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**Knockdown of *DDX4* decreases the number of germ cells in male and female chicken embryonic gonads**

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Short title: Effects of *DDX4* KD on chicken embryonic gonads

Key words: primordial germ cells, sex differentiation, ovary, testis

## 20    **Abstract**

21    DEAD-box helicase 4 (*DDX4*, also known as *vasa*) is essential for the proper formation and  
22    maintenance of germ cells. Although *DDX4* is conserved in a variety of vertebrates and  
23    invertebrates, its roles differ between species. This study investigated the function of *DDX4* in  
24    chicken embryos by knocking down its expression using retroviral vectors that encoded  
25    *DDX4*-targeting microRNAs. *DDX4* was effectively depleted *in vitro* and *in vivo* via this  
26    approach. Male and female gonads of *DDX4*-knockdown embryos contained a decreased  
27    number of primordial germ cells, indicating that *DDX4* is essential to maintain a normal level of  
28    these cells in chicken embryos of both sexes. *DMRT1* and *SOX9*, which are involved in testis  
29    determination and differentiation, were expressed as normal in male gonads of  
30    *DDX4*-knockdown embryos. By contrast, expression of *CYP19A1*, which encodes aromatase  
31    and is essential for ovary development, was significantly decreased in female gonads of  
32    *DDX4*-knockdown embryos. Expression of *FOXL2*, which plays an important role in ovary  
33    differentiation, was also slightly reduced, but this was not statistically significant. *FOXL2* was  
34    previously hypothesized to regulate aromatase expression based on several pieces of evidence.  
35    These results indicate that aromatase expression is also regulated by several additional  
36    pathways.

## 37    **Introduction**

38    DEAD-box helicase 4 (DDX4, also called vasa) is a member of the DEAD-box protein family,  
39    which is important for the proper formation of germ cells. The roles of DDX4 are reported to  
40    differ between species. Expression of DDX4 has been depleted in *Drosophila melanogaster*  
41    (Styhler et al, 1998), *Caenorhabditis elegans* (Roussell and Bennett, 1993; Gruidl et al, 1996;  
42    Kawasaki et al, 1998; Kuznicki et al, 2000; Spike et al, 2008), mice (Tanaka et al, 2000), and  
43    zebrafish (Braat et al, 2001) via gene knockout (KO), RNA interference (RNAi), and  
44    morpholino treatment. DDX4 is essential for oogenesis in most species and is necessary for  
45    male germ cells to progress through spermatogenesis in several species, including mice (Tanaka  
46    et al, 2000; Kuramochi-Miyagawa et al, 2010; Ewen-Campen et al, 2013). These diverse  
47    functions of DDX4 evolved for reproductive adaptation.

48        The chicken *DDX4* homolog (also called chicken vasa homologue, *CVH*) was first cloned  
49    by Tsunekawa et al. (2000). This gene encodes 663 amino acids and is located on the Z  
50    chromosome (male: ZZ, female: ZW). DDX4 protein localizes to the cytoplasm of germ cells,  
51    including presumptive primordial germ cells (PGCs) in uterine-stage embryos and spermatids  
52    and oocytes in adult gonads. These findings indicate that the chicken germ lineage is maternally  
53    predetermined (Tsunekawa et al 2000). Taylor et al. (2017) efficiently knocked out the *DDX4*

locus via homologous recombination mediated by transcription activator-like effector nucleases. Targeted male PGCs were injected into surrogate host chicken embryos. Seventeen targeted G1 offspring were generated following mating of a host male cockerel with wild-type (WT) hens. G1 females (hemizygous KO chickens) were sterile and did not exhibit detectable follicles post-hatching. The germ cell lineage was initially formed in early embryos, but PGCs were subsequently lost during meiosis. These findings indicate that *DDX4* plays an important role in the female reproductive system. By contrast, homozygous (null) male KO chickens could not be generated because it was impossible to cross G1 female and male KO chickens (Taylor et al, 2017). Therefore, the function of *DDX4* in male PGCs remains unclear in chickens.

Herein, we show that *DDX4* is essential for PGCs in chicken embryos of both sexes. We knocked down *DDX4* using retroviral vectors that encoded *DDX4*-targeting microRNAs (miRNAs). There was a decreased number of PGCs in male and female gonads of *DDX4*-knockdown (KD) embryos. *DMRT1* and *SOX9*, which are involved in testis determination and differentiation, were expressed as normal in male gonads of *DDX4*-KD embryos. By contrast, expression of *CYP19A1*, which encodes aromatase, was significantly decreased in female gonads of *DDX4*-KD embryos.

## 71    **Materials and Methods**

### 72    ***Animals and ethics statement***

73    Fertilized chicken eggs (*Gallus gallus domesticus*) were purchased from Takeuchi Hatchery  
74    (Nara, Japan). The Hy-Line Maria chicken strain was used in this study. Fertilized eggs were  
75    incubated at 37.8°C. The sex of each embryo was determined by PCR genotyping using  
76    genomic DNA as the template (Fridolfsson & Ellegren 1999).

77            All animal experiments described in this study were approved by the Institutional Animal  
78    Care and Use Committee of National University Corporation Hokkaido University and were  
79    performed in accordance with the Guidelines for the Care and Use of Laboratory Animals  
80    issued by Hokkaido University. This study did not involve any human participants or  
81    specimens.

### 83    ***Construction of RNAi vectors***

84    Two miRNAs targeting different regions of chicken *DDX4* mRNA and two non-targeting  
85    (scrambled) controls were designed using BLOCK-iT RNAi Designer (Thermo Fisher Scientific,  
86    MA) and siRNA Wizard v3.1 (Thermo Fisher Scientific) (Table 1). The miRNA constructs were  
87    cloned as described previously (Tanaka et al., 2017). pRFPRNAi.C and RCASRNAi were

provided by ARK-Genomics, The Roslin Institute (Das et al, 2006). The vectors encoding *DDX4*-targeting and scrambled miRNAs were named RCASA.miRNA.DDX4.eGFP and RCASA.miRNA.Sc.eGFP, respectively.

To test the KD efficiency using DF-1 cells, RCASA.miRNA.DDX4 and RCASA.miRNA.Sc, which lacked the eGFP sequence, were constructed. A vector expressing *DDX4* fused to GFP was also generated for *in vitro* experiments. A forward primer containing a *Cla*I site and a reverse primer containing a *Spe*I site were designed to amplify the coding DNA sequence of *DDX4* (Table 1). Following digestion with *Cla*I and *Spe*I, the PCR product was cloned into the eGFP-harboring RCAS.B vector that had been digested with the same restriction enzymes. This construct was named RCASB.DDX4\_eGFP. RCAS.B proviral DNA, RCAS.A, and RCAS.B were kindly provided by Dr. Hughes, National Cancer Institute (Hughes et al, 1987).

#### ***Preparation and injection of viruses***

Endotoxin-free proviral DNA was prepared using a PureYield™ Plasmid Miniprep Kit (Promega, WI). DF-1 cells were transfected with this DNA using Lipofectamine 2000 (Thermo Fisher Scientific ) according to the manufacturer's protocol, transferred to 10-mm dishes

(surface area of 78.5 cm<sup>2</sup>), and cultured to sub-confluency in Dulbecco's Modified Eagle's Medium supplemented with 10% (v/v) fetal bovine serum. Active viruses in the pooled medium were concentrated. Briefly, 100 ml of medium was centrifuged overnight at 6,000 × *g*, and then the supernatant was carefully decanted to leave a small pellet in 200 µl of medium. The resuspended viral solution was aliquoted and stored at -80°C until use. The viral titer was determined as described previously (Nakata et al, 2013).

Embryos were injected as described previously (Nakata et al, 2013) with minor modifications. Approximately 3 µl of concentrated viruses containing 0.025% Fast Green tracking dye was injected into the subgerminal cavity of blastoderms at day 0 (Hamburger and Hamilton stage X; Hamburger & Hamilton 1951). Viruses carrying RCASA.miRNA.DDX4 were injected into 189 eggs to KD *DDX4*. The eggs were sealed and incubated until embryonic day (E)8.5. The number of embryos developed at ~E8.5 are 18 (ZW) and 15 (ZZ).

#### ***Reverse-transcription polymerase chain reaction (RT-PCR) and quantitative real-time polymerase chain reaction (qRT-PCR)***

For RT-PCR, total RNA was extracted from DF-1 cells infected with RCASB.DDX4\_eGFP or with RCASA.miRNA.DDX4 followed by RCASB.DDX4\_eGFP. For qRT-PCR, total RNA was

extracted from gonads of male and female *DDX4*-KD embryos infected with RCASA.miRNA.*DDX4*.eGFP and from gonads of male and female WT embryos at E8.5. Total RNA was extracted using an RNeasy Kit (Invitrogen) according to the manufacturer's instructions. RNA was treated with DNase I and reverse-transcribed using SuperScript III reverse transcriptase (Invitrogen) and an oligo(dT) primer. qRT-PCR was performed using Power SYBR® Green PCR Master Mix (Thermo Fisher Scientific) and an ABI 7300 Fast Real-Time PCR System (Thermo Fisher Scientific). To ensure the amplification efficiency was maximal in all reactions, the amplicon size was restricted to 90–101 bp. Reactions were performed in triplicate using 96-well plates in a reaction volume of 10 µl. Data were analyzed using the  $\Delta\Delta C_t$  method, and the mRNA level of each target gene was normalized against that of beta-actin (*ACTB*). The primers used to quantify *DDX4* expression are listed in Table 1, while those used to measure the expression levels of other genes were previously described (Nakata et al, 2013).

### ***Immunohistochemistry***

Urogenital tissues of chicken embryos were fixed in formalin for 1 h at room temperature. Paraffin sections (10 µm thick) were cut along the dorsal-ventral axis and thaw-mounted onto

139 MAS-coated glass slides. The experimental conditions of immunohistochemistry were  
140 described previously (Nakata et al, 2013). The anti-DDX4 antibody was kindly provided by Dr.  
141 Hattori, Kyushu University (Aramaki et al, 2009).

142

### 143 ***Alkaline phosphatase (AP) staining***

144 Urogenital tissues of chicken embryos were fixed in 4% paraformaldehyde prepared in  
145 phosphate-buffered saline (PBS). Serial paraffin sections (10 µm thick) were cut along the  
146 anterior-posterior axis and mounted onto MAS-coated glass slides. De-waxed slides were  
147 treated with PBS containing 0.5% Triton X-100 for 20 min and incubated with the NBT/BCIP  
148 substrate (Promega).

149

### 150 ***In situ hybridization***

151 A fragment of DND microRNA-mediated repression inhibitor 1 (*DND1*, also known as *CDH*)  
152 was amplified by RT-PCR using cDNA obtained from gonads at E8.5 as the template. The  
153 primers are listed in Table 1. RNA extraction and cDNA synthesis were performed as described  
154 earlier. The PCR product of 568 bp was subcloned using a pGEM T-Easy vector system  
155 (Promega). cDNA clones were labeled using Digoxigenin RNA Labeling Mix (Roche, Basel,

156 Switzerland) and T7 or SP6 RNA polymerase (MAXIscript™; Thermo Fisher Scientific).  
157 Hybridization to serial frozen sections was performed as described previously (Nakata et al,  
158 2013). The images were captured using a cooled CCD camera (DS-Ri1, Nikon corporation,  
159 Tokyo, Japan) mounted on a Nikon ECLIPSE E800 microscope, and were analyzed with the  
160 NIS ELEMENTS application program of Nikon corporation.

161

## 162 ***Statistical analysis***

163 The data are presented as mean  $\pm$  standard deviation. Statistical comparison was made by  
164 Student *t* test.  $P < 0.005$  was considered statistically significant.

165

## 166 **Results**

### 167 ***DDX4-targeting miRNAs efficiently deplete DDX4***

168 We first examined the efficiency of *DDX4* KD using *DDX4*-targeting miRNAs in DF-1 cells.  
169 These cells were infected with RCASB.DDX4\_eGFP (Fig. 1a, b), an RCAS vector expressing  
170 *DDX4* fused to eGFP, or with RCASA.miRNA.Sc followed by RCASB.DDX4\_eGFP (Fig. 1c,  
171 d) as controls. eGFP was observed in both cases. However, eGFP was not detected in DF-1 cells  
172 infected with RCASA.miRNA.DDX4, which encoded two *DDX4*-targeting miRNAs, prior to

173 RCASB.DDX4\_eGFP (Fig. 1e, f). In addition, *DDX4* was detected by RT-PCR in DF-1 cells  
174 infected with RCASB.DDX4\_eGFP, but not in those infected with RCASA.miRNA.DDX4  
175 prior to RCASB.DDX4\_eGFP (Fig. 1g).

176 We performed immunohistochemical staining of *DDX4* in gonads of *DDX4*-KD embryos  
177 at E8.5. *DDX4* staining was reduced in female (Fig. 2a–d) and male (Fig. 2e–h) gonads of  
178 *DDX4*-KD embryos. *DDX4* expression in gonads of WT and *DDX4*-KD embryos at E8.5 was  
179 quantified by qRT-PCR. This demonstrated that *DDX4* expression was significantly reduced in  
180 *DDX4*-KD embryos of both sexes (Fig. 2i). These results demonstrate that *DDX4*-targeting  
181 miRNAs efficiently deplete *DDX4*.

182

### 183 ***The number of PGCs is decreased in male and female gonads of DDX4-KD embryos***

184 We performed AP staining and *DND1* *in situ* hybridization to label PGCs in gonads of  
185 *DDX4*-KD embryos. PGCs exhibit AP staining in many species. *DND1* co-localizes with *DDX4*  
186 in germ cell lines generated from chicken embryonic gonads (Kito et al 2010) and is thus a  
187 marker of PGCs.

188 PGCs were detected by AP staining in female and male gonads of WT embryos at E8.5  
189 (Fig. 3a, b). However, AP staining was not clearly observed in *DDX4*-KD embryos of either sex

(Fig. 3c, d). The *DND1 in situ* hybridization experiments yielded similar results (Fig. 3e–h).

The total number of PGCs was counted in serial sections subjected to AP staining (Fig. 3i) and *DND1 in situ* hybridization (Fig. 3j). Based on AP staining and *DND1 in situ* hybridization, the number of PGCs was decreased by 74% and 79% in *DDX4*-KD female embryos, respectively, and by 61.4% and 62.2% in *DDX4*-KD male embryos, respectively. The detection sensitivity of each method is different: the sensitivity of AP staining is relatively higher than that of *DND1 in situ* hybridization. Although total number of PGCs detected by AP-staining is larger than *DND1 in situ* hybridization, the reduction rate of number of PGCs is almost same between both methods.

#### ***KD of DDX4 decreases CYP19A1 expression***

We quantified expression of *DMRT1*, *SOX9*, *FOXL2*, and *CYP19A1*, which are involved in gonadal differentiation, in gonads of WT and *DDX4*-KD embryos at E8.5 (Fig. 4). In both WT and *DDX4*-KD embryos, expression of *DMRT1* and *SOX9* was high in male gonads, but low in female gonads (Fig. 4), consistent with previous reports (Smith et al, 1999a; 1999b). Expression of *DMRT1* and *SOX9* did not significantly differ between WT and *DDX4*-KD embryos.

Expression of *FOXL2* and *CYP19A1* was reported to be high in female embryonic gonads,

but extremely low in male embryonic gonads (Govoroun et al, 2004). We confirmed this finding in WT embryos. However, *CYP19A1* expression in female gonads was significantly lower in *DDX4*-KD embryos than in WT embryos (Fig. 4). Expression of *FOXL2* was slightly lower in female gonads of *DDX4*-KD embryos than in those of WT embryos, but this difference was not statistically significant.

## Discussion

The *DDX4* gene is conserved in a variety of vertebrates and invertebrates. Although *DDX4* is essential for the development and maintenance of germ cells in a wide range of species, its roles differ between species. A null mutation of *DDX4* in *D. melanogaster*, in which this gene was initially identified, causes female sterility due to severe defects in oogenesis, while males remain fertile (Styhler et al, 1998). However, male *DDX4*-KO mice exhibit a reproductive deficiency; male homozygotes do not produce sperm in the testes, and the proliferation of PGCs is hampered (Tanaka et al, 2000). By contrast, disruption of *DDX4* does not affect the development of female germ cells in mice, despite the fact that this gene is expressed in both male and female germ cells of mice (Tanaka et al, 2000). The current study demonstrated that *DDX4*-targeting miRNAs effectively suppressed *DDX4* expression *in vitro* and *in vivo* (Fig. 1,

2). The number of proliferative cells detected by EdU is significantly reduced in female gonads of *DDX4*-KO chicken embryos at E10.5, indicating that germ cells form at an early developmental stage and are subsequently lost (Taylor et al, 2017). Our results confirmed this previous finding and demonstrated that the number of PGCs was also reduced in male gonads of *DDX4*-KD embryos (Fig. 3). Therefore, *DDX4* is also involved in the proliferation of PGCs in males, suggesting it is important for both male and female fertility in chickens.

We investigated the expression levels of genes involved in gonadal differentiation and development in *DDX4*-KD embryos (Fig. 4). Expression of *DMRT1* and *SOX9*, which are markers of testis determination and differentiation, were quantified. *DMRT1* is located on the Z chromosome in birds, is a major avian male-determining factor, and begins to be expressed at ~E3.5. Expression of *DMRT1* is detected in the medulla of gonads and is higher in males than in females. Upon high *DMRT1* expression, *SOX9* is continuously expressed for testis development in male chicken embryos after E6.5. Expression of *DMRT1* and *SOX9* was higher in male gonads than in female gonads of both *DDX4*-KD and WT embryos (Fig. 4). This suggests that testis differentiation and development occurred as normal in male gonads with a decreased number of PGCs.

By contrast, expression of *CYP19A1* was significantly lower in female gonads of

241 *DDX4*-KD embryos than in those of WT embryos (Fig. 4). Gonadal sex differentiation in female  
242 birds is sensitive to the sex steroid hormone estradiol. This hormone is only detected in female  
243 embryonic gonads and is required and sufficient for ovarian development (Elbrecht & Smith  
244 1992). Aromatase converts androgens into estradiol. Consequently, *CYP19A1* is essential for the  
245 female reproductive system. Indeed, treatment with aromatase inhibitors, such as fadrozole, can  
246 effectively inhibit to feminization in ZW embryos (Elbrecht & Smith 1992). Aromatase  
247 localizes to the cytoplasm of somatic cells in the medulla of female embryonic gonads from  
248 E6.5 onwards and its expression increases during ovarian development (Govoroun et al, 2004;  
249 Smith et al. 2005). *DDX4* is thought to increase the translation of several proteins required for  
250 meiotic progression and the assembly of cytoplasmic granules in germ cells. The  
251 ATP-dependent catalytic activity of RNA helicases regulates the translation of multiple mRNAs  
252 (Carrera et al, 2000). Considering its expression pattern and function, *DDX4* is not thought to  
253 directly regulate expression of *CYP19A1*. Repression of the *DDX4* expression and/or a  
254 decreased number of PGCs might indirectly affect aromatase expression.

255 Expression of *FOXL2*, another marker of ovary differentiation, was slightly lower in  
256 female gonads of *DDX4*-KD embryos than in those of WT embryos; however, this difference  
257 was not statistically significant (Fig. 4). *FOXL2* is an essential factor that is widely conserved in

vertebrates, including chickens (Loffler et al, 2003; Wang et al, 2007; Pisarska et al, 2011). The expression patterns of *FOXL2* and *CYP19A1* highly correlate with each other in the developing ovary after 4.7–12.7 days of incubation (Govoroun et al. 2004). The proteins encoded by these genes co-localize at the nuclei of medullar cord cells in female embryonic gonads. *FOXL2* is expressed just prior to *CYP19A1*, suggesting it directly or indirectly regulates transcription of *CYP19A1*, at least in embryos. This hypothesis is supported by the finding that *FOXL2* expression is induced in aromatase-overexpressing male embryos (Lambeth et al, 2013). *FOXL2*-binding sites (5'-CACAACA-3') are located in the promoter region of *CYP19A1* (-668 and -3453 bp in the antisense strand and -3642 bp in the sense strand; Govoroun et al, 2004). However, *FOXL2* fails to upregulate *CYP19A1* expression in luciferase assays (Wang and Gong, 2017). The expression patterns of *FOXL2* and aromatase differ in the ovaries of adult chickens, with the former protein localizing to the granulosa cell layer and the latter protein localizing to the theca cell layers, indicating that the regulatory mechanism of aromatase differs between embryos and adults (Wang and Gong, 2017). Our data suggest that several pathways regulate expression of *CYP19A1*, in addition to that involving *FOXL2*. Furthermore, treatment with an aromatase inhibitor reduces *FOXL2* expression *in vivo*, suggesting that a regulatory feedback loop exists between *FOXL2* and *CYP19A1* (Hudson et al, 2005). Therefore, the slight reduction

275 in *FOXL2* expression observed in female gonads of *DDX4*-KD embryos might be due to the  
276 substantial decrease in *CYP19A1* expression.

277 In summary, this study demonstrates that *DDX4* is important for the female and male  
278 reproductive systems of chickens. Furthermore, our data indicate repression of the *DDX4*  
279 expression and/or a decreased number of PGCs affect the regulation of *CYP19A1* expression.

280

#### 281 **Declaration of interest**

282 The authors declare that no competing interests exist.

283

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287

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290 culture experiments, and M.-A. Hattori for providing the anti-*DDX4* antibody.

291

292 **Authors' contributions**

293 NA performed all the experiments and analyzed all the data. HI constructed the retroviral  
294 vectors. AK conceived and designed the study and SM and AK wrote the manuscript. All  
295 authors read and approved the final manuscript.

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393

## Figure legends

### Figure 1 *DDX4* expression is efficiently suppressed *in vitro*

DF-1 cells were infected with (a, b) RCASB.DDX4\_eGFP alone, (c, d) RCASA.miRNA.Sc and RCASB.DDX4\_eGFP, or (e, f) RCASA.miRNA.DDX4 and RCASB.DDX4\_eGFP. Although eGFP was observed in both cases (a-d), eGFP was not detected in DF-1 cells infected with RCASA.miRNA.DDX4, which encoded two *DDX4*-targeting miRNAs, prior to RCASB.DDX4\_eGFP (e, f). Scale bar, 100  $\mu$ m. (g) RT-PCR analysis of *DDX4* in DF-1 cells infected with RCASB.DDX4\_eGFP alone or together with RCASA.miRNA.DDX4. The *DDX4* expression is not detected in DF-1 cells infected with RCASA.miRNA.DDX4 prior to RCASB.DDX4\_eGFP (g). M: molecular marker.

### Figure 2 *DDX4* expression is efficiently suppressed *in vivo*

(a–h) Immunohistochemical staining of *DDX4* in the left gonads of (a–d) female and (e–h) male embryos at E8.5 was performed to examine the KD efficiency of *DDX4*-targeting miRNAs. (a, b) Female and (e, f) male gonads of WT embryos at E8.5 exhibited normal expression of *DDX4*, indicating that PGCs localized to the cortical and interior of the embryonic gonad, respectively. *DDX4* expression was suppressed in (c, d) female and (g, h) male embryos infected with

411 RCASA.miRNA.DDX4. FITC fluorescence is shown in a, c, e, g. Hoechst staining is shown in  
412 B, D, F, H. Scale bar, 100  $\mu$ m. (i) qRT-PCR analysis of *DDX4* in gonads at E8.5. Delivery of  
413 *DDX4*-targeting miRNAs effectively suppressed *DDX4* expression. Black, dark gray, white, and  
414 light gray bars correspond to WT female, *DDX4*-KD female, WT male, and *DDX4*-KD male  
415 embryos, respectively. The data are presented as mean  $\pm$  standard deviation. \* $P < 0.005$ ;  $n \geq 3$ .

416

417 **Figure 3 The number of PGCs is decreased in gonads of *DDX4*-KD embryos**

418 (a–d) AP staining in gonads of WT and KD embryos at E8.5. Gonad sections (dorsal-ventral  
419 axis) of (a) WT female, (b) WT male, (c) *DDX4*-KD female, and (d) *DDX4*-KD male embryos.

420 Marked staining was observed in female and male gonads of WT embryos (a, b), but not in  
421 those of *DDX4*-KD embryos (c, d). Arrows indicate AP staining. Scale bar, 100  $\mu$ m. (e–g)

422 *DND1* *in situ* hybridization in frozen gonad sections of WT and KD embryo at E8.5. Gonad

423 sections of (e) WT female, (f) WT male, (g) *DDX4*-KD female, and (h) *DDX4*-KD male

424 embryos. Marked staining was observed in female and male gonads of WT embryos (E, F), but

425 not in those of *DDX4*-KD embryos (g, h). Arrows indicate *DND1* staining. Scale bar, 100  $\mu$ m.

426 (i) AP staining and (j) *DND1* *in situ* hybridization. Black, dark gray, white, and light gray bars

427 show the numbers of positively stained cells in gonads of WT female, *DDX4*-KD female, WT

428 male, and *DDX4*-KD male embryos at E8.5, respectively.

429

430 **Figure 4 KD of *DDX4* decreases *CYP19A1* expression in female embryonic gonads**

431 qRT-PCR analysis of *CYP19A1*, *FOXL2*, *DMRT1*, and *SOX9* expression in gonads of WT

432 female (black bars), *DDX4*-KD female (dark gray bars), WT male (white bars), and *DDX4*-KD

433 male (light gray bars) embryos at E8.5. Expression of *DMRT1* and *SOX9* did not significantly

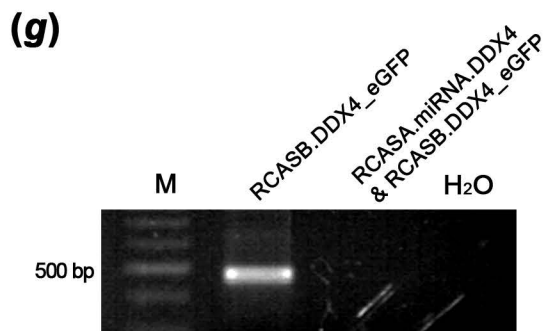
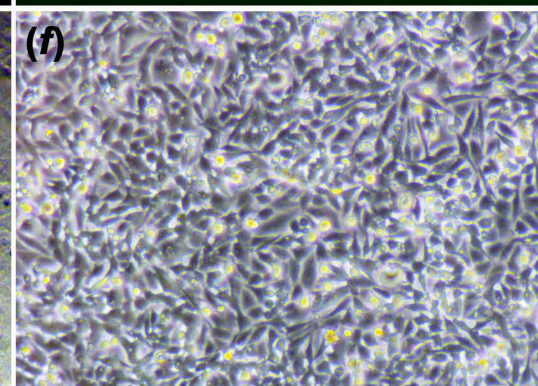
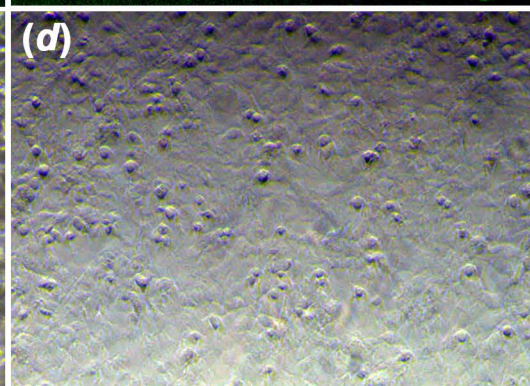
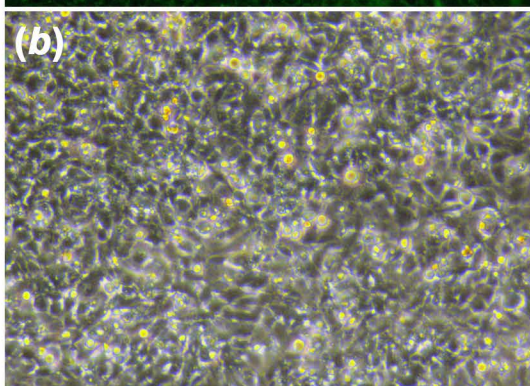
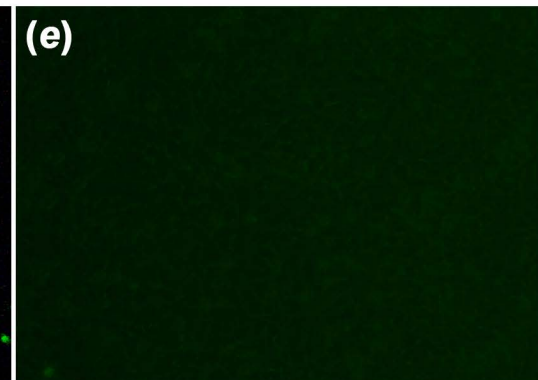
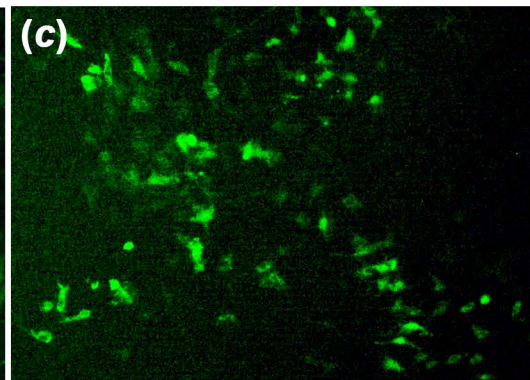
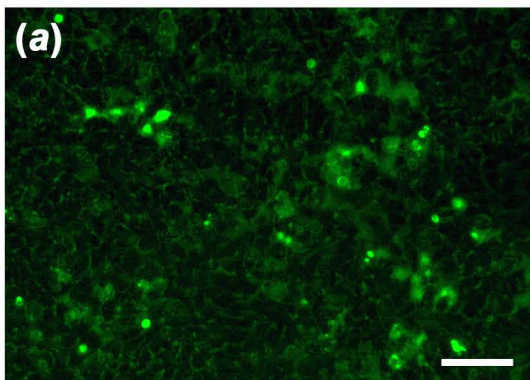
434 differ between WT and *DDX4*-KD embryos. *CYP19A1* expression in female gonads was

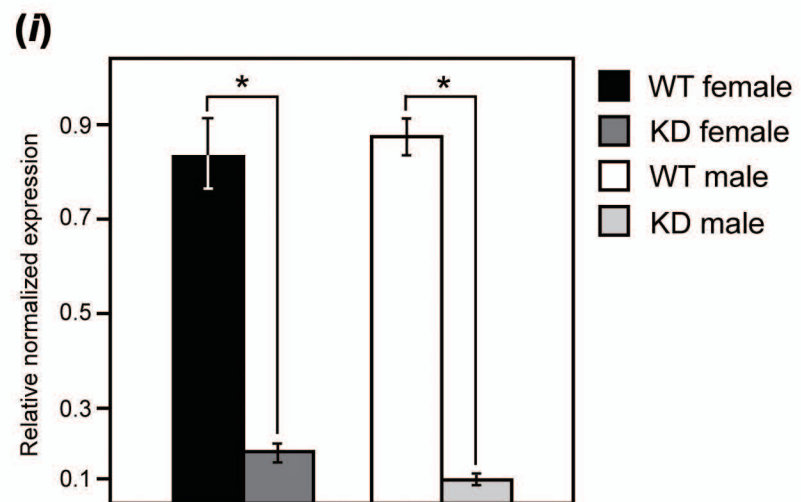
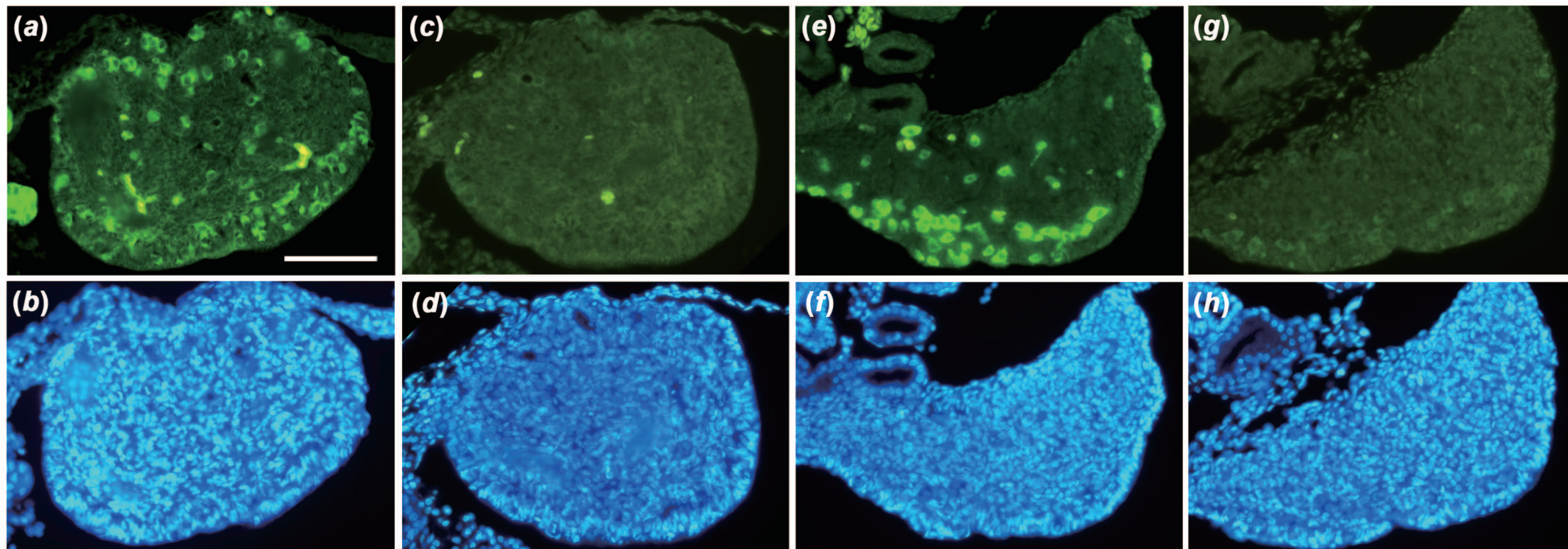
435 significantly lower in *DDX4*-KD embryos than in WT embryos. Expression of *FOXL2* was

436 slightly lower in female gonads of *DDX4*-KD embryos than in those of WT embryos, but this

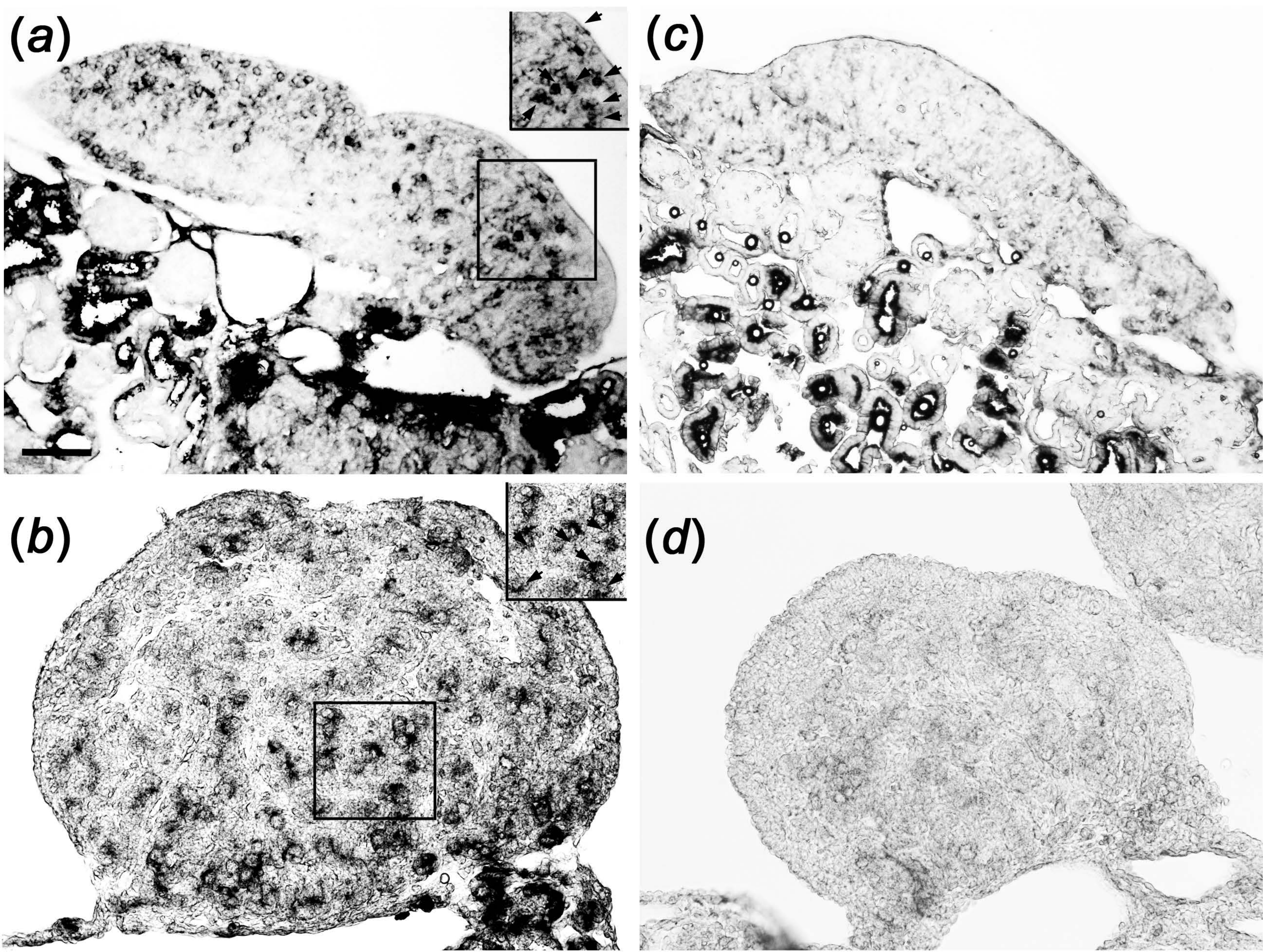
437 difference was not statistically significant. The data are presented as mean  $\pm$  standard deviation.

438 \* $P < 0.005$ ;  $n \geq 3$ .

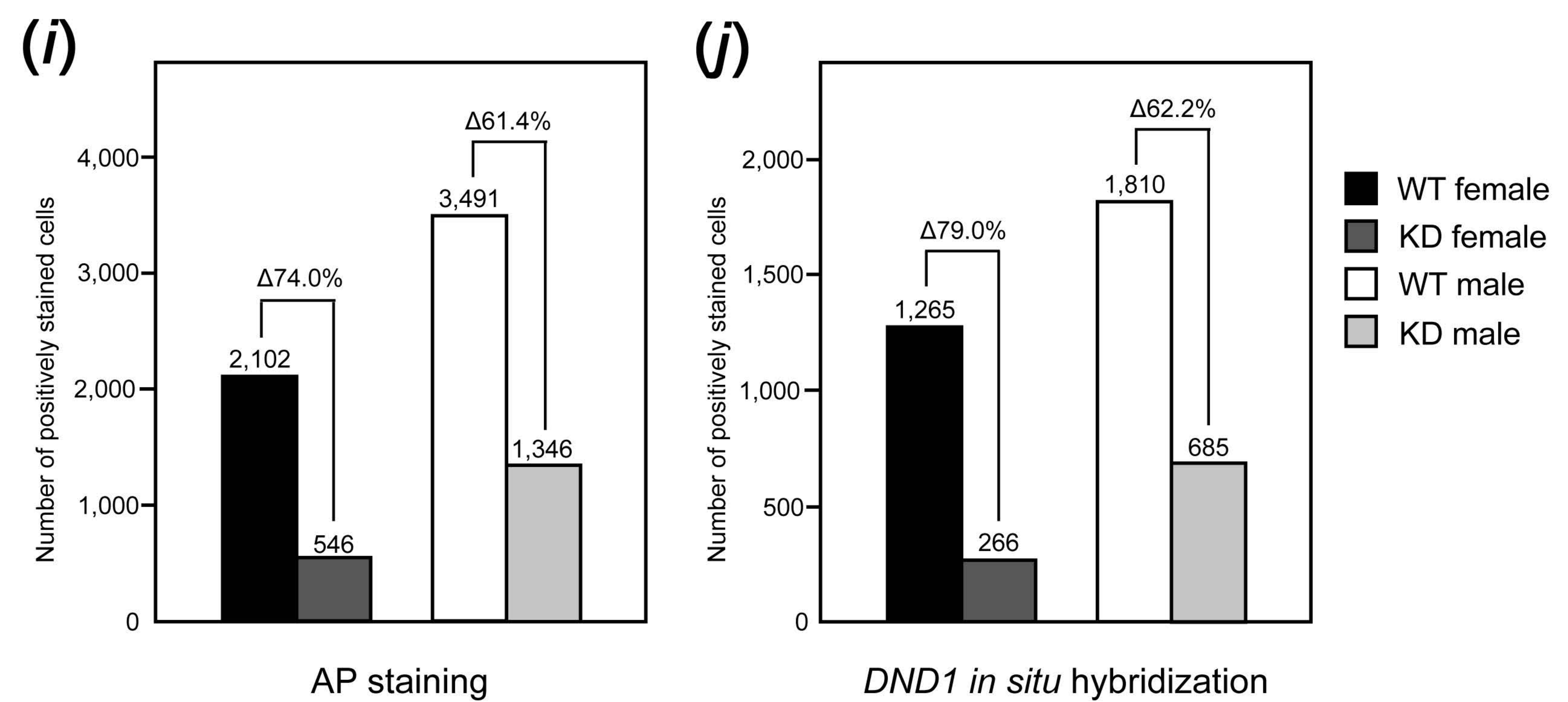
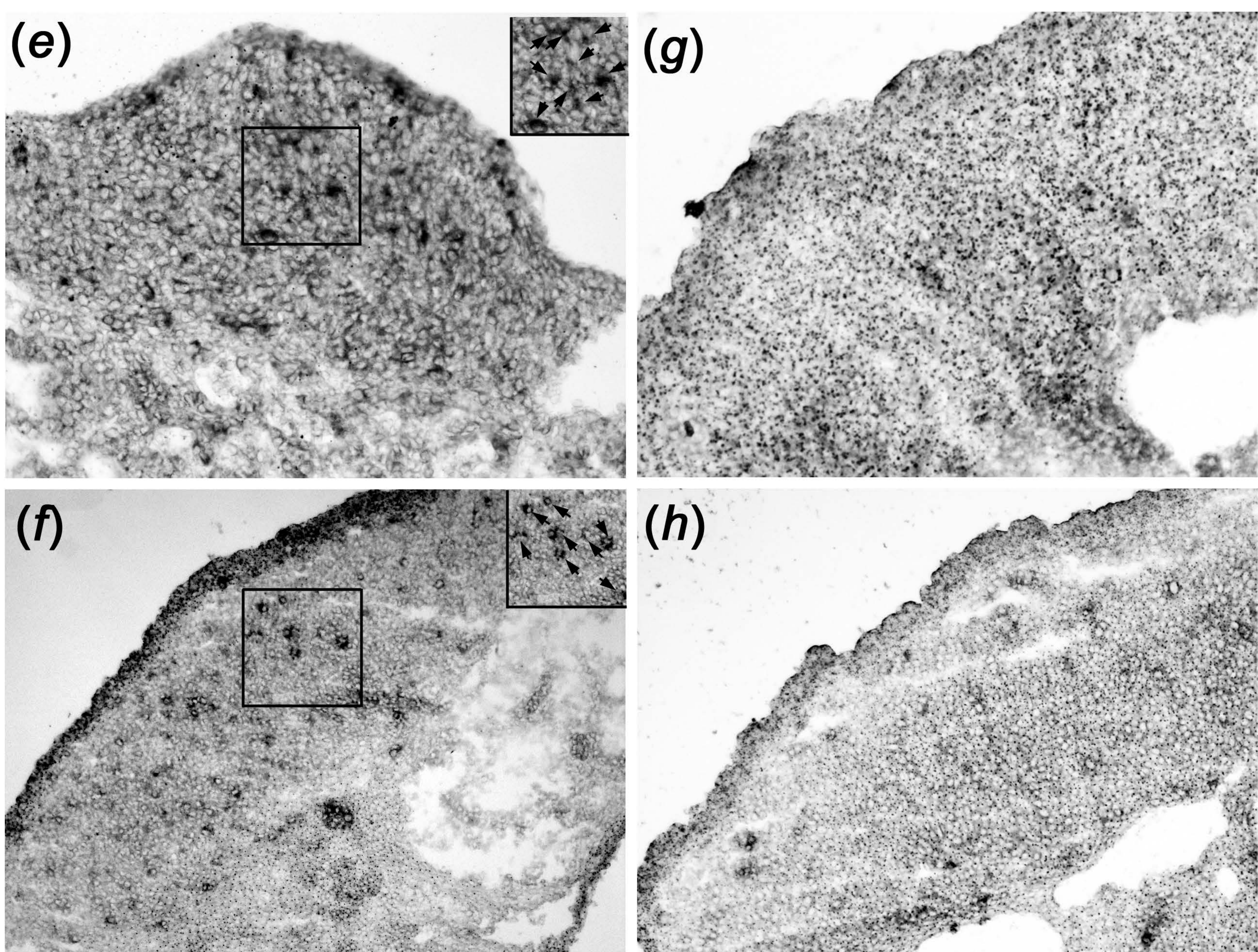




AP staining



*DND1* in situ hybridization



■ WT female    ■ KD female    □ WT male    ■ KD male

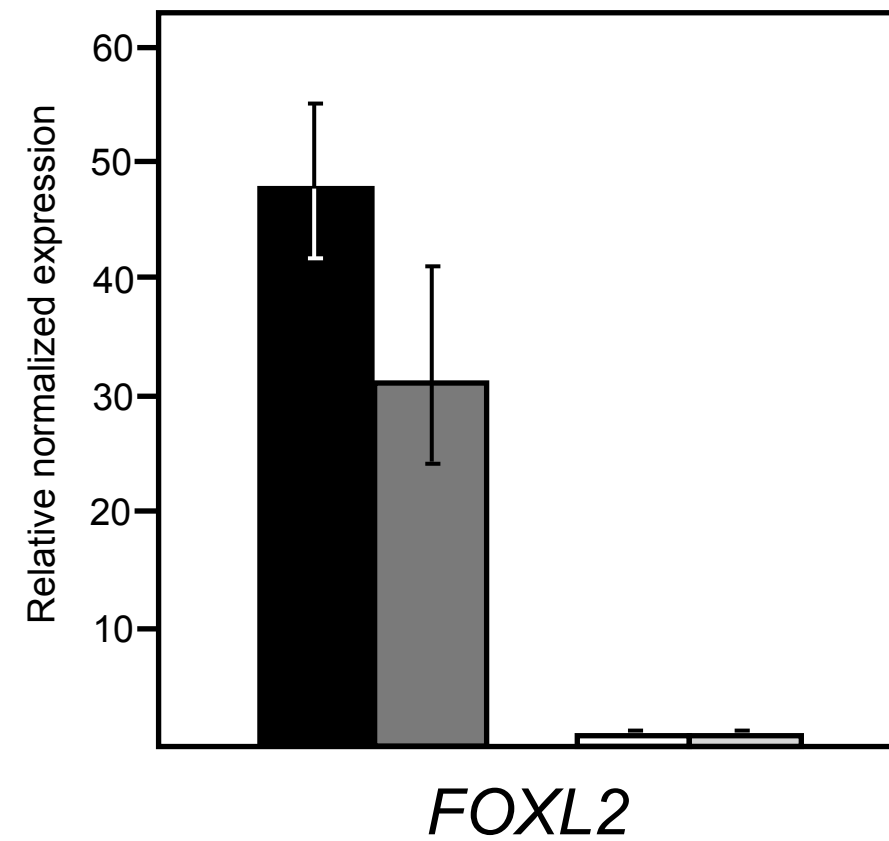
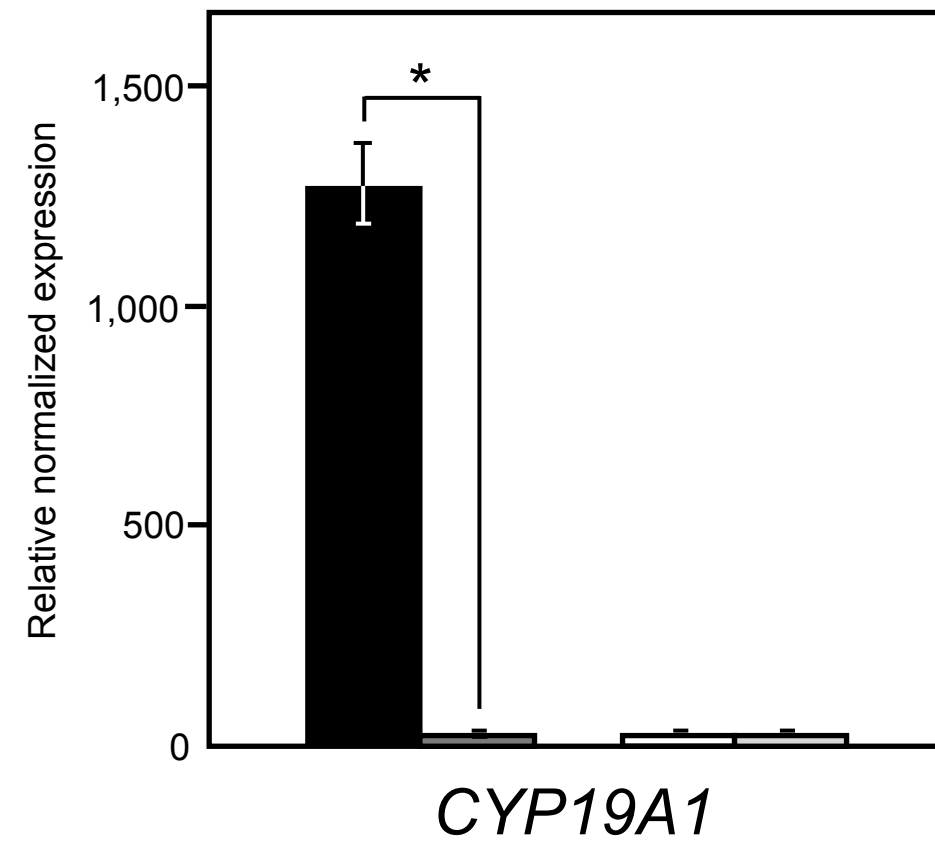
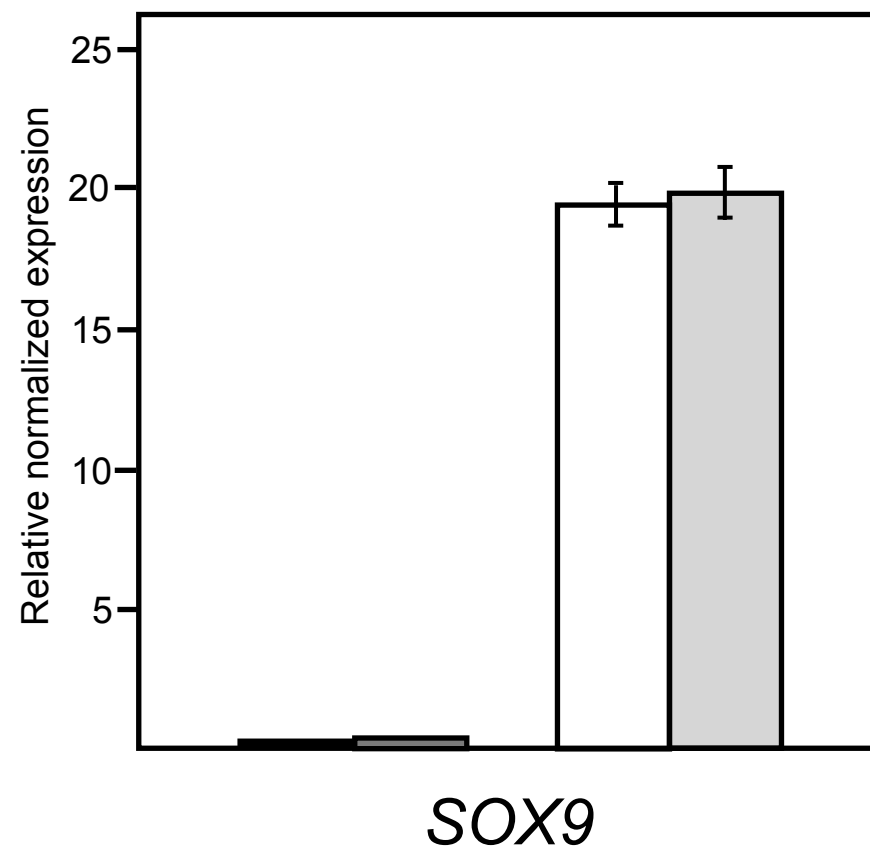
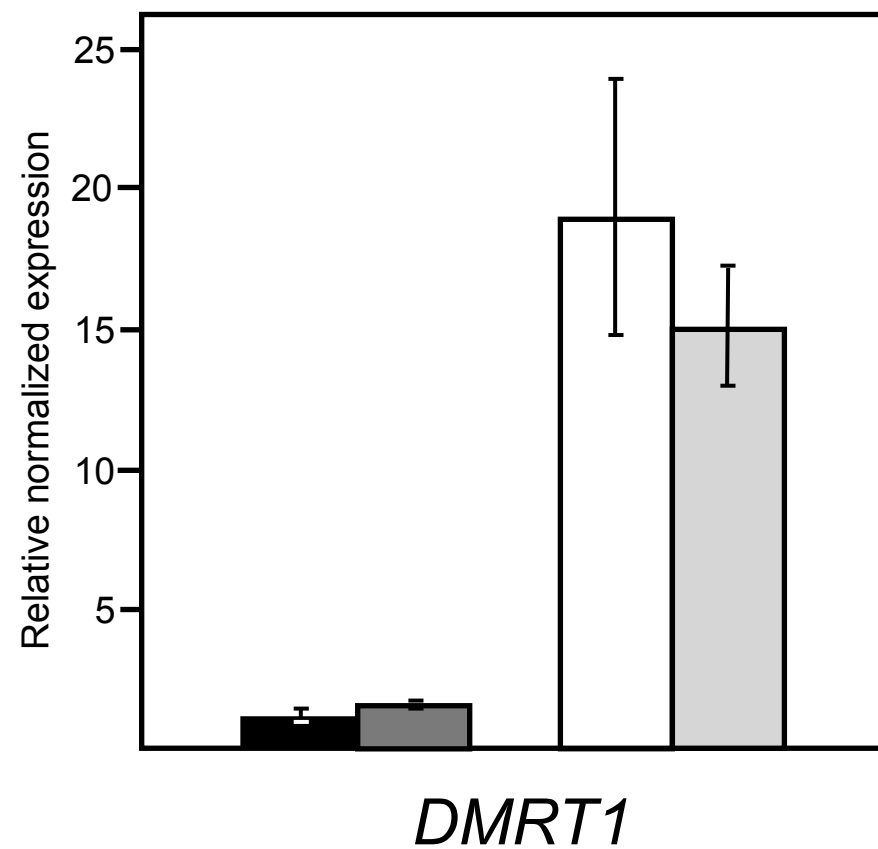


Table 1 Primer sequences

| Experiment                                               | Direction of primer | Sequence                                                                                                                                                                     |
|----------------------------------------------------------|---------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Construction of miRNA vectors<br>DDX4-targeting miRNA #1 | F<br>R              | 5'- GAG AGG TGC TGC TGA GCG AGA TCC TGG TAT GCA AGA TCA TAG TGA AGC CAC AGA TGT A -3'<br>5'- ATT CAC CAC CAC TAG GCA TGA TCC TGG TAT GCA AGA TCA TAC ATC TGT GGC TTC ACT -3' |
| DDX4-targeting miRNA #2                                  | F<br>R              | 5'- CTG GTT CCT CCG TGA GCG AGG TGC TAA TGA AGG ACT TAA TAG TGA AGC CAC AGA TGT A -3'<br>5'- CCT GAA GAC CAG TAG GCA TGG TGC TAA TGA AGG ACT TAA TAC ATC TGT GGC TTC ACT -3' |
| Scrambled miRNA #1                                       | F<br>R              | 5'- GAG AGG TGC TGC TGA GCG AAC CTT CTC AAT TCT CAT CAT TAG TGA AGC CAC AGA TGT A -3'<br>5'- ATT CAC CAC CAC TAG GCA GAC CTT CTC AAT TCT CAT CAT TAC ATC TGT GGC TTC ACT -3' |
| Scrambled miRNA #2                                       | F<br>R              | 5'- CTG GTT CCT CCG TGA GCG ACA TGT TCC CTC GTC ACT TTA TAG TGA AGC CAC AGA TGT A -3'<br>5'- CCT GAA GAC CAG TAG GCA GCA TGT TCC CTC GTC ACT TTA TAC ATC TGT GGC TTC ACT -3' |
| Construction of the DDX4_GFP vector                      | F<br>R              | 5'- TAA TCG ATG GAG GAG GAC TGG G -3'<br>5'- GCA CTA GTC TCC CAT GAC TTA AAT GTT G -3'                                                                                       |
| DND1 in situ hybridization probe                         | F<br>R              | 5'- CAA CCG GAC CAA TAA GAT GG -3'<br>5'- ATT CCC TTC CAC CAG AGC TT -3'                                                                                                     |
| qRT-PCR analysis of DDX4                                 | F<br>R              | 5'- GTA GCA TCA AGA GGC CTG GA -3'<br>5'- ACG ACC AGT TCG TCC AAT TC -3'                                                                                                     |