



HOKKAIDO UNIVERSITY

Title	Clinical efficacy of the combined treatment of anti-PD-L1 rat-bovine chimeric antibody with a COX-2 inhibitor in calves infected with Mycoplasma bovis
Author(s)	Shinya, Goto; Satoru, Konnai; Yuki, Hirano et al.
Citation	Japanese Journal of Veterinary Research, 68(2), 77-90
Issue Date	2020-05
DOI	https://doi.org/10.14943/jjvr.68.2.77
Doc URL	https://hdl.handle.net/2115/78601
Type	departmental bulletin paper
File Information	JJVR68-2_77-90_ShinyaGoto.pdf



Clinical efficacy of the combined treatment of anti-PD-L1 rat-bovine chimeric antibody with a COX-2 inhibitor in calves infected with *Mycoplasma bovis*

Shinya Goto¹⁾, Satoru Konnai^{1,2*)}, Yuki Hirano³⁾, Junko Kohara³⁾, Tomohiro Okagawa²⁾, Naoya Maekawa²⁾, Yamato Sajiki¹⁾, Kei Watari¹⁾, Erina Minato⁴⁾, Atsushi Kobayashi⁴⁾, Satoshi Gondaira⁵⁾, Hidetoshi Higuchi⁵⁾, Masateru Koiwa⁵⁾, Motoshi Tajima⁵⁾, Eiji Taguchi⁶⁾, Masaru Ishida⁶⁾, Ryoko Uemura⁷⁾, Shinji Yamada⁸⁾, Mika K. Kaneko⁸⁾, Yukinari Kato^{8,9)}, Keiichi Yamamoto^{2,10)}, Mikihiro Toda^{2,10)}, Yasuhiko Suzuki^{2,12,13)}, Shiro Murata^{1,2)} and Kazuhiko Ohashi^{1,2)}

¹⁾ Department of Disease Control, Faculty of Veterinary Medicine, Hokkaido University, Sapporo, Japan

²⁾ Department of Advanced Pharmaceutics, Faculty of Veterinary Medicine, Hokkaido University, Sapporo, Japan

³⁾ Animal Research Center, Agriculture Research Department, Hokkaido Research Organization, Shintoku, Japan

⁴⁾ Department of Veterinary Clinical Medicine, Faculty of Veterinary Medicine, Hokkaido University, Sapporo, Japan

⁵⁾ School of Veterinary Medicine, Rakuno Gakuen University, Ebetsu, Japan

⁶⁾ Shibetsu Animal Hospital, Shibetsu, Japan

⁷⁾ Department of Veterinary Medical Science, Faculty of Agriculture, University of Miyazaki, Miyazaki, Japan

⁸⁾ Department of Antibody Drug Development, Graduate School of Medicine, Tohoku University, Sendai, Japan

⁹⁾ New Industry Creation Hatchery Center, Tohoku University, Sendai, Japan

¹⁰⁾ Research and Development Center, Fuso Pharmaceutical Industries, Ltd., Osaka, Japan

¹¹⁾ New Business and International Business Development, Fuso Pharmaceutical Industries, Ltd., Osaka, Japan

¹²⁾ Division of Bioresources, Research Center for Zoonosis Control, Hokkaido University, Sapporo, Japan

¹³⁾ Global Station for Zoonosis Control, Global Institution for Collaborative Research and Education (GI-CoRE), Hokkaido University, Sapporo, Japan

Received for publication, December 9, 2019; accepted, December 20, 2019

Abstract

Mycoplasma bovis (*M. bovis*) is a highly contagious pathogen and *M. bovis*-associated diseases, particularly pneumonia, occur predominantly as herd enzootics, causing considerable economic losses because of calf mortality, weight loss in surviving calves. In our previous studies, the programmed death-1 (PD-1)/ PD-ligand 1 (PD-L1) pathway and prostaglandin E₂ (PGE₂) were shown to be involved in the immunosuppression during chronic infectious diseases in cattle. In this study, the efficacy of dual blockade of the PD-1/PD-L1 pathway and PGE₂ in *M. bovis* infection *in vivo* was investigated using anti-bovine PD-L1 rat-bovine chimeric antibody, Boch4G12, and cyclooxygenase 2 (COX-2) inhibitor, meloxicam. The calves treated with Boch4G12 and meloxicam significantly enhanced *M. bovis*-specific IFN- γ response after the administration. On the other hand, IFN- γ response was not activated in the controls and cattle treated with meloxicam alone throughout the experimental period. Interestingly, bacterial loads in nasal discharge and bronchoalveolar lavage fluid among calves treated with Boch4G12 with or without meloxicam were significantly decreased. These results suggest that the combination of anti-PD-L1 antibody with a COX-2 inhibitor is a candidate for therapeutic applications in calves infected with *M. bovis*.

Key Words: *Mycoplasma bovis*, Immunotherapy, PD-L1, PGE₂, COX-2 inhibitor,

* Corresponding author: Department of Disease Control, Faculty of Veterinary Medicine, Hokkaido University, Sapporo, 060-0818 Japan. Phone: +81-11-706-5216. Fax: +81-11-706-5217. E-mail: konnai@vetmed.hokudai.ac.jp
doi: 10.14943/jjvr.68.2.77

Introduction

Mycoplasma bovis (*M. bovis*) was first isolated in the United States in 1961 from a severe case of mastitis in a dairy herd experiencing an outbreak, affecting more than 30% of the animals¹⁷. *M. bovis* has emerged as an important cause of not only mastitis but also other disease manifestations include pneumonia, arthritis and tenosynovitis, otitis media in dairy calves and beef calves^{5,14,15}. *M. bovis* is a highly contagious pathogen and *M. bovis*-associated diseases, particularly pneumonia, occur predominantly as herd enzootics, causing considerable economic losses because of calf mortality, weight loss in surviving calves^{5,14,15}. Unfortunately, no effective vaccines have been developed against *M. bovis* infection. The treatment with antibiotics is potentially effective against *M. bovis*, but strains which resistant to antibiotics have recently been spreading^{1,24,37}. Therefore, a novel alternative strategy for the control of *M. bovis* infection is required.

Recently, several reports have demonstrated that immunoinhibitory molecules are one of the mechanisms of immunosuppression and contribute to the disease progression during various human chronic infections and cancers^{9,22,38,43}. Cell surface immunoinhibitory receptors, such as programmed death-1 (PD-1), are negative regulators of T cell activation. After binding to its ligand, PD-ligand 1 (PD-L1), PD-1 suppresses the activation signaling mediated by T cell receptor and inhibits the effector functions of T cells such as cytokine production and cell proliferation⁴². This dysfunction of T cells is called T cell exhaustion. Thus, the PD-1/PD-L1 pathway is recognized as a common mechanism of immunosuppression in chronic infections and cancers. Interestingly, the blockade of the PD-1/PD-L1 pathway by specific antibodies against either the receptor or ligand restores the function of exhausted T cells⁴². Indeed, anti-PD-1/PD-L1 antibodies are recognized as candidates for the treatment of several human cancers^{9,22,38,43}. As

examples, fully humanized mAbs targeting PD-1, BMS-936558 (known as Nivolumab), and PD-L1, BMS-936559, have shown striking response against melanoma and other cancers³.

In the veterinary field, during bovine chronic infection, such as *Mycobacterium avium* subsp. *paratuberculosis* infection (Johne's disease), bovine leukemia virus (BLV) infection and bovine anaplasmosis, the expression of PD-1 is upregulated on CD4⁺ and CD8⁺ T cells and PD-L1 is also upregulated on several cells in line with disease progression^{19,20,21,31,32,34,35}. Furthermore, prostaglandin E₂ (PGE₂), a product of cyclooxygenase (COX) 2, induces immune check point molecules including PD-L1 and contributes to the progression of chronic infectious diseases in cattle^{34,35}. Previously, we established anti-bovine PD-1 and PD-L1 rat-bovine chimeric Abs (chAbs) and conducted clinical studies of these chAbs in cattle infected with BLV^{30,33,35}. Inoculation of cattle with the anti-PD-1 or PD-L1 chAb restored T cell functions, such as IFN- γ response and the proliferation of anti-BLV-specific CD4⁺ T cells^{30,33,35}. Moreover, the chAb treatment significantly reduced BLV provirus loads, clearly demonstrating that this treatment induced antiviral activities^{30,33,35}. More interestingly, combined treatment of chAb with a COX-2 inhibitor substantially reduced the BLV provirus loads more effectively in cattle infected with BLV³⁵. Therefore, the blockade of the PD-1/PD-L1 pathway and/or PGE₂ can be applied for the treatment of bovine chronic infection (Supplemental figure 1).

In *M. bovis* infection in cattle, immunosuppressive effects were observed, such as the inhibition of bovine peripheral blood mononuclear cell (PBMC) proliferation, induction of bovine lymphocyte apoptosis, and delay of bovine monocyte apoptosis, along with the suppression of IFN- γ and TNF- α productions^{28,39,40}. However, the mechanisms involved in immunosuppression had not been fully elucidated. In a previous report, we had indicated that the immunosuppression was associated with the

Table I. The information of calves used in clinical studies.

Cattle	#1	#2	#3	#4	#5	#6	#7
Age	2 months old	7 months old	6 months old				
Species	Holstein	Holstein	Holstein	Holstein	Holstein	Holstein	Holstein
Sex	Male	Male	Male	Male	Male	Male	Male
Weight (kg)	85	252	213				
Respiratory symptom	Cough and watery discharge	Cough and watery discharge	Cough and watery discharge	Cough and watery discharge	Cough and watery discharge	Cough and watery discharge	Cough and watery discharge
Serum PGE ₂ concentration at day 0 (pg/mL)	242	2,057	1,226	762	1,657	160	2,681
Inoculation dose(Boch4G12)	Saline 100mL, <i>i. v.</i>	Saline 100mL, <i>i. v.</i>		2 mg/kg, <i>i. v.</i>	2 mg/kg, <i>i. v.</i>	2 mg/kg, <i>i. v.</i>	2 mg/kg, <i>i. v.</i>
Administration dose(Meloxicam)	-		0.5 mg/kg, once a week, totally 2 times, <i>s. c.</i>			0.5 mg/kg, once a week, totally 2 times, <i>s. c.</i>	0.5 mg/kg, once a week, totally 2 times, <i>s. c.</i>

PD-1/PD-L1 pathway during *M. bovis* infection¹⁶⁾. More specifically, it was elucidated that T cell exhaustion caused by the PD-1/PD-L1 pathway suppressed IFN- γ production against *M. bovis* and *in vitro* treatment with anti-bovine PD1/PD-L1 mAbs restored decreased *M. bovis*-specific IFN- γ response during *M. bovis* infection¹⁶⁾. However, the clinical efficacy of blockade of the PD-1/PD-L1 pathway and/or PGE₂ on *M. bovis*-infected calves remains to be elucidated. Thus, in this study, the efficacy of combined treatment of anti-bovine PD-L1 chAb with a COX-2 inhibitor was examined using *M. bovis*-infected calves. The findings in this study indicated that novel biomedicines targeting the PD-1/PD-L1 pathway and PGE₂ are potentially applicable for immune therapies for the control of *M. bovis* infection.

Materials and methods

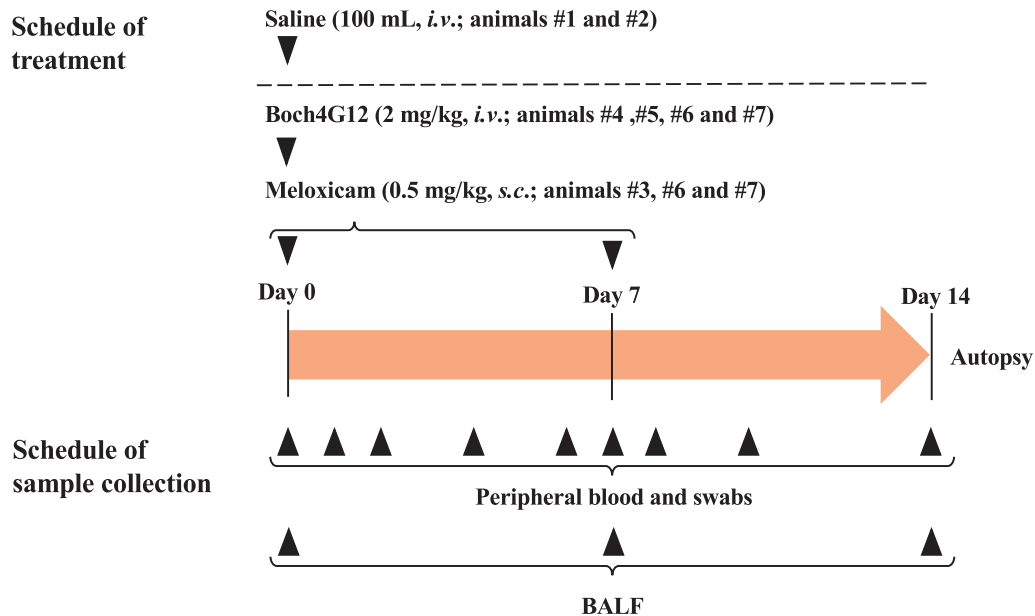
Animals

Calves, which clinical tests were performed in, were seven calves naturally infected by *M. bovis*, positive for respiratory mycoplasmas (nasal swab culture) obtained from farms at Hokkaido, Japan. *M. bovis* infection was confirmed by polymerase chain reaction (PCR) using clinical

samples as previously described¹⁸⁾. Animals were launched for this experiment and kept separately at the isolated rooms in a biosafety level I animal facility at the Animal Research Center, Agricultural Research Department, Hokkaido Research Organization (Shintoku, Hokkaido, Japan). The details of used cattle are shown in Table 1.

Administration of anti-bovine PD-L1 chimeric antibody (chAb) and a COX-2 inhibitor to *M. bovis*-infected calves

To investigate the effect of the PD-1/PD-L1 blockade and COX-2 inhibition against *M. bovis* in vivo, 2 mg/kg of anti-PD-L1 chAb (Boch4G12)³⁰⁾ was administered intravenously in four *M. bovis*-infected calves with pneumonia (animals #4-#7), and 0.5 mg/kg of meloxicam (Boehringer Ingelheim, Ingelheim, Germany) was co-administered by subcutaneous injection twice at a weekly interval in animals #3, #6 and #7. As negative controls, animals #1 and #2 were administered with 100 mL of saline (Fuso Pharmaceutical Industries, Ltd. Osaka, Japan). After the treatments, peripheral blood samples, nasal discharge and bronchoalveolar lavage fluid (BALF) were then collected from these calves according to the schedule of sample collection as

**Fig.1**

Schedule of treatment and sample collection in clinical study. 2 mg/kg of anti-PD-L1 chAb (Boch4G12) was administered intravenously in four *M. bovis*-infected calves with pneumonia (animal No #4-#7) and 0.5 mg/kg of meloxicam was co-administered by subcutaneous injection twice at a weekly interval in animals (animal No #3, #6 and #7). As controls, animals (animal No #1 and #2) were administered 100 mL of saline. Peripheral blood and nasal swabs were collected at days 0, 1, 2, 4, 6, 7, 8, 10 and 14. BALF was collected at days 0, 7 and 14.

shown in Fig. 1. At 14 days after the treatments the inoculation of Boch4G12, animals #1, #4 and #6 were euthanized and subjected to autopsy.

Kinetic analysis of serum PGE₂ by ELISA

To confirm the kinetics of PGE₂ in treated calves, the serum PGE₂ levels were measured using the Prostaglandin E₂ Express ELISA kit (Cayman Chemical, Ann Arbor, MI, USA), according to the manufacturer's instructions. The optical density was measured at 450 nm in a microplate reader (Corona Electronics, Tokyo, Japan).

In vivo efficacy of anti-PD-L1 chAb and a COX-2 inhibitor on IFN-γ response

To investigate the efficacy of the PD-1/PD-L1 blockade on *M. bovis*-specific immune response, peripheral blood samples were collected and PBMCs were purified by density gradient centrifugation on Percoll (GE Healthcare, Little

Chalfont, England, UK). The IFN-γ response against the heat-killed *M. bovis* antigen was investigated as previously described¹⁶. Briefly, to examine the IFN-γ production in treated calves, PBMCs were incubated with heat-killed *M. bovis* antigen (1.5 ng/mL) in RPMI 1640 medium (Sigma-Aldrich, St. Louis, MO, USA) containing 10% fetal calf serum (Thermo Fisher Scientific, Waltham, MA, USA) and 100 IU/mL penicillin, 100 µg/mL streptomycin and 2 mM L-glutamine (Thermo Fisher Scientific) at 37°C under 5% CO₂ for 5 days. Collected culture supernatants were assayed for IFN-γ using a commercial ELISA kit (Mabtech, Nacka Strand, Sweden), in accordance with the manufacturer's instructions. Data are presented as means of triplicate samples.

The efficacy of anti-PD-L1 chAb and a COX-2 inhibitor on bacterial load in nasal discharge and bronchoalveolar lavage fluid from infected calves

To investigate the efficacy of the PD-1/PD-

L1 blockade against *M. bovis* infection, bacterial load was measured in nasal discharge and bronchoalveolar lavage fluid (BALF) from treated calves using real-time PCR. Nasal discharges were collected by using sterilized P200 tips (Douo Rika Co., Sapporo, Japan) and 1.5-mL micro tubes (Fukaekasei, Tokyo, Japan). With regard to the collection of BALF from each animal, 100 mL of saline (Fuso Pharmaceutical Industries, Ltd.) were injected into the trachea using a cannula tube (Fukaekasei) and collected the injected saline as BALF by aspiration. DNA was extracted from total nasal discharge or 2 mL of BALF using GenElute Bacterial Genomic DNA Kits (Sigma–Aldrich, St. Louis, MO) according to the manufacturer's instructions. Bacterial load was evaluated by using real-time PCR with Thunderbird SYBR qPCR Mix (Toyobo, Tokyo, Japan) and Cica genus *Mycoplasma bovis* detection kit (Kanto Chemical, Tokyo, Japan). Real-time PCR was performed in triplicate using a thermal cycler (LightCycler 480 system II; Roche Diagnostic, Mannheim, Germany).

Statistics

Statistical significance was determined by Dunnett's test (vs. day 0 for each result) using the statistical analysis program MEPHAS (<http://www.gen-info.osaka-u.ac.jp/MEPHAS/>). A p-value < 0.05 was considered statistically significant.

Ethics statement

The animal experiments were approved by the Ethics Committee of the Animal Research Center, Agricultural Research Department, Hokkaido Research Organization (No. 1905), and the Ethics Committee of the Faculty of Veterinary Medicine, Hokkaido University (No. 17-0024).

Results

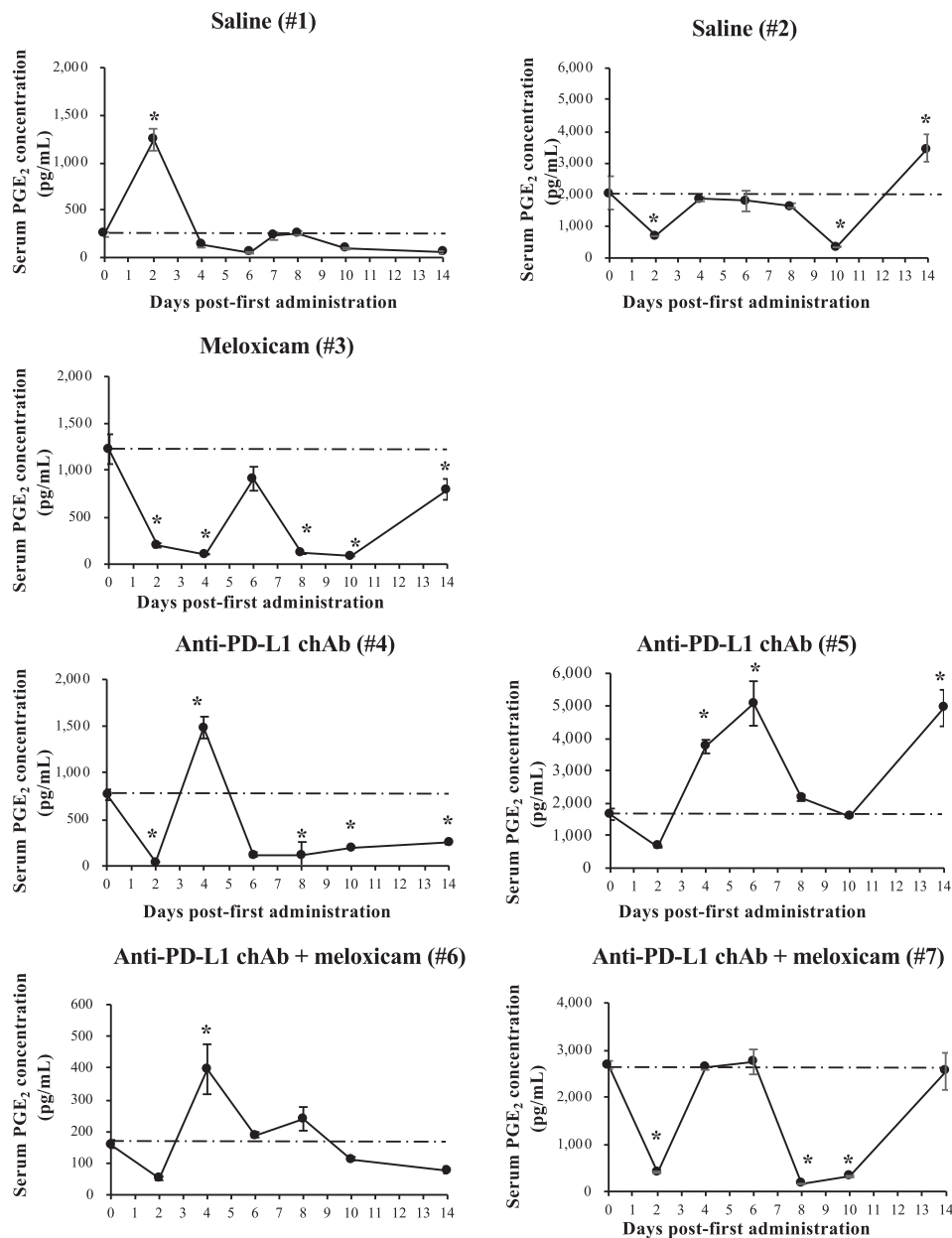
Serum PGE₂ concentration in the treated calves

To investigate the efficacy of the treatment with Boch4G12 and/or meloxicam on the PGE₂

levels in peripheral blood from treated calves, serum PGE₂ concentration was measured by ELISA. The controls (animal No. #1 and #2) and one of two calves treated with Boch4G12 and meloxicam (animal No. #6) did not change serum PGE₂ concentration throughout the experimental period (Fig. 2). On the other hand, serum PGE₂ concentration in calves treated with meloxicam alone (animal No. #3), Boch4G12 alone (animal No. #4) and both Boch4G12 and meloxicam (animal No. #7) significantly decreased after the administration. Serum PGE₂ concentration in animal No. #5 calf treated with Boch4G12 alone significantly increased after the administration.

Clinical efficacy of M. bovis specific-IFN-γ response in the treated calves

We previously reported that meloxicam activates Th1 responses in the treated cattle³⁵. To examine whether meloxicam activates the *M. bovis*-specific Th1 responses, PBMCs from meloxicam-treated calf (animal No. #3) was cultured in the presence of heat-killed *M. bovis*, and IFN-γ production was measured by ELISA (Fig. 3). However, unexpectedly, IFN-γ production was not enhanced in the meloxicam-treated calf (animal No. #3) as well as those of control cattle (animal No. #1 and #2). On the other hand, we also investigated the efficacy of the treatment with Boch4G12 alone/and meloxicam on the IFN-γ production because we previously reported that the blockade of the PD-1/PD-L1 pathway activates IFN-γ production *in vitro* from PBMCs of *M. bovis*-infected cattle¹⁶. Although the enhancement of IFN-γ production was not observed in one of two calves treated with Boch4G12 alone, IFN-γ production against *M. bovis* antigen was significantly higher than before administration in calves treatment with Boch4G12 alone (animal No. #4)/and meloxicam (animal No. #6 and #7) (Fig. 3). Interestingly, IFN-γ production against *M. bovis* antigen was strongly enhanced and maintained in calves treated with combinations of Boch4G12 and meloxicam (animal No. #6 and #7) (Fig. 3)

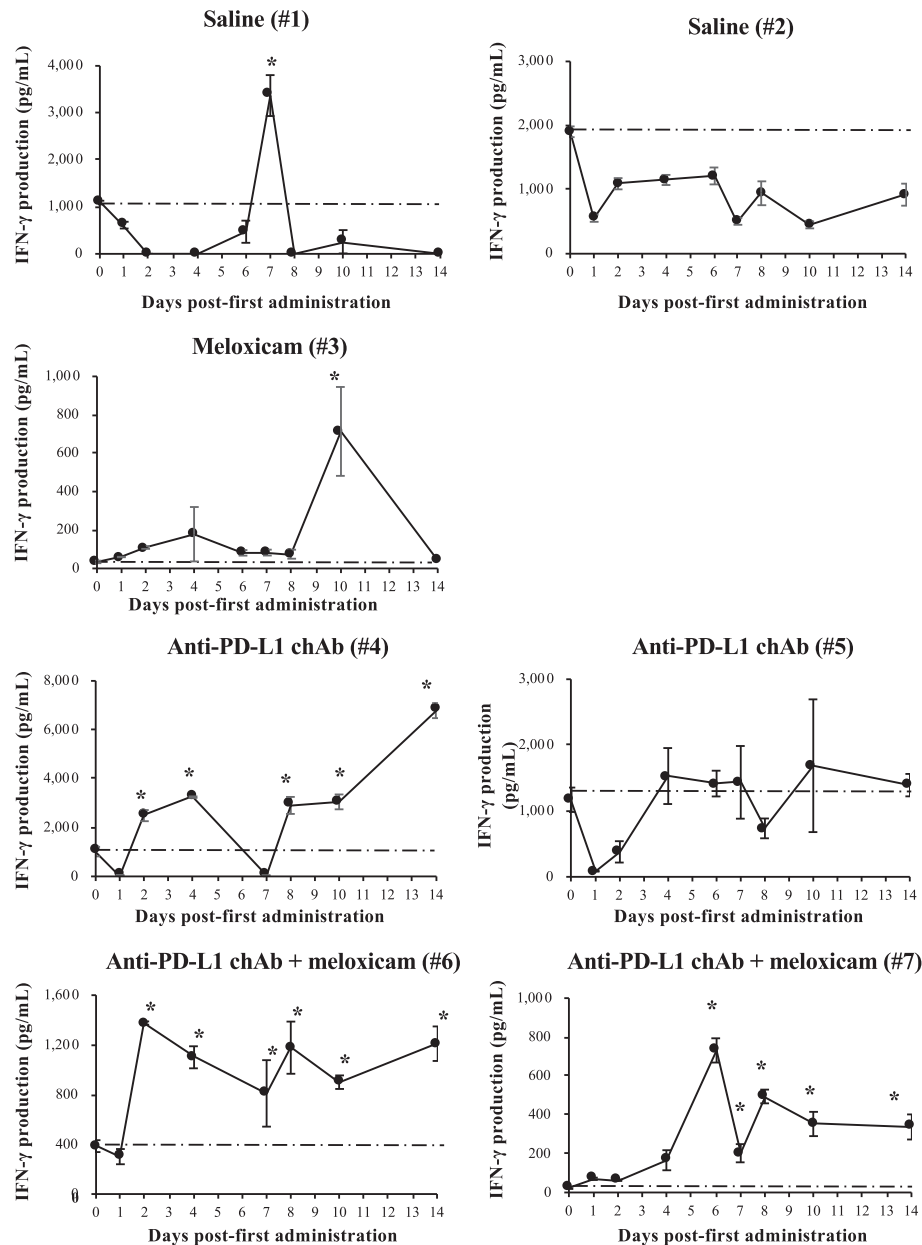
**Fig.2**

Kinetic analysis of serum PGE₂ in *M. bovis*-infected calves treated with anti-PD-L1 chAb (Boch4G12) and meloxicam. *M. bovis*-infected calves were intravenously administered 2 mg/kg of purified Boch4G12 (animal No #4-#7) or saline (control, animal No #1 and #2). In addition, animals (animal No #3, #6 and #7) were subcutaneously administered 0.5 mg/kg of meloxicam twice with a weekly interval. Serum PGE₂ was measured by ELISA. Statistical significance was determined by Dunnett's test. **P* < 0.05 (vs. day 0 for each result).

Clinical efficacy of bacterial loads in the treated calves

To investigate the efficacy of the combined treatment of boch4G12 and meloxicam, bacterial load of *M. bovis* in nasal discharges and BALFs

was measured by real-time PCR. Bacterial load in nasal discharges and BALFs from negative control was not changed throughout the experimental period (animal No. #1) (Fig. 4), but those from treated cattle were significantly

**Fig.3**

Activation of IFN- γ responses in *M. bovis*-infected calves treated with Boch4G12 and meloxicam. *M. bovis*-infected calves were intravenously administered 2 mg/kg of purified Boch4G12 (animal No. #4-#7) or saline (control, animal No. #1 and #2). In addition, animals (animal No.#3, #6 and #7) were subcutaneously administered 0.5 mg/kg of meloxicam twice with a weekly interval. PBMCs from treated cattle were incubated in the presence of heat-killed *M. bovis* (1.5 ng/mL). IFN- γ production in culture supernatants was measured by ELISA. Statistical significance was determined by Dunnett's test. * $P < 0.05$ (vs. day 0 for each result).

decreased after the treatment of Boch4G12 with or without meloxicam (animal No #4 and #6) (Fig. 4). In other tested animals (animal No #2, #3, #5 and #7), the bacterial loads in nasal discharges and BALFs were not detectable by real-time PCR

due to low bacterial loads (data not shown).

Gross lung lesions in the treated calves

To examine the pathologic findings of pneumonia in the treated calves, *M. bovis*-

detected calves (animal No #1, #4 and #6) were dissected at 14 days after administration. Saline-inoculated calf as control was observed severe findings of typical mycobacterial pneumonia such as pleural adhesion, multifocal white nodules and atelectasis in the cranioventral areas of the lungs⁴⁾ (Fig. 5). On the other hand, although mild atelectasis was partially observed, the severe findings including pleural adhesion and multifocal white nodules were not confirmed in the calves treated with Boch4G12 alone (animal No. #4)/ and meloxicam (animal No. #6) (Fig. 5).

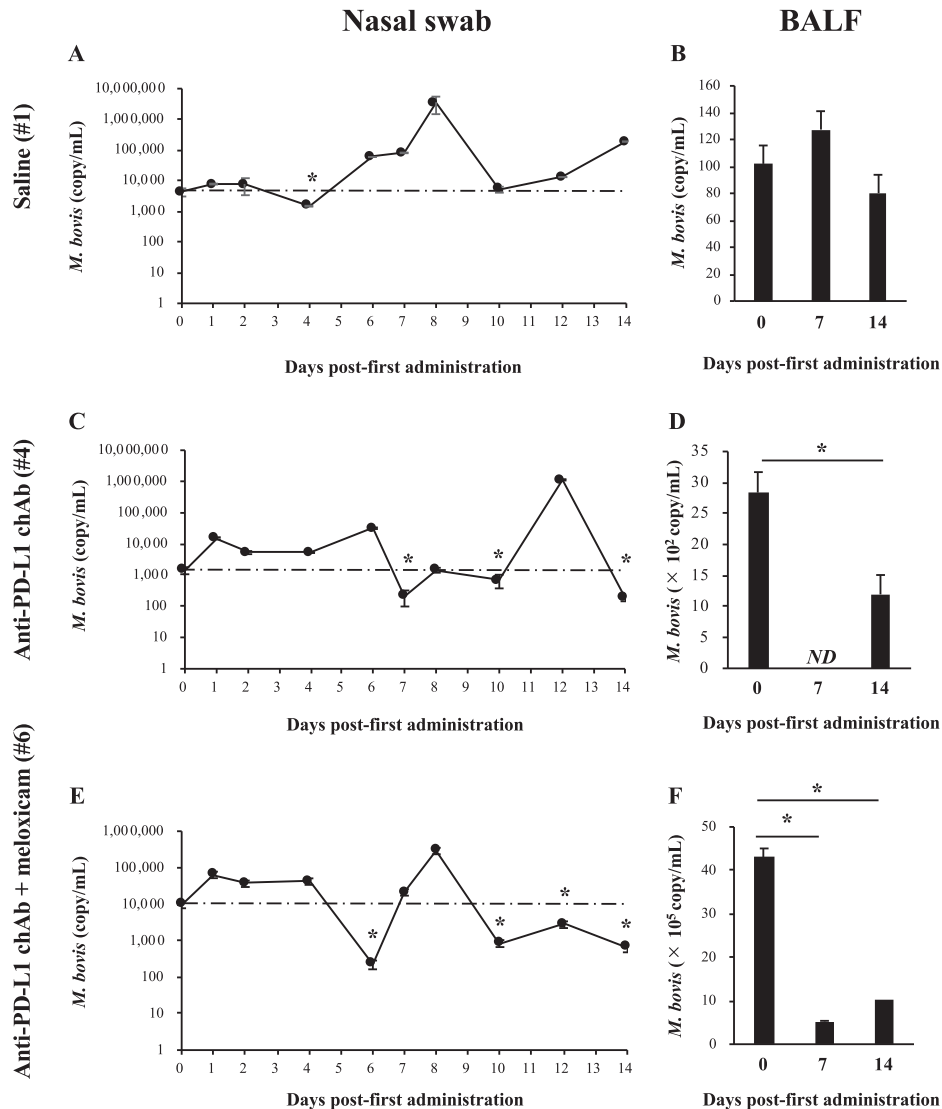
Discussion

Pneumonia is a typical symptom of *M. bovis* infection in calves, and tends to be chronic progression^{4,5)}. The chronicity and the persistence of *M. bovis* infection imply that the immune response is insufficient in eliminating the pathogen. In our previous studies, it was found that immunoinhibitory molecules, PD-1/PD-L1, and PGE₂ caused the immunosuppression during several bovine chronic infectious diseases including BLV infection, Johne's disease and bovine TB and these molecules had the potential as targets for control of these infection^{34, 35)}. During *M. bovis* infection, T cell exhaustion caused by upregulation of PD-1/PD-L1 expression on immune cells and blockade of PD-1/PD-L1 pathway using anti-bovine PD-1 or PD-L1 mAb restored *M. bovis*-specific immunoresponse *in vitro*¹⁶⁾. In this study, to examine the efficacy of the treatment of anti-PD-L1 chAb with or without COX-2 inhibitor on *M. bovis* infection *in vivo*, the clinical study was performed using *M. bovis*-infected cattle with pneumonia.

Firstly, to evaluate the efficacy of the blockade of the PD-1/PD-L1 pathway and PGE₂ *in vivo*, IFN- γ response against *M. bovis* was analyzed. Although IFN- γ response was not activated in controls and cattle treated with meloxicam, cattle treated with Boch4G12 or both Boch4G12 and meloxicam showed the reactivation

of the IFN- γ response against *M. bovis* antigen (Fig. 3). This result suggests that the treatment of Boch4G12 and meloxicam enhances *M. bovis*-specific immune response *in vivo*. However, IFN- γ response against *M. bovis* in animal (animal No. #5) was not reactivated by anti-PD-L1 Ab alone. The reason for this observation is unclear but this might be due to the high level of serum PGE₂ (Table 1). Several reports indicate that the treatment with anti-PD-1/PD-L1 Abs reactivates T cells, and that the production of Th1 cytokine such as TNF- α ¹³⁾. TNF- α is one of the inducers for PGE₂ production by the activation of COX-2 via NF- κ B^{26,29)}. Thus, the low IFN- γ response observed in animal No. #5 might be due to the increased-PGE₂ production resulted from the enhancement of immune activity by Boch4G12. Indeed, serum PGE₂ concentration of animal No. #5 was consistently higher compared to those of other tested calves (Fig. 2). Interestingly, *M. bovis*-specific IFN- γ response of animal No. #7, which decreased serum PGE₂ concentration after the meloxicam administration, was significantly enhanced throughout the experimental period. These findings might suggest an association between IFN- γ response correlated with PGE₂ concentration. Further studies on the relationship between Th1 cytokine and PGE₂ production might help to understand the unresponsiveness of immune reaction by the treatment with anti-PD-L1 Ab.

Bacterial loads in nasal discharge and BALF among calves treated with Boch4G12 with or without meloxicam were decreased (Fig. 3). The calves decreased bacterial loads clearly indicated IFN- γ production after the treatments as described above. In contagious bovine pleuropneumonia caused by *Mycoplasma mycoides* subsp. *mycoides* SC (MmmSC), a positive correlation was found between the number of MmmSC-specific IFN- γ -secreting CD4⁺ T cells and disease recovery^{10,11)}. IFN- γ promotes the activation and proliferation of T cells and macrophages, which play vital roles in host defenses in the lungs. Previous studies showed that macrophage dysfunction is induced

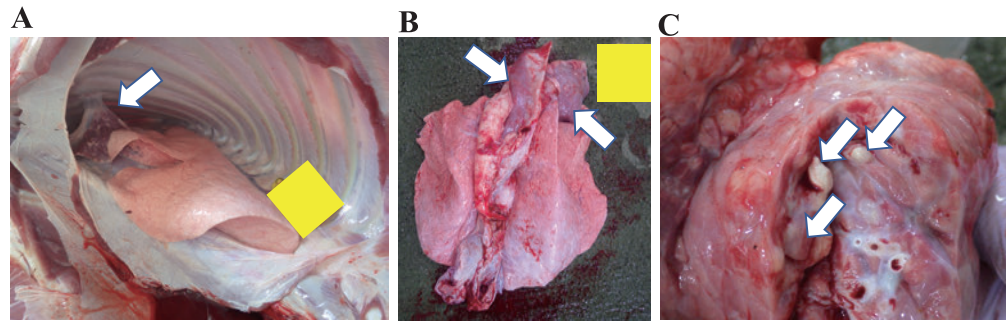
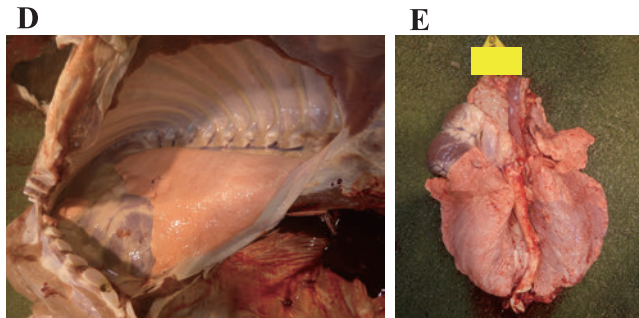
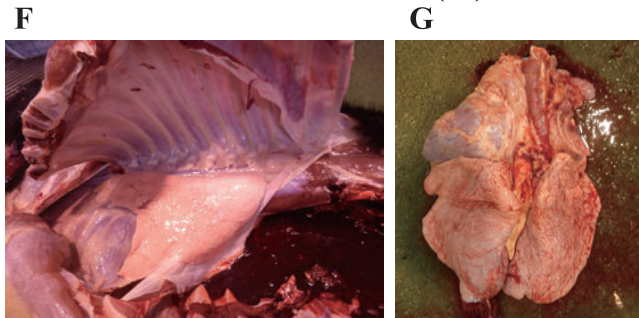
**Fig.4**

Bacterial load in nasal discharge and BALF from *M. bovis*-infected calves treated with Boch4G12 and meloxicam. *M. bovis*-infected calves were intravenously administered 2 mg/kg of purified Boch4G12 (animal No. #4 and #6) or saline (control, animal No. #1). In addition, animal (animal No. #6) was subcutaneously administered 0.5 mg/kg of meloxicam twice with a weekly interval. Bacterial load in nasal discharge and BALF was measured by real-time PCR (A and B: data of animal No. #1, C and D: data of animal No. #4, E and F: data of animal No. #6). Statistical significance was determined by Dunnett's test (A-F). * $P < 0.05$ (vs. day 0 for each result). ND; not detected.

by *Mycoplasma* spp., including *M. bovis*, and promotes persistent infection^{12,23,36}. Therefore, macrophages activated by IFN- γ might have been involved in the decreased bacterial loads in nasal discharge and BALF.

The pathogenesis of severe pneumonia during *M. bovis* infection is still unclear. Recently,

several studies reported that type 17 helper T cell (Th17), which mainly produced proinflammatory cytokine, IL-17A, worsens the disease by making animal susceptible to mycoplasma infection including *M. bovis*^{7,13}, *M. pneumoniae* which causes human pneumonia⁴¹ and *M. pulmonis* which causes mice pneumonia²⁷. IL-17A activates

Saline (animal No. #1)**Anti-PD-L1 chAb (animal No. #4)****Anti-PD-L1 chAb + meloxicam (#6)****Fig.5**

Gross lung lesions of calves treated with Boch4G12 and meloxicam. *M. bovis*-infected calves were intravenously administered 2 mg/kg of purified Boch4G12 (animal No. #4 and #6) or saline (control, animal No. #1). In addition, animal (animal No. #6) was subcutaneously administered 0.5 mg/kg of meloxicam twice with a weekly interval. At 14 days after administration, autopsy was conducted (A, B, and C: pictures of animal No. #1, D and E: pictures of animal No. #4, F and G: pictures of animal No. #6). Arrows indicate the typical lung lesions in animal No. #1 (A: pleural adhesion, B: atelectasis, C: multifocal white nodules). Yellow boxes in the figures hide the ear tags of each animal.

tissue-damaging inflammatory cascades²⁵). Several studies demonstrated that PGE₂ promotes the differentiation and function of human and murine Th17 via EP2 and EP4 signaling^{2,8,44}). In addition, a previous study reported a relationship between the PD-1/PD-L1 pathway and Th17. In patients of pulmonary fibrosis, the number of PD-1⁺ Th17 was increased in peripheral blood. Furthermore, the blockade of the PD-1/PD-L1 pathway in mouse model of fibrosis resulted in reductions of IL-17A expression from CD4⁺ T cells and in improved fibrosis symptoms⁶). The present study found that the moderate lung lesion was

observed in the treatment animals treated with Boch4G12 and meloxicam when compared to that of untreated cattle (Fig. 5). Taken together, it is hypothesized that the efficacy might be due to the suppression of Th17 by the treatment with Boch4G12 and meloxicam. Further investigation on Th17 is required to examine this hypothesis and it might support to understand the mechanisms of action of the combination of PD-1/PD-L1 blockade with COX-2 inhibition.

In conclusion, dual blockade of the PD-1/PD-L1 pathway and PGE₂ *in vivo* significantly restored *M. bovis*-specific IFN- γ response. In

addition, bacterial loads in nasal discharge and BALF among treated calves were decreased. Thus, this study indicates that the combination therapy of anti-bovine PD-L1 chAb with a COX-2 inhibitor has a potential for the control of *M. bovis* infection. This pilot study could help developing potential novel methods for the control of *M. bovis* infection. Further experiments using a large number of infected cattle are required to confirm the efficacy of this novel treatment approach for clinical application.

Acknowledgements

We are grateful to Dr. Hideyuki Takahashi, Dr. Tomio Ibayashi and Dr. Yasuyuki Mori for valuable advice and discussions. This work was supported by Grants-in-Aid for Scientific Research from the Japan Society for the Promotion of Science, and by grants from the Project of the NARO, Bio-oriented Technology Research Advancement Institution (Research Program on Development of Innovative Technology 26058 BC [to S.K.] and Special Scheme Project on Regional Developing Strategy, Grant 16817557 [to S.K.]), and by AMED under Grant Number: JP19am0101078 (Y.K.). We thank Enago (<http://www.enago.jp>) for the English language review.

Supplemental data

Supplemental data associated with this article can be found, in the online version, at <http://dx.doi.org/10.14943/jjvr.68.2.77>

References

- 1) Amram E, Mikula I, Schnee C, Ayling RD, Nicholas RA, Rosales RS, Harrus S, Lysnyansky I. 16S rRNA gene mutations associated with decreased susceptibility to tetracycline in *Mycoplasma bovis*. *Antimicrob Agents Chemother* 59, 796-802, 2015.
- 2) Boniface K, Bak-Jensen KS, Li Y, Blumenschein WM, McGeachy MJ, McClanahan TK, McKenzie BS, Kastelein RA, Cua DJ, de Waal Malefyt R. Prostaglandin E2 regulates Th17 cell differentiation and function through cyclic AMP and EP2/EP4 receptor signaling. *J Exp Med* 20, 535-548, 2009.
- 3) Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, Drake CG, Camacho LH, Kauh J, Odunsi K, Pitot HC, Hamid O, Bhatia S, Martins R, Eaton K, Chen S, Salay TM, Alaparthi S, Grosso JF, Korman AJ, Parker SM, Agrawal S, Goldberg SM, Pardoll DM, Gupta A, Wigginton JM. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. *N Engl J Med* 366, 2455-2465, 2012.
- 4) Caswell JL, Archambault M. *Mycoplasma bovis* pneumonia in cattle. *Anim Health Res Rev* 8, 161-86, 2007.
- 5) Caswell JL, Bateman KG, Cai HY, Castillo-Alcala F. *Mycoplasma bovis* in respiratory disease of feedlot cattle. *Vet Clin North Am Food Anim Pract* 26, 365-379, 2010.
- 6) Celada LJ, Kropski JA, Herazo-Maya JD, Luo W, Creecy A, Abad AT, Chioma OS, Lee G, Hassell NE, Shaginurova GI, Wang Y, Johnson JE, Kerrigan A, Mason WR, Baughman RP, Ayers GD, Bernard GR, Culver DA, Montgomery CG, Maher TM, Molyneaux PL, Noth I, Mutsaers SE, Prele CM, Peebles RS Jr, Newcomb DC, Kaminski N, Blackwell TS, Van Kaer L, Drake WP. PD-1 up-regulation on CD4⁺ T cells promotes pulmonary fibrosis through STAT3-mediated IL-17A and TGF- β 1 production. *Sci Transl Med* 10, 460, 2018.
- 7) Chao J, Han X, Liu K, Li Q, Peng Q, Lu S, Zhu X, Hu G, Dong Y, Hu C, Chen Y, Chen J, Khan FA, Chen H, Guo AA. Calves Infected with Virulent and Attenuated *Mycoplasma bovis* Strains Have Upregulated Th17 Inflammatory and Th1 Protective Responses,

- Respectively. Genes (Basel) 28, 10, 2019.
- 8) Chizzolini C, Chicheportiche R, Alvarez M, de Rham C, Roux-Lombard P, Ferrari-Lacraz S, Dayer JM. Prostaglandin E₂ synergistically with interleukin-23 favors human Th17 expansion. Blood 112, 3696-3703, 2008.
 - 9) Day CL, Kaufmann DE, Kiepiela P, Brown JA, Moodley ES, Reddy S, Mackey EW, Miller JD, Leslie AJ, DePierres C, Mncube Z, Duraiswamy J, Zhu B, Eichbaum Q, Altfeld M, Wherry EJ, Coovadia HM, Goulder PJ, Klenerman P, Ahmed R, Freeman GJ, Walker BD. PD-1 expression on HIV-specific T cells is associated with T-cell exhaustion and disease progression. Nature 443, 350-354, 2006.
 - 10) Dedieu L, Balcer-Rodrigues V, Yaya A, Hamadou B, Cisse O, Diallo M, Niang M. Gamma interferon-producing CD4 T cells correlate with resistance to *Mycoplasma mycoides* subsp. *mycoides* S.C. infection in cattle. Vet Immunol Immunopathol 107, 217–233, 2005.
 - 11) Dedieu L, Balcer-Rodrigues V, Cisse O, Diallo M, Niang M. Characterization of the lymph node immune response following *Mycoplasma mycoides* subsp. *Mycoides* SC infection in cattle. Vet Res 37:579-591, 2006.
 - 12) Deeney AS, Maglennon GA, Chapat L, Crussard S, Jolivet E, Rycroft AN. *Mycoplasma hyopneumoniae* evades phagocytic uptake by porcine alveolar macrophages *in vitro*. Vet Res 50, 51, 2019.
 - 13) Dulos J, Carven GJ, van Bortel SJ, Evers S, Driessen-Engels LJ, Hobo W, Gorecka MA, de Haan AF, Mulders P, Punt CJ, Jacobs JF, Schalken JA, Oosterwijk E, van Eenennaam H, Boots AM. PD-1 blockade augments Th1 and Th17 and suppresses Th2 responses in peripheral blood from patients with prostate and advanced melanoma cancer. J Immunother 35, 169-178, 2012.
 - 14) Fox LK. *Mycoplasma* mastitis: causes, transmission, and control. Vet Clin North Am Food Anim Pract 28, 225-237, 2012.
 - 15) Gagea MI, Bateman KG, Shanahan RA, van Dreumel T, McEwen BJ, Carman S, Archambault M, Caswell JL. Naturally occurring *Mycoplasma bovis*-associated pneumonia and polyarthritis in feedlot beef calves. J Vet Diagn Invest 18, 29–40, 2006.
 - 16) Goto S, Konnai S, Okagawa T, Nishimori A, Maekawa N, Gondaira S, Higuchi H, Koiwa M, Tajima M, Kohara J, Ogasawara S, Kato Y, Suzuki Y, Murata S, Ohashi K. Increase of cells expressing PD-1 and PD-L1 and enhancement of IFN- γ production via PD-1/PD-L1 blockade in bovine mycoplasmosis. Immun Inflamm Dis 5, 355-363, 2017.
 - 17) Hale HH, Helmboldt CF, Plastringe WN, Stula EF. Bovine mastitis caused by a *Mycoplasma* species. Cornell Vet 52, 582-591, 1962.
 - 18) Higuchi H, Iwano H, Kawai K, Ohta T, Obayashi T, Hirose K, Ito N, Yokota H, Tamura Y, Nagahata H. A simplified PCR assay for fast and easy *mycoplasma* mastitis screening in dairy cattle. J Vet Sci 12, 191–193, 2011.
 - 19) Ikebuchi R, Konnai S, Shirai T, Sunden Y, Murata S, Onuma M, Ohashi K. Increase of cells expressing PD-L1 in bovine leukemia virus infection and enhancement of anti-viral immune responses *in vitro* via PD-L1 blockade Vet Res 42, 103, 2011.
 - 20) Ikebuchi R, Konnai S, Okagawa T, Yokoyama K, Nakajima C, Suzuki Y, Murata S, Ohashi K. Blockade of bovine PD-1 increases T cell function and inhibits bovine leukemia virus expression in B cells *in vitro*. Vet Res 44, 59, 2013.
 - 21) Ikebuchi R, Konnai S, Okagawa T, Yokoyama K, Nakajima C, Suzuki Y, Murata S, Ohashi K. Influence of PD-L1 cross-linking on cell death in PD-L1-expressing cell lines and bovine lymphocytes. Immunology 142, 551–561, 2014.
 - 22) Inman BA, Sebo TJ, Frigola X, Dong H, Bergstralh EJ, Frank I, Fradet Y, Lacombe L, Kwon ED. PD-L1 (B7-H1) expression by urothelial carcinoma of the bladder and BCG-

- induced granulomata: associations with localized stage progression. *Cancer* 109, 1499-1505. 2007.
- 23) Lai JF, Zindl CL, Duffy LB, Atkinson TP, Jung YW, van Rooijen N, Waites KB, Krause DC, Chaplin DD. Critical role of macrophages and their activation via MyD88-NF κ B signaling in lung innate immunity to *Mycoplasma pneumoniae*. *PLoS One* 5, e14417, 2010.
 - 24) Lerner U, Amram E, Ayling RD, Mikula I, Gerchman I, Harrus S, Teff D, Yogev D, Lysnyansky I. Acquired resistance to the 16-membered macrolides tylosin and tilmicosin by *Mycoplasma bovis*. *Vet Microbiol* 168, 365-371, 2014.
 - 25) Li X, Bechara R, Zhao J, McGeachy MJ, Gaffen SL. IL-17 receptor-based signaling and implications for disease. *Nat Immunol* 20, 1594-1602, 2009.
 - 26) Lin CC, Hsiao LD, Chien CS, Lee CW, Hsieh JT, Yang CM. Tumor necrosis factor- α -induced cyclooxygenase-2 expression in human tracheal smooth muscle cells: involvement of p42/p44 and p38 mitogen-activated protein kinases and nuclear factor- κ B. *Cell Signal* 16, 597-607, 2004.
 - 27) Mize MT, Sun XL, Simecka JW. Interleukin-17A exacerbates disease severity in BALB/c mice susceptible to lung infection with *Mycoplasma pulmonis*. *Infect Immun* 86, 9, 2018.
 - 28) Mulongo M, Prysliak T, Scruten E, Napper S, Perez-Casal J. *In vitro* infection of bovine monocytes with *Mycoplasma bovis* delays apoptosis and suppresses production of gamma interferon and tumor necrosis factor alpha but not interleukin-10. *Infect Immun* 82, 62-71, 2013.
 - 29) Nakao S, Ogtata Y, Shimizu E, Yamazaki M, Furuyama S, Sugiya H. Tumor necrosis factor alpha (TNF- α)-induced prostaglandin E₂ release is mediated by the activation of cyclooxygenase-2 (COX-2) transcription via NF κ B in human gingival fibroblasts. *Mol Cell Biochem* 238, 11-18, 2002.
 - 30) Nishimori A, Konnai S, Okagawa T, Maekawa N, Ikebuchi R, Goto S, Sajiki Y, Suzuki Y, Kohara J, Ogasawara S, Kato Y, Murata S, Ohashi K. *In vitro* and *in vivo* antiviral activity of an anti-programmed death-ligand 1 (PD-L1) rat-bovine chimeric antibody against bovine leukemia virus infection. *PLoS One* 12, e0174916, 2017.
 - 31) Okagawa T, Konnai S, Deringer JR, Ueti MW, Scoles GA, Murata S, Ohashi K, Brown WC. Cooperation of PD-1 and LAG-3 contributes to T cell exhaustion in *Anaplasma marginale*-infected cattle. *Infect Immun* 84, 2779-2790, 2016.
 - 32) Okagawa T, Konnai S, Nishimori A, Ikebuchi R, Mizorogi S, Nagata R, Kawaji S, Tanaka S, Kagawa Y, Murata S, Ohashi K. Bovine immunoinhibitory receptors contribute to the suppression of *Mycobacterium avium* subsp. *paratuberculosis*-specific T cell responses. *Infect Immun* 84, 77-89, 2016.
 - 33) Okagawa T, Konnai S, Nishimori A, Maekawa N, Ikebuchi R, Goto S, Nakajima C, Kohara J, Ogasawara S, Kato Y, Suzuki Y, Murata S, Ohashi K. Anti-Bovine Programmed Death-1 Rat-Bovine Chimeric Antibody for Immunotherapy of Bovine Leukemia Virus Infection in Cattle. *Front Immunol* 8, 650, 2017.
 - 34) Sajiki Y, Konnai S, Okagawa T, Nishimori A, Maekawa N, Goto S, Ikebuchi R, Nagata R, Kawaji S, Kagawa Y, Yamada S, Kato Y, Nakajima C, Suzuki Y, Murata S, Mori Y, Ohashi K. Prostaglandin E₂ Induction Suppresses the Th1 Immune Responses in Cattle with Johne's Disease. *Infect Immun* 86, 5, 2018.
 - 35) Sajiki Y, Konnai S, Okagawa T, Nishimori A, Maekawa N, Goto S, Watari K, Minato E, Kobayashi A, Kohara J, Yamada S, Kaneko MK, Kato Y, Takahashi H, Terasaki N, Takeda A, Yamamoto K, Toda M, Suzuki Y, Murata S, Ohashi K. Prostaglandin E₂-Induced Immune Exhaustion and

- Enhancement of Antiviral Effects by Anti-PD-L1 Antibody Combined with COX-2 Inhibitor in Bovine Leukemia Virus Infection. *J Immunol* 203, 1313-1324, 2019.
- 36) Shaw BM, Simmons WL, Dybvig K. The Vsa shield of *Mycoplasma pulmonis* is antiphagocytic. *Infect Immun* 80, 704-709, 2012.
 - 37) Sulyok KM, Kreizinger Z, Wehmann E, Lysnyansky I, Bányai K, Marton S, Jerzsele Á, Rónai Z, Turcsányi I, Makrai L, Jánosi S, Nagy SÁ, Gyuranecz M. Mutations associated with decreased susceptibility to seven antimicrobial families in field and laboratory-derived *Mycoplasma bovis* strains. *Antimicrob Agents Chemother* 61, 2, 2017.
 - 38) Thompson RH, Gillett MD, Cheville JC, Lohse CM, Dong H, Webster WS, Krejci KG, Lobo JR, Sengupta S, Chen L, Zincke H, Blute ML, Strome SE, Leibovich BC, Kwon ED. Costimulatory B7-H1 in renal cell carcinoma patients: Indicator of tumor aggressiveness and potential therapeutic target. *Proc Natl Acad Sci U S A* 101, 17174-17179, 2004.
 - 39) Van der Merwe J, Prysliak T, Perez-Casal J. Invasion of bovine peripheral blood mononuclear cells and erythrocytes by *Mycoplasma bovis*. *Infect. Immun* 78, 4570-4578, 2010.
 - 40) Vanden Bush TJ, Rosenbusch RF. *Mycoplasma bovis* induces apoptosis of bovine lymphocytes. *FEMS Immunol Med Microbiol* 32, 97-103, 2002.
 - 41) Wang X, Chen X, Tang H, Zhu J, Zhou S, Xu Z, Liu F, Su C. Increased Frequency of Th17 Cells in Children With *Mycoplasma pneumoniae* Pneumonia. *J Clin Lab Anal* 30, 1214-1219, 2016.
 - 42) Wherry EJ. T cell exhaustion. *Nat Immunol* 12, 492-499, 2011.
 - 43) Wu C, Zhu Y, Jiang J, Zhao J, Zhang XG, Xu N. Immunohistochemical localization of programmed death-1 ligand-1 (PD-L1) in gastric carcinoma and its clinical significance. *Acta Histochem* 108, 19-24, 2006.
 - 44) Yao C, Sakata D, Esaki Y, Li Y, Matsuoka T, Kuroiwa K, Sugimoto Y, Narumiya S. Prostaglandin E₂-EP4 signaling promotes immune inflammation through Th1 cell differentiation and Th17 cell expansion. *Nat Med* 15, 633-640, 2009.