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博士学位論文

Studies on the multisensory integration in cricket's
auditory system

(コオロギ聴覚系における多感覚統合に関する研究)

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学位論文内容の要旨

博士の専攻分野の名称 博士（生命科学） 氏名 染谷 真琴

学位論文題名

Studies on the multisensory integration in cricket's auditory system

(コオロギ聴覚系における多感覚統合に関する研究)

動物は異なる感覚器官で受容された複数のモダリティの感覚刺激の情報を中枢神経系で統合する。この過程は多感覚統合と呼ばれ、それによる神経活動の変化は物体の知覚や刺激によって誘導される行動の修飾をもたらす。多感覚統合は捕食者の検出感度の向上や、より正確な位置や距離の把握に有効であり、生存のための回避行動にも利益をもたらすと推測される。昆虫の中枢神経系には捕食者の接近に関連した刺激を検出するいくつかのニューロンが同定されているが、これらのニューロンが複数モダリティの感覚入力を統合しうるのかは不明であった。本学位論文では、捕食者であるコウモリが発する超音波エコーに応答するコオロギの聴覚ニューロン AN2 を対象として、やはり捕食者の接近を検知する気流感覚器である尾葉からの機械感覚入力と前肢鼓膜器官からの聴覚入力の多感覚統合に着目し、この問題に取り組んだ。

第一章では、AN2 を含む聴覚ニューロンが尾葉からの機械感覚入力を受けるかを検証した。尾葉は空気流の変化によって捕食者の接近を検出し、コオロギに逃避行動を引き起こす。まず、頸部腹側縦連合からの細胞外記録により、音刺激および気流刺激に応答する上行性神経活動を検証し、次に細胞内記録法により前胸神経節内における多感覚ニューロンを探索した。その結果、すでに聴覚ニューロンとして同定されている AN2, ON1 を含むいくつかの前胸介在ニューロンが気流刺激に対してスパイク応答を示した。気流応答は尾葉を切除すると消失したことから、鼓膜器官で受容された聴覚入力と尾葉で受容された気流感覚入力が、前胸神経節内の感覚情報処理の初期段階において統合されることが示唆された。

第二章では、聴覚および気流感覚入力が AN2 のスパイク発火に与える影響を検証した。音と気流刺激の同時提示刺激に対する応答はそれぞれの刺激の単独

提示に対する応答より有意に大きく、多感覚統合によって応答が増大されることがわかった。しかし、同時提示に対する応答は、基本的に単独提示に対する応答の和にほぼ等しく、AN2 は音と気流入力を線形に加算していた。単独提示応答の線形和よりも大きな加算は、スパイク閾値に近い弱い強度の刺激を同時提示した場合にのみ観察された。したがって、AN2 の多感覚統合における非線形加算は、興奮性シナプス後電位 (EPSP) からスパイクへ変換する閾値特性を反映したものであると考えられる。

AN2 は音源の位置、音圧および音の長さといった情報を高頻度のスパイク発火(バースト発火)で符号化することが報告されている。音と気流の同時提示は、単独提示よりも高い頻度でバースト発火を誘発したことから、多感覚統合は音情報表現へも影響することが示唆された。しかし、単独提示した音刺激の強度を増大した場合にも同様にバースト発火は増大し、同程度の大きさのスパイク活動に含まれるバースト数は単独提示と同時提示に違いは見られなかった。したがって、バースト発火の増加は単にスパイク応答の増大によるもので、多感覚統合特異的なものではないと考えられる。したがって、AN2 は刺激のモダリティによらず興奮性入力を統合しており、その統合過程は一般的なシナプス入力加算のメカニズムによるものであると推測された。

第三章では、AN2 におけるバースト発火をもたらす神経基盤を理解する手がかりとして、感覚刺激のモダリティ、およびバースト発火と閾値以下の膜電位応答の関連を検証した。細胞内記録した AN2 の膜電位応答からスパイク成分を取り除き、閾値下の興奮性シナプス後電位 (EPSP) を推定した。第二章においてほとんどのバースト発火は一発目の活動電位に続いて生じていたため、この初発スパイク後の EPSP に注目したところ、同時提示刺激に対する応答は単独提示刺激に対する応答よりも脱分極がより持続していた。この期間の脱分極の高とその持続時間は、いずれもバースト発火の発生および個々のバースト発火の大きさと相関を示したが、その関係性に刺激モダリティによる違いはみられなかった。以上の結果から、閾値以下の膜電位変化はバーストの発生およびその大きさを決定する重要な要因であるが、それに対する影響は入力強度の増大によるものであり多感覚統合特異的ではないことがわかった。

AN2 はコオロギの捕食者であるコウモリが発する超音波を検出し、そのバー

スト発火は飛行中のコオロギに音源と反対方向に飛行する逃避行動を引き起こすことから、捕食者検出ニューロンとして働くと考えられている。本研究によって、AN2 が尾葉への気流刺激に対しても興奮性の応答を示し、鼓膜からの聴覚入力と線形に加算したスパイク出力をすることによってバースト発生を増大させることがわかった。本研究結果は、感覚情報処理の初期に単一ニューロン内で複数のモダリティの感覚入力統合され、逃避行動に利用されている可能性を示すものである。

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General Discussion

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Acknowledgment

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General Introduction

The central nervous systems of animals integrate various modalities of signals detected by distinct sensory organs in a process known as multisensory integration (Pouget et al., 2002; Fetsch et al., 2013). Multisensory integration modulates the neural activity, which leads to improvement of sensory perception and enhancement of the behavioral responses to cross-modal stimuli compared to those of unimodal stimuli (Stein et al., 1988; Alais and Burr, 2004; Gu et al., 2008; McMeniman et al., 2014). For example, Human accurately perceives an object by integrating the visual and tactile signals (Ernst and Banks, 2002).

Thus, multisensory integration offers some advantages in detection of predators and in successful escaping because the attack of predator gives rise to multiple sensory cues. Several sensory interneurons in insects have been identified as detectors of predators' signals, and their activity elicits avoidance behaviors. For example, the visual interneurons, such as LPLC2 in fruit flies and DCMD in locusts are sensitive to looming motion stimuli that are perceived as an approaching predator. If the spiking responses of these neurons exceed a certain threshold, escape behavior is triggered (Fotowat and Gabbiani, 2007; Herberholz and Marquart, 2012; Klapoetke et al., 2017). The several identified auditory interneurons in crickets, moths, and mantids are also sensitive to ultrasound, such as the echolocation call of bats that prey on these insects, and activity of these neurons evoked by these stimuli elicits

23 defensive steering in flight (Hoy et al., 1989; Pollack, 2015). However, it
24 remains unknown whether these key neurons are sensitive to multiple
25 sensory cues relevant to a predator's attack. If these neurons have
26 multimodal sensitivity, how are their spiking outputs altered by
27 simultaneous sensory inputs of different modalities? To address these
28 questions, which are main concerns of my dissertation, I testes multimodal
29 sensitivity in prothoracic auditory neurons in the crickets, and focused on the
30 auditory neuron 2 (AN2) that was identified as one of the multimodal neurons
31 sensitive to both ultrasound and airflow.

32 **Chapter I**

33 **Identification of Multimodal Neurons of Crickets**

1.1 Introduction

Crickets process information on aero dynamics with two distinct sensory systems, cercal and auditory systems, which have the distinctly different sensory organs and frequency ranges of aero dynamics (Fig. 1-1A). Both sensory systems provide crucial signals of the predator attack and trigger the escape behavior.

The cerci are abdominal appendages covered with 500-750 filiform hairs that sense surrounding air particle displacement (Palka and Olberg, 1977; Miller et al., 2011). Information about airflow dynamics is processed by a local circuit within the terminal abdominal ganglion and transmitted by several ascending giant interneurons (GIs) (Fig. 1-1B) (Miller et al., 1991; Theunissen and Miller, 1991). The airflow signals detected by cerci are considered to be perceived as a sign of a predator's attack for crickets standing on the ground, because short air puff stimuli elicit running or jumping escapes (Tauber and Camhi, 1995; Oe and Ogawa, 2013; Sato et al., 2017).

On the other hand, sound with the higher frequency than 3 kHz (Imaizumi and Pollack, 1999) is detected by auditory receptor neurons of tympanal organ located at tibia of the front legs. The auditory receptor neurons project their afferent axons into the prothoracic ganglion (Hennig, 1988; Imaizumi and Pollack, 1999, 2005). Several prothoracic neurons directly receive excitatory synaptic inputs from auditory receptor afferents (Hennig, 1988). Sound information is conveyed by several kinds of ascending

projection neurons to brain (Hennig, 1988). The acoustic signals mediated by the ascending auditory neurons elicit escape steering during the flight (Nolen and Hoy, 1984; Schildberger and Hörner, 1988; Hedwig, 2006; Marsat and Pollack, 2012).

The cercal GIs project large long axon to brain and have axon collaterals arborizing into each segmental ganglion including the prothoracic ganglion (Hirota et al., 1993), suggesting the possibility that prothoracic auditory neurons are also sensitive to the airflow via the cercal system. Actually, it has been revealed that some prothoracic auditory neurons respond to wind stimuli (Atkins and Pollack, 1987). However, the wind sensitivity of auditory neurons related to avoidance behaviors has not been tested.

In this chapter, I explored whether the auditory neurons, which play key roles in the sensory coding relevant to biologically important events, receive the airflow inputs from cerci without the neural pathways through protocerebrum.

1.2 Materials and Methods

Animals

Adult female crickets (*Gryllus bimaculatus*) were used for the experiments. The crickets were bred and housed on a 12-h light/dark cycle at a constant temperature of 28°C.

Extracellular recording

For preparation of the extracellular recording, crickets were pinned dorsal side up on a silicone platform after removing the middle and hind legs. The connective nerve cords between the prothoracic and subesophageal ganglia were exposed by removing the gut and surrounding fat and were surgically cut to block the descending signals from the cephalic ganglia. Ascending spikes were extracellularly recorded using glass suction electrodes filled with isotonic saline placed proximally on the cut end of the nerve cord (Fig. 1-1A). For recording all ascending signals as multiple spike units, the whole nerve cord was suctioned by a glass micro-pipettes, while a part of ascending activities was recorded from a piece of split nerve cord for the single-unit recording. Signals were digitized at 20 kHz with a Powerlab 4/30 analog-digital converter (ADInstruments) and filtered with a 150–3,000 Hz band pass implemented in Chart version 7 software (ADInstruments).

Recorded spikes were sorted using a commercially available software (Spike Taro, Chinou Jouhou Shisutemu), which uses a sorting algorithm

based on the following two criteria: peak detection to separate individual units from a compound spike, and classification based on the correlation between separated units in terms of spike waveform and height. This algorithm has been used to analyze extracellular recordings of insect receptors (Kidokoro-Kobayashi et al., 2012). Spikes with heights in the range of $\pm 35\%$ and waveforms with a correlation coefficient of >0.90 were considered to be generated by a single neuron. Units that fired more than once on average in a trial were selected for analysis. In the extracellular recording of the whole nerve cord, 134 units were sorted from 4 recordings.

Stimulation for extracellular recording experiments

The airflow stimulus consisted of short puffs of nitrogen gas from a nozzle connected to a PV820 pneumatic picopump (World Precision Instruments, Sarasota, FL, USA). Eight nozzles were arranged around the cerci on the same horizontal plane with their ends positioned 45° apart at a distance of 40 mm from the cerci. The velocity of airflow was 1.12 m/s as measured at the cerci. The duration of the airflow stimulus was 200 ms.

For sound stimulation, a 10-kHz pure tone was generated by RPsEx software (Tucker Davis Technologies, Alachua, FL, USA) and presented through 1.5 inch full-range sealed loudspeaker (MM-SPS2, Sanwa Supply, Okayama, Japan). Eight speakers were arranged around the cerci on the same horizontal plane with their ends positioned 45° apart. The duration of

sound pulses was 200 ms with symmetrical rise and fall times of 2 ms. The sound intensity was 70 dB SPL in the recording from whole nerve cord and 76 dB SPL in the recording from a split nerve cord.

Single airflow or sound stimulus was provided independently from various directions for multi-unit recording or from ipsilateral side to the nerve cord for single-unit recording. To confirm whether the airflow was not detected by the tympanum, responses to airflow and sound stimuli were collected again after both frontal legs were ablated or both tympanums were gummed up by adhesion bond.

Intracellular recording

For preparation of the intracellular recording, the wings and hind legs were removed and the crickets were positioned ventral side-up on a block of clay and restrained with small metal clamps. The prothoracic ganglion was exposed and stabilized on a stainless-steel platform using a silver wire ring. To eliminate descending signals from the cephalic ganglion, the connective nerve cords between the prothoracic and subesophageal ganglia were cut. In the ablation experiment, both cerci were removed to confirm that the auditory neurons received the sensory signals from the cerci.

Glass microelectrodes (40-60 M Ω) were filled with 8% Lucifer yellow CH lithium salt (Sigma-Aldrich, RRID:SCR_008988) in 200-mM LiCl or 2-mM Oregon Green BAPTA-1 hexapotassium salt in 200-mM potassium acetate.

Lucifer yellow was used for most of the recordings and both kinds of the electrode solution have no influence on the spontaneous activities and neural responses. Glass microelectrode was inserted into dendrites or axons of auditory neurons and membrane potentials of recorded neurons were measured with current clamp mode. Electrophysiological signals were digitized at 20 kHz with a Powerlab 4S analog-to-digital converter (ADInstruments) and recorded with Chart version 7 software (ADInstruments). To load the fluorescent dye, a hyperpolarizing current of -1 to -2 nA was injected during recording. To identify the recorded neuron, the prothoracic ganglion was dissected after recording, fixed in 4% paraformaldehyde in 200-mM phosphate buffer over night, dehydrated for 15 min each in a series of 50, 70, 90, 100, and 100% ethanol, and cleared in methyl-salicylate.

Stimulation in the intracellular recordings

The airflow stimulus consisted of short puffs of nitrogen gas from a nozzle connected to an IM-300 microinjector (Narishige, Japan). For stimulation, the nozzle ends were positioned 15 mm away from the cerci on either side of the cricket. The duration of the airflow stimulus was 200 ms. The velocity of airflow was from 0.17 to 0.43 m/s (see each figure legend). For finding the auditory neurons, 5- or 15-kHz pure tone was generated by R PvdsEx software

(Tucker Davis Technologies) and presented through an ATH-CM707 headphone speaker (audio-technica, Tokyo, Japan). A pair of speakers were positioned 25 mm away from the tympanum on either side. The duration of sound pulses was 30 ms with symmetrical rise and fall times of 2 ms. The sound intensity was 60 dB SPL.

Data analysis

Spike traces were binned at 20-ms resolution through all figures in this chapter. To analyze subthreshold voltage traces, raw traces of the recordings were filtered by a median filter with a 5-ms window (Shi et al., 2017). Then, the resting level of membrane potential was subtracted from the filtered potentials to indicate the stimulus-evoked postsynaptic potentials. The latency of EPSP was defined as the duration from the stimulus onset to the points when the filtered potential exceeded 3 mV, which corresponded to 2 SD (2.89 mV) of the spontaneous fluctuation in the resting potentials of AN2 and omega neuron 1, from the resting potential.

MATLAB software (MathWorks, RRID:SCR_001622) was used for data processing and analysis, and R programming software v3.4.0 (R Core Team 2013) was used for statistical analysis. To avoid pseudoreplication, the data obtained from the same samples were averaged for each individual. All values are reported as mean $\pm$ standard error of the mean. A p -value of < 0.05 was considered to be statistically significant.

1.3 Results

1.3.1 Exploring multimodal neurons by extracellular recordings

First, I examined existence of ascending multimodal neurons in the thoracic or abdominal ganglia that responded to the acoustic and airflow stimuli. The spiking responses of the ascending neurons to the sound stimulus applied to tympanums and the airflow stimuli applied to cerci were extracellularly recorded from the distal cut end of a nerve cord between the subesophageal and prothoracic ganglia (Fig. 1-2A, B). Individual spikes were sorted into multiple units based on their wave shapes of spikes (see Methods). I found multi-modal units which showed firing responses to both sound and airflow stimuli (Fig. 1-3A, B). These multimodal units accounted for 46% of all sorted units (Fig. 1-3C). Thirty percent of units were sensitive to only airflow stimuli, but no units responded to only sound stimuli. The multimodal units showed remarkable increase in spiking activity in responses to the both stimuli (Fig. 1-3D). The auditory responses completely disappeared after the tympanums were removed (Fig. 1-3D). After the tympanum removal, 96% of them showed no significant increase in spiking responses to sound (unpaired t-test). These results indicate that a part of ascending neurons is sensitive to both airflow and sound stimuli.

However, the multi-modal units might contain multiple cellular activities derived from auditory-specific and cercal-specific neurons or activity of a single cell might split into multiple units. To confirm the

existence of multimodal neurons, single unit recording was performed from a split nerve cord (Fig. 1-4A). In the single unit recording, the multimodal spike unit was recorded, which showed spiking responses to both sound and airflow stimuli (Left 2 panels in Fig. 1-4B, C). After both tympanums were covered with adhesion bond, the units showed no response to the sound while it still responded to the airflow. This result also supported the existence of ascending multimodal neurons in the thoracic or abdominal ganglia.

1.3.2 Multimodal neurons in the prothoracic ganglion

To identify the multimodal neurons observed with extracellular recordings, I performed intracellular recording in the prothoracic ganglion to monitor the membrane potential responses to auditory and airflow stimuli and to stain the recorded neuron with fluorescent dye. I found that ascending neuron identified as AN2 and local interneuron identified as omega neuron 1 (ON1), both of which have been identified as the auditory ascending neuron (Wohlers and Huber, 1982), were sensitive to the airflow stimuli to cerci (Fig. 1-5A, B). Their responses to the airflow were significantly larger in the firing rate than the spontaneous activities (Fig. 1-5C). In addition to AN2 and ON1, omega neurons 2 (ON2), T-shape neuron 1 (TN1) and descending neuron 1 (DN1) also showed distinct spiking responses to airflow stimuli. However, their airflow-evoked responses were no significance statistically because of small size of samples. In contrast, the auditory ascending neuron 1 (AN1) showed

no response to the airflow stimuli (Fig. 1-6A, B), of which firing rate and membrane potential during the airflow stimulation were not different from their spontaneous activity (Fig. 1-6C, D). My results demonstrated that certain types of auditory neurons including ascending, descending, and local interneurons have bi-modal sensitivity.

1.3.3 Response property of the auditory neurons to airflow stimulation

Next, I examined the response properties of AN2 and ON1 to airflow stimuli (Fig. 1-7A). In both AN2 and ON1, the airflow stimulus evoked transient firing (Fig. 1-7B). The latency from stimulus onset to the first spike in AN2 was shorter than that in ON1 although this time difference was not significant (Fig. 1-7C). I observed no difference in their firing rate in the transient spiking responses (Fig. 1-7D) and during the later phase following the transient response (Fig. 1-7E).

There was significant difference in the time course of the subthreshold membrane potential changes between AN2 and ON1 (Fig. 1-8A). Both AN2 and ON1 showed transient depolarization of their membrane potential in response to airflow stimuli. In AN2, the depolarized membrane potential quickly returned to the resting level. In contrast, ON1's membrane potential depolarization was sustained even after the stimulus termination. The latency of EPSP of AN2 was shorter than that of ON1 (Fig. 1-8B). There was no difference in EPSP amplitude during the transient phase (Fig. 1-8C),

246 whereas the EPSP during the later phase following the transient
247 depolarization was larger in ON1 than AN2 (Fig. 1-8D). Therefore, although
248 both AN2 and ON1 show transient depolarization in response to airflow
249 stimuli, ON1's EPSP was longer lasting and evoked sustained firing. This
250 difference in time course of the responses suggests that different neural
251 pathways convey the airflow signals to the ascending and local interneurons.

252 ON1 shows directional selectivity for the auditory stimulus and plays a
253 prominent role in producing the directionality of AN1 and AN2 (Selverston et
254 al., 1985; Horseman and Huber, 1994). Thus, I investigated the directional
255 selectivity for airflow stimulus in ON1. ON1 tended to generate more spikes
256 in response to the airflow stimuli from ipsilateral to its soma than to the
257 contralateral stimuli (Fig. 1-9A, B). There was no difference in the latency
258 from stimulus onset to the first spike between the stimulus directions (Fig. 1-
259 9C). Since ON1 prefers the ipsilateral stimulus to its soma in its auditory
260 response (Wohlers and Huber, 1978), the directional preference in the airflow
261 response coincided with that for auditory stimuli.

262 Finally, I examined whether the cercal system mediated the airflow-evoked
263 excitatory synaptic inputs to AN2 and ON1. As shown in Fig. 1-7A and 1-8A,
264 AN2 and ON1 showed remarkable EPSPs and spiking responses to airflow
265 stimuli. In the individuals of which both cerci were ablated, both AN2 and
266 ON1 showed no change in their membrane potential in response to the airflow
267 stimuli (Fig. 1-10), meaning that the mechanosensory signals to AN2 and

268 ON1 are mediated by the cercal system.

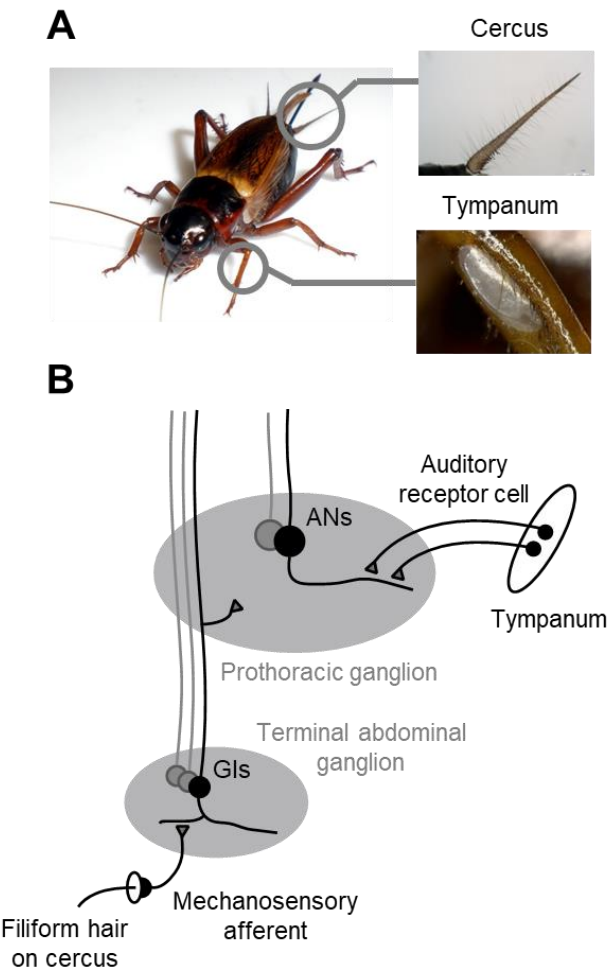
1.4 Discussion

In this study, I demonstrated that certain types of prothoracic auditory neurons including AN2 were sensitive to mechanosensory stimuli received via the cercal organ. On the other hand, AN1 considered to be a key neuron to elicit positive phonotaxis (Schildberger and Hörner, 1988) was insensitive to the airflow. AN1 responds to calling songs of male crickets, of which carrier frequency is about 5 kHz (Horseman and Huber, 1994), and induces oriented walking behavior in female attracted by the male's song (Schildberger and Hörner, 1988). In contrast, AN2 responds to ultrasound like echolocation call of bats that prey on the crickets. Thus, our results suggest that the airflow signals would influence the processing of auditory information relevant not to mating but to predators. Further, I revealed that ON1 was also sensitive to mechanosensory stimuli. This local interneuron inhibits activities of the contralateral ON1, AN1 and AN2, resulting in the enhancement of the directional selectivity in AN1 and AN2 (Selverston et al., 1985; Horseman and Huber, 1994). The airflow-driven activity of ON1 might modulate the directional selectivity of AN1 and AN2.

The interneurons AN2, ON1, ON2, TN1 and DN1, which were examined in this study, have been identified as auditory neurons in the prothoracic ganglion (Wohlers and Huber, 1982). Some previous studies have reported that these auditory interneurons are also affected by other modalities of sensory inputs. For example, AN2 and ON1 receive inhibitory inputs from

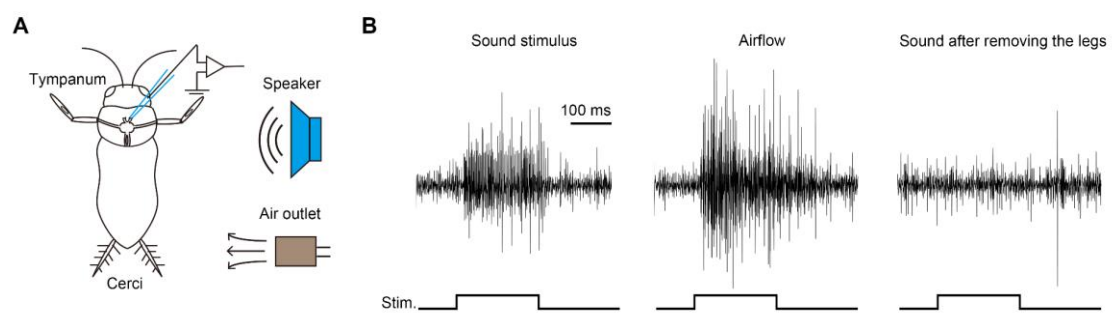
mechanoreceptors within the subgenual organ of legs which detects mechanical vibration (Wiese, 1981; Kühne et al., 1984). TN1 and DN1 responds to wind stimuli (Atkins and Pollack, 1987). In addition to these interneurons, certain T-shaped neurons, which extend both ascending and descending axons, and descending neurons are sensitive to sound, wind and light stimuli (Atkins and Pollack, 1987). These findings indicate that the auditory information is not processed by modality-specific pathways. Rather, the sensory systems within the prothoracic ganglion process multiple sensory modalities. Notably, AN2 directly receive excitatory inputs from receptor neurons in the tympanum (Hennig, 1988). Considering this previous finding and my results together, inputs from multiple sensory modalities are integrated in the early stage of sensory processing.

I demonstrated that the airflow inputs to AN2 and ON1 was mediated by the cercal system. Neural pathways from the cerci to wind sensitive auditory interneurons remain unclear. However, the wind-sensitive giant interneurons (GIs), which project their ascending axons to the brain, arborize their axon-collaterals into the thoracic ganglion (Hirota et al., 1993). Most of the GIs show directional selectivity and prefer airflow from ipsilateral side to their axons (Jacobs et al., 2008). ON1 also had directional preference for the airflow from ipsilateral side to its soma, which suggests that GIs with their axons ipsilateral to ON1's cell body provide airflow inputs to ON1.



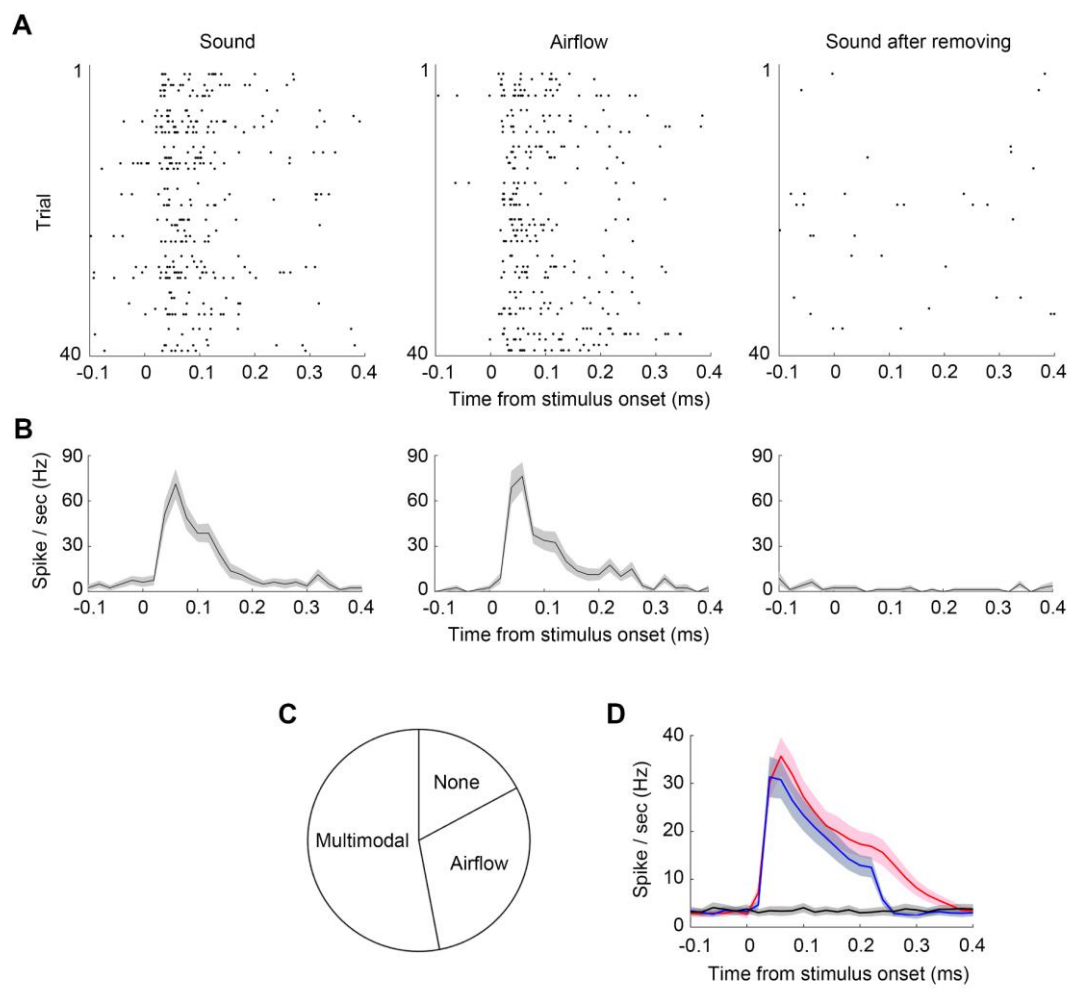
313 Figure 1-1. Auditory and cercal sensory systems of crickets. A) Two sensory
314 organs, tympanum and cercus. B) Diagram indicating the auditory and
315 cercal-sensory pathways. GIs, wind-sensitive giant interneurons; ANs,
316 auditory ascending neurons.

317 Figure 1-2

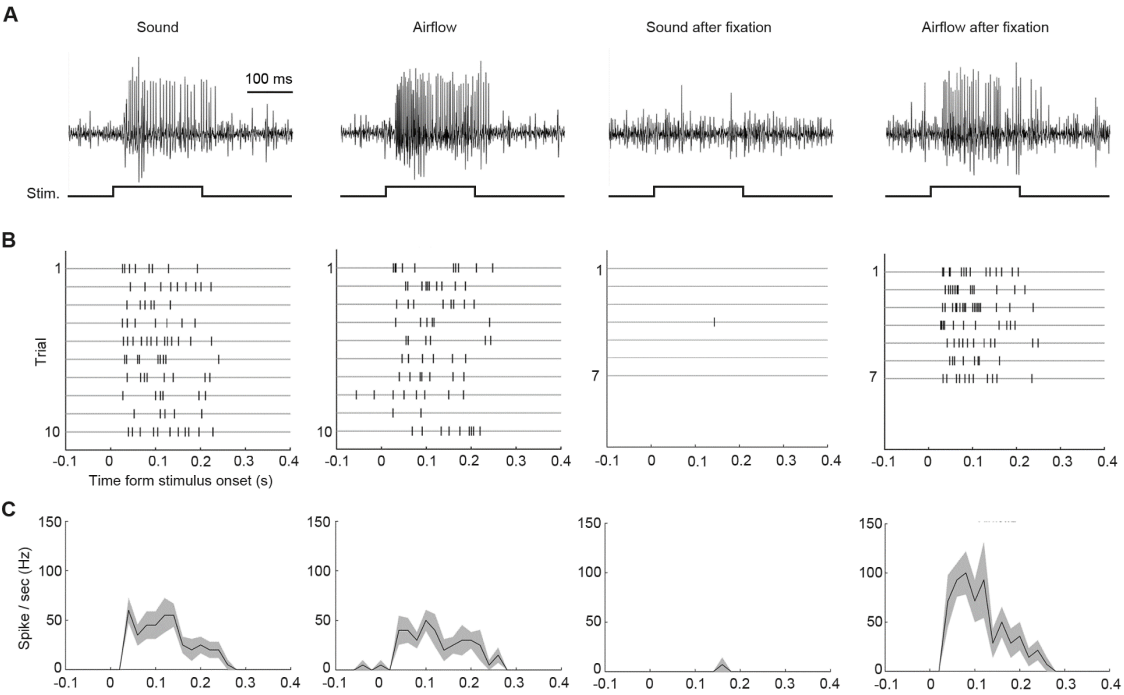


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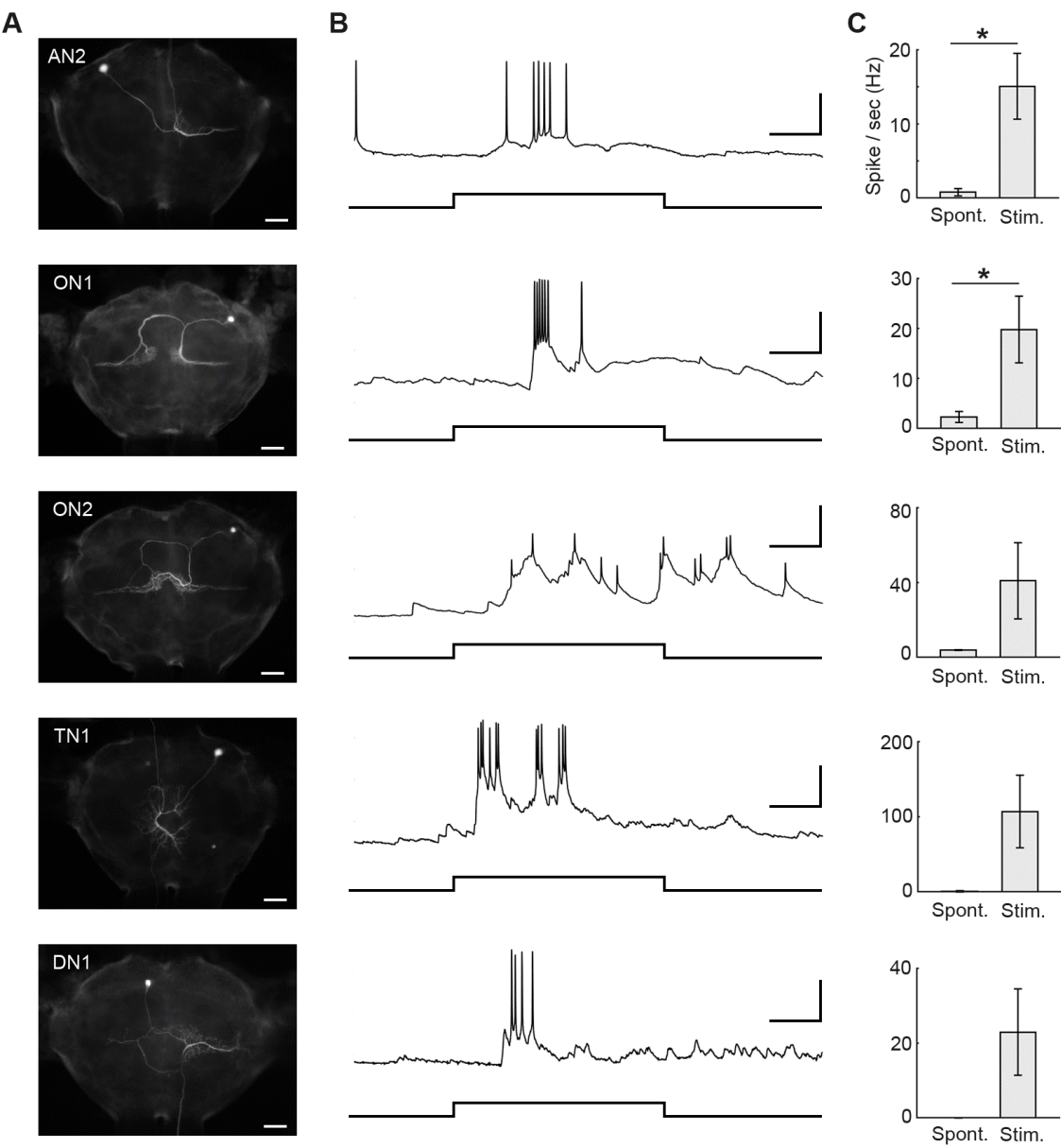
319 Figure 1-2. Extracellular recordings of ascending neurons. A) Experimental
320 setup for extracellular recording and stimulation. B) Typical ascending
321 activities evoked by sound (left panel) and airflow (middle). Right panel shows
322 sound-evoked response after both frontal legs were removed. Lower
323 rectangular traces indicate the stimulation.



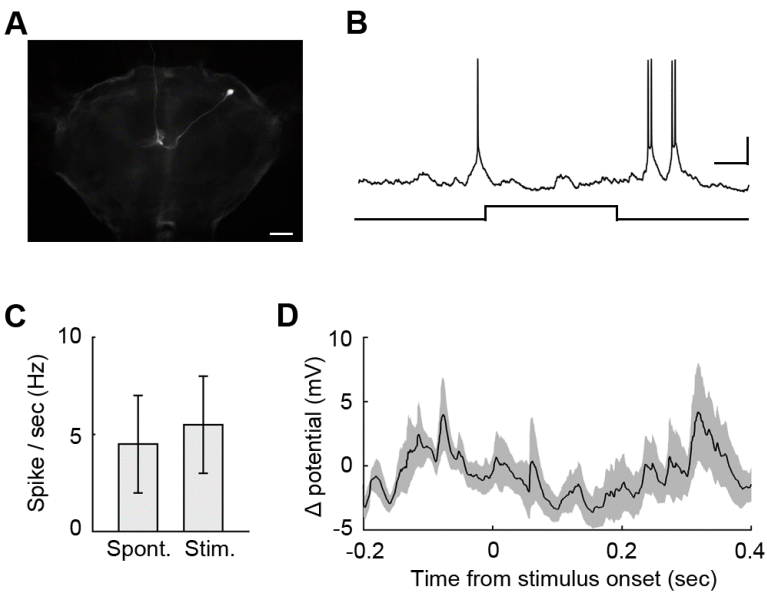
326 Figure 1-3. Ascending spike units responding to airflow and sound stimuli. A)
327 Raster plot of a typical multimodal unit sensitive to sound and airflow. Each
328 dot indicates individual spike for sound (left), airflow (middle) and sound
329 stimulations after the frontal legs were removed (right). B) Time course of
330 firing rates of the unit shown in A. Stimulation in each column corresponds
331 to that in Fig. 1-3A. C) Fraction of multimodal, airflow-specific, and non-
332 responsive spike units in all recorded units. Each unit was labeled whether
333 its firing rate during the stimulation was significantly different from
334 spontaneous firing rate (paired t-test and Holm-Bonferroni correction). D)
335 Time course of mean firing rate of all multimodal units for sound (blue),
336 airflow (red), and sound stimulation after leg removal (black).



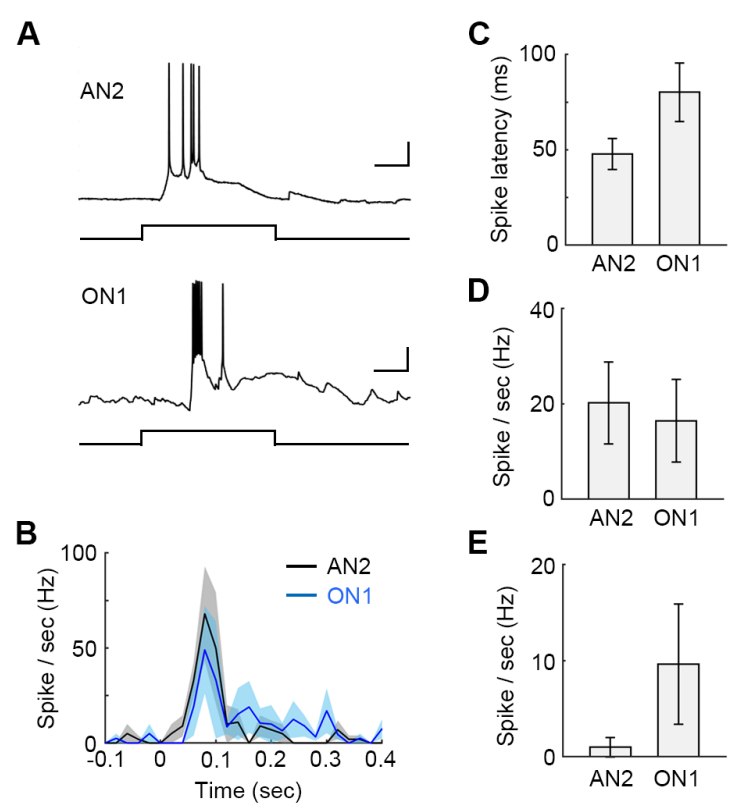
339 Figure 1-4. Single-unit recording of ascending neurons from the split nerve
340 cord. A) Raw traces of typical ascending activities for sound and airflow
341 stimulations in naïve (left two panels) and auditory-blocked (right two panels)
342 crickets in which both tympanums were gummed up. B), C) Raster plots (B)
343 and time course of the firing rate (C) of typical multimodal unit shown in Fig.
344 1-4B. Stimulation for each column corresponds to that in Fig. 1-4A.



346 Figure 1-5. Airflow-evoked membrane-potential responses in identified
347 auditory neurons. A) Fluorescent images of auditory neuron AN2, ON1, ON2,
348 TN1 and DN1 recorded intracellularly. Scale bars indicate 100 μ m. B) Typical
349 traces of membrane potential during the airflow stimulation. Lower
350 rectangular traces indicate the airflow stimulation. Vertical and horizontal
351 scale bars indicate 20 mV and 50 ms, respectively. C) Mean firing rate during
352 the spontaneous activity (Spont) and airflow stimulation (Stim), each of
353 which was measured as mean value for 200 ms before or during the airflow
354 stimulation. Sample sizes were 11 AN2s, 8 ON1s, 2 ON2s, 3 TN1s and 3 DN1s
355 (paired t-test, $p_{AN2} = 0.06 \times 10^{-1}$, $p_{ON1} = 0.04$, $p_{TN1} = 0.16$ and $p_{DN1} = 0.18$).
356 The cell type in each row corresponds to that in Figure 1-5A. Velocity of
357 airflow stimulus was from 0.17 to 0.43 m/s.

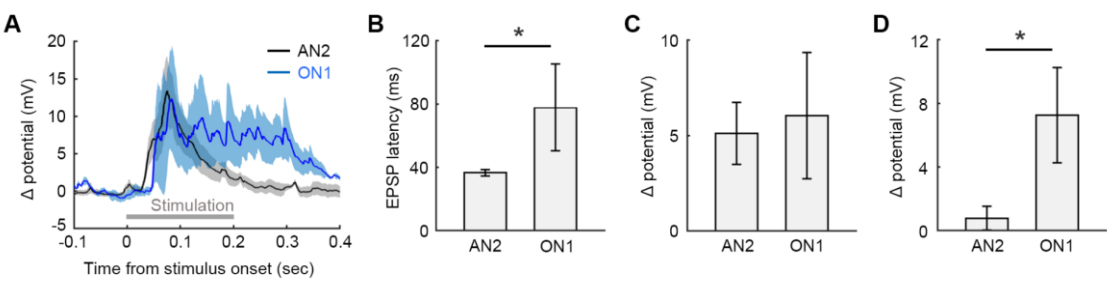


360 Figure 1-6. Neural response of AN1 to airflow stimulus. A) Fluorescent image
361 of AN1. Scale bar indicates 100 μm . B) Typical trace of membrane potential
362 in AN1 for airflow stimulation. Vertical and horizontal scale bars indicate 10
363 mV and 50 ms, respectively. C) Firing rate during spontaneous activity
364 (Spont) and stimulation (Stim) ($n = 2$ AN1s). D) Subthreshold change in the
365 membrane potential in AN1. After action potentials were removed using a
366 median filter with 5-ms window, the difference in the filtered membrane
367 potential from the average of resting potentials before stimulation was
368 calculated. Velocity of airflow stimulus was 0.43 m/s, and stimuli were
369 delivered from contralateral side to AN1's soma.



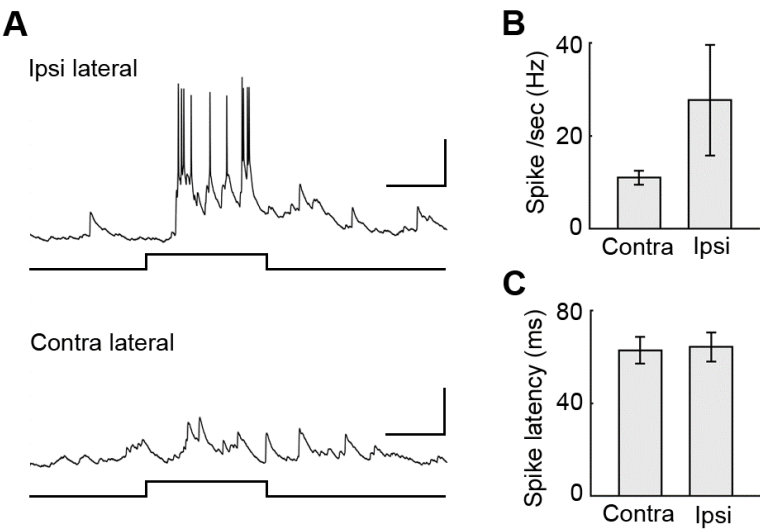
371 Figure 1-7. Firing-response properties of AN2 and ON1 for airflow
372 stimulation. A) Typical traces of membrane potentials in AN2 and ON1.
373 Vertical and horizontal scale bars indicate 10 mV and 50 ms, respectively. B)
374 Time course of firing rate in AN2 and ON1 ($n = 5$ neurons respectively
375 through Fig. 1-7). C) Latency from stimulus onset to the evoked first spike
376 (Wilcoxon rank sum test, $p = 0.10$). D) Firing rates in the earlier phase within
377 200 ms after the stimulus onset (unpaired t-test, $p = 0.77$). E) Firing rates in
378 the later phase which was time period from 200 to 300 ms after the stimulus
379 onset (unpaired t-test, $p = 0.24$). Velocity of airflow stimulus was 0.18 m/s.
380 Stimuli were delivered from ipsilateral side to ON1's soma and from
381 contralateral side to AN2's soma.

382 Figure 1-8



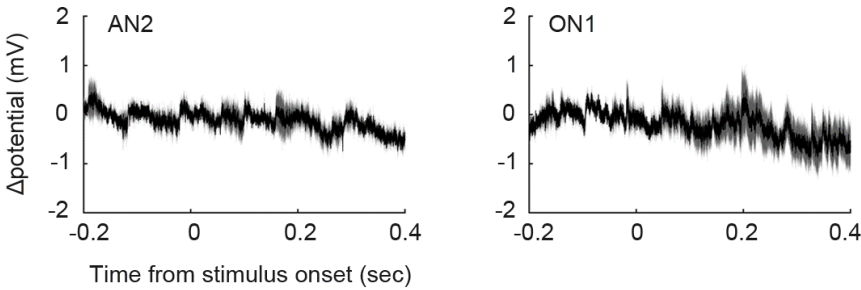
383

384 Figure 1-8. Subthreshold membrane-potential response properties of AN2
385 and ON1 for airflow stimulation. A) Time courses of subthreshold activity in
386 AN2 (black; $n = 5$ neurons) and ON1 (blue; $n = 3$ neurons). Gray line indicates
387 the airflow stimulation. B) Latency of EPSPs evoked by airflow stimulus
388 (sample size was the same as shown in A through B-D; Wilcoxon rank sum
389 test, $p = 0.04$). C) Mean postsynaptic potentials in the earlier phase within
390 200 ms after the stimulus onset (unpaired t-test, $p = 0.78$). D) Mean
391 potentials in the later phase which was time period from 200 to 300 ms after
392 the stimulus onset (unpaired t-test, $p = 0.04$). Velocity and direction of
393 stimulus are the same as shown in Fig. 1-7.



398 Figure 1-9. Directional selectivity to airflow stimulation in ON1. A) Typical
399 traces of membrane potentials in ON1 during airflow stimuli from ipsil-
400 (upper) or contralateral (lower) side to its soma. B) Mean firing rates in
401 responses to ipsil- (Ipsi) or contralateral (Contra) stimulation ($n = 3$ neurons;
402 paired t-test, $p = 0.29$). C) Latency from the stimulus onset to the first spike
403 for ipsil- or contralateral stimulation ($n = 3$ neurons; Wilcoxon signed-rank
404 test, $p = 0.75$). Velocity of airflow stimulus was 0.18 m/s.

405 Figure 1-10



406

407 Figure 1-10. Airflow-evoked responses in AN2 and ON1 without cerci. Time
408 courses of difference in the raw membrane potentials in AN2 and ON1 for
409 airflow stimulation after the both cerci were ablated. Velocity of airflow
410 stimulus was 0.17 m/s.

411 **Chapter II**

412 **Multisensory Integration in an Auditory Neuron AN2**

2.1 Introduction

In the previous chapter, I demonstrated that AN2 was sensitive to airflow stimuli to cerci. AN2 is one of the primary auditory interneurons that receives direct synaptic inputs within the prothoracic ganglion from auditory receptor neurons of the tympanal organ located in the tibia of the front legs (Wohlers and Huber, 1982; Hennig, 1988). This neuron is sensitive to high-frequency sounds (Wohlers and Huber, 1978; Popov and Markovich, 1982), and its high-frequency firing responses can trigger an avoidance steering response in flying crickets (Nolen and Hoy, 1986). Therefore, AN2 is considered to be a neuron that detects the presence of bats. The location of the sound source and its intensity and duration are encoded by ‘burst firing’ of AN2 with short inter-spike intervals, the behavioral functions of which have been well studied (Marsat and Pollack, 2005, 2006, 2010). My previous finding of the airflow-sensitivity of AN2 suggests that AN2 could detect other modality of sensory clue of a bat’s attack, such as strong turbulent airflow caused by the bat’s wingbeats.

It has been reported that the simultaneous presentation of different modal stimuli, termed as cross-modal stimulation, elicits enhancement or depression of spiking responses comparing to those responses to each component stimulus (Meredith and Stein, 1983). Furthermore, enhancement in cross-modal stimulation results in nonlinear (supra- or sub-linear) or linear summation of unimodal responses (Stein and Stanford, 2008). In this chapter,

435 I examined multisensory integration in the spiking outputs of AN2 in
436 response to the cross-modal stimulus combining the sound and airflow.

437 In addition, cluster of spikes in short intervals, called bursts, plays an
438 important role in the sensory signaling by AN2. The burst firing of AN2 can
439 improve the detection of sound stimuli and the representation of sound-source
440 location (Marsat and Pollack, 2006). The fine temporal structure of the burst
441 firing represents additional stimulus-related information such as intensity
442 and duration of sound (Marsat and Pollack, 2010). Thus, I also examined
443 effect of the multisensory integration on burst firings in AN2.

2.2 Materials and Methods

See the method in Chapter I about *Animals* and *Intracellular recordings*. Throughout the experiment in this chapter, 4 or 8% Lucifer yellow CH lithium salt in 200-mM LiCl was used for intracellular recordings.

Stimulation

See Methods in Chapter I about the apparatuses for airflow and sound stimulations. The velocity of airflow was 0.17 m/s at the position of cerci and the duration was 200 ms. For sound stimulation, only 15-kHz pure tone was used.

For unimodal stimulation of the cerci, single air puffs were used (Fig. 2-1). Different manners of stimulation were used for experiments with repetitive stimulation or various auditory intensities. In the repetitive-stimulation experiments that were performed to test multisensory integration, 9 sound pulses at 60 dB were given with 500-ms inter-stimulus interval, and airflow was provided coincident with the 4th sound pulse. To analyze multisensory effects, I compared the responses to the 3rd sound pulse with those to the combined stimuli. All responses to sound pulses (1st-3rd and 5th-9th) were used for comparing burst firing with non-burst firing. In the experiments with varying auditory intensities that were performed to test the effects of input amplitude, single sound pulses of 50, 60, 70, 80, or 90 dB were applied for the unimodal stimulation, and a single air puff combined with

various intensities of sound pulses was applied for the cross-modal stimulation. All stimuli were delivered contralateral to the soma of the recorded AN2. For the cross-modal stimuli with both types of stimulation, differences in the stimulus onset between the sound pulse and airflow were < 15 ms, and the onset of the sound stimulus was defined as the stimulus onset for cross-modal stimuli.

Data analysis

In the repetitive stimulation experiments, neural responses to sound pulses, airflow, and combined stimuli were recorded from 8 neurons in different crickets. In the various intensities experiments, the responses to sound pulses were recorded from 4 neurons in different crickets, and those to airflow and combined stimuli were obtained from 3 out of these 4 neurons. In addition to these experiments, responses to airflow stimuli were also recorded from 5 neurons. In the cerci ablation experiments, neural responses to airflow stimuli were recorded from 2 neurons in 2 crickets. Spikes were detected using custom-written MATLAB routines (R2016a, MathWorks, RRID:SCR_001622). Raw traces of membrane potential were filtered with a 150-Hz high-pass filter, and spike peaks were detected based on the filtered traces using the findpeaks function in MATLAB. To analyze the time course of the firing rate, spike trains were binned at a size of 1 ms and convolved with a moving-average filter with a 20-ms window. Summation of the

488 responses (r_{sum}) to the uni-modal stimulus was defined as

489
$$r_{sum} = r_{sound} + r_{airflow}$$

490 where r_{sound} and $r_{airflow}$ indicate the firing rate in response to sound or
491 airflow stimulus, respectively. Linearity of the multisensory integration was
492 defined as

493
$$Linearity = \frac{r_{cross}}{r_{sum}}$$

494 where r_{cross} indicates the firing rate in the response evoked by the cross-
495 modal stimulus. Histogram of inter-spike intervals (ISI) was fitted to a kernel
496 probability distribution using normal kernel smoothing with a 0.3-ms
497 bandwidth. To test the ISI distribution results from a Poisson process, the ISI
498 histogram was also fitted to a single exponential curve. A burst was defined
499 as a cluster of spikes separated by short intervals (< 6.5 ms; Marsat and
500 Pollack 2006). The first and last spike within a burst were defined as the
501 onset and offset of the burst, respectively. Size of the burst was defined as a
502 product of the burst duration and the number of spikes within the burst
503 (Marsat and Pollack, 2010).

2.3 Results

2.3.1 Multisensory enhancement in AN2 spiking activity

To examine the multisensory effects of simultaneous inputs with different modalities on the spiking activities of AN2, I compared neural responses to single air puffs, the 3rd sound pulses, and cross-modal stimulation combining an air puff with the 4th sound pulse (Fig. 2-2A). Cross-modal stimuli elicited significantly larger numbers of spikes than unimodal stimuli did, but the amplitudes of the responses to the cross-modal stimuli were similar to the arithmetic sum of the responses to the 3rd sound pulse and air puff that were separately presented (Fig. 2-2A, B). The linearity of the responses to cross-modal stimuli was examined using various intensities of sound combined with airflow, the velocity of which was constant (Fig. 2-2C). Higher intensities of sound stimuli elicited greater spiking responses in both uni- and bimodal stimulus conditions, but the responses to cross-modal stimuli were close to the arithmetic sum of the responses to airflow and to sound pulses regardless of the sound intensity. To verify the effects of the sound intensity on the linearity of the summation in the cross-modal responses, I examined the relationship between the linearity of summation and firing rates in the summed responses to unimodal stimuli (Fig. 2-2D). Linearity depended on the magnitude of the summed responses, and supra-linear summation was observed only in small responses in which the firing rate was lower than 10 Hz. This suggests that AN2 linearly and additively integrates auditory and

mechanosensory input, and supra-linear summation occurred in the integration of small sensory inputs evoked by stimuli around the threshold.

The linearity of summation in the cross-modal responses could depend on the stimulus intensity (Meredith and Stein, 1986; Stein and Stanford, 2008) but also on the latency from stimulus onset (Rowland and Stein, 2007; Rowland et al., 2007). Thus, I examined the temporal profile of the summation linearity. The cross-modal responses were larger in their firing rate than sound-evoked responses (Fig. 2-3A) and the arithmetic sum of unimodal responses (Fig. 2-3B) during the period of 30–60 ms after stimulus onset. However, there was no significance in the firing rates in this period between the cross-modal and summed responses ($n = 8$ neurons, paired t-test, $p = 0.07$, the firing rates were 85.05 ± 10.44 Hz for cross-modal and 66.61 ± 13.00 Hz for the summed responses, respectively). In addition, even if I focused on this period, the supra-linear summation occurred only in the small cross-modal responses (Fig. 2-3C). The supra-linear summation would simply result from the fact that summed inputs of the subthreshold unimodal stimuli were larger than the spike threshold and evoked action potentials. Our results suggest that AN2 linearly sums auditory and mechanosensory inputs and that supra-linearity cross-modal summation would arise from the spike threshold.

2.3.2 Multisensory enhancement in burst firing of AN2

In previous studies (Marsat and Pollack, 2006, 2007), the responses of AN2 to 30-kHz long-duration tones with Gaussian amplitude envelopes showed a characteristic distribution of inter-spike intervals (ISI), with a peak at short intervals and broad tail of longer intervals. Based on this ISI distribution, the spike trains with intervals shorter than 6.5 ms are classified into ‘bursts,’ which are used for encoding precise ultrasound information (Marsat and Pollack 2006, 2010). In the current study, ISI distribution of the multiple spikes that were evoked in 58% of responses (170/294 in 17 individuals) to 15-kHz tone pulses at 60 dB showed similar features, which included a peak at short intervals and a peaky but broad tail at longer intervals (Fig. 2-4A). In addition, the bimodal peaks in ISI distribution in our data could be also separated at 6.5 ms intervals. To confirm that the peak at the shorter intervals indicated the distribution of the burst firing, I fitted a single exponential curve to the whole ISI histogram. If the peaks surpass this curve, these activities were considered to be burst firings different from the ISI distribution expected by Poisson process (Bastian and Nguyenkim, 2001; Marsat and Pollack, 2012). The distinct peak at the intervals shorter than 6.5 ms was higher than the exponential curve, indicating that the 15-kHz tone pulse stimulus enabled AN2 to generate the burst firings, which could be separated from the isolated spikes using the same criteria as in previous studies.

I examined effects of cross-modal stimuli on generation of burst firing.

Burst firing responses were more frequently evoked by cross-modal stimuli than by unimodal stimuli (Fig. 2-5A). To clarify whether this enhancement of burst firing activity was specific to multisensory integration or simply resulted from increased firing rates due to linear summation, I evaluated the effects of response magnitude on the probability of burst occurrence among cross- and unimodal responses. The larger the spiking responses were, the higher the probability of burst occurrence was regardless of the sensory modality (Fig. 2-5B). Further, there was little difference among the uni- and cross-modal responses in the dynamic ranges of the curves, that indicated a relationship between the response magnitude and the bursting probability. This means that multisensory integration does not specifically affect the generation of burst firing. Rather, more frequent burst firing would result simply from an increase in spiking responses via linear summation of auditory and mechanosensory inputs.

I also focused on temporal aspects of burst generation (Fig. 2-5C). Most of the bursts occurred following the first action potentials evoked by the auditory and cross-modal stimuli. In contrast, the bursts evoked by the airflow stimuli followed several isolated spikes to appear in the middle of spike trains. Taken together, these results illustrate that AN2 integrates sound and airflow inputs and enhances the response magnitude, resulting in burst firing activity at the beginning of responses. However, the bursts were also enhanced by stronger unimodal stimuli, and no difference in the timing

of burst generation was observed between the auditory and cross-modal responses.

2.3.3 Effect of temporal coincidence of the stimuli on the burst firing

In multisensory integration, the neural responses to the cross-modal stimulus were enhanced when the stimuli were presented at the same time and/or provided from the same location (Holmes and Spence, 2005). This spatio-temporal coincidence detection is considered as a mechanism to precisely detect a single event or object presenting multiple sensory cues. To examine whether AN2 responses to the cross-modal stimulus depended on the timing of stimulus presentation, the sound and airflow stimuli were provided in various intervals between them (Fig. 2-6A). The number of spikes evoked by stimulus did not depend on the temporal difference in the stimulus onset (Fig. 2-6B). However, the cross-modal stimuli elicited a single burst when the airflow and sound stimuli were presented at approximately the same time (Fig. 2-6C). When the stimulus onsets separated over 40 ms, AN2 generated multiple bursts. To focus on the temporal structure of burst, I measured the duration and size, which is product of number of spikes within burst and duration of burst and is positively correlated with magnitude of the behavioral response (Marsat and Pollack, 2010). Figures 2-6D and 2-6E show the relationships between the time lag between sound and airflow stimuli and the maximum duration or maximum size of the burst, both of which were

614 measured from the largest burst in the response if multiple bursts were
615 evoked. Both duration and size became larger as the difference of stimulus
616 interval was shorter (Fig. 2-6D, E). Not only the maximum duration and size
617 of the individual burst, but the summation of size in all bursts elicited by a
618 single stimulation also depended on the difference of stimulus onset (Fig. 2-
619 6F, G). These results suggest that AN2 represents the coincidence of the
620 stimulus onsets by using the temporal structure of burst.

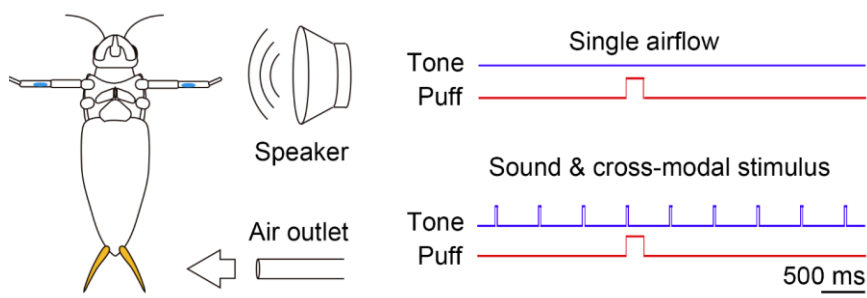
2.4 Discussion

I demonstrated that the cross-modal stimulation enhanced the magnitude of firing activity. Further, simply put, the spike outputs evoked in AN2 by cross-modal stimuli were equal to linear summation of those evoked by the respective unimodal stimuli. Supra-linear summation in spiking activity was observed only when the unimodal stimuli were as weak as the threshold for generating spikes, suggesting that supra-linear summation would result from the non-linear transduction from graded potentials to action potentials by threshold properties of voltage-gated channels (Holmes and Spence, 2005). Our result that supra-linear summation was observed only around spike threshold is consistent with that in recent studies of the MSTd (dorsal medial superior temporal) area of macaques (Morgan et al. 2008) and the superior colliculus of cats (Perrault et al., 2003, 2005; Stanford, 2005). Because the supra-linearity of the spike threshold is based on intrinsic active membrane properties, the supra-linear summation of spike responses of AN2 would not be specific to multisensory integration.

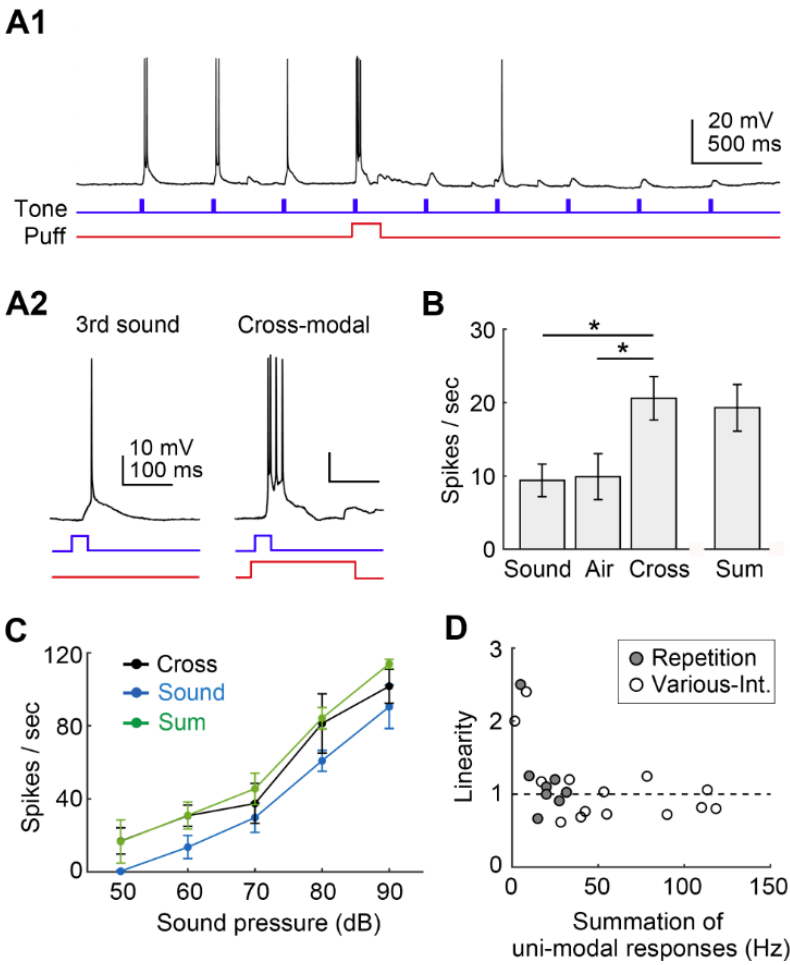
As well as the mean firing rate of the response, the burst firing was also enhanced by the cross-modal stimulation. Considering that AN2 uses bursts to encode the sound information such as intensity, duration, and location (Marsat and Pollack, 2006, 2010), airflow stimulus would have some impacts on acoustic representation in AN2. Although it is unclear the role of cercal signals on the acoustic representation in AN2, multisensory integration may

643 improve neural representation of the component stimulus (Ibrahim et al.,
644 2016; Atilgan et al., 2018).

645 Figure 2-1



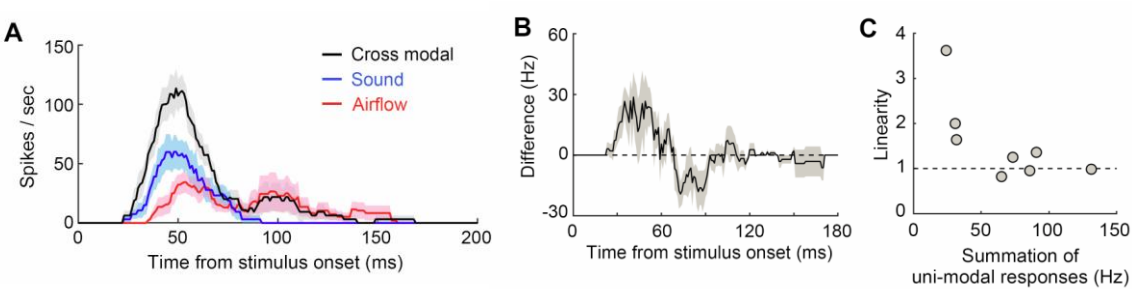
646 Figure 2-1. Experimental setup (left) and stimulation protocol of repetitive-
647 stimulation experiments (right). Single short durations of airflow were
648 delivered separately (upper). Nine tone pulses were repetitively applied and
649 single airflow stimuli were combined with the 4th pulse (lower).



652 Figure 2-2. Multisensory enhancement of AN2 firing activity. A) Typical
 653 responses to sound and cross-modal stimuli in the repetitive stimulation
 654 experiments (A1), and the enlarged views of the responses to the 3rd sound
 655 pulse and cross-modal stimulus combining the 4th sound pulse with airflow
 656 (A2). B) Firing rates of responses to 3rd sound, airflow, and cross-modal
 657 stimuli for 200 ms after stimulus onset. Cross-modal stimuli caused
 658 significant increases in the firing rate than unimodal stimuli ($n = 8$ neurons,
 659 one-way repeated measures ANOVA, $p = 0.02$; paired t-tests and Holm-
 660 Bonferroni correction, cross vs. sound, $p = 0.03$; cross vs. airflow, $p =$
 661 0.09×10^{-2} ; sound vs. air, $p = 0.91$). Right bar indicates the arithmetic sum
 662 (Sum) of the firing responses to sound and airflow. There was no significant
 663 difference in the firing rate between the sum of the unimodal responses and
 664 the cross-modal responses ($n = 8$ neurons, paired t-test, $p = 0.39$). C) Sound
 665 intensity-dependent curves of the responses to the sound (blue, $n = 4$ neurons),
 666 cross-modal stimuli (black, $n = 3$ neurons excluding the point at 80 dB [$n = 2$
 667 neurons]) and those predicted from the arithmetic sum of the unimodal
 668 responses (green, $n = 3$ neurons). For cross-modal stimulation, the constant
 669 velocity (0.17 m/s) airflow was combined with sound at various intensities (50
 670 to 90 dB). D) Relationship between linearity of the cross-modal response and
 671 the firing rate in summation of the unimodal responses to sound and airflow.
 672 Filled circles indicate the mean value of data measured from repetitive
 673 stimulation experiments for each individual ($n = 8$ neurons). Open circles

674 indicate the data of the response measured from various intensities of
675 stimulation experiments ($n = 14$ data points from 3 neurons). 'Linearity = 1'
676 shown by a dashed line indicates that the cross-modal response was equal to
677 the linear summation of the unimodal responses. 'Linearity > 1 ' or ' < 1 '
678 indicates supra- or sub-linear summation, respectively.

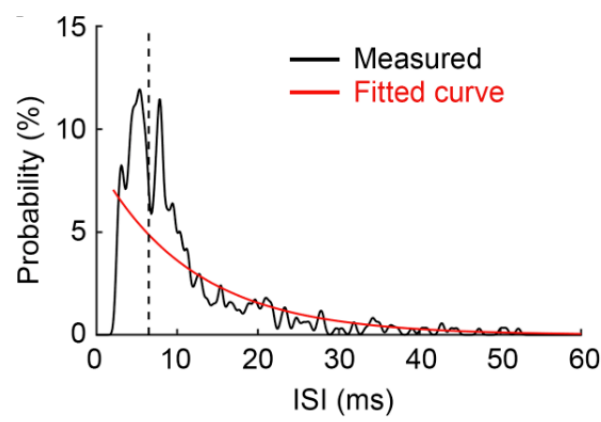
679 Figure 2-3



680

681 Figure 2-3. Dependency of linearity on latency from stimulus onset. A)
682 Variation in the mean firing rate over time in response to cross-modal (black),
683 sound (blue), and airflow (red) stimuli in the repetitive stimulation
684 experiments ($n = 8$ neurons). B) Difference in firing rate between cross-modal
685 responses and the summation of auditory and cercal responses ($n = 8$ neurons).
686 C) Relationship between linearity of the cross-modal response and the firing
687 rates in the summation of unimodal responses during the period of 10–60 ms
688 after the stimulus onset, which corresponds to the period of 30–60 ms shown
689 in F.

690 Figure 2-4



691 Figure 2-4. Distribution of ISIs measured from all spiking responses to 15-
692 kHz sound pulses of 60 dB (black; n = 424 ISIs from 16 neurons). A single
693 exponential curve was fitted to the whole distribution (red; $f(x|\mu) = \frac{1}{\mu} e^{\frac{-x}{\mu}}$,
694 $\mu = 11.87$). Dashed line indicates 6.5 ms ISI that was used for criterion of the
695 burst.

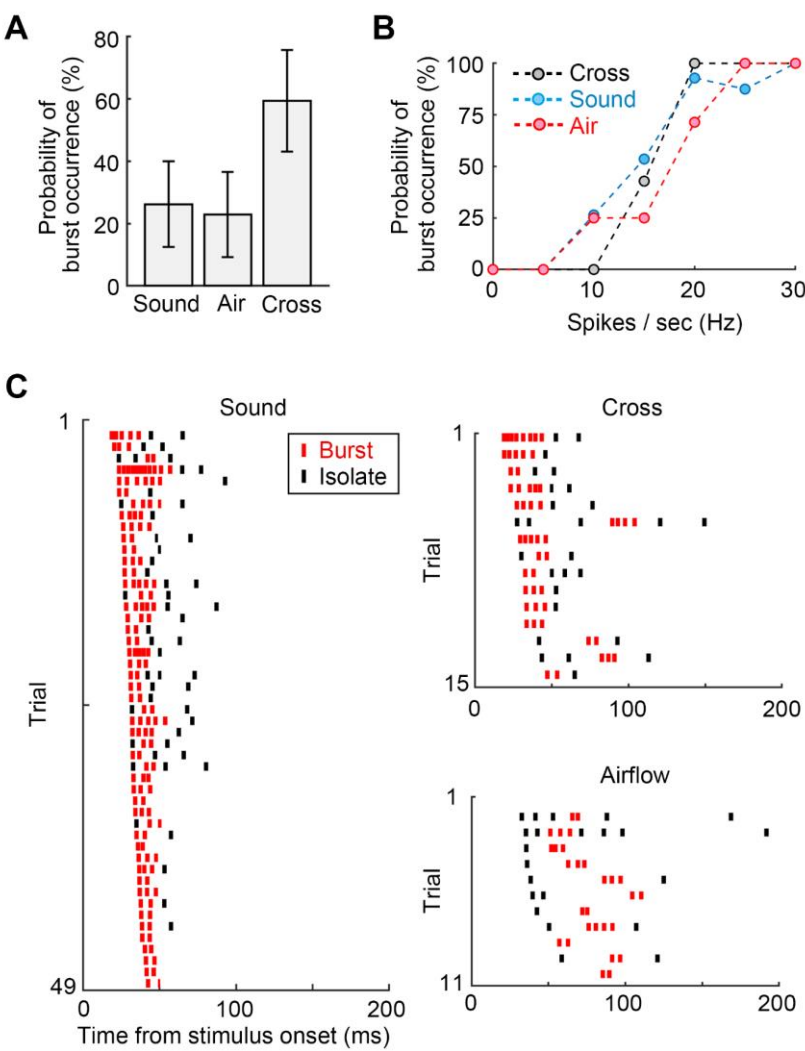
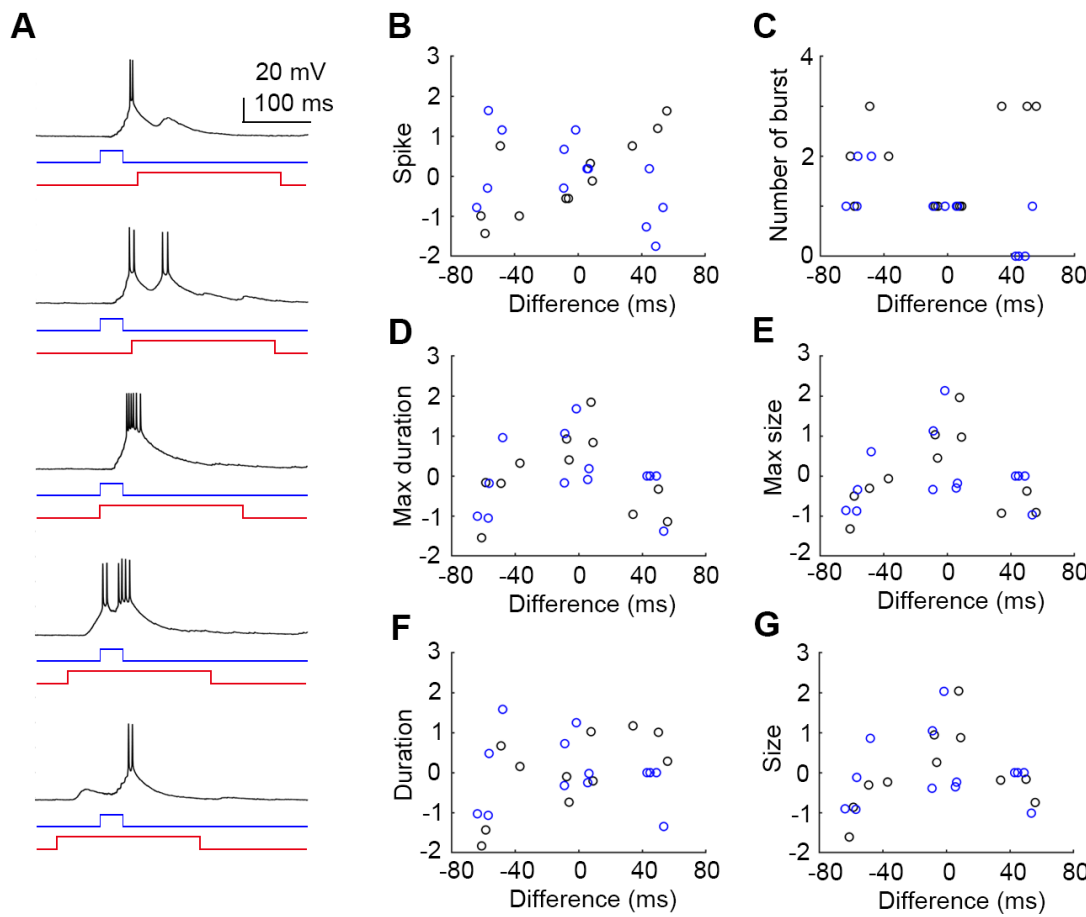


Figure 2-5. Multisensory enhancement of bursting. A) Probability of burst occurrence in the unimodal and cross-modal responses in the repetitive stimulation experiments ($n = 8$ neurons, Friedman test, $p = 0.04$; Wilcoxon signed-rank test, cross vs. sound, $p = 0.13$; cross vs. air, $p = 0.06$; air vs. sound, $p = 1.00$). B) Dependency of the probability of burst generation on the firing rate in uni- and cross-modal responses. The data of both uni- and cross-modal responses were obtained from all samples in the repetitive stimulation and various intensities stimulation experiments ($n = 11$ neurons for sound and cross-modal) and the single airflow stimulation ($n = 16$ neurons for airflow). The firing rates were measured for 200 ms after the stimulus onset. C) Timing of burst firing in response to sound (left), cross-modal (upper right), and airflow (lower right) stimuli. Red and black lines indicate bursting and isolated spikes for individual trials, respectively. Each trial was sorted by the latency of the first spike. The same samples as in C were used.



713 Figure 2-6. Dependency of the burst structures on temporal coincidence of
714 stimulus. A) Traces of membrane potentials in AN2 in response to the
715 combination of sound (blue) and airflow (red) stimuli with various inter-
716 stimulus intervals. Positive value means that the onset of sound precedes the
717 onset of airflow. B–F) Relationships between the time lag between the sound
718 and airflow stimulus onset and firing rates normalized as Z-score (B), number
719 of bursts (C), maximum burst duration (D), or maximum burst size (E). The
720 values of the burst structure in (E) and (E) were measured from the largest
721 burst in the response if multiple bursts were evoked. F, G) Summation of
722 duration (F) and size (G) of all evoked bursts. Color indicates the data
723 recording from different individuals through Fig. 2-6 from B to G. The max.
724 duration and size of the burst were normalized as Z-score through D to G.

725 **Chapter III**

726 **Neural Mechanisms Underlying Multisensory Enhancement in AN2**

3.1 Introduction

In the previous chapter, I demonstrated the effect of multisensory integration on the spiking outputs of AN2. The cross-modal stimulus combining the sound pulse and airflow enhanced spiking activities including bursting activity. And, the linearity of summation of the multisensory integration depended on the magnitude of the evoked responses. As discussed in the previous chapter, it was possible that supra-linear summation would result from the non-linear transduction from graded potentials to action potentials. To reveal the cellular mechanisms relevant to multisensory enhancement in AN2, I focused on sub-threshold changes in the membrane potential evoked by uni- and cross-modal stimuli. Also, I examined the modulation of spike threshold that affects spike outputs. In several sensory neurons, fast membrane depolarization lowers the spike threshold, which is known as a dynamic threshold (Azouz and Gray, 2003; Wilent, 2005; Jeanne and Wilson, 2015). The dynamic threshold helps neurons to detect a coincidence of inputs (Azouz and Gray, 2003; Jeanne and Wilson, 2015).

In this chapter, I investigated the difference of subthreshold activity among modalities, and between bursting and isolated spikes. And I showed the modulation of spike threshold by temporal pattern of action potentials.

3.2 Materials and Methods

Data, which was obtained by the experiments shown in Chapter II, was used for the analysis. See Methods in Chapter II about *Animals*, *Intracellular recordings* and *Stimulation*.

Data analysis

A burst was defined as a cluster of spikes separated by short intervals (< 6.5 ms; Marsat and Pollack, 2006). This criterion was applicable to my data as shown by figure 2-4. The first and last spike within a burst were defined as the start and end of the burst, respectively. To analyze subthreshold voltage traces, raw traces of the recordings were filtered by a median filter with a 5-ms window (Shi et al., 2017). To focus on the time variation of EPSPs, the extracted post-synaptic potentials were normalized by the potentials at the peak time of the first spike.

I adopted the following procedure to detect spike thresholds (Jeanne and Wilson, 2015). After filtering using a moving-average filter with 150- μ m window, the spike threshold was defined as the membrane potential immediately preceding a spike that was detected as the inflection point in the plot of membrane potential and the slope of the potential, dV/dt . The inflection point was defined as the point where the d^2V/dt^2 fell below the 10% maximum of d^2V/dt^2 in the stereotypical spike trajectory. The rate of depolarization was calculated as the slope of the filtered voltage trace for 1.5

769 ms before the spike threshold. The inter-spike interval (*ISI*) of the n^{th} spike
770 within a spike train was calculated from the interval between the $(n - 1)^{th}$
771 and n^{th} spike.

772 MATLAB software (MathWorks, RRID:SCR_001622) was used for data
773 processing and computation. All values are reported as mean $\pm$ standard error
774 of the mean. A p -value of < 0.05 was considered to be statistically significant.

3.3 Results

3.3.1 Cross-modal stimulation maintains depolarization following initial spiking

First, I compared evoked EPSPs in response to cross- and unimodal stimuli. It was technically difficult to suppress the spike generation of AN2 by injecting minus current through a sharp electrode because of cell size and geometry. Instead, the EPSPs were extracted from traces of membrane potential changes using a 5-ms median filter (Fig. 3-1A). For EPSPs evoked by airflow, the membrane potential was transiently depolarized with longer latency and decreased for 150 to 200 ms after stimulus onset even though airflow was continuously applied (Fig. 3-1B). Cross-modal stimuli evoked larger EPSPs than either unimodal stimulus types did (Fig. 3-1B), but there was no significant difference in the peak value of EPSPs among the stimulus modalities (peak potentials for cross-modal, sound, and airflow stimulation were 16.04 ± 2.11 , 13.10 ± 2.28 , and 12.78 ± 1.60 mV, respectively; $n = 8$ neurons, one-way repeated measures ANOVA, $p = 0.31$). In contrast, the EPSPs evoked by cross-modal stimuli were significantly larger in their mean during the transient elevation (0-150 ms after the stimulus onset) than those evoked by the sound and airflow stimuli (Fig. 3-1C). However, EPSP evoked by the cross-modal stimuli were much smaller in both peak and mean of potential changes than the arithmetic sum of the EPSPs evoked by sound and airflow stimuli, implying that the subthreshold potentials were sub-linearly

summed in AN2.

As shown in figure 2-5C, the cross-modal stimuli enhanced spiking activity to evoke burst firing more frequently at the beginning of responses compared to unimodal stimuli, suggesting differences in depolarization following the first spikes in response to cross-modal versus unimodal stimuli. To examine this possibility, I aligned the EPSP traces with the peak of their first spikes and found that EPSPs evoked by cross-modal stimuli lasted longer and had more sustained depolarization compared to EPSPs evoked by sound or airflow alone (Fig. 3-2A). To quantify this, I measured the average value of normalized changes in membrane potential over 6.5 ms following the first spike (Fig. 3-2B). This corresponded to a border line of ISIs for separating bursts and isolated spikes and probably affected the burst firing and offset of the sustained peak (Fig. 3-2C), defined as the time required EPSPs to decrease from 100% to 90% of the normalized potential at the first spike (shown by the dashed line in Fig. 3-2A). Cross-modal stimuli evoked slightly larger depolarizations after the first spike compared to unimodal stimuli, but this difference was not significant (Fig. 3-2B). In contrast, the offset of peak for cross-modal stimuli was significantly delayed compared to those for unimodal stimuli (Fig. 3-2C). These results demonstrate that cross-modal stimuli evoke longer-lasting EPSPs than unimodal stimuli.

The followings two factors could be considered as causes resulting in the elongation of EPSP following the first spikes: the first was the time lag in the

response latency to the sound and airflow, the other was mechanical fluctuation of the stimulus onsets of the airflow. To test the first possibility, I measured the latency of the first spikes for each type of unimodal stimuli. The latency of the airflow-evoked spike was significantly longer than sound- and cross-modal stimulus-evoked spikes (Fig. 3-3A), meaning that the airflow responses were delayed to the auditory response. From this quantification of the response latencies, the time shift between the sound-evoked and airflow-evoked responses was 24.10 ± 15.81 ms ($n = 6$ neurons), which was longer than the offset of peak in EPSP evoked by the cross-modal stimuli (13.88 ± 3.61 ms shown in 4F, $n = 8$ neurons), but no significance (Wilcoxon rank sum test, $p = 0.57$). In addition, the variance of the peak offsets in the cross-modal responses was much smaller than that of the time shift between the auditory- and airflow-evoked first spikes (104.31 ms for the peak offset and 1500.18 ms for the time lag, respectively). These facts suggest that the long-lasting peak of the cross-modal responses would be partly resulted from the time difference of auditory and cercal inputs, but this would not be all cause for the EPSP elongation. In regard to the second possibility, I found no relationship between the time lag between the sound and airflow stimulus onsets and the depolarization size (Fig. 3-3B) or peak offset (Fig. 3-3C) in the EPSP evoked by cross-modal stimuli, indicating that mechanical lag of the unimodal stimuli had no effects on the EPSP shape.

3.3.2 Subthreshold activity accounts for burst firing in response to cross-modal stimuli in AN2.

Our results revealed differences in the time course of depolarization following the first spike in response to cross-modal and unimodal stimuli. However, it was unclear whether differences in EPSPs accounted for differences in burst firing. To investigate this hypothesis, I compared EPSPs accompanied by burst firing and by non-burst firing (isolated spikes). In responses to unimodal (sound) and cross-modal stimuli, both of which evoked bursts at the beginning of their responses (Fig. 2-5C), EPSPs accompanying bursts were larger than those accompanying isolated spikes (Fig. 3-4A1, A2). As shown in Fig. 3-2A, subthreshold potentials were normalized to the voltage at the base of the first spike and aligned with the peak time of the first spike (Fig. 3-4B1, B2), and the mean potentials over 6.5 ms following the first spike as well as the peak offset were measured. There were significant differences in mean potential changes between burst firing and isolated spikes evoked by unimodal and cross-modal stimuli (Fig. 3-4C). The peak offset for burst firing was also significantly delayed compared to that for isolated spikes in response to the unimodal and cross-modal stimuli (Fig. 3-4D). These results suggest that longer-lasting depolarization following the first spike accounted for the generation of burst firing regardless of sensory modality.

I further tested whether these parameters of the subthreshold potential are also correlated with the temporal size of bursts. Both the mean potential

change after the first spike and the peak offset were significantly correlated with burst duration. The correlation coefficient between the peak offset and the burst duration was higher (0.71) than that between the mean potential change and burst duration (0.33) (Fig. 3-5A, B). However, distribution of the plots for auditory and cross-modal stimuli overlapped each other. These results indicated that the size and persistence of the depolarization after individual spikes were key factors in determining burst generation, and that the persistence of depolarization could affect the duration of the burst firing. Further, both the size and persistence of EPSPs were enhanced not only by multisensory integration but also by higher stimulus intensity regardless of sensory modality.

3.3.3 Inter-spike interval modulates spike threshold but modality don't.

Spike output depends on the spike threshold in addition to the magnitude of EPSPs. It has been reported that in some sensory systems such as *Drosophila* olfactory neurons and mammalian visual cortical neurons, rapid membrane depolarization decreases the spike threshold, creating a dynamic threshold (Azouz and Gray, 2000, 2003; Jeanne and Wilson, 2015). To examine whether cross-modal stimuli modulates the spike threshold in AN2, I compared spike thresholds between the cross-modal and uni-modal stimulus conditions. I found that the spike threshold was dynamically altered, especially during burst firing (Fig. 3-6A, B); however, there was no difference in first spike

thresholds among the different stimulus modalities (Fig. 3-6C). Next, I examined which factors are related to the spike threshold. The rate of depolarization immediately preceding individual spikes was poorly correlated with the spike threshold (Fig. 3-6D). In contrast, inter-spike interval (ISI) was negatively correlated with the spike threshold regardless of the stimulation modality (Fig. 3-6E). In summary, spike threshold was dynamically altered in accordance with ISI in AN2, whereas the sensory modality had no effect on the spike threshold.

3.4 Discussion

I demonstrated that bimodal stimuli combining sound and airflow evoked linearly or supra-linearly summed spike outputs. In contrast, different modalities of sensory inputs were sub-linearly summed as reflected by EPSPs. Many studies on linearity in synaptic responses to cross-modal stimulation have revealed various types of computation including supra-linear, sub-linear, and linear summations. In neurons in the optic tectum of *Xenopus*, multimodal inputs are supra-linearly summed in the evoked EPSPs that result from NMDA receptor activation (Truszkowski et al., 2017). The linear summation of EPSPs is observed in the superior colliculus of rats (Skaliora et al., 2004), and sub-linear summation has been reported in the lamprey optic tectum and mouse neocortex (Olcese et al., 2013; Kardamakis et al., 2016). Thus, linearity of summation of EPSPs evoked by multimodal stimulation varies among the brain regions and animals. In previous studies, linear summation was observed as changes in membrane potential of a few millivolts from resting potentials (Skaliora et al. 2004), whereas sub-linear summation has been observed when multimodal stimuli evoke larger EPSPs close to the spike threshold (Olcese et al. 2013). In the present study, the analyzed EPSPs that were extracted from spiking responses were ‘over-threshold’ depolarizations by over 10 mV. Therefore, sub-linear summation in the synaptic potentials of AN2 might result from active properties of the cell membrane, which are mediated by voltage-gated cation channels.

In addition, I found the elongation of EPSP evoked by cross-modal stimuli that might result from time difference of auditory- and cercal-sensory inputs. AN2 receives direct excitatory inputs from the auditory receptor afferents (Hennig 1988) whereas cercal sensory inputs are mediated by at least 2 synapses that include synapses from mechanoreceptor afferents to cercal projection neurons such as GIs and that from those projection neurons to AN2. It is possible that the cercal sensory pathway may involve in other prothoracic interneurons. AN2 might apply the time difference of multisensory inputs to elongate the compound EPSP generating the burst firings.

The modulation of spike threshold has been observed in some sensory neurons (Azouz and Gray, 2003; Wilent, 2005). In these neurons, the rate of depolarization modulates the spike threshold. In AN2, the rate of depolarization and sensory modality did not affect the spike threshold whereas the spike threshold was modulated by the inter-spike interval. This intrinsic property might prevent the generation of inappropriate bursts.

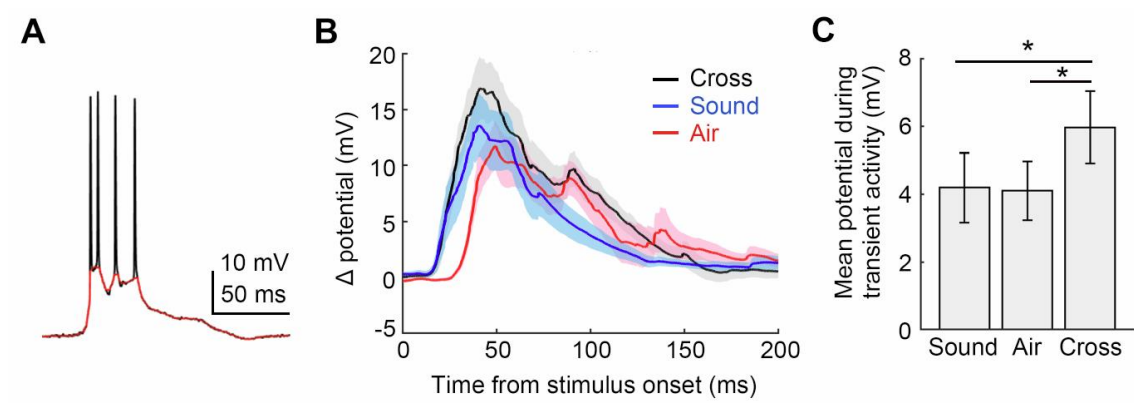
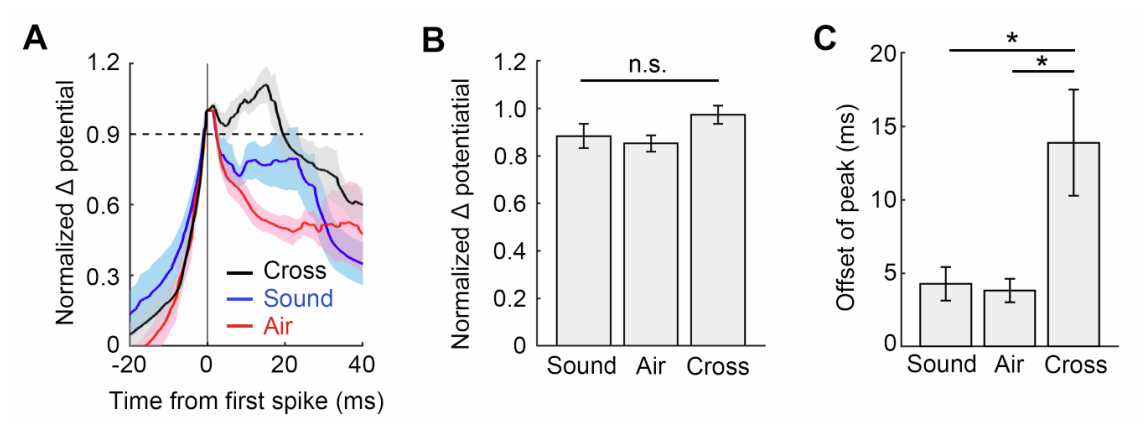


Figure 3-1. Excitatory post-synaptic potentials (EPSPs) evoked by cross-modal and unimodal stimuli. A) Extracted PSPs. Action potentials were removed from the raw trace of membrane potential (black trace) using a 5-ms median filter to estimate the graded potential of PSPs (red trace). B) Time variation in the membrane potential changes in EPSPs evoked by cross-modal (black), sound (blue), and airflow (red) stimuli in repetitive stimulation experiments (n = 8 neurons). C) Mean changes of membrane potential in EPSPs for 150 ms after the stimulus onset (n = 8 neurons, Friedman test, $p = 0.01$; Wilcoxon signed-rank test with Holm-Bonferroni correction, cross vs. sound, $p = 0.47 \times 10^{-1}$; cross vs. air, $p = 0.02$; sound vs. air, $p = 0.84$).



944 Figure 3-2. EPSPs relevant to first spike for stimulus. A) Time variation in
945 graded potential changes in EPSPs following the first spike. EPSP traces
946 were temporally aligned to peak time of the first spike and normalized in
947 magnitude by a value of the graded potentials at that moment ($n = 8, 7$, and
948 7 neurons for cross-modal, sound, and airflow, respectively). The dashed line
949 indicates 90% of the EPSP at the time of the first spike (0 ms). E, F) Mean
950 values of the normalized changes in membrane potential over 6.5 ms
951 following the first spike (E; $n = 8, 7$, and 7 neurons for cross-modal, sound,
952 and airflow, respectively. Kruskal-Wallis test, $p = 0.14$) and the offset of peak
953 EPSPs (F; $n = 8, 7$, and 7 neurons for cross-modal, sound, and airflow,
954 respectively, One-way factorial ANOVA, $p = 0.01$; unpaired t-test with Holm-
955 Bonferroni correction, cross vs. sound, $p = 0.02$; cross vs. air, $p = 0.02$; sound
956 vs. air, $p = 0.47$) evoked by different modalities of stimuli. The offset of peak
957 was measured as the time when EPSPs were $> 90\%$ of the normalized EPSP
958 indicated by the broken line in D.

959 Figure 3-3.

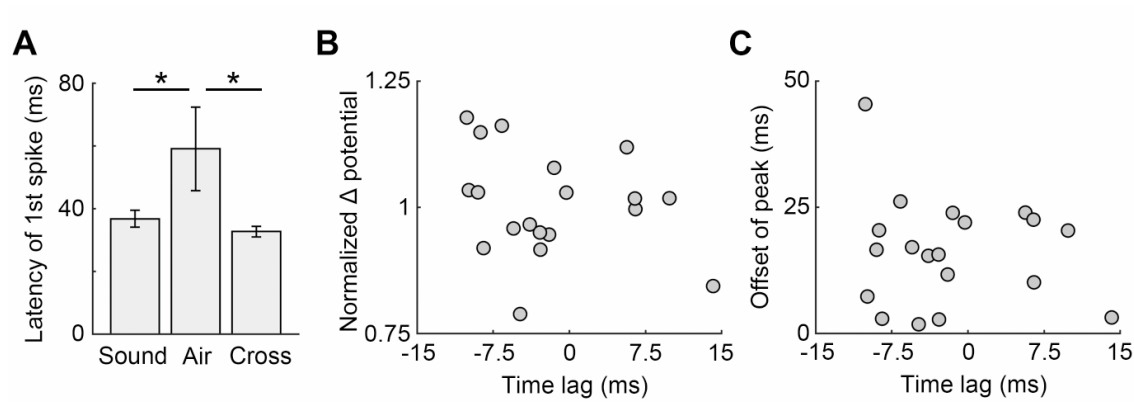


Figure 3-3. Effects of the time difference in response latency and in stimulus onset on EPSP's properties. A) Latency of the first spikes evoked by different types of stimulation from the stimulus onset. The spike latency of the airflow response was significantly longer than those of sound and cross-modal responses ($n = 7, 7$, and 8 neurons for sound, airflow and cross-modal stimulations, respectively, Kruskal-Wallis test, $p = 0.04 \times 10^{-1}$; Wilcoxon rank sum test with Holm-Bonferroni correction, sound vs. air, $p = 0.04$; cross vs. air, $p = 0.04 \times 10^{-1}$; sound vs. cross, $p = 0.28$). B, C) Relationships between the time lag between the sound and airflow stimulus onset and the normalize membrane potentials within 6.5 ms after the 1st spike peak (B) or the offset of peak (C) in EPSPs evoked by cross-modal stimuli ($n = 19$ responses from 8 neurons, Spearman's rank correlations, $r = -0.32$, $p = 0.18$ for B, $r = -0.04$, $p = 0.89$ for C). Negative value of the time lag indicates that the airflow preceded the combined sound stimulus onset.

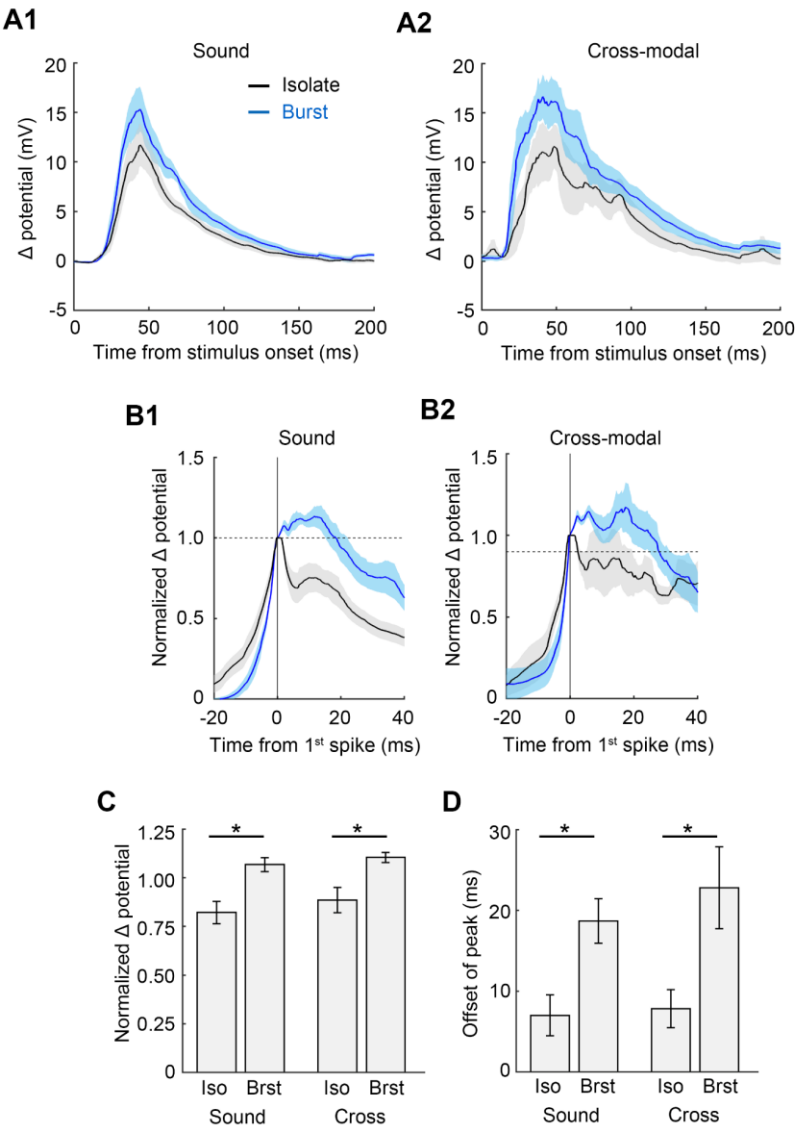
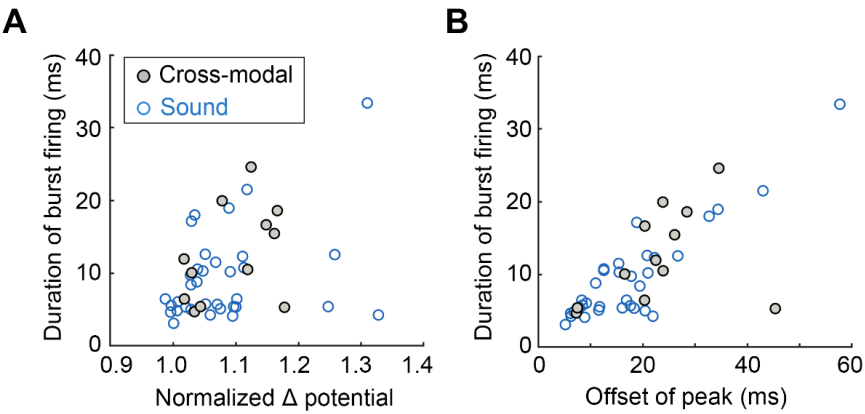


Figure 3-4. Excitatory post-synaptic potential (EPSP) characteristics affecting burst firing. A) Extracted EPSPs accompanied by isolated spikes (black) or burst firing (blue) in response to sound (A1) and cross-modal (A2) stimuli. All data used for Figure 3-4 were obtained from the responses to uni- and cross-modal stimuli using 60-dB sound in both the repetitive stimulation and various intensities of stimulation experiments. Sample sizes used for A to D were $n = 8$ and 5 neurons for isolated spikes and bursts evoked by sound stimuli, and $n = 9$ and 7 neurons for isolated spikes and bursts evoked by cross-modal stimulation, respectively. B) Time variation of the graded potential changes in EPSPs following the first spike evoked by sound (B1) or cross-modal (B2) stimuli. Colors of lines correspond to those in A. C, D) Mean values of the normalized changes in membrane potential for 6.5 ms after the first spike (C; Wilcoxon rank sum test, isolated vs. burst for sound stimuli, $p = 0.03 \times 10^{-1}$; isolated vs. burst for cross-modal stimuli, $p = 0.05 \times 10^{-1}$) and of the offset of peak in EPSPs (D; unpaired t-test, isolated vs. burst for sound stimuli, $p = 0.01$; isolated vs. burst for cross-modal stimuli, $p = 0.01$) accompanied by isolated spikes or bursts in response to sound and cross-modal stimuli.



995 Figure 3-5. EPSP characteristics relevant to structure of burst. A, B)
996 Relationship between the duration of burst firing and the mean of normalized
997 potential changes (A; $n = 46$ burst responses from 10 neurons, Spearman's
998 rank correlation, $r = 0.33$, $p = 0.02$) or the offset of peak EPSPs (B; sample
999 size was the same as in A, Spearman's rank correlation, $r = 0.71$; $p =$
1000 0.03×10^{-6}). Blue open circles represent data for sound stimuli and gray filled
1001 circles represent data for cross-modal stimuli.

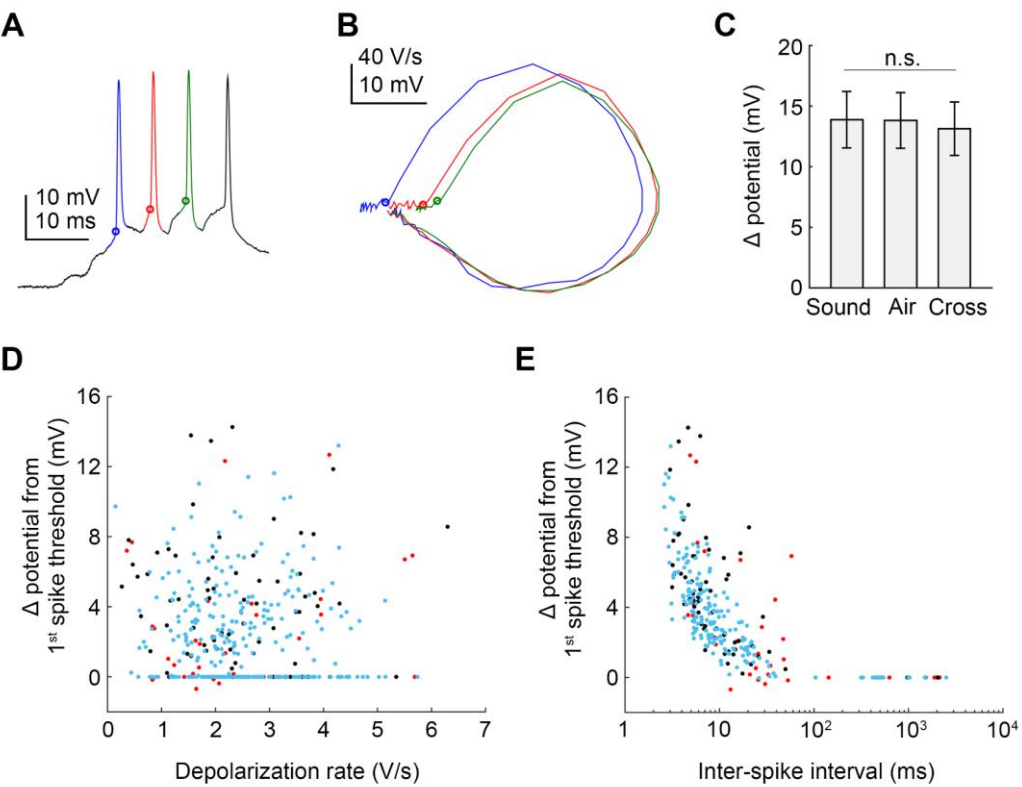


Figure 3-6. Modulation of the spike threshold in AN2. A) Trace of a typical firing response and estimated spike threshold potentials. Blue, red, and green circles indicate the threshold for the first, second, and third spikes, respectively. B) Phase portraits (dV/dt versus V) for the 3 spikes shown in A. Colors of traces correspond to those in A. C) Thresholds of the first spikes evoked by different modalities of stimuli ($n = 7, 7$ and 8 neurons; One-way factorial ANOVA, $p = 0.81$). Delta potentials of spike thresholds were the differences between the threshold potential and resting membrane potential before stimuli. D, E) Relationship between spike threshold and the preceding depolarization rate (D; Spearman's rank correlation, $r_{cross} = -0.20$, $r_{sound} = -0.07$, and $r_{airflow} = 0.05$; $p_{cross} = 0.08$, $p_{sound} = 0.22$, and $p_{airflow} = 0.73$) or inter-spike interval (E; Spearman's rank correlation, $r_{cross} = -0.84$, $r_{sound} = -0.92$, and $r_{airflow} = -0.68$; $p_{cross} = 1.59 \times 10^{-21}$, $p_{sound} = 1.98 \times 10^{-119}$, and $p_{airflow} = 4.00 \times 10^{-7}$). Sample size shown in D was the same as shown in E ($n = 293, 44$ and 78 spike thresholds from $8, 7$ and 8 neurons in sound, air and cross-modal stimulations respectively). The depolarization rate was measured as the slope of the change in membrane potential for 1.5 ms prior to each spike-threshold point. Delta (Δ) potential from first spike threshold was measured as the difference between the threshold potential and the first spike threshold. Colors of dots in D and E indicate data for cross-modal (black), sound (blue), and airflow (red) stimuli.

General Discussion

Multisensory integration is fundamental computation observed in various sensory systems and different brain regions (Meredith and Stein, 1986; Ghazanfar and Schroeder, 2006; Deeg et al., 2009; Zahar et al., 2009; Kardamakis et al., 2016). Multisensory integration is performed in not only higher processing stage for association but also earlier stages close to sensory surface such as the primary auditory cortex of macaque and the primary visual cortex of mouse (Schroeder and Foxe, 2005; Ghazanfar and Chandrasekaran, 2007; Lakatos et al., 2007; Ibrahim et al., 2016). In *Drosophila* larvae, certain sensory neurons directly receive sensory inputs from receptor cells of different modalities (Ohyama et al., 2015). The prothoracic auditory circuit of crickets is also such a sensory system to integrate various modalities in the early stage as discussed in Chapter I. These findings imply that multisensory integration is fundamental mechanisms in early stage of sensory processing.

Finally, I refer to the behavioral relevance of multisensory integration in AN2. AN2 is considered to play a crucial role in the detection of predatory bats, because it is sensitive to ultrasound-like echolocation calls of bats, and its firing activity is necessary and sufficient to trigger avoidance steering in the opposite direction of the sound source (Nolen and Hoy, 1984, 1986). Notably, bursting of AN2 correlates with abdominal movement in the avoidance steering behavior (Marsat and Pollack 2006, 2010). I demonstrated

that AN2 generates bursting in response to 15-kHz tone pulses of which the carrier frequency was lower than that used in previous studies (Marsat and Pollack 2006, 2007, 2010), and that cross-modal stimuli combining the tone pulse with airflow enhanced the generation of bursting. AN2 represents directionality, intensity, and duration of sound stimuli with its bursting (Marsat and Pollack, 2006, 2010). Our results, therefore, suggest that multisensory integration in AN2 may enable quicker and more precise identification of alert signals via simultaneous detection of the searching echo and air flow caused by flying bats.

The avoidance steering driven by the bursts of the AN2 are observed only in response to ultrasound during flight. So, it remains unclear whether flying crickets use wind stimuli as a sign of a bat's attack. Several types of insects, such as mantids and cockroaches, detect the predator's attack by their cercal organs and execute avoidance steering during flight (Ganihar et al., 1994; Tribblehorn and Yager, 2006). It is possible that the wind sensitivity of AN2 might be useful for successful escape via multisensory integration with auditory inputs. On the other hand, crickets standing on the ground exhibit escape behaviors such as running or jumping in response to wind stimuli (Tauber and Camhi, 1995; Oe and Ogawa, 2013; Sato et al., 2017). AN2 is also involved in the phonotaxis via detection of conspecific calls (Schildberger and Hörner, 1988), meaning that AN2 could respond to ultrasound and also to airflow in standing crickets. Recently, I reported that

1069 10- or 15-kHz pure tones that will activate AN2 modulate wind-elicited escape
1070 behavior in crickets (Fukutomi et al., 2015; Fukutomi and Ogawa, 2017).
1071 Taken together, the findings in this study suggest that AN2 is an excellent
1072 model for understanding the neuroethological significance of multisensory
1073 integration.

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1288 Someya M, Ogawa H (2017) Coincident multisensory inputs enhance burst

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