, the CD8⁺ subset of NK1.1⁺ T cells, a rather rare subset [28], increased in adipose tissue in WT and Ja18^{-/-} mice. Although we have not analyzed TCR V β chain usage, Nishimura described them as V β 7 and V β 10b [6]. Since V β 7 is one of the preferred V β chains of NKT cells [29], it is possible that these cells or a portion of these cells include NKT cells. Very recently, Mantell et al. reported that no protection against metabolic abnormalities was noted in CD1d^{-/-} mice fed an HFD [30]. They demonstrated no significant differences between WT and CD1d^{-/-} mice in physiologic, metabolic, and inflammatory parameters. Nevertheless, since there was an early and reversible

decrease in liver NKT cells, mice appeared to respond to an HFD. Specific reasons for these divergent findings will need to be explored in future studies. One possibility might be the colonization of mice from different animal facilities with different flora. An important role of intestinal flora in the process of obesity has emerged [31], and the differential dominance of certain species of microbes has been demonstrated in obese or anorexic human individuals [32]. Furthermore, in toll-like receptor 5 knockout (TLR5^{-/-}) mice, metabolic syndrome was induced by altered gut microbiota with hyperphagic traits [33]. Moreover, these metabolic changes could be conferred to WT, germ-free mice upon transfer of the gut microbiota derived from TLR5^{-/-} mice. Although adaptive immunity is not directly related to the process of obesity in TLR5^{-/-} mice, it should be noted that the development of adiposity could be controlled by balancing the complex microbiota. Since bacterial colonization in the intestine is regulated by a CD1d-dependent, NKT cell-mediated mechanism [34], it is possible that NKT cell deficiency may alter the intestinal flora, and thus influence either an adipogenic or anti-adipogenic host milieu.

The recruitment of M ϕ towards visceral fat, also known as adipose tissue M ϕ (ATM ϕ ; CD11b⁺F4/80⁺), has also been thought to be a hallmark of obesity [1,2]. The F4/80⁺ fraction can be subdivided into the F4/80^{high} and F4/80^{low} populations [35]. In the present study, ATM ϕ of WT and Ja18^{-/-} mice had an increased F4/80^{high} population, whereas CD1d^{-/-} mice predominantly had an increased percentage and number of the

F4/80^{low} population. The F4/80^{high} population exhibits higher expression of TLR-4, TNF- α , and PPAR γ than the F4/80^{low} population. Therefore, this subset represents pro-inflammatory M ϕ (M1) and accumulates in adipose tissue of obese mice [36]. On the other hand, the F4/80^{low} population exhibits higher IL-4 expression and represents anti-inflammatory M ϕ [36]. Furthermore, elimination of the F4/80^{high} population is associated with improved insulin sensitivity [37]. Our studies suggest that the predominance of F4/80^{high} ATM ϕ is likely dependent on the presence of type II NKT cells, since it was observed in WT and J α 18^{-/-} mice. Consistently, SVF that contained more F4/80^{low} M ϕ from CD1d^{-/-} mice produced more IL-10 upon stimulation with LPS (Figure 8). The anti-inflammatory milieu induced by M ϕ polarization may ameliorate the development of obesity in CD1d^{-/-} mice. It should also be noted that F4/80^{high} ATM ϕ expressed higher levels of CD1d than F4/80^{low} M ϕ in WT and J α 18^{-/-} mice, suggesting that NKT cells might better polarize these cells to cause adipose tissue inflammation. Alternatively, F4/80^{high} ATM ϕ might express higher levels of CD1d as a result of the interaction with type II NKT cells.

The present study suggests that type II NKT cells may serve as a potential target for controlling the volume of visceral fat and insulin resistance. A possible approach may be the administration of a CD1d-binding ligand to modulate NKT cell function. Indeed, Ilan's group has demonstrated that the administration of glucocerebroside (β -glucosylceramide; β -GlcCer) or the direct transfer of NKT cells ameliorated metabolic aberration and steatohepatitis of leptin-deficient *ob/ob* mice [38,39]. They demonstrated that the therapeutic effects of β -GlcCer were associated with a Th2-shift (decreased IFN- γ and increased IL-10) and the redistribution of NKT cells in the liver to the periphery. Furthermore, β -GlcCer modulated the lipid rafts of NKT cells, thus altering the expression of molecules of cellular signaling, such as flotillin-2, Lck, and STAT1 [40]. Although the mode of action appears to be dependent on iNKT cells, it remains possible that these CD1d-binding non-stimulatory ligands may simply function as antagonists for all CD1d-restricted NKT cells.

Of special interest, Lynch et al. demonstrated that NKT cells and CD1d⁺ cells are abundantly detected in the omentum tissue in humans [41]. Half of the NKT (CD3⁺CD56⁺) cells stained with the 6B11 mAb [42], indicating that they were iNKT cells, but the remaining cells likely are type II NKT cells. They also reported a net decrease of iNKT cells in severely obese individuals with a relative increase of CD56⁺ T cells (healthy, 49.9%; obese, 61.9%) and discussed that the relative immunocompromised status in obesity due to the decreases in iNKT cells might be related to sensitivity to infections [40]. On the other hand, the omentum may be a source of not only iNKT cells but also type II NKT cells that are capable of inducing adiposity in the abdominal cavity.

In conclusion, we showed that deficiency of CD1d-restricted NKT cells suppressed the development of obesity, likely because inflammation of adipose tissue and liver were ameliorated and reduced levels of insulin resistance were induced. NKT cells, especially the type II subset, may be the initial subset of T cells that recognizes and responds to lipid excess. Thus, CD1d-restricted, type II NKT cells may be a novel target for therapeutic

intervention in the metabolic syndrome. Searching for regulatory ligands and novel ways of regulation that include the selective elimination of pathogenic type II NKT cells might permit the control of not only metabolic syndrome but also various diseases related to lipid inflammation mediated by NKT cells [43].

Supporting Information

Figure S1 BW and insulin resistance of mice administered α -GalCer. (A) BW was determined weekly in WT mice given either vehicle or α -GalCer (0.1 μ g/g BW). (B) ITT was performed for WT mice injected with vehicle or α -GalCer (n = 5 female mice in each group). Representative data of two similar experiments are shown. The results are expressed as mean $\pm$ s.d. Statistical analysis was performed according to the Student's *t*-test. **p* < 0.05.

(TIFF)

Figure S2 Serum T-cholesterol, HDL-cholesterol, and TG levels. (A–C) Serum T-cholesterol, HDL-cholesterol, and TG level after an 18 wk feeding and a 16 h fasting period (n = 6–8 female mice in each group). Representative data of three similar experiments are shown. The results are expressed as mean $\pm$ s.d. Statistical analysis was performed according to the Tukey-Kramer test. **p* < 0.05, ***p* < 0.01.

(TIFF)

Figure S3 Dynamics of NKT cells in liver and adipose tissue during the early phase of feeding. The cell number of iNKT cells and NK1.1⁺TCR β ⁺ cells in liver (A) and in adipose tissue (B) at 1, 2, 3, and 18 wk of HFD-feeding (n = 3–6 female mice in each group). Representative data of two similar experiments are shown. The results are expressed as mean $\pm$ s.d. (TIFF)

Figure S4 Effects of adoptive transfer of J α 18^{-/-} HMNC to CD1d^{-/-} mice. (A) CD1d^{-/-} mice received HMNC (1 $\times$ 10⁶) from J α 18^{-/-} mice since 8 wk of age on an HFD. BW was determined weekly. (B, C) IPGTT and ITT were performed at 14 wk of HFD feeding. (D) The number of NK1.1⁺TCR β ⁺ cells (a) and the CD4/8 proportion in liver and adipose tissue (b, c) were analyzed by flow cytometry (n = 5 male mice in each group). Representative data of two similar experiments are shown. The results are expressed as mean $\pm$ s.d. Statistical analysis was performed according to Student's *t*-test. **p* < 0.05.

(TIFF)

Acknowledgments

We would like to thank Dr. Luc Van Kaer for sharing preliminary data and critically reading the manuscript. We are also grateful to Kirin Brewery Co., Ltd for kindly providing α -GalCer.

Author Contributions

Conceived and designed the experiments: YA MS NI HT KO KI. Performed the experiments: MS YA CSC HO NI KI KE. Analyzed the data: MS YA SF NH KE. Contributed reagents/materials/analysis tools: TN MT KE. Wrote the paper: MS KI.

References

1. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, et al. (2003) Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest* 112: 1796–1808.
2. Xu H, Barnes GT, Yang Q, Tan G, Yang D, et al. (2003) Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J Clin Invest* 112: 1821–30.
3. Kintscher U, Hartge M, Hess K, Forst-Ludwig A, Clemenz M, et al. (2008) T-lymphocyte infiltration in visceral adipose tissue: a primary event in adipose tissue inflammation and the development of obesity-mediated insulin resistance. *Arterioscler Thromb Vasc Biol* 28: 1304–10.
4. Waki H, Tontonoz P (2007) Endocrine functions of adipose tissue. *Annu Rev Pathol Mech Dis* 2: 31–56.

5. Hotamisligil GS (2006) Inflammation and metabolic disorders. *Nature* 444: 860–867.
6. Nishimura S, Manabe I, Nagasaki M, Eto K, Yamashita H, et al. (2009) CD8⁺ T effector T cells contribute to macrophage recruitment and adipose tissue inflammation in obesity. *Nat Med* 15: 914–20.
7. Winer S, Chan Y, Palster G, Truong D, Tsui H, et al. (2009) Normalization of obesity-associated insulin resistance through immunotherapy. *Nat Med* 15: 921–9.
8. Feuerer M, Herrero L, Cipolletta D, Naaz A, Wong J, et al. (2009) Lean, but not obese, fat is enriched for a unique population of regulatory T cells that affect metabolic parameters. *Nat Med* 15: 930–9.
9. Van Kaer L (2007) NKT cells: T lymphocytes with innate effector functions. *Curr Opin Immunol* 19: 354–364.
10. Kawano T, Cui J, Koezuka Y, Taura I, Kaneko Y, et al. (1997) CD1d-restricted and TCR-mediated activation of V α 14 NKT cells by glycosylceramides. *Science* 278: 1626–9.
11. Matsuda JL, Mallevaey T, Scott-Browne J, Gapin L (2008) CD1d-restricted iNKT cells, the ‘Swiss-Army knife’ of the immune system. *Curr Opin Immunol* 20: 358–68.
12. Gumperz JE, Roy C, Makowska A, Lum D, Sugita M, et al. (2000) Murine CD1d-restricted T cell recognition of cellular lipids. *Immunity* 12: 211–21.
13. Exley MA, Koziel MJ (2004) To be or not to be NKT: Natural killer T cells in the liver. *Hepatology* 40: 1033–40.
14. Caspar-Bauguil S, Cousin B, Galinier A, Segafredo C, Nibbelink M, et al. (2005) Adipose tissues as an ancestral immune organ: Site-specific change in obesity. *FEBS Lett* 579: 3487–92.
15. Tupin E, Nicoletti A, Elhage R, Rudling M, Ljunggren HG, et al. (2004) CD1d-dependent activation of NKT cells aggravates atherosclerosis. *J Exp Med* 199: 417–422.
16. Nakai Y, Iwabuchi K, Fujii S, Ishimori N, Dashtsoodil N, et al. (2004) Natural killer T cells accelerate atherogenesis in mice. *Blood* 104: 2051–2059.
17. Major AS, Wilson MT, McCaleb JL, Ru Su Y, Stanic AK, et al. (2004) Quantitative and qualitative differences in proatherogenic NKT cells in apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol* 24: 2351–2357.
18. Ohmura K, Ishimori N, Ohmura Y, Tokuhara S, Nozawa A, et al. (2010) Natural killer T cells are involved in adipose tissues inflammation and glucose intolerance in diet-induced obese mice. *Arterioscler Thromb Vasc Biol* 30: 193–199.
19. Hansen TH, Huang S, Arnold PL, Fremont DH (2007) Patterns of nonclassical MHC antigen presentation. *Nat Immunol* 8: 563–568.
20. Mendiratta SK, Martin WD, Hong S, Boesteanu A, Joyce S, et al. (1997) CD1d1 mutant mice are deficient in natural T cells that promptly produce IL-4. *Immunity* 6: 469–477.
21. Cui J, Shin T, Kawano T, Sato H, Kondo E, et al. (1997) Requirement for V α 14 NKT cells in IL-12-mediated rejection of tumors. *Science* 278: 1623–1626.
22. Watanabe H, Ohtsuka K, Kimura M, Ikarashi Y, Ohmori K, et al. (1992) Details of an isolation method for hepatic mononuclear cells in mice. *J Immunol Methods* 146: 145–154.
23. Ogawa Y, Masuzaki H, Hosoda K, Aizawa-Abe M, Suga J, et al. (1999) Increased glucose metabolism and insulin sensitivity in transgenic skinny mice overexpressing leptin. *Diabetes* 48: 1822–9.
24. Wu D, Molofsky AB, Liang HE, Ricardo-Gonzalez RR, Jouihan HA, et al. (2011) Eosinophils sustain adipose alternatively activated macrophages associated with glucose homeostasis. *Science* 332: 243–7.
25. Winer DA, Winer S, Shen L, Wadia PP, Yantha J, et al. (2011) B cells promote insulin resistance through modulation of T cells and production of pathogenic IgG antibodies. *Nat Med* 17: 610–7.
26. Miyazaki Y, Iwabuchi K, Iwata D, Miyazaki A, Kon Y, et al. (2008) Effect of western diet on NKT cell function and NKT cell-mediated regulation of Th1 response. *Scand J Immunol* 67: 230–237.
27. Jahng A, Maricic I, Aguilera C, Cardell S, Halder RC, et al. (2004) Prevention of autoimmunity by targeting a distinct, noninvariant CD1d-reactive T cell population reactive to sulfatide. *J Exp Med* 199: 947–957.
28. Schümann J, Voyle RB, Wei BY, MacDonald HR (2003) Influence of the TCR V β domain on the avidity of CD1d: α -galactosylceramide binding by invariant V α 14 NKT cells. *J Immunol* 170: 5815–9.
29. Hammond KJ, Pelikan SB, Crowe NY, Randle-Barrett E, Nakayama T, et al. (1999) NKT cells are phenotypically and functionally diverse. *Eur J Immunol* 29: 3768–81.
30. Mantell BS, Stefanovic-Racic M, Yang X, Dedousis N, Sipula IJ, et al. (2011) Mice lacking NKT cells but with a complete complement of CD8 T-cells are not protected against the metabolic abnormalities of diet-induced obesity. *PLoS One* 6: e19831.
31. Bäckhed F, Ding H, Wang T, Hooper LV, Koh GY, et al. (2004) The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci U S A* 101: 15718–23.
32. Armougom F, Henry M, Vialettes B, Raccach D, Raoult D (2009) Monitoring bacterial community of human gut microbiota reveals an increase in *Lactobacillus* in obese patients and *Methanogens* in anorexic patients. *PLoS One* 4: e7125.
33. Vijay-Kumar M, Aitken JD, Carvalho FA, Cullender TC, Mwangi S, et al. (2010) Metabolic syndrome and altered gut microbiota in mice lacking Toll-like receptor 5. *Science* 328: 228–231.
34. Nieuwenhuis EE, Matsumoto T, Lindenbergh D, Willemsen R, Kaser A, et al. (2009) Cd1d-dependent regulation of bacterial colonization in the intestine of mice. *J Clin Invest* 119: 1241–50.
35. Bassaganya-Riera J, Misyak S, Guri AJ, Hontecillas R (2009) PPAR γ is highly expressed in F4/80^{hi} adipose tissue macrophages and dampens adipose-tissue inflammation. *Cell Immunol* 258: 138–46.
36. Guri AJ, Hontecillas R, Ferrer G, Casagran O, Wankhade U, et al. (2008) Loss of PPAR γ in immune cells impairs the ability of abscisic acid to improve insulin sensitivity by suppressing monocyte chemoattractant protein-1 expression and macrophage infiltration into white adipose tissue. *J Nutr Biochem* 19: 216–228.
37. Lumeng CN, Bodzin JL, Saltiel AR (2007) Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *J Clin Invest* 117: 175–84.
38. Elinav E, Pappo O, Sklair-Levy M, Margalit M, Shibolet O, et al. (2006) Adoptive transfer of regulatory NKT lymphocytes ameliorates non-alcoholic steatohepatitis and glucose intolerance in ob/ob mice and is associated with intrahepatic CD8 trapping. *J Pathol* 209: 121–8.
39. Margalit M, Shalev Z, Pappo O, Sklair-Levy M, Alper R, et al. (2006) Glucocerebroside ameliorates the metabolic syndrome in OB/OB mice. *J Pharmacol Exp Ther* 319: 105–10.
40. Lalazar G, Ya'acov AB, Lador A, Livovsky DM, Pappo O, et al. (2008) Modulation of intracellular machinery by β -glycolipids is associated with alteration of NKT lipid rafts and amelioration of concanavalin-induced hepatitis. *Mol Immunol* 45: 3517–25.
41. Lynch L, O'Shea D, Winter DC, Geoghegan J, Doherty DG, et al. (2009) Invariant NKT cells and CD1d⁺ cells amass in human omentum and are depleted in patients with cancer and obesity. *Eur J Immunol* 39: 1893–1901.
42. Exley MA, Hou R, Shaulov A, Tonti E, Dellabona P, et al. (2008) Selective activation, expansion, and monitoring of human iNKT cells with a monoclonal antibody specific for the TCR α -chain CDR3 loop. *Eur J Immunol* 38: 1756–66.
43. Roy KC, Maricic I, Khurana A, Smith TR, Halder RC, et al. (2008) Involvement of secretory and endosomal compartments in presentation of an exogenous self-glycolipid to type II NKT cells. *J Immunol* 180: 2942–50.