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Interactive effect of leaf age and ozone on mesophyll conductance in Siebold's beech

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Mesophyll conductance (G_m) is one of the most important factors determining
photosynthesis. Tropospheric ozone (O_3) is known to accelerate leaf senescence
and causes a decline of photosynthetic activity in leaves. However, the effects of
age-related variation of O_3 on G_m have not been well investigated, and we,
therefore, analysed leaf gas exchange data in a free-air O_3 exposure experiment on
Siebold's beech with two levels (ambient and elevated O_3 : 28 and 62 $nmol\ mol^{-1}$
as daylight average, respectively). In addition, we examined whether O_3 -induced
changes on leaf morphology (leaf mass per area, leaf density and leaf thickness)

may affect CO₂ diffusion inside leaves. We found that O₃ damaged the photosynthetic biochemistry progressively during the growing season. The G_m was associated with a reduced photosynthesis in O₃-fumigated Siebold's beech in August. The O₃-induced reduction of G_m was negatively correlated with leaf density, which was increased by elevated O₃, suggesting that the reduction of G_m was accompanied by changes in the physical structure of mesophyll cells. On the other hand, in October, the O₃-induced decrease of G_m was diminished because G_m decreased due to leaf senescence regardless of O₃ treatment. The reduction of photosynthesis in senescent leaves after O₃ exposure was mainly due to a decrease of maximum carboxylation rate (V_{cmax}) and/or maximum electron transport rate (J_{max}) rather than diffusive limitations to CO₂ transport such as G_m . A leaf age×O₃ interaction of photosynthetic response will be a key for modelling photosynthesis in O₃-polluted environments.

Abbreviations

C_c	CO ₂ concentration inside the chloroplast envelope (μmol mol ⁻¹)
C_i	Sub-stomatal CO ₂ concentration (μmol mol ⁻¹)
ETR	Rate of electron transport (μmol m ⁻² s ⁻¹)
Φ	Initial slope of photosynthetic light response curve (mol mol ⁻¹)
Φ_{CO_2}	Quantum efficiency of carbon gain (dimensionless)
Φ_{PSII}	Quantum yield of PSII in the light (dimensionless)
Γ	Light compensation point (μmol mol ⁻¹)
Γ^*	CO ₂ compensation point to photorespiration (μmol mol ⁻¹)
G_m	Mesophyll conductance to CO ₂ (mol m ⁻² s ⁻¹ bar ⁻¹)
G_{sCO_2}	Stomatal conductance for CO ₂ (mol m ⁻² s ⁻¹)
G_{sH_2O}	Stomatal conductance for water vapour (mol m ⁻² s ⁻¹)

53	G_{totCO_2}	Total conductance to CO_2 ($\text{mol m}^{-2} \text{s}^{-1}$)
54	J_{max}	Maximum rate of electron transport required for RuBP regeneration (μmol
55	$\text{m}^{-2} \text{s}^{-1}$)	
56	l_b	Relative biochemical limitation to photosynthesis (%)
57	LD	Leaf density (g cm^{-3})
58	l_b	Relative mesophyll conductance limitation to photosynthesis (%)
59	LMA	Leaf mass per area (g cm^{-2})
60	l_b	Relative stomatal limitation to photosynthesis (%)
61	LT	Leaf thickness (mm)
62	P_N	Net photosynthetic rate ($\mu\text{mol m}^{-2} \text{s}^{-1}$)
63	$P_{N\text{sat}}$	Light-saturated net photosynthetic rate ($\mu\text{mol m}^{-2} \text{s}^{-1}$)
64	R_d	Respiration in the light ($\mu\text{mol m}^{-2} \text{s}^{-1}$)
65	R_n	Respiration in the dark ($\mu\text{mol m}^{-2} \text{s}^{-1}$)
66	R_{PR}	Photorespiration ($\mu\text{mol m}^{-2} \text{s}^{-1}$)
67	V_{cmax}	Maximum rate of RuBP carboxylation ($\mu\text{mol m}^{-2} \text{s}^{-1}$)
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Introduction

Anthropogenic emissions of air pollutants such as nitrogen oxides (NO_x) and volatile organic compounds have elevated the ground-level concentration of ozone (O₃) since pre-industrial times (Mills et al. 2018). Ozone is phytotoxic to plants, adversely affecting physiological and biochemical processes contributing to a decline in plant growth (Matyssek et al. 2013, Grulke and Heath 2020).

In cool-temperate forests in Japan, deciduous broadleaved Siebold's beech (*Fagus crenata*) is the dominant tree species (Koike et al. 1998). Previous experimental studies have shown that Siebold's beech is highly sensitive to O₃ (e.g. Yamaguchi et al. 2011, Izuta 2017). Watanabe et al. (2012) estimated that the current concentration of O₃ in Japan may reduce the growth of Siebold's beech from 3.2% to a maximum of 9.7%. This O₃ induced reduction in tree growth is considered to be related to lower photosynthetic activity (Watanabe et al. 2014a). The exact mechanisms underlying the effects of O₃ on photosynthesis are still unclear. However, previous studies have suggested that O₃ causes biochemical limitations associated with reduced ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) activity in leaves (Fiscus et al. 2005, Hoshika et al. 2013, Watanabe et al. 2014a, 2014b, Bagard et al. 2015). The loss of Rubisco activity is commonly inferred from decreases in the maximum rate of carboxylation (V_{cmax}), as indicated by a decrease in the initial slope of a P_N/C_i curve (net photosynthetic rate versus estimated sub-stomatal CO₂ concentration; e.g. Matyssek et al. 2012, Watanabe et al. 2013). Any loss in the biochemical efficiency of CO₂ assimilation induced by O₃ exposure may also be evident in a decline in the maximum rate of electron transport required for ribulose-1,5-bisphosphate (RuBP) regeneration (J_{max} ; Hoshika et al. 2013). The traditional calculation of photosynthetic parameters from P_N/C_i curves assumes an infinite mesophyll

conductance to CO₂ (G_m ; e.g. Long and Bernacchi 2003). However, G_m may exert similar limits to P_N as stomatal resistance to CO₂ transport (Terashima et al. 1995, Lauteri et al. 2014, Killi and Haworth 2017, Gago et al. 2019, Veromann-Jürgenson et al. 2020). This indicates that previous observations of the O₃-induced reduction of V_{cmax} and J_{max} parameters calculated from P_N/C_i curves assuming infinite G_m incorporate not only biochemical limitations but also the effect of G_m (Warren et al. 2007).

The generation of reactive oxygen species during O₃ exposure could disrupt the physical integrity and biochemistry of mesophyll cells (Kangasjärvi et al. 2005). This would suggest that the exposure to O₃ would impair both the physical and biochemical transport of CO₂ from the sub-stomatal leaf air-space to chloroplasts. In fact, a reduction in G_m was observed in O₃-sensitive snapbean (*Phaseolus vulgaris*) exposed to 60 nmol mol⁻¹ O₃, while no effect was observed on the G_m values of resistant varieties (Flowers et al. 2007). Xu et al. (2019) found that the O₃-induced reduction of photosynthesis was mainly due to a decreased G_m in a hybrid poplar clone. In addition, Watanabe et al. (2018a) reported that O₃ exposure reduced G_m in Siebold's beech without any effect on C_c -based V_{cmax} . However, European beech (*F. sylvatica*) grown in an O₃ FACE system showed no effect of a doubling of ambient O₃ concentration on G_m (Warren et al. 2007). Likewise, fumigation of localized areas of Holm oak (*Quercus ilex*) and pubescent oak (*Q. pubescens*) leaves with O₃ between 50 to 350 nmol mol⁻¹ did not reduce G_m (Velikova et al. 2005).

Mesophyll conductance is generally lower in leaves with thicker cell walls and greater leaf density (Adachi et al. 2013, Ivanova et al. 2018) that are conditions limiting the diffusion of CO₂ in the leaf liquid phase (Tomás et al. 2013, Onoda et al. 2017). Ozone may affect leaf morphology, which is likely to modify the CO₂

diffusion inside leaves (Heath 1994, Moura et al. 2018). The white poplar (*Populus alba*) grown in an O₃-enriched atmosphere (150 nmol mol⁻¹) developed thicker and denser leaves than that grown in clean air (Fares et al. 2006). Likewise, leaf density (LD) was increased by O₃ in European silver birch (*Betula pendula*; Oksanen 2005) and European beech (Bussotti et al. 2005). In contrast, the leaves of Pima cotton (*Gossypium barbadense*) formed in O₃-enriched air showed lower mesophyll density (Grantz and Shrestha 2006), probably due to damage and disruption of the mesophyll cells (Günthardt-Goerg et al. 1993). Watanabe et al. (2018a) did not find any effect of O₃ on leaf morphology in Siebold's beech, although plants were exposed to relatively low O₃ concentrations (29 to 47 nmol mol⁻¹ as daylight average). The density of cells in the mesophyll layer is closely related to leaf mass per area (LMA; Haworth and Raschi 2014). Meta-analysis suggests that the effect of O₃ on LMA is not pronounced or uni-directional (Poorter et al. 2009), raising the likelihood that O₃ effects on leaf structure depend on species or varieties (Pääkkönen et al. 1995).

Previous works have shown that G_m is variable with leaf age (Loreto et al. 1994, Niinemets et al. 2009, Tosens et al. 2012, Marino et al. 2020). After the onset of leaf senescence, G_m reduced due to age-dependent variation of leaf structure and physiological activity (Niinemets et al. 2009). It is known that leaf senescence is accelerated by O₃ (Grulke and Heath 2020). As a matter of fact, O₃-induced structural changes in chloroplasts appear to be similar to those during autumn senescence (e.g. Vollenweider et al. 2019), which may be associated with a reduced mesophyll CO₂ diffusion. The question was therefore raised if O₃ and leaf age could interactively affect G_m , but the age-related change of O₃ effect on G_m has not been addressed in previous studies.

To examine a leaf age×O₃ interaction on G_m , we measured leaf gas exchange

three times (June, August, October) in a free-air O₃ exposure experiment on Siebold's beech. The aim of the present study was to: (1) characterise the effect of O₃ fumigation on diffusive resistances to P_N , in particular the impact on G_m values derived from the variable J method (Harley et al. 1992, Loreto et al. 1992), while comparing with those derived from the P_N/C_i curve fitting approach (Ethier and Livingston 2004); (2) assess the impact of O₃ exposure on the efficiency of photosynthetic light capture; and (3) analyse the interaction between O₃ fumigation and leaf morphology (LMA, LD and leaf thickness), and the impact upon photosynthetic physiology throughout the growing season.

Materials and Methods

Experimental site and plant material

Measurements were conducted at a free-air O₃ exposure experiment facility located in the Sapporo Experimental Forest, Hokkaido University, in northern Japan (43°04' N, 141°20' E, 15 m a.s.l., mean annual temperature: 8.9°C, mean total annual precipitation: 1107 mm, the snow-free period was late April to mid-November). The details of the system are described in Watanabe et al. (2013). The soil is a brown forest soil (Dystric Cambisols). Two plots were established, one for ambient O₃ (AA) and the other for elevated O₃ (eO₃). The size of each plot was 5.5 m × 7.2 m. The distance between the O₃-enhanced plot and the ambient plot was about 20 m, which was enough distance for the O₃-enriched air not to affect the AA plot (Watanabe et al. 2013). Ten seedlings of two-year-old Siebold's beech (*Fagus crenata*) were obtained from a nearby nursery and planted in each plot in May 2003 and then grown under ambient conditions from 2003 to 2010. The plants were ten years old and well-established when fumigation with O₃ began in 2011. The 11-year-old Siebold's beech saplings analysed in this study were around 3.4 m in height in 2012. We confirmed that photosynthetic parameters, plant height and diameter were not different between treatments before O₃ exposure (in July 2011, data not shown). The target O₃ concentration was 60 nmol mol⁻¹ during daylight hours. The enhanced O₃ treatment was employed during daylight hours to

ten Siebold's beech saplings from August to November in 2011, and from May to November in 2012. Ozone concentrations at canopy height were recorded continuously by an O₃ monitor (Mod. 202, 2B Technologies). The daytime hourly mean of O₃ concentration in AA and eO₃ were 28 nmol mol⁻¹ and 62 nmol mol⁻¹, respectively, during the 2012 experimental period. The level of O₃ concentration in eO₃ was equal to the environmental standard value in Japan (= 60 nmol mol⁻¹), and has been frequently observed in many Asian regions including forested areas (Izuta 2017). The soil moisture was measured in the root layer (at depth 20 cm) by 10HS sensors equipped with an EM5b data logger (Decagon Devices). The average soil moisture (volumetric soil water content) was $28.1 \pm 2.8\%$ during these measurements. These values were equivalent to the field capacity of the soil. Previous studies have established that no reduction in P_N due to soil water content occurred during the growing season for Siebold's beech in this experiment (Hoshika et al. 2012, 2013, Watanabe et al. 2014a).

Measurement of leaf gas exchange

Leaf gas exchange was measured in fully expanded leaves exposed to full sun at the top of the canopy ($n = 6$ trees in each O₃ treatment) using a Li6400 portable infra-red gas analyzer (Li-Cor instruments). Only first flush leaves (flushed in May) were used for these measurements, because the number of second flush leaves was very small (0.16% of the total leaf number taken over all saplings). Measurements were undertaken on 15-16 June, 26-31 August and 9-16 October on days with clear sky between 8:00 and 16:00 h. Conditions within the leaf cuvette were set to maintain a leaf temperature of 25°C and the leaf-to-air vapour pressure deficit (VPD) within a range of 1.0-1.8 kPa. The Li6400 was connected to a 6400-40 leaf chamber fluorimeter cuvette with red and blue LEDs when conducting the P_N – photosynthetic photon flux density (PPFD) response curves. The response of P_N to PPFD was measured at 11 light intensities (1500, 1100, 800, 500, 400, 300, 200, 100, 75, 50, 0 $\mu\text{mol m}^{-2} \text{s}^{-1}$). At each stage of the P_N /PPFD curve, P_N , stomatal conductance ($G_{\text{H}_2\text{O}}$), and the quantum yield of PSII in the light (Φ_{PSII}) were recorded. The P_N /PPFD response curves were utilised to calculate the light saturated rate of P_N ($P_{N\text{sat}}$), initial slope of photosynthetic light response curve (Φ), light compensation point (Γ) and the quantum efficiency of carbon gain (Φ_{CO_2}) following Kaipiainen et al. (2009). The Kok (1948) method was used to estimate

respiration in the light (R_d). The effect of variation in the sub-stomatal concentration of $[\text{CO}_2]$ (C_i) on R_d was corrected using the iterative method of Kirschbaum and Farquhar (1987). Respiration in the dark (R_n) was measured by switching off the LED light unit after the Kok protocol, shading the area of the plant being measured and recording the rate of CO_2 efflux from the leaf, after values had remained stable for 5-10 min. Mesophyll conductance was calculated at $P_{N\text{sat}}$ using the variable J method as described by Loreto et al. (1992):

$$G_m = \frac{P_{N\text{sat}}}{C_i - \frac{\Gamma^* \cdot [ETR + 8 \cdot (P_N + R_d)]}{ETR - 4 \cdot (P_N + R_d)}} \quad (1),$$

where the electron transport rate (ETR) is calculated from fluorescence (Genty et al. 1989) as following:

$$ETR = \text{PPFD} \cdot \phi\text{PSII} \cdot \alpha \cdot \beta \quad (2),$$

where the partitioning factor (β) between photosystems I and II was considered to be 0.5 and leaf absorbance ($\alpha = 0.85$; Laik and Loreto 1996). The CO_2 compensation point to photorespiration (Γ^*) was calculated using the Rubisco specificity factor of Galmes et al. (2005). Total conductance to CO_2 (G_{totCO_2}) was calculated as:

$$G_{\text{totCO}_2} = [G_{s\text{CO}_2} \cdot G_m] / [G_{s\text{CO}_2} + G_m] \quad (3).$$

where $G_{s\text{CO}_2}$ is the stomatal conductance for CO_2 . The concentration of $[\text{CO}_2]$ inside the chloroplast envelope (C_c) was calculated using the value of G_m produced by the variable J method as:

$$C_c = C_i - \frac{P_{N\text{sat}}}{G_m} \quad (4).$$

Photorespiration (R_{PR}) was determined following Sharkey (1988):

$$R_{PR} = \frac{P_{N\text{sat}} + R_d}{\frac{C_c}{\Gamma^*} - 1} \quad (5).$$

Hoshika et al. (2013) previously reported the P_N/C_i curves of the Siebold's beech exposed to O₃-FACE, based on the approach by Long and Bernacchi (2003) that assumed infinite G_m . Here the P_N/C_i data were re-analysed using the protocol of Ethier and Livingston (2004) to consider G_m values using the curve-fitting approach. The P_N/C_i curves were obtained by measuring P_N over 12 [CO₂] steps (C_a , 380, 300, 220, 140, 60, 300, 380, 500, 800, 1100, 1400, 1700 $\mu\text{mol mol}^{-1}$). The response of P_N to C_i was then used to calculate the carboxylation capacity of ribulose-1,5-bisphosphate carboxylase/ oxygenase (Rubisco; $V_{c\text{max}}$) and the maximum rate of electron transport required for ribulose-1,5-bisphosphate (RuBP) regeneration (J_{max}), assuming that G_m were constant at all [CO₂] steps. All gas exchange data were corrected for leaks from gaskets using an empty cuvette and the entire chamber head was enclosed in a plastic bag to minimise the effect of leak (Flexas et al. 2007).

According to Grassi and Magnani (2005), the relative limitations in $P_{N\text{sat}}$ (stomatal limitation, l_s ; mesophyll conductance limitation, l_m ; biochemical limitation, l_b) were determined as follows:

$$l_s = \frac{G_{\text{totCO}_2}/G_{s\text{CO}_2} \cdot \delta P_{N\text{sat}}/\delta C_c}{G_{\text{totCO}_2} + \delta P_{N\text{sat}}/\delta C_c} \quad (6),$$

$$l_m = \frac{G_{\text{totCO}_2}/G_m \cdot \delta P_{N\text{sat}}/\delta C_c}{G_{\text{totCO}_2} + \delta P_{N\text{sat}}/\delta C_c} \quad (7),$$

$$l_b = \frac{G_{\text{totCO}_2}}{G_{\text{totCO}_2} + \delta P_{N\text{sat}}/\delta C_c} \quad (8),$$

where $\delta P_{N\text{sat}}/\delta C_c$ is an initial slope of P_N/C_c curves (C_c : over a range of 0–150 $\mu\text{mol mol}^{-1}$).

Measurements of leaf traits

After the measurement of gas exchange rate, leaves were collected for the determination of the leaf mass per unit area (LMA), leaf density (LD) and leaf thickness (LT). LT was measured avoiding the mid-vein by using a hand-held micrometer (Digimatic micrometer, Mitsutoyo). The projected area (one side of the leaf) was determined with image-processing software (LIA 32, Nagoya University,

Japan). Leaves were then dried in an oven at 70°C for 1 week and were then weighed. The LMA was calculated as the ratio of the dry mass to the area of the leaves. The LD was calculated as the leaf dry mass per unit leaf volume.

Data analysis

Statistical analyses were performed using R 3.5.1 (R Core Team 2018). To test the effects of O₃ fumigation treatment and measurement months, a two-way ANOVA was used to assess differences in variance between samples. Linear regression was used to investigate possible relationships between P_N and diffusive limitations to CO₂ transport and leaf traits. For these analyses, we used G_m values derived from variable J method. Principle component analysis (PCA) was used to investigate patterns in the photosynthetic capacity, light harvesting and diffusive components of photosynthesis in Siebold's beech after fumigation with O₃. Results were considered significant at $P < 0.05$.

Results

Leaf density is modulated by ozone and season

No significant effects of O₃ or measurement month on LMA and LT were found (Table 1). On the other hand, LD was significantly increased by O₃. In addition, LD increased with season as it was relatively smaller in June (0.45 g cm⁻³ in AA and 0.44 g cm⁻³ in eO₃) compared to in October (0.48 g cm⁻³ in AA and 0.54 g cm⁻³ in eO₃).

Photosynthetic parameters are progressively decreased by ozone

Most photosynthetic parameters of Siebold's beech were significantly affected by O₃ exposure (Fig. 1, Table 2). Ozone decreased P_{Nsat} of Siebold beech trees in

August (-28%) and October (-46%), although $P_{N_{sat}}$ values in both O₃ treatments were similar in June (-7% in eO₃ compared to in AA; Fig. 1A). Significant increases of R_n were induced by O₃ (+70% in August and +88% in October; Fig. 1B). There were no significant changes in R_{PR} associated with elevated O₃ in either June or August (Fig. 1C). However, O₃ decreased R_{PR} by 56% relative to AA plants in October. Ozone decreased G_{sH_2O} , G_m and G_{totCO_2} , although significant differences were not observed between the treatments in October (Fig. 1D-F). In addition, the measurements by fluorimeter indicated that the PSII electron transport rate (ETR) and actual quantum efficiencies of PSII (Φ_{PSII}) were significantly decreased by O₃ (Table 2, Fig. 2H,L). The eO₃ treatment significantly reduced $P_{N_{sat}}$ and values of V_{cmax} and J_{max} from the P_N/C_c curves (Table 2, Fig. S1). A significant increase in the J_{max}/V_{cmax} ratio was found in October due to a larger reduction in V_{cmax} than J_{max} after O₃ exposure. Estimated values of G_m derived from the P_N/C_i response curves approach were matched with those calculated by the variable J method (Fig. S2).

Rates of $P_{N_{sat}}$ were positively correlated to the measures of CO₂ conductance under AA conditions (Fig. 3A-C). However, in eO₃, $P_{N_{sat}}$ was not associated with G_{sH_2O} (Fig. 3A). In addition, no significant relationship between G_m and G_{sH_2O} was found in eO₃ (Fig. 3D).

The light-saturated net photosynthetic rate ($P_{N_{sat}}$) and diffusive conductance parameters (G_{sH_2O} , G_m and G_{totCO_2}) were not correlated with LD under the AA condition (Fig. 4A-D). On the other hand, in eO₃, $P_{N_{sat}}$, G_m and G_{totCO_2} showed negative relationships with LD (Fig. 4A,C,D), while there was no significant correlation between LD and G_{sH_2O} (Fig. 4B). LMA and LT did not correlate to those diffusive conductance parameters in both O₃ treatments (Figs S3 and S4).

Relative limitations to photosynthesis are affected by ozone and season

The relative limitations to photosynthesis (l_s , l_m and l_b) are shown in Table 3. Relative limitation to photosynthesis by stomata (l_s) decreased with season. In particular, in eO₃, the reduction in l_s was remarkable in the end of the growing season (l_s : -39% in August, -54% in October compared to June). On the other hand, the l_b increased over the season, and this was highlighted in eO₃ according to the post-hoc Tukey test. A relative limitation to photosynthesis by mesophyll conductance (l_m) was rather constant among the sampling campaigns (approximately 30 to 40% over the season).

Principal component analysis revealed the interaction of leaf age and ozone on photosynthetic response

Principal component analysis of parameters derived from analyses of the photosynthetic capacity (P_N/C_c curves), light harvesting ($P_N/PPFD$ curves) and diffusive limitations to CO₂ transport (G_{sH_2O} , G_m and G_{totCO_2}) of Siebold's beech indicated a high degree of overlap in the multi-variate space occupied by AA- and eO₃-fumigated plants when measured in June (Fig. 5; student's t-test of PCA scores: component 1, $P=0.215$; component 2, $P=0.646$). However, the measurement month affected variations in component 1 and component 2, while O₃ was a significant factor affecting only component 1 (two-way ANOVA of PCA scores: component 1, O₃: $P<0.001$, Month: $P<0.001$, O₃×Month: $P=0.002$; component 2, O₃: $P=0.164$, Month: $P=0.029$, O₃×Month: $P=0.059$). In fact, eO₃-treated plants did not overlap those treated by AA in August and October. In addition, Siebold's beech measured during October occupied distinct multivariate space in comparison to previous analyses in June and August. In this analysis, component 1 accounts for 49.2% of the variance within the dataset and component 2 accounts for 14.8%

of variance.

Discussion

Ozone effects on mesophyll conductance and other photosynthetic parameters

We analysed leaf gas exchange data in Siebold's beech exposed to elevated O_3 to assess whether O_3 affected G_m . We found that O_3 caused a significant reduction of G_m in August accompanied by a reduction of P_{Nsat} (Fig. 3B). On the other hand, no significant relationship between G_{sH_2O} and P_{Nsat} was found in eO_3 (Fig. 3A). As a result, G_m could better explain the observed reduction of photosynthetic rate rather than G_{sH_2O} in eO_3 (Table 3). In addition, the present study indicates that eO_3 caused a decoupling of G_{sH_2O} from G_m (Fig. 3D), although a significant relationship has been previously found between G_{sH_2O} and G_m (Lloyd et al. 1992, Lauteri et al. 2014). Although stomata generally regulate CO_2 uptake, several studies have shown evidence that chronic O_3 exposure impairs efficient stomatal control of gas exchange (Paoletti 2005, Hoshika et al. 2019, 2020).

In the previous report by Hoshika et al. (2013), O_3 was found to decrease C_i -based V_{cmax} of Siebold's beech leaves in August and October based on the calculation from P_N/C_i curves assuming an infinite mesophyll conductance to CO_2 (Long and Bernacchi 2003). Such a decrease of C_i -based V_{cmax} is, however, attributed to a decreased G_m or degradation of Rubisco activity (Loreto et al. 1994). In the present study, O_3 decreased G_m in August while C_c -based V_{cmax} was not significantly altered (Fig. 1E, Table 2), which is supported by previous findings in Siebold's beech (Watanabe et al. 2018a). In addition, we found that O_3 exposure decreased C_c -based V_{cmax} and J_{max} in October while no reduction was found in G_{sH_2O} , G_m and G_{totCO_2} . The results suggest that a reduction in G_m may be a primary response in O_3 -induced decrease of photosynthesis of Siebold's beech.

Leaf age effects on mesophyll conductance and other photosynthetic parameters

Mesophyll conductance (G_m) decreased with leaf ageing, in association with a reduction of the leaf photosynthetic rate (Fig. 1A, E). Leaf morphological traits were not significantly correlated with G_m during the ageing process in control condition (AA). This is probably because the change in leaf morphology was small during leaf ageing in AA, although LD statistically significantly increased with season. Tosens et al. (2012) similarly reported the slow increase in LD and LMA with leaf ageing after leaf maturity in *Populus tremula*, while LT remained constant. The observed reduction of G_m in senescent leaves may be attributed to age-dependent alterations such as changes of chloroplasts number and size, the location of chloroplasts, photosynthetic enzyme content, and aquaporin status (Evans and Vellen 1996, Niinemets et al. 2009). In addition to G_m , most of photosynthetic parameters decreased with leaf ageing (Fig. 1, Table 2). Interestingly, the relative contribution to photosynthesis by mesophyll conductance (I_m) was rather constant over the season (Table 3). Additionally, the stomatal limitation to photosynthesis (I_s) decreased with leaf ageing, while the relative biochemical limitation (I_b) increased in the end of the growing season. It is known that stomata in senescent leaves remain open even in low light conditions, called the “dull leaf” phenomenon. They are, therefore, unable to regulate leaf gas exchange efficiently (Terashima, 2002). On the other hand, the low biochemical photosynthetic capacity in senescent leaves was due to the decline of leaf nitrogen (Watanabe et al., 2018b), which may further limit photosynthesis in autumn. These results suggest that photosynthesis was primarily determined by diffusive limitations of CO₂ transport during summer in Siebold’s beech, while biochemical limitations were relatively important in

senescent leaves in autumn. This seasonal change is consistent with the PCA analysis for photosynthetic characteristics (Fig. 5).

Interactive effects of ozone and leaf age on leaf gas exchange

Leaf senescence is often accelerated by O₃ exposure (Grulke and Heath 2020). The effects of O₃ on leaf gas exchange were similar to those of leaf senescence, i.e. significant reductions in the photosynthetic capacity, light harvesting and diffusive components of photosynthesis, but the reduction of photosynthetic capacity was observed earlier in eO₃ than in AA (Fig. 1A-F, Table 2). Mesophyll conductance (G_m) showed a sharp decrease after O₃ exposure in August and no further change in October. Meanwhile, LD increased with leaf age especially at eO₃ (Table 1). Previous studies similarly reported that O₃ altered leaf cell ultrastructure in Siebold's beech (Yonekura et al. 2001) and increased LD in European beech (Bussotti et al. 2005). A larger LD may decrease CO₂ diffusion in the liquid phase and, thus, G_m (Niinemets 2015). As a result, a significant relationship between G_m and LD over the season was found at eO₃ (Fig. 4C), suggesting that the reduction of G_m was accompanied by changes in the physical structure of mesophyll cells. It is known that some leaf structural changes are highlighted only when senescence is accelerated by environmental stress (Vollenweider et al. 2019). In fact, as a leaf age×O₃ interaction, inner cell wall thickening has been observed in aged leaves after O₃ exposure due to progressing cell injury in eO₃ (Günthardt-Goerg and Vollenweider 2007). However, we should note that the correlation between LD and G_m was not strong ($R^2 = 0.24$). This suggests that there may also be other factors causing a reduced G_m after O₃ exposure. For example, aquaporins are involved in the regulation of G_m (e.g. Hanba et al. 2004) and water across the bundle sheath (Shatil-Cohen et al. 2011). Ozone and the resultant reactive oxygen species (ROS)

reach the cell membrane (Vainonen and Kangasjärvi 2015), which may affect G_m through direct effects on aquaporins. In fact, O_3 may decrease leaf hydraulic conductance (Zhang et al. 2018), which is related to vein architecture in association with aquaporins (Scoffoni and Sack 2017). This hypothesis needs to be tested in further studies.

Limiting factors for the reduction of P_N after O_3 exposure may vary with leaf age. In fact, diffusive resistances to CO_2 transport were dominant factors for photosynthesis at e O_3 in August, while the reduction of P_N after O_3 exposure in October was mainly due to the co-limitation of V_{cmax} and J_{max} (Table 3). At the same time, O_3 progressively increased the J_{max}/V_{cmax} ratio with leaf age (Table 2). This increase in J_{max}/V_{cmax} ratio suggests that the photosynthetic rate was limited more by Rubisco carbon fixation than RuBP regeneration by electron transport. Such a selective degradation of Rubisco quantity or activity was also observed in a poplar clone exposed to O_3 (e.g. Bagard et al. 2008).

Rubisco activity is closely related to the capacity of photorespiration (R_{PR}) (Farquhar et al. 1980). In fact, R_{PR} in Siebold's beech was decreased by enhanced O_3 treatments with leaf ageing (Fig. 1C). The reduced R_{PR} is equivalent to reduced photoprotective function that, alongside a lower energy usage for photochemistry, would produce ROS, damaging photosynthetic biochemistry (Kangasjärvi et al., 2005). In addition, the plant photorespiratory system requires the glycine decarboxylase (GDC) reaction cycle in the mitochondria, which requires the oxidized form of nicotinamide adenine dinucleotide (NAD^+) and generates NADH (the reduced form of NAD; Obata et al. 2016). This decrease in R_{PR} may result in a reduced NADH/ NAD^+ ratio stimulating mitochondrial respiration (Dizengremel et al. 2012) as confirmed here in the O_3 -induced increase of dark respiration rate (Fig. 1B).

Conclusion

Our results indicate that O₃ damaged photosynthetic mechanisms in Siebold's beech leaves. Mesophyll conductance was associated with a reduced photosynthesis in O₃-fumigated Siebold's beech in August. This O₃-induced reduction of G_m was negatively correlated with leaf density, which was increased by O₃ exposure. This suggests that the reduction of G_m in August was accompanied by changes in the physical structure of mesophyll cells. However, the response of photosynthetic parameters to O₃ was different in the later growing season. In October, the O₃-induced reduction of photosynthesis was mainly due to the co-limitation of V_{cmax} and J_{max} rather than diffusive resistances to CO₂ transport. Mesophyll conductance (G_m) decreased in October due to leaf senescence regardless of O₃ treatments. In consequence, the O₃-induced decrease of G_m was diminished. These results suggest that seasonal changes of the photosynthetic response to O₃ will be a key to modelling photosynthesis in an O₃-polluted environment.

Author contributions

Y.H. conceived the study and methodology, and analysed leaf gas exchange data from the ozone FACE experiment performed by Y.H., M.W. and T.K.; M.H. co-conceived the study and contributed to the analysis of light-response curve. All authors were involved in writing the paper, although Y.H. took a lead role.

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Data availability statement

Data sharing is not applicable to this article as all new created data is already contained within this article.

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Supporting information

Additional Supporting Information may be found in the online version of this article:

Fig. S1. Responses of net photosynthetic rate (P_N) to the sub-stomatal concentration of CO₂ (C_i) or the concentration of CO₂ inside the chloroplast envelope (C_c).

Fig. S2. Relationship between mesophyll conductance (G_m) values derived from the variable J method and those derived from the P_N/C_i curve fitting method.

Fig. S3. Relationships between leaf gas exchange parameters and leaf mass per

area (LMA) of Siebold's beech saplings.

Fig. S4. Relationships between leaf gas exchange parameters and leaf thickness (LT) of Siebold's beech saplings.

Figure legends

Figure 1. Photosynthetic parameters of Siebold's beech grown under ambient (AA) and elevated O_3 (eO_3). (A) Light-saturated net photosynthetic rate [P_{Nsat}], (B) dark respiration rate [R_N], (C) photorespiration [R_{PR}], (D) stomatal conductance to water vapor [G_{sH_2O}], (E) mesophyll conductance to CO_2 [G_m], (F) total conductance to CO_2 [G_{totCO_2}]. Each value is the mean $\pm$ standard error (n=6). Two-way ANOVA: * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, ns denotes not significant. Different letters show significant differences among treatments ($P < 0.05$, Tukey test).

Figure 2. Responses of net photosynthetic rate (P_N), stomatal conductance (G_{sH_2O}), mesophyll conductance (G_m) and electron transport rate (ETR) to photosynthetic photon flux density (PPFD). The P_N /PPFD curves were obtained by measuring P_N over 11 PPFD steps (1500, 1100, 800, 500, 400, 300, 200, 100, 75, 50, 0 $\mu mol\ m^{-2}\ s^{-1}$) in Siebold's beech exposed to different O_3 concentrations (ambient [AA] or elevated O_3 [eO_3]). Each value is the mean $\pm$ standard error (n=6).

Figure 3. Relationships between (A) light-saturated net photosynthetic rate [P_{Nsat}] and stomatal conductance for water vapor [G_{sH_2O}], (B) P_N and mesophyll conductance to CO_2 [G_m], (C) P_N and total conductance to CO_2 [G_{totCO_2}], (D) G_m and G_{sH_2O} , in Siebold's beech. Linear regression analysis was used to assess the significance of the relationship in ambient (AA) and elevated O_3 (eO_3). * $P < 0.05$, *** $P < 0.001$, ns denotes not significant.

Figure 4. Relationships between leaf gas exchange parameters (light-saturated net photosynthetic rate [P_{Nsat}], stomatal conductance [G_{sH_2O}], mesophyll conductance

[G_m] and total conductance to CO_2 [G_{totCO_2}]) and leaf density (LD) of Siebold's beech saplings under different O_3 concentrations (ambient [AA] or elevated O_3 [eO_3]). Simple linear regression analysis was applied to assess the relationships. * $P < 0.05$, ns denotes not significant.

Figure 5. Principal component analysis of parameters derived from analyses of the photosynthetic capacity (P_N/C_c curves), light harvesting (P_N/PPFD curves) and diffusive limitations to CO_2 transport (G_{sCO_2} , G_m and G_{totCO_2}) of Siebold's beech grown under different O_3 concentrations (ambient [AA] or elevated O_3 [eO_3]). Ellipses represent 95% confidence intervals for measurements conducted in June, August and October. The correlation circle is also shown. Measurements of AA-treated plants in June and August and eO_3 -treated plants in June occupy the same position in multi-variate space. On the other hand, eO_3 -treated plants analysed in October occupy statistically distinct multi-variate space. Component 1 accounts for 49.2% of the variance within the dataset and component 2 accounts for 14.8% of variance.

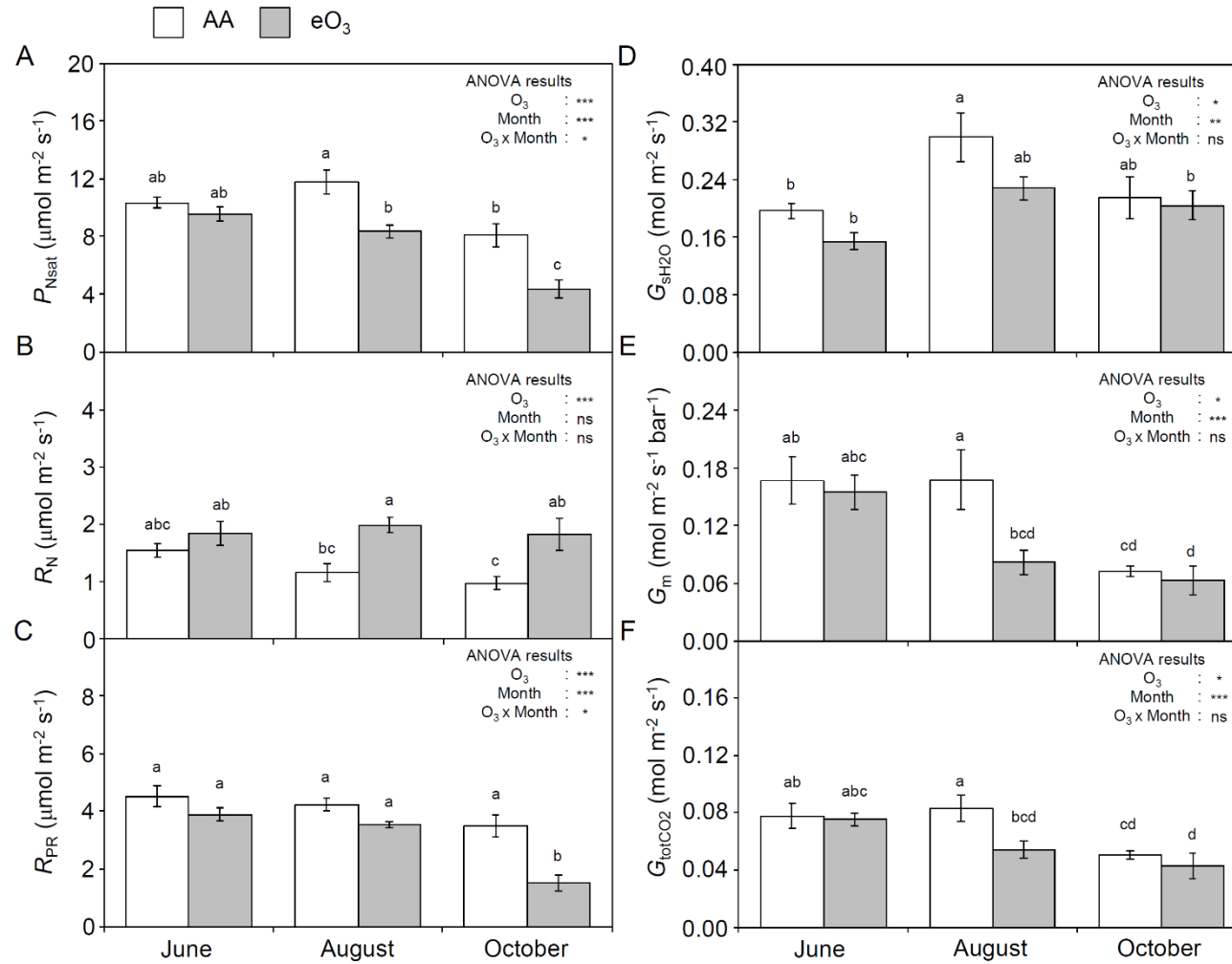


Figure 1. Photosynthetic parameters of Siebold's beech grown under ambient (AA) and elevated O_3 (eO_3): (A) Light-saturated net photosynthetic rate [P_{Nsat}], (B) dark respiration rate [R_N], (C) photorespiration [R_{PR}], (D) stomatal conductance to water vapor [G_{SH_2O}], (E) mesophyll conductance to CO_2 [G_m], (F) total conductance to CO_2 [G_{totCO_2}]. Each value is the mean $\pm$ standard error ($n = 6$). Two-way ANOVA: * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, ns denotes not significant. Different letters show significant differences among treatments ($p < 0.05$, Tukey test).

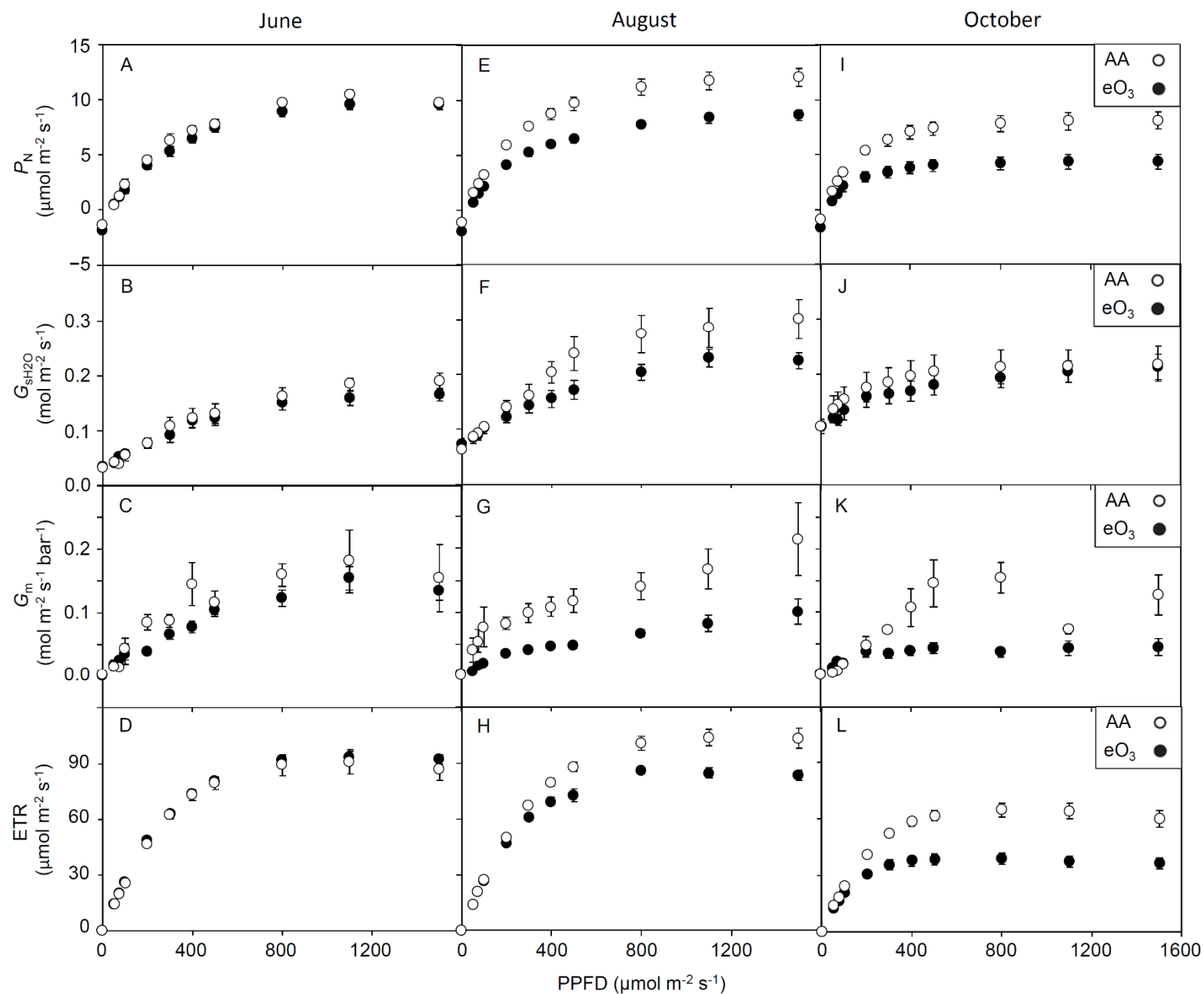


Figure 2. Responses of net photosynthetic rate (P_N), stomatal conductance (G_{SH2O}), mesophyll conductance (G_m) and electron transport rate (ETR) to

photosynthetic photon flux density (PPFD). The P_N /PPFD curves were obtained by measuring P_N over 11 PPFD steps (1500, 1100, 800, 500, 400, 300, 200, 100, 75, 50, 0 $\mu\text{mol m}^{-2} \text{s}^{-1}$) in Siebold's beech exposed to different O_3 concentrations (ambient [AA] or elevated O_3 [e O_3]). Each value is the mean $\pm$ standard error (n = 6).

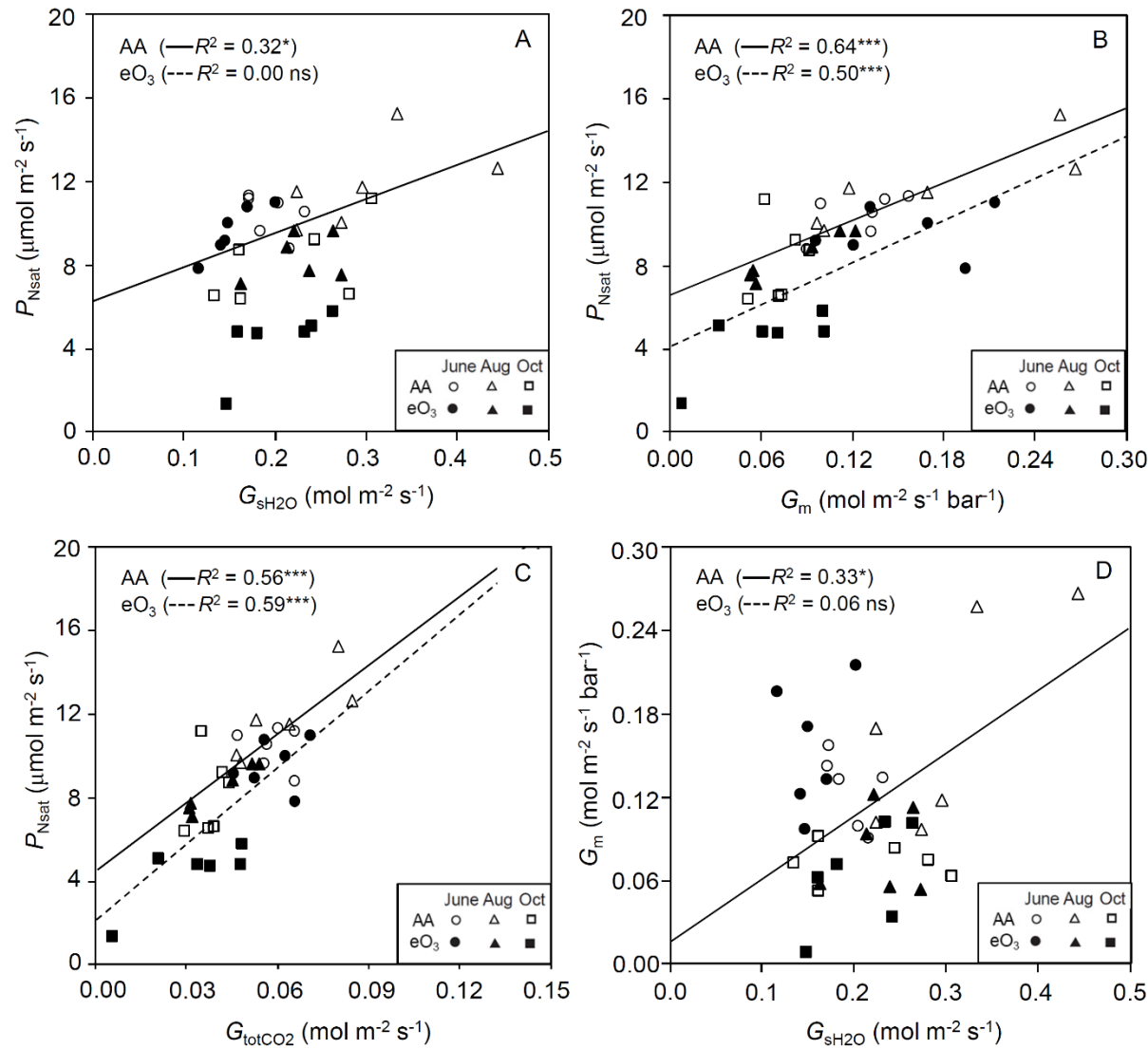


Figure 3. Relationships between (A) light-saturated net photosynthetic rate [P_{Nsat}] and stomatal conductance for water vapor [G_{sH2O}], (B) P_N and mesophyll conductance to CO₂ [G_m], (C) P_N and total conductance to CO₂ [G_{totCO2}], (D) G_m and G_{sH2O} , in Siebold's beech. Linear regression analysis was used to assess

the significance of the relationship in ambient (AA) and elevated O₃ (eO₃). * $p < 0.05$, *** $p < 0.001$, ns denotes not significant.

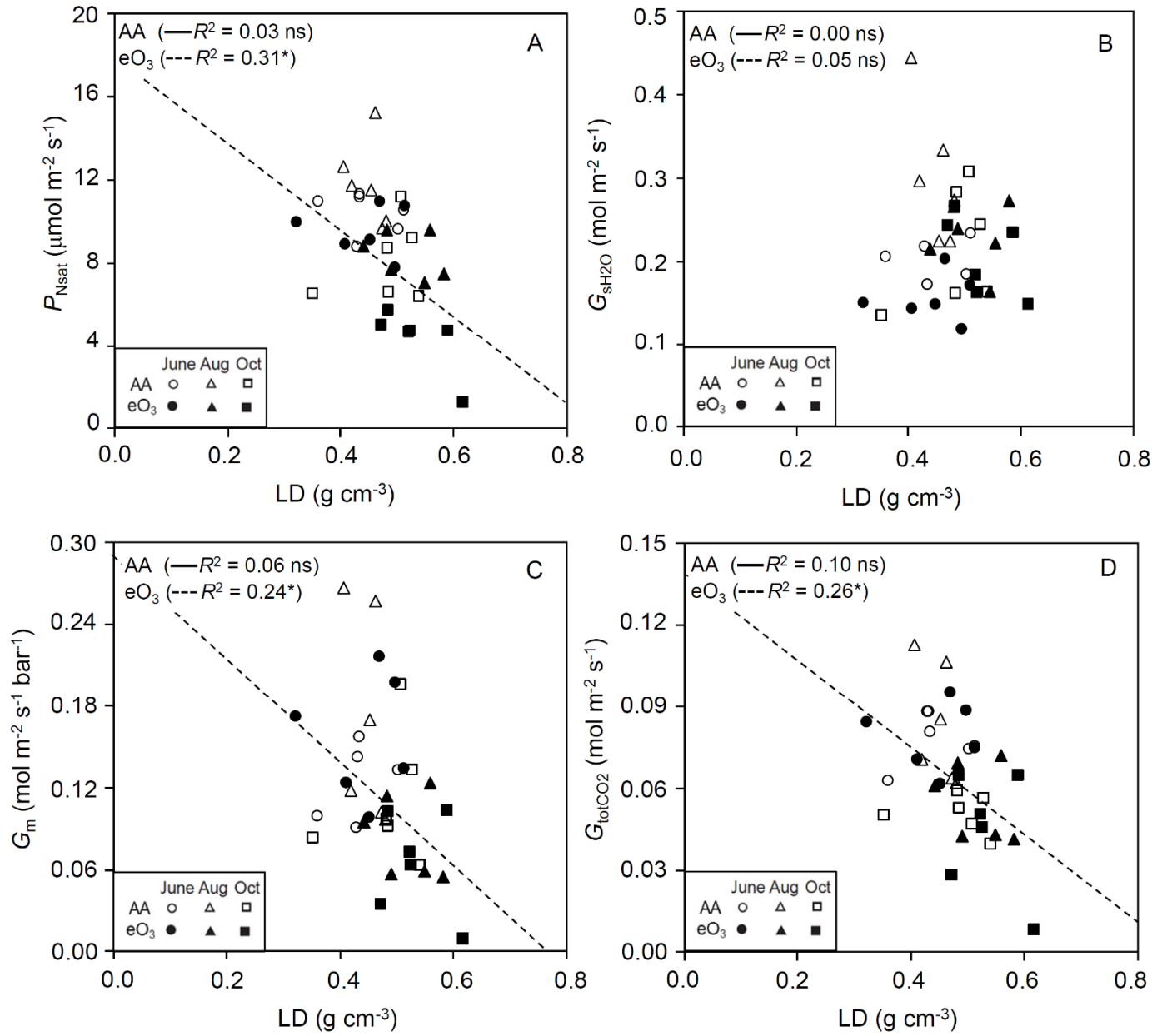


Figure 4. Relationships between leaf gas exchange parameters (A: light-saturated net photosynthetic rate [P_{Nsat}], B: stomatal conductance [$G_{\text{SH}_2\text{O}}$], C: mesophyll

conductance [G_m] and D: total conductance to CO₂ [G_{totCO_2}]) and leaf density (LD) of Siebold's beech saplings under different O₃ concentrations (ambient [AA] or elevated O₃ [eO₃]). Simple linear regression analysis was applied to assess the relationships. * $p < 0.05$, ns denotes not significant.

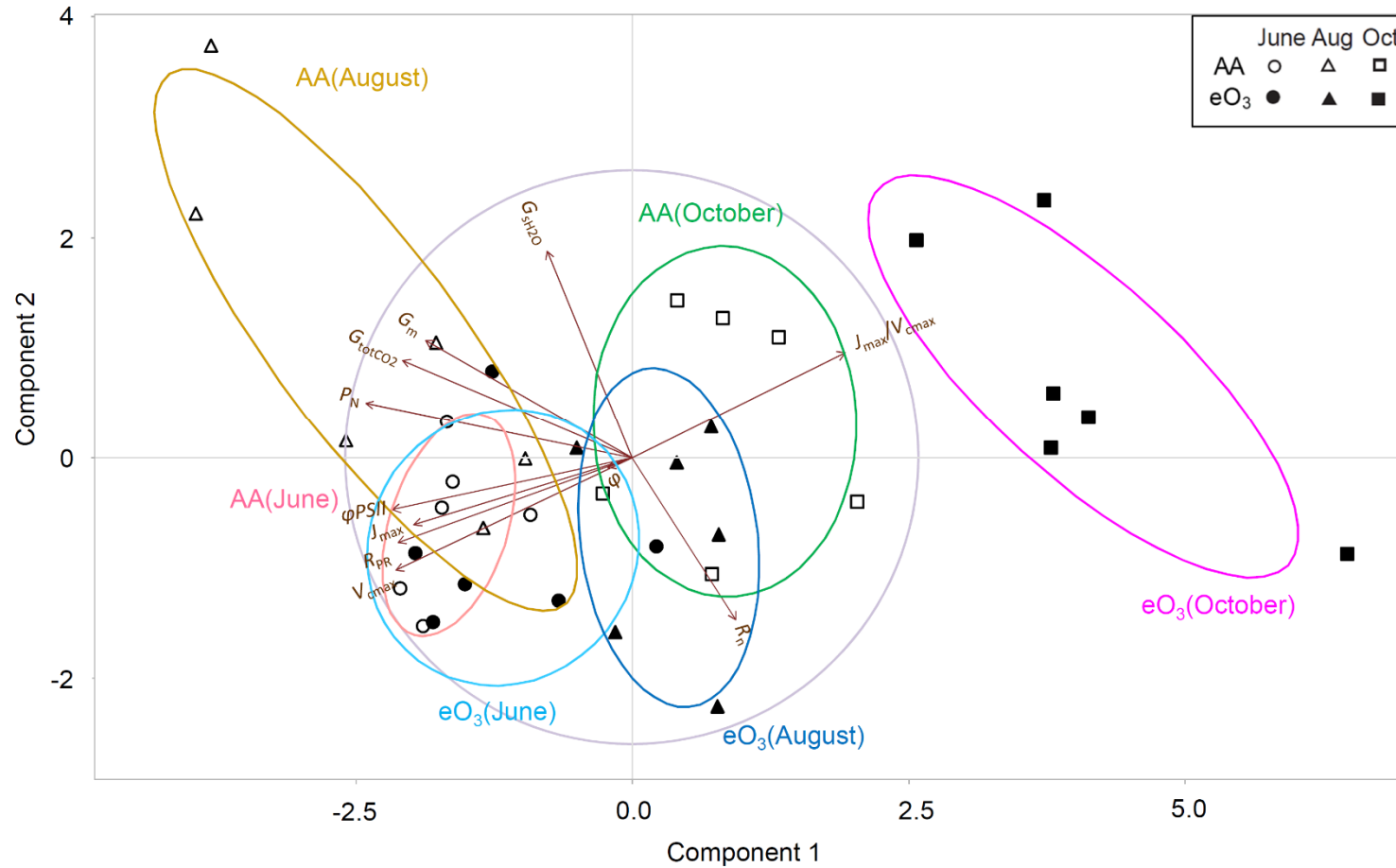


Figure 5. Principal component analysis of parameters derived from analyses of the photosynthetic capacity (P_N/C_c curves), light harvesting ($P_N/PPFD$ curves) and diffusive limitations to CO_2 transport (G_{sCO_2} , G_m and G_{totCO_2}) of Siebold's beech grown under different O_3 concentrations (ambient [AA] or elevated O_3 [eO₃]). Ellipses represent 95% confidence intervals for measurements conducted in June, August and October. The correlation circle is also shown. Measurements of AA-treated plants in June and August and eO₃-treated plants in June occupy the same position in multi-variate space. On the other hand, eO₃-treated plants analysed in October occupy statistically distinct multi-variate space. Component 1 accounts for 49.2% of the variance within the dataset and

component 2 accounts for 14.8% of variance.

Table 1 Leaf mass per area (LMA), leaf density (LD) and leaf thickness (LT) of Siebold's beech saplings grown under ambient (AA) and elevated O₃ (eO₃) measured in 2012. Each value is the mean $\pm$ standard error (n = 6). Two-way ANOVA: * $p < 0.05$, ns denotes not significant. Different letters show significant differences among treatments ($p < 0.05$, Tukey test).

	LMA (g m ⁻²)				LD (g cm ⁻³)				LT (mm)			
	AA		eO ₃		AA		eO ₃		AA		eO ₃	
June	90.9 $\pm$ 7.3	a	83.0 $\pm$ 6.0	a	0.45 $\pm$ 0.02	ab	0.44 $\pm$ 0.03	a	0.20 $\pm$ 0.01	a	0.19 $\pm$ 0.01	a
August	81.3 $\pm$ 3.0	a	92.0 $\pm$ 2.6	a	0.45 $\pm$ 0.01	ab	0.52 $\pm$ 0.02	bc	0.18 $\pm$ 0.01	a	0.18 $\pm$ 0.00	a
October	90.8 $\pm$ 8.0	a	96.6 $\pm$ 4.4	a	0.48 $\pm$ 0.03	abc	0.54 $\pm$ 0.02	c	0.19 $\pm$ 0.01	a	0.18 $\pm$ 0.00	a
<i>ANOVA results</i>												
O ₃		ns				*				ns		
Month		ns				*				ns		
O ₃ $\times$ Month		ns				ns				ns		

Table 2 Parameters derived from analyses of light harvesting (P_N /PPFD curves: initial slope of photosynthetic light response curve [Φ], quantum yield of photosystem II photochemistry [ϕ PSII], electron transport rate [ETR]) and photosynthetic capacity (P_N /Cc curve: maximum rate of carboxylation [V_{cmax}], maximum rate of electron transport for RuBP regeneration [J_{max}], the ratio of J_{max} to V_{cmax} [$J_{\text{max}}/V_{\text{cmax}}$ ratio]) of Siebold's beech grown under ambient (AA) and elevated O_3 (e O_3). Each value is the mean $\pm$ standard error (n = 6). Two-way ANOVA: * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, ns denotes not significant. Different letters show significant differences among treatments ($p < 0.05$, Tukey test).

	Φ (mol mol ⁻¹)				ϕ PSII				ETR ($\mu\text{mol m}^{-2} \text{s}^{-1}$)			
	AA		e O_3		AA		e O_3		AA		e O_3	
June	0.042 $\pm$ 0.003	a	0.037 $\pm$ 0.002	a	0.20 $\pm$ 0.01	ab	0.20 $\pm$ 0.01	ab	91.0 $\pm$ 6.0	ab	93.4 $\pm$ 3.2	ab
August	0.043 $\pm$ 0.002	a	0.041 $\pm$ 0.001	a	0.22 $\pm$ 0.01	a	0.18 $\pm$ 0.01	b	103.8 $\pm$ 4.6	a	84.8 $\pm$ 2.7	b
October	0.044 $\pm$ 0.001	a	0.038 $\pm$ 0.004	a	0.14 $\pm$ 0.01	c	0.08 $\pm$ 0.01	d	64.3 $\pm$ 4.0	c	37.6 $\pm$ 2.9	d
<i>ANOVA results</i>												
O_3		ns				***				***		
Month		ns				***				***		
$\text{O}_3 \times \text{Month}$		ns				**				**		

	V_{cmax} ($\mu\text{mol m}^{-2} \text{s}^{-1}$)				J_{max} ($\mu\text{mol m}^{-2} \text{s}^{-1}$)				$J_{\text{max}}/V_{\text{cmax}}$ ratio			
	AA		e O_3		AA		e O_3		AA		e O_3	
June	66.8 $\pm$ 6.9	a	62.7 $\pm$ 6.1	a	118.0 $\pm$ 7.8	ab	114.9 $\pm$ 10.1	ab	1.8 $\pm$ 0.1	a	1.9 $\pm$ 0.1	a
August	63.2 $\pm$ 2.5	a	54.5 $\pm$ 6.0	a	113.5 $\pm$ 1.8	ab	87.9 $\pm$ 6.4	bc	1.8 $\pm$ 0.1	a	1.7 $\pm$ 0.2	a
October	47.2 $\pm$ 4.9	a	24.8 $\pm$ 2.4	b	95.2 $\pm$ 3.4	abc	70.1 $\pm$ 6.1	c	2.1 $\pm$ 0.2	a	2.9 $\pm$ 0.1	b
<i>ANOVA results</i>												
O_3		**				**				*		
Month		***				***				***		
$\text{O}_3 \times \text{Month}$		ns				ns				**		

Table 3 Relative contribution of stomatal, mesophyll conductance and biochemical limitations (l_s , l_m and l_b , respectively) to light-saturated net photosynthesis (P_{Nsat}) of Siebold's beech grown under ambient (AA) and elevated O_3 (e O_3). Each value is the mean $\pm$ standard error (n = 6). Two-way ANOVA: * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, ns denotes not significant. Different letters show significant differences among treatments ($p < 0.05$, Tukey test).

	l_s (%)				l_m (%)				l_b (%)			
	AA		e O_3		AA		e O_3		AA		e O_3	
June	27.9 $\pm$ 0.8	ab	35.1 $\pm$ 2.1	a	35.2 $\pm$ 4.7	a	29.6 $\pm$ 4.4	a	36.9 $\pm$ 5.1	ab	35.3 $\pm$ 2.9	a
August	23.9 $\pm$ 1.4	b	21.3 $\pm$ 1.4	bc	27.2 $\pm$ 4.1	a	34.1 $\pm$ 4.0	a	48.9 $\pm$ 4.9	ab	44.6 $\pm$ 4.9	ab
October	22.2 $\pm$ 2.7	bc	16.0 $\pm$ 1.8	c	36.3 $\pm$ 3.2	a	30.3 $\pm$ 4.8	a	41.5 $\pm$ 2.3	ab	53.7 $\pm$ 4.9	b
ANOVA results												
O_3	ns				ns				ns			
Month	***				ns				*			
$O_3 \times$ Month	**				ns				ns			

Supplementary materials

Interactive effect of leaf age and ozone on mesophyll conductance in Siebold's beech

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Supplementary figures

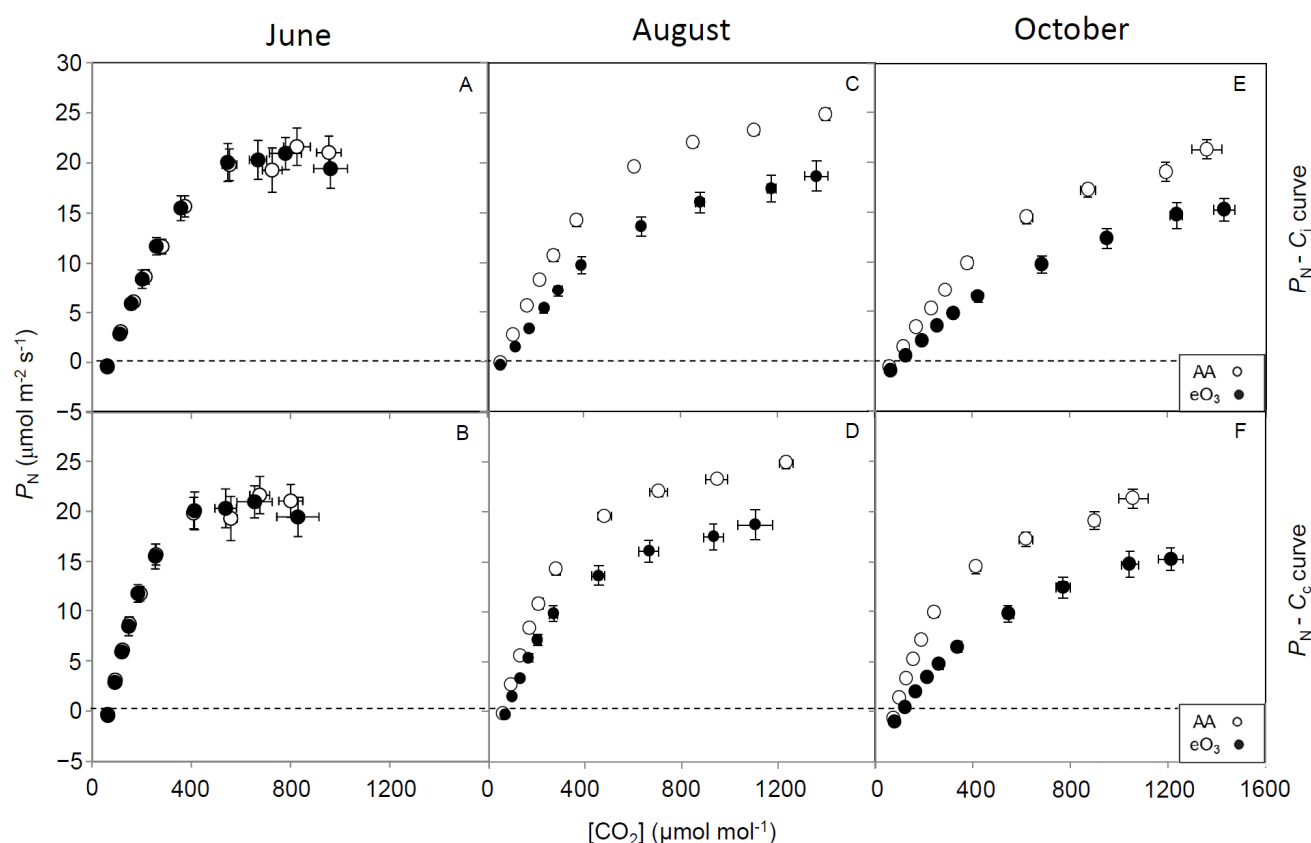


Figure S1. Responses of net photosynthetic rate (P_N) to the sub-stomatal concentration of CO_2 (C_i) (upper figures) or the concentration of CO_2 inside the chloroplast envelope (C_c) (bottom figures) at different ages. The P_N/C_i curves were obtained by measuring P_N over 12 ambient CO_2 concentration steps (C_a , 380, 300, 220, 140, 60, 300, 380, 500, 800, 1100, 1400, 1700 $\mu\text{mol mol}^{-1}$). Values of G_m derived from the curve fitting method (Ethier and Livingston, 2004) were used to calculate the P_N/C_c curves. Each value is the mean $\pm$ standard error (n = 6).

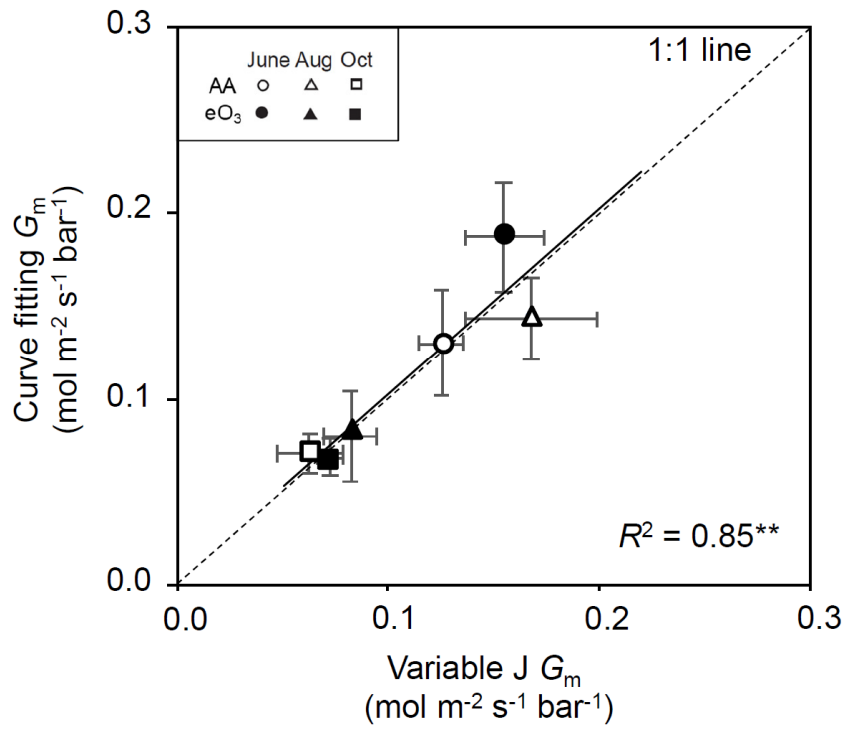


Figure S2. Relationship between mesophyll conductance (G_m) values derived from the variable J method and those derived from the P_N/C_i curve fitting method in Siebold's beech exposed to different O_3 concentrations (ambient [AA] or elevated O_3 [eO_3]). Each value is the mean $\pm$ standard error (n = 6). A linear regression analysis: $**p < 0.01$.

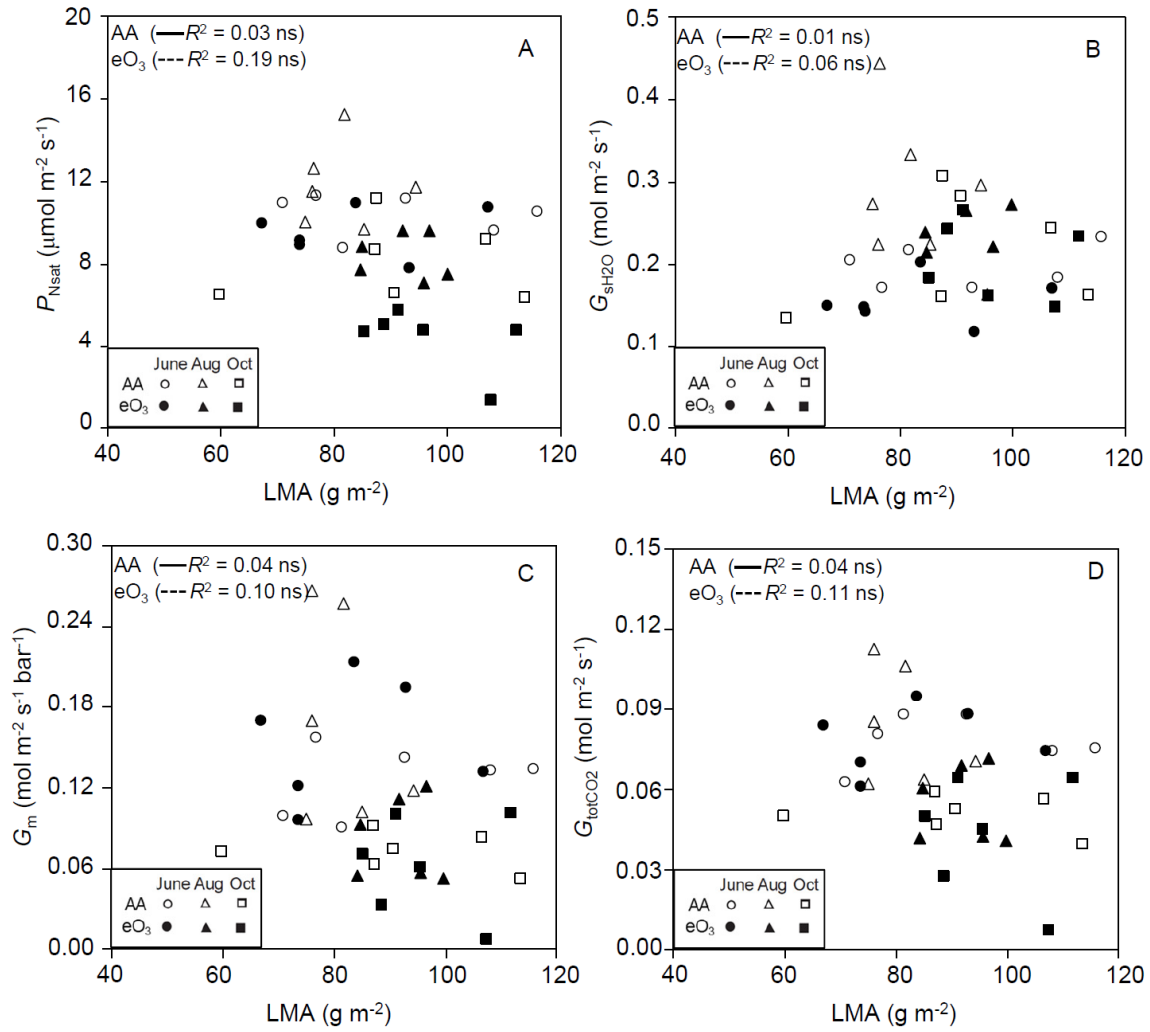


Figure S3. Relationships between leaf gas exchange parameters (A: light-saturated net photosynthetic rate [P_{Nsat}], B: stomatal conductance [G_{sH_2O}], C: mesophyll conductance [G_m] and D: total conductance to CO₂ [G_{totCO_2}]) and leaf mass per area (LMA) of Siebold's beech saplings under different O_3 concentrations (ambient [AA] or elevated O_3 [eO₃]). Simple linear regression analysis was applied to assess the relationships. ns denotes not significant.

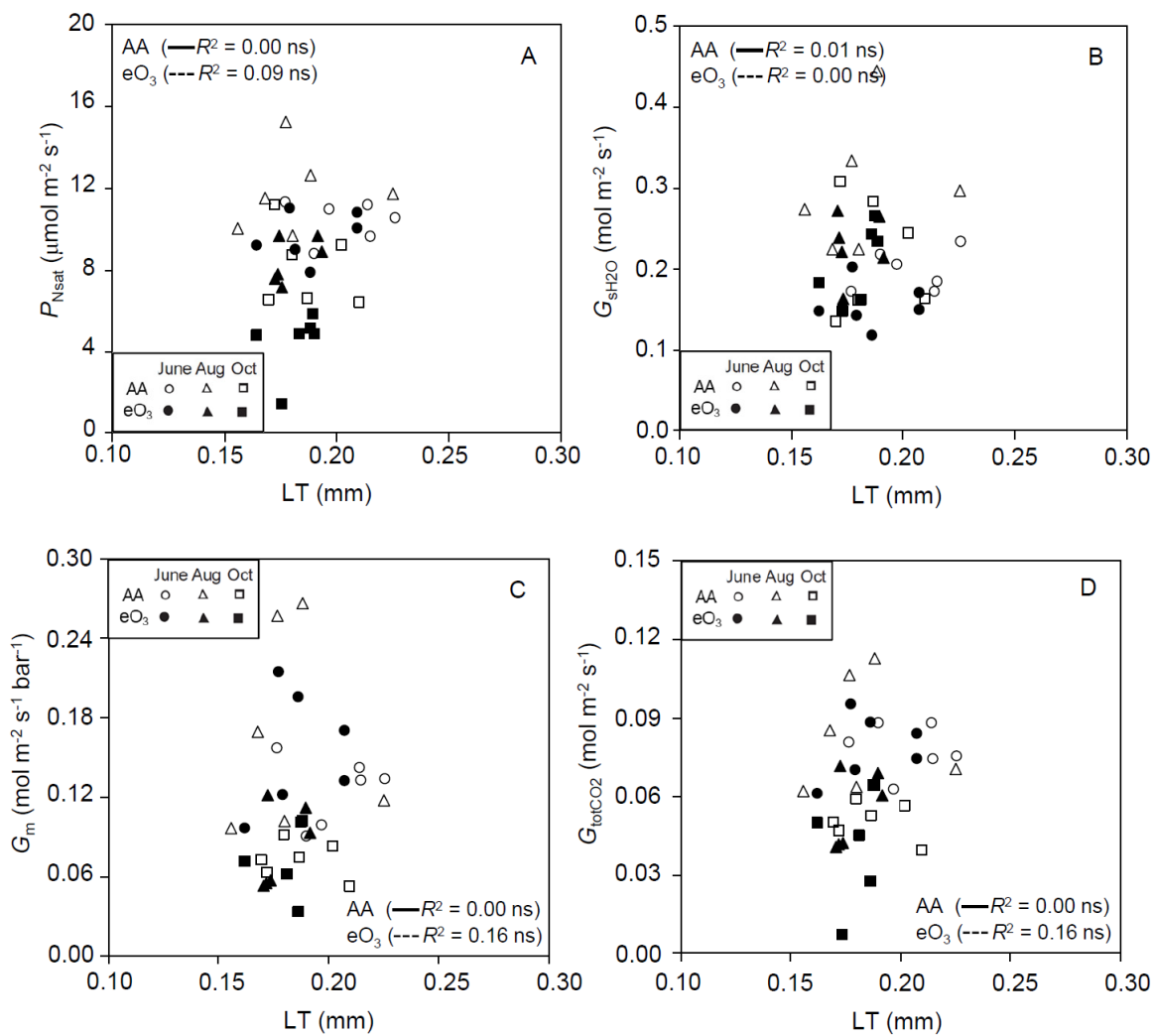


Figure S4. Relationships between leaf gas exchange parameters (A: light-saturated net photosynthetic rate [P_{Nsat}], B: stomatal conductance [G_{sH_2O}], C: mesophyll conductance [G_m] and D: total conductance to CO_2 [$G_{tot\text{CO}_2}$]) and leaf thickness (LT) of Siebold's beech saplings under different O_3 concentrations (ambient [AA] or elevated O_3 [eO_3]). Simple linear regression analysis was applied to assess the relationships. ns denotes not significant.